# Provided for non-commercial research and education use. Not for reproduction, distribution or commercial use.



The journal of Toxicology and pest control is one of the series issued twice by the Egyptian Academic Journal of Biological Sciences, and is devoted to publication of original papers related to the interaction between insects and their environment.

The goal of the journal is to advance the scientific understanding of mechanisms of toxicity. Emphasis will be placed on toxic effects observed at relevant exposures, which have direct impact on safety evaluation and risk assessment. The journal therefore welcomes papers on biology ranging from molecular and cell biology, biochemistry and physiology to ecology and environment, also systematics, microbiology, toxicology, hydrobiology, radiobiology and biotechnology.

www.eajbs.eg.net

Egypt. Acad. J. Biolog. Sci., 9(3):135-147 (2017) Egyptian Academic Journal of Biological Sciences F. Toxicology & Pest control **ISSN: 2090 - 0791** www.eajbs.eg.net



**Biochemical Parameters, Toxicity, Development and Reproductive Effects of** Two Novel Insecticides on Spodoptera littoralis (Lepidoptera: Noctuidae)

Hatem, A. E., Sorour, H. A. and Hassan, A. T.

Plant Protection Research Institute, Agriculture Research Center, Giza, Egypt. E-mail:- a\_hattem@yahoo.com and hamsor98@hotmail.com

#### **ARTICLE INFO**

**Article History** Received: 3/9/2017 Accepted:10/11/2017

Key words:

Sublethal concentration Fecundity egg viability adult longevity biochemical parameters

#### ABSTRACT

The response of the cotton leafworm, Spodoptera littoralis (Boisduval) (Lepidoptera: Noctuidae), to lethal, sublethal and biochemical parameters effects of clothianidin and metaflumizone were determined by a leaf dipping technique bioassays. Mortality of newly molted 4<sup>th</sup> instar larvae increased with the concentration resulting LC<sub>50</sub> values of 70.24 ppm and 20.41 ppm, for clothiandin and metaflumizone respectively.

Sublethal effects were studied by treatment of 4<sup>th</sup> instar larvae with a concentration equivalent to LC50. The larval development time, from treatment until pupation, of the survivors were significantly increased in both insecticides, the pupation period shorted significantly for both sex but shorted insignificantly in male pupa treated with clothianidin, and the weighty of pupa was reduced significantly in both insecticides. However, no significant differences were found in the oviposition and oviposition days viability and percentage of egg viability, but significant differences were found in the preoviposition days when larvae treated with metaflumizone, fecundity percentage decreased in both insecticides and adult longevity was shorted in either both sex or both insecticides.

Biochemical analysis indicated that while treatment of 4th instar larvae with LC50 of both pesticides had no significant effect on acetylcholine esterase activity (AChE) in larval homogenate, it significantly decreased larval contents of total protein and total carbohydrates, reduced alkaline phosphatase (ALP), and esterase (EST) activities, and increased glutathione-s-transferase (GST) acitivity.

#### **INTRODUCTION**

The Egyptian cotton leafworm, Spodoptera littoralis (Boisduval) (Lepidoptera: Noctuidae) is a key pest of cotton and other many crops in the Mediterranean area and Middle Eastern countries (Gómez and Rivero 1951; Campion et al. 1977; Nasr et al. 1984; Ahmad 1988; Domínguez 1993 and Belda et al., 1994). The fact that the insect infests more than 112 host plants belonging to 44 families (Moussa et al. 1960; Brown and Dewhurst 1975; Hatem et al. 2015 and Hatem et al. 2016) makes it a model of serious polyphagous pests. The control of this pest is focused to the searching of new insecticides with biological and ecological qualities (e.g. clothianidin and metaflumizone).

One of these strategies, the using of new pesticide group such as neonicotinoids like neonicotinoids are relatively new class of pesticides which belong to the fastestgrowing class of pesticides in modern pest protection followed after the conventional pesticide group (Ahmed and Matsumura 2012).

Interestingly, neonicotinoids have a unique mode of action as compared to other classes of insecticides in that they act as the agonists to the nicotinic acetylcholine receptor (nAChRs) (Shao et al. 2013 and Sandor et al. 2015). Furthermore, their mammalian toxicities are generally low and also show low acute toxicities to birds, and fish, but displayed significant toxicity to bees (Ahmed 2011). Thus, the use of neonicotinoid pesticides is recommended to be increased in future to control the lepidopterous pests (Lorenzo and David 2015). Metaflumizone, was discovered by Nihon Nohyaku in the early 1990's and belongs to the new class of semicarbazone insecticides. Metaflumizone is derived from the pyrazoline chemistry and acts by the voltagedependent sodium channels in insects binding. Metaflumizone produces а relaxed paralysis in a broad range of important pest insects (Lidia and Justin 2007and Kristopher et al. 2010).

The lethal effects are the most frequently evaluated parameters. Pesticides at sublethal concentrations have a strong impact on insects physiologically and behaviorally (Haynes 1988). Although sublethal effects on surviving insects should also be considered; for instance, insects not dying by product-insect interaction may their have physiological processes affected resulting in development and reproduction alterations (Desneux et al. 2007). Sublethal doses of insecticides may be potentially toxic to different instars and stages of insects through diverse effects such as interfering with the function of glutathione S-transferases carboxylesterase, and (GST), other metabolic enzymes, or changing the behavioural patterns associated with feeding, migration, reproduction, and/or the exchange of chemical information (Haynes 1988).

Furthermore, the extensive usage of these pesticides in cotton leafworm control has been lead to the emergence of pesticides resistance (Mahmoud et al. 2009 and Abdel Rahman et al. 2014). Thus, it is very considerable to use new effective countermeasure strategy in order to avoid the development of resistance particularly among major insect pests of great health concern. Kullik et al. (2011) reported that the clothianidin at a rate of 25 mg kernel-1 on Bt corn increased larval mortality and reduced larval weight of freshly molted third instar of black cutworm, Agrotis ipsilon (Hufnagel) (Lepidoptera: Noctuidae). There is little information on lethal and sublethal effects of metaflumizone in S. littoralis (Boisduval) (Lepidoptera: Noctuidae). Feng-Juan et al. (2012) showed that the pupation rate, emergence rate and total number of eggs laid by one female of Spodoptera exigua (Hübner) (Lepidoptera: Noctuidae), in both generations were significantly lower in the LC15 or LC25 groups than in the control group. Zhang et al. (2012) reported that pupation rate, pupal period and pupal weight were significantly declined comparing with the control Plutella xylostella (L.) (Lepidoptera: Plutellidae) third instar larvae were exposed to  $LC_{15}$ and  $LC_{25}$ of metaflumizone, the fecundity lagged behind control group. Han et al. (2012) found that the metaflumizone at the sublethal concentration (0.09 mg/L) tested decreased the pupation rate and pupal weight, prolonged the pupal duration and decreased the emergence rate, fecundity, oviposition duration and longevity of adults in the treated generation of Plutella xylostella (L.) (Lepidoptera: Plutellidae).

Sublethal effects may be as important as lethal effects in crop protection programs as a result of feeding suppression, delayed development, and reduced reproductive potential of survivos. Thus, sublethal effects of clothianidin and metaflumizone may be great importance in regulating population of a target species, particulary in polivoltine species.

The objectives of this study were to determine: lethal effects of (1)clothianidin and metaflumizone on S. littoralis larvae. (2) sublethal effects of both insecticides on development and reproduction, and (3) sublethal effects of clothianidin and metaflumizone as a biological control agent on some biochemical parameters such as total protein total carbohydrates and enzyme activity in 4<sup>th</sup> larval instars of *S. littoralis*.

#### MATERIALS AND METHODS Insects:

A laboratory colony of S. littoralis continuously was reared away from insecticidial contamination in Plant Protection Research Institute. Rearing of insects were conducted following the technique described by (El- Defrawi et al. 1964). Larvae were fed on fresh castor bean (Ricinus communis L.) leaves until pupation. Moths were fed on 10% sugar solution soaked a piece of cotton tissue. Each jar was provided with branches of tafla, Nerium oleander L. as an oviposition site. The insects were maintained at 25±2 °C, 65±5% RH and a photoperiod of 16:8 h (L:D).

# **Bioinsecticides:**

- Clothianidin 48% SC wt/vol SC (Supertox-1): was obtained from Jiangsu Flag chemical industry Co., China.

- Metaflumizone 24% SC wt/vol SC (Alverde): was obtained from BASF, France.

# **Bioinsecticidal effects:**

Different concentrations of clothianidin (20, 40, 60, 80 and 100 ppm for *S. littoralis* and metaflumizone (5, 10, 20, 40 and 80 ppm were assayed against *S. littoralis* 4<sup>th</sup> larvae by using leaf dipping technique. Fresh castor bean leaves were dipped in each concentration for 10 second. Each concentration was

replicated three times and each replicate contained 30 larvae. Control larvae were fed on castor bean leaves immersed in distilled water. Mortality was recorded every 24 hrs after treatment.

# Sublethal effects on reproductive:

Newly molted  $\overline{4}^{th}$  instar of S. littoralis larvae was treated with LC<sub>50</sub>s ppm clothianidin 70.24 for and metaflumizone 20.41 ppm. Larvae fed on treated leaves during 24-48 hrs. Control larvae were fed on caster bean leaves immersed in distilled water. Males from the nighty after emergence were paired with 1 or 2-day-old virgin females in jars (6 cm in diameter and 12 cm high) which were supplied with leaves of N. oleander as an oviposition site, one pair per jar and on 15% honey solution fed and maintained in the containers until they died. Egg production was recorded daily and eggs were allowed to hatch. All bioassays were conducted at 25±2 °C. Mortality was recorded every 24hrs.

#### **Biochemical parameters:**

# Sample preparation for biochemical analysis:

The 4th instar larvae of *S. littoralis* were immobilized by chilling on ice for 15 minutes. The larvae were cleaned with distilled water, weighed and homogenized in cold distilled water. The homogenates were centrifuged at 5000 rpm for 10 minutes and the supernatants obtained after centrifugation were used for determination of total protein, total carbohydrates and enzyme activity assays.

# Total carbohydrates assay:

Total carbohydrates were extracted, prepared for assay and estimated in samples by the phenol-sulphuric acid reaction according to procedure of Dubois *et al.* (1956) and Crompton and Birt (1967). Total carbohydrate is expressed as mg glucose/gm larval fresh weight.

#### Total protein assay:

Total protein concentration was determined by using Coomassie Brilliant

blue G-250 reagent and bovine albumin as a standard according to the method of Bradford (1976). Total protein is expressed as mg/g larval fresh weight.

# Enzyme activity assays:

Alpha esterases ( $\alpha$ -EST) and beta esterases ( $\beta$ -EST) were determined according to Van Asperen (1962) using  $\alpha$ -naphthyl acetate or  $\beta$ -naphthyl acetate as substrates, respectively. The activity of the EST was presented as ug  $\alpha$ nphthol/min/g.b.wt.

Acetylcholinesterase (AchE) activity was measured according to the method described by Simpson *et al.* (1964) using acetylcholine bromide (AchBr) as substrate and expressed as ug AchBr/min/g.b.wt.

Alkaline phosphatase (ALP) activity was measured according to the method described by Powell and Smith (1954) using p-nitrophenyl phosphate (p-NP) as substrate. the enzyme activity is expressed by unit (U), where 1 unit represents amount of enzyme that hydrolyze 1.0 u mole of p-nitophenyl phosphate per minute at 37°C, at pH 10.4.

Glutathione S-transferase (GST) activity was measured using 1-chloro-2, 4-dinitrobenzene (CDNB) as substrate based on the methods of Habig *et al.* (1974). The activity of the GST was presented as m.mole sub.conj/min/g.b.wt.

# Analysis of data:

Median lethal concentrations (LC50,s) were determined by linear regression analysis and a test was made for parallelism according to the relative potency estimation method (Finney 1971) using the microcomputer program POLO-PC (Russell et al. 1977). The larval pupal development, and preoviposition, oviposition period, total fecundity. egg viability and adult longevity data were analysed bv ANOVA and comparison of means by the least significant difference test (LSD) were calculated. The assays for total protein, total carbohydrates and enzyme activity were performed in 3 replicates, and the data were subjected to one way ANOVA. The means were separated by Multiple Tukey Range Test for significance (p<0.05) using costat software for windows (Costat 2007).

#### RESULTS

# **Bioinsecticidal effects:**

The mortality percentage increased with the concentration of Clothianidin and Metaflumizone. The larvae died from 1 to 2 days after treatment, for both insecticides (Table 1) but the most of them died in the same instar of treatment for both insecticides.

Tractment	Concentration (nom)	NIQ	Mortality	
Treatment	Concentration (ppm)	IN	n°	%
	0	90	0	0
	20	90	10	11.11
Clathianidin	40	90	20	22.22
Ciotmanidin	60	90	44	48.89
	80	90	52	57.79
	100	90	55	61.11
Metaflumizone	0	90	0	0
	5	90	30	27.78
	10	90	44	48.89
	20	90	46	51.11
	40	90	49	54.44
	80	90	60	66 67

Table 1: Mortality of *Spodoptera littoralis* larvae treated in fourth instar with clothianidin and metaflumizone.

N°=Treated larvae number. n°=Died larvae number.

Analysis by probit regression line revealed the following equations and  $LC_{50}$ 's (with 95% confidence limits): y=2.36X+4.13;  $\chi^2=3.35$  (3df) and 70.42 ppm (62.16±80.30) for Clothianidin; y=0.71X+6.65;  $\chi^2=3.94$  (3df) and 20.41 ppm (13.56±30.96) for Metaflumizone. The adjustment was acceptable using the test  $\chi^2$ .

### Sublethal effects on reproductive:

Development period of the male and female larvae treated with Clothianidin (8.56 and 7.62 davs respectivily) and Metaflumizone (12.19 and 12.00 days respectivily) were prolonged significantly for male and femal (F=369; df=2; p=0.0000 and F=416; df=2; p=0.0000) respectivily (Table 2). Development period of the

pupae treated with each insecticide was shorter than the control pupae, while males tended to develop slower than females too (Table 3). Pupal development of females was significantly (F=32.5; df=2; p=0.0000 and F=242; df=2; p=0.0000) decreased but the male pupae were not affected by the Pupal weights differed treatments. significantly depending on the insecticide on which the larvae were treated and differed significantly between females and males when they treated with both insecticide (F=12.70; df=2; p=0.0000 and F=47.60; df=2; p=0.0000) (Table 4). The female pupae were generally heavier than their male, and heavier in control than treatment.

Table 2: Effects of clothianidin and metaflumizone on larval development of Spodoptera littoralis.

Sex	NIO	Treatment	]	Time (days)	
	1	Treatment	Mean	Range	S.E.
	34	Control	7.29 a	7-9	0.14
4	29	Clothianidin	7.62 a	7-8	0.15
	43	Metaflumizone	12.00 b	11-14	0.12
	36	Control	8.11 a	7-9	0.12
3	34	Clothianidin	8.56 b	7-9	0.12
	42	Metaflumizone	12.19 c	11-13	0.11
	Sex P 3	$\begin{array}{c c} Sex & N^{\circ} \\ \hline & 34 \\ \hline & 29 \\ \hline & 43 \\ \hline & 36 \\ \hline & 34 \\ \hline & 42 \\ \hline \end{array}$	SexN°Treatment $\begin{array}{c} \begin{array}{c} 34 \\ 29 \end{array}$ Control $\begin{array}{c} 43 \end{array}$ Metaflumizone $\begin{array}{c} 36 \end{array}$ Control $\begin{array}{c} 34 \end{array}$ Clothianidin $\begin{array}{c} 42 \end{array}$ Metaflumizone	$\begin{array}{c c c c c c c c c c c c c c c c c c c $	$\begin{array}{c c c c c c c c c c c c c c c c c c c $

Means followed by the same letter are not significantly different (LSD, p = 0.05). S.E. = Standard error.

Table 3: Effects of clothianidin and metaflumizone c	n pup	al develo	pment of S	podo	ptera littoral	is
--	-------	-----------	------------	------	----------------	----

Concentration	Sov	NIO	Traatmont		Time (days)	
(ppm)	Sex	IN	Treatment	Mean	Range	S.E.
0.0		34	Control	9.50 a	9-11	0.13
70.24	4	29	Clothianidin	9.03 b	8-10	0.14
20.41		43	Metaflumizone	8.14 c	7-9	0.11
0.0		36	Control	11.47 a	10-12	0.11
70.24	3	34	Clothianidin	11.26 a	10-13	0.11
20.41		42	Metaflumizone	8.59 b	8-10	0.10

Means followed by the same letter are not significantly different (LSD, p=0.05). S.E=Standard error.

Table 4: Effects of clothianidin and metaflumizone on pupal wieghty of Spodoptera littoralis.

				1 0 7	1 1	
Concentration	Sou	NIO	Traatmant	weighty (grams)		
(ppm)	Sex	IN	Treatment	Mean	Range	S.E.
0.0		34	Control	0.26 a	0.2-0.36	0.005
70.24	Ŷ	29	Clothianidin	0.24 b	0.21-0.29	0.006
20.41		43	Metaflumizone	0.23 c	0.20-0.33	0.005
0.0		36	Control	0.26 a	0.20-0.31	0.005
70.24	8	34	Clothianidin	0.20 b	0.16-0.25	0.005
20.41		42	Metaflumizone	0.20 b	0.16-0.25	0.005

Means followed by the same letter are not significantly different (LSD, p=0.05). S.E.=Standard error

Most of females were mated and most of them were fecund and fertil, independently of treatment (Table 5). Mean number of eggs laid per female Clothianidin surviving to and Metaflumizone were decreased in all mating combinations respect to the control combination, but this difference were statistically significant (p=0.0000). The mean percentage of viability per female for both insecticide bioassays decreased in most of the combinations related to the control, but the differences were not statistically significant

(p=0.5378) (Table 6). Mean number of laid eggs and laid eggs viable days did not differ significanly among treated and control (p=0.2244 and p=0.2041). Mean days of preovipostion were longer in larvae treated with Metaflumeziron than either Clothianidian or control with significant differences (p=0.0000) (Table 7). The mean of adults longevity for females males and were shorted in treated larvae than significantly control (p=0.0000 and p=0.0000) (Table 8).

 Table 5: Response of Spodoptera littoralis females came from larvae treated with clothianidin and metaflumizone according to different mating combinations.

Combination	NIO	N° females			
Comonitation	IN	Eggs laid	Eggs viable laid		
$\begin{array}{l} \bigcirc \\ \bigcirc \\ \end{array}$ control X $\begin{array}{c} \bigcirc \\ \bigcirc \\ \end{array}$ control	15	10	10		
$\mathcal{P}_{\mathbf{c}} \mathbf{X} \mathcal{O}_{\mathbf{c}}$	16	12	12		
$\begin{array}{c} \bigcirc \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $	15	13	13		

 $N^{\circ}$  = Mated female number. c = clothianidin treatment. m = metaflumizone treatment.

Table 6: Reproductive potential of *Spodoptera littoralis* larvae treated with clothianidin and metaflumizone according to different mating combinations.

		Fecu	ndity	% V	iability
Combination	N°	Mean	Range	Mean	Range
$\stackrel{\bigcirc}{=}_{\text{control}} \mathbf{X} \stackrel{\frown}{\odot}_{\text{control}}$	10	1485.0 a	1075-2050	76.03 a	56-100
$\begin{array}{c} \bigcirc \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ \\ $	12	721.2 b	200-1550	73.32 a	42.86-100
$\mathcal{L}_{\mathbf{m}} \mathbf{X} \mathcal{T}_{\mathbf{m}}$	13	655.8 b	125-1350	64.97 a	20-97.41

Means followed by the same letter are not significantly different (LSD, p=0.05). N° = Mated female number.

c = clothianidin treatment. m = metaflumizone treatment.

 Table 7: Laid periods of Spodoptera littoralis females came from larvae treated with clothianidin and metaflumizone according to different mating combinations.

Combination	N	Nº Preoviposition days		Ovipos	N° ition days	N	N <sup>o</sup> Oviposition days viability	
Combination	IN <sub>1</sub>	Mean	Range	Mean	Range	IN <sub>2</sub>	Mean	Range
$\stackrel{\bigcirc}{+}_{\text{control}} \mathbf{X} \stackrel{\bigcirc}{\circ}_{\text{control}}$	10	3.10 a	1-5	3.8 a	2-5	10	3.4 a	1-5
$\begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} $	12	2.00 a	1-3	3.0 a	1-5	12	2.5 a	1-4
$\begin{array}{c} \bigcirc \\ + \\ m \end{array} \mathbf{X}  \stackrel{\frown}{\bigcirc} \\ \mathbf{m} \end{array}$	13	3.23 b	2-4	3.1 a	1-5	13	2.92 a	1-5

Means followed by the same letter are not significantly different (LSD, p= 0.05). N1 = Fecundity female number.

N2 = Fertility female number. c= clothianidin treatment. m= metaflumizone treatment

Table 8: Adults longevity of *Spodoptera littoralis* came from larvae treated with clothianidin and metaflumizone according to different mating combinations.

		Males		Females	
Combination	N°	Mean	Range	Mean	Range
$\mathcal{Q}_{\text{control}} \mathbf{X} \mathcal{O}_{\text{control}}$	10	11.2 a	6-14	9.5 a	6-11
$\mathcal{Q}_{\mathbf{c}} \mathbf{X} \mathcal{O}_{\mathbf{c}}$	12	5.7 b	4-11	5.8 b	4-9
$\begin{array}{c} \bigcirc \\ + \\ m \end{array} \mathbf{X}  \bigcirc \\ \mathbf{m} \end{array}$	13	7.1 b	5-10	6.9 b	5-9

Means followed by the same letter are not significantly different (LSD, p=0.05). N° = Mated female number. c = clothianidin treatment.

(Table 9) shows the results of biochemical analysis of S. littoralis 4th instar larvae treated with LC<sub>50</sub> of Clothianidin and Metaflumizone compared with untreated control larvae. Treatment with Clothianidin and Metaflumizone resulted in slight reduction of larval total protein contents (32.1±0.8 and 34.2±0.7 mg /g.b.wt, respectively) compared with the untreated control ( $40.9\pm1.4$  mg/g.b.wt).

Total carbohydrates also significantly decreased from  $11.8\pm0.9$  mg/g.b.wt for untreated larvae to  $9.5\pm0.4$  and  $6.1\pm0.5$  mg/g.b.wt for larvae treated with Clothianidin and Metaflumizone, respectively. The activity of  $\alpha$ -EST was significantly lower in Clothianidin treated larvae than control (249.0 $\pm$ 8.0 Vs 326.3 $\pm$ 15.5 respectively) while both insecticides causes significant reduction in  $\beta$ -EST and ALP activities.

Table 9: Amounts of total protein, total carbohydrates and the activities of selected enzymes in *S*. *littoralis* 4th instar larvae treated with  $LC_{50}$  of metaflumizone and clothianidin compared with untreated control larvae (mean  $\pm$  SD).

· · · · · · · · · · · · · · · · · · ·	Control	Clothianidin	Metaflumizone
Total protein (mg /g.b.wt)	40.9±1.4 a	32.1±0.8 b	34.2±0.7b
Total carbohydrates (mg /g.b.wt)	11.8±0.9 a	9.5±0.4 b	6.1±0.5b
α-EST (ug α-nphthol/min /g.b.wt)	326.3±15.5 a	249.0±8.0 b	327.7±5.5a
$\beta$ -EST (ug $\beta$ -nphthol/min /g.b.wt)	163.0±9.8 a	116.7±4.2 b	143±6.1c
AchE (ug AchBr/min/g.b.wt)	251.3±8.1 a	238.3±7.6 a	267.7±8.9a
ALP $(U \times 10^3 / g.b.wt)$	433±27.4 a	278.7±6.8 b	271.7±7.4b
GST (m mole sub.conj/min/g.b.wt)	25.3±1.7 a	30.4±0.6 b	31.2±1.1b

Means in the same raw with different letters are significantly different (LSD, p < 0.05).

The GST activity was obviously and higher Clothianidin in Metaflumizone treated larvae than control larvae  $(30.4\pm0.6)$ untreated  $31.2\pm1.1$  and  $25.3\pm1.7$  respectively). Table 9 also shows that AChE activity does not change significantly in larvae exposed to  $LC_{50}$  of Both insecticides compared with control

#### DISCUSSION

## **Bioinsecticidal effects:**

The laboratory colony of S. littoralis susceptible tested to was both insecticides. Metaflumizone and Clothianidian were present a high toxicity for some lepidopteran species like Agrotis segetum (Denis & Schiffermuller) (Lepidoptera: Noctuidae) was evaluated through leaf dipping method of bioassay against third and sixth instar larvae of A. segetum. The Clothianidin was more toxic for the third instar larvae and the LD50 was 1.6 ng a.i./larva but for sixth instar larvae LD<sub>50</sub> was 3.5 ng a.i./larva (Nikhil and Verma

2015). The LC<sub>50</sub> value obtained in S. *litttoralis* was similar to the  $LC_{50}$ reported by Tofangsazi et al. (2015) in the larvae of Herpetogramma phaeopteralis (Guenée 1854) (Lepidoptera: Crambidae) (46.6 ppm), treated with clothianidin by topical application of insecticides against medium-sized (third and fourth instar) *H*. phaeopteralis.

Khakame et al. (2013) evaluated the toxicity to Diamondback moth, Plutella xylostella (L.) (Lepidoptera: Plutellidae) to third instar larvae using the leaf dipping bioassays. The concentrations were calculated in milligrams per liter (ppm). The result showed that Metaflumizone toxic to P. The populations of *P*. xvlostella. xylostella from different 14 geographical areas in China displayed a narrow variation in LC50 ranged from 1.34 to 8.16 ppm. Metaflumizone could provide an effective alternative insecticide for Diamondback moth management. Tian et al. (2014)reported that the Metaflumizone was toxic to beet Spodoptera armyworm, exigua. Emmanouil et al. (2013) were used in a leaf-dipping bioassay for second-instar larvae of Tomato borer Tuta absoluta (Meyrick) (Lepidoptera: Gelechiidae) from Greece. All assays were performed in a 32- cell replication dish (RT32W; Bioserve, USA, www.insectrearing.com). The  $LC_{50}$  ranged from 31.8 to 159.5 mgL-1, to the second instar T. absoluta. The LC<sub>50</sub> values for Metaflumizone were different in the Ankara-Beypazarı 52.7 ppm and Antalya-Serik 21.02 ppm population of T. absoluta L2 larvae. The experiment was conducted using a leafdipping bioassay method with L2 larvae. This indicates that the susceptibility levels of these populations to this insecticide are different. However it is difficult to decide using only two populations without а susceptible population (Karaagac 2012).

# Sublethal effects on reproductive:

results showed that the Our development period of the male and female larvae treated with Clothianidin Metaflumizone were prolonged and significantly. Pupation period of S. littoralis males was about two days longer than females, either treated or nontreated larvae, have as been previously reported by Vargas Osuna (1985). For this reason the study of pupation period was made considering the males and females separately. As general the development period of the pupae treated with each insecticide was shorter than the control pupae, while males tended to develop slower than females too. Pupal development of females was significantly decreased but the male pupae were not affected by the treatments. Pupal weights differed significantly depending on the insecticide on which the larvae were treated and differed significantly between females and males when they treated with both insecticide. The female pupae were generally heavier than their male, and

heavier in control than treatment. Kullik et al. (2011)reported that the Clothianidin at a rate of 25 mg kernel-1 on Bt corn increased larval mortality and reduced larval weight of freshly molted third instar of Black cutworm, Agrotis (Hufnagel) (Lepidopter: ipsilon Noctudiae). There are little information on lethal and sublethal effects of Metaflumizone in S. littoralis. (Feng-Juan et al. 2012) showed that the pupation rate and emergence rate of Spodoptera exigua (Hübner) (Lepidopter: Noctudiae) in both generations were significantly lower in the  $LC_{15}$  or  $LC_{25}$  groups than in the control group. Zhang et al. (2012) reported that pupation rate, pupal period and pupal weight were significantly declined comparing with the control P. xylostella third instar larvae were exposed to  $LC_{15}$ and  $LC_{25}$ of Metaflumizone. Han et al. (2012) found that the Metaflumizone at the sublethal concentration (0.09)mg/L) tested decreased the pupation rate and pupal weight, prolonged the pupal duration and decreased the emergence rate, in the treated generation of *P. xylostella*.

The reproduction of *S. littoralis* has been affected with some of the treatments studied but in different form according to the insecticide. Mean number of eggs laid per female surviving to Clothianidin and Metaflumizone were decreased in all mating combinations respect to the control combination but this difference was statistically significant. The mean percentage of viability per female for both insecticide bioassays decreased in most of the combinations related to the control but the differences were not statistically significant. Mean number of laid eggs and laid eggs viable days did not differ significanly among treated and control. Mean days of preovipostion were longer in larvae treated with Metaflumeziron than either Clothianidian or control with significant differences. Adults longevity of S. littoralis was not

affected when treated with the treated insecticides. The mean of adults longevity for males and females were shorted significantly in treated larvae than control.

Feng-Juan et al. (2012) showed that the emergence rate and total number of eggs laid by one female of S. exigua (Hübner) in both generations were significantly lower in the  $LC_{15}$  or  $LC_{25}$ groups than in the control group. Zhang et al. (2012) reported that the fecundity of P. xylostella third instar larvae were exposed to  $LC_{15}$ and LC25 of Metaflumizone lagged behind control group. Han et al. (2012) found that the Metaflumizone at the sublethal concentration (0.09)mg/L) tested decreased the emergence rate, fecundity, oviposition duration and longevity of adults in the treated generation of P. xvlostella.

Metaflumizone is a voltagedependent sodium channel blockers that selectively target voltage-gated sodium channels in the slow inactivated state by binding at or near the local an aesthetic receptor within the sodium channel pore (Stein and Soderlund 2012 and Tian et al. 2014). Clothianidin is new insecticide to imidacloprid, and similar other neonicotinoids acting selectively on insect nicotinic acetylcholine receptors (nAChRs) as an activator of postsynaptic acetylcholine receptors but do not inhibit acetylcholine esterase (AChE) (Thany 2009). The two compounds are without ester bond in their chemical structure, which means that hydrolysis by esterases might not the major mechanism for their detoxification by insects. This explain why treatment of S. *littoralis* larvae with  $LC_{50}$  of the two insecticides did not affect the activity of AChE compared with untreated control as it act on ester bond. However alpha and beta esterase also act on ester bond and the significant decrease in alpha and beta esterase activities in larvae treated with both insecticides may be due to their

inhibition by sequestration. Esterases have been reported to function primarily as sequestration proteins in several insect species including Myzus persicae (Sulzer) (Hemiptera: Aphididae), Culex quinquefasciatus (Say) (Diptera: Culicidae), Culex pipiens (L.) (Diptera: Culicidae) and Leptinotarsa decemlineata (Coleoptera: (Say) Chrysomelidae) (Lee and Clark 1998 and Tian et al. 2014). Sayyed and Wright (2006) confirmed that resistance to indoxacarb, another voltage dependent sodium channel blockers in a field population of diamondback moth was esterase associated. Similarly Smirle et al. (2010) and Jemec et al. (2007) reported significant decrease in the activity of esterase in Aphis pomi (de Geer, 1773) (Hemiptera: Aphididae), Aphis spiraecola (Patch.) (Hemiptera: Aphididae) and *Daphnia magna* (Straus 1820) (Cladocera: Daphniidae) exposed of imidacloprid, to  $LC_{50}$ another neonicotinoids similar to Clothianidin. Our results also showed significant decrease in alkaline phosphatase activity due to treatment with both insecticides. The decrease in level of alkaline phosphatase activity in response to treatments indicates insecticide's harmful effect on insect's digestive system and development as ALP; play an important physiological role in digestive system (Bharat et al. 2017). On the other hand S. *littoralis* larvae treated with LC<sub>50</sub> of both Clothianidin and Metaflumizone were found to possess significantly higher GST activity compared to the untreated control. These results reflect the role of important GST in the of detoxification Clothianidin and Metaflumizone. In accordance with our results Yalcin et al. (2015) confirmed an increased enzyme activity of T. absoluta due to Metaflumizone treatment. Also GST activity was noticeably increased during exposure of Helix aspersa (Müller (Stylommatophora: 1774) Helicidae) (Radwan and Mohamed 2013) and

zebrafish (Ge et al. 2015) to the neonicotinoid imidacloprid . Results of this study also showed that treatment of S. littoralis larvae with Clothianidin and Metaflumizone affected not only the investigated enzymes, but also significantly reduced larval contents of total carbohydrate and proteins, which reflects generally impaired а physiological state of an organism. The decreased amounts of total carbohydrates and proteins confers to the antifeedant nature of the investigated compounds where the larvae unable to metabolise the diet provided for its survival, growth, and reproduction (Jemec et al. 2007 and Bharat et al. 2017).

Further studies will therefore aim to investigate the histology involved (e.g., midgut, ovary, testes,...etc) in these effects on development and reproduction of the surviving insects. These results were valuable for the practical use of the Clothianidin and Metaflumizone in Integrated Pest Management (IPM) programmes.

#### REFERENCES

- Abdel Rahman, M.H.E.; Mohamed, A.I.A, and Samy, H.M. (2014). Toxicological Evaluation of Selected Biopesticides and One Essential Oil in comparison with Indoxacarb Pesticide on Cotton leafworm, Spodoptera littoralis (Boisd.) (Lepidoptera: Noctuidae) under Laboratory Conditions. American-Eurasian Journal of Sustainable Agriculture, 8(2): 58-64.
- Ahmad, T.R. (1988). Field Studies on Sex Pheromone Trapping of Cotton Leafworm *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). J. Appl. Ent., 1(5):212-215.
- Ahmed, M.A.I. (2011). Biochemical and Molecular Biological Studies on the Action of Formamidines in Synergizing Insecticidal Action of Pyrethroids and Neonicotinoids in *Aedes aegypti* Mosquitoes. Ph.D Thesis. UCDavis, CA, USA.
- Ahmed, M.A.I. and Matsumura, F. (2012). Synergistic Action of Formamidine

Insecticides on the Activity of Pyrethroids and Neonicotinoids against *Aedes aegypti* (Diptera: Culicidae). Journal of Medical Entomology, 49(6):1405-1410.

- Belda, J.; Casado E.; Gómez, V.; Rodríguez, M.D, and Sáez, E. (1994). Plagas y Enfermedades de los Cultivos Hortícolas Intensivos. Phytoma, 57:9-40.
- Bharat, N.; Senigala, K.J.; Gayatridevi, S, and Kuruba, S. (2017). Imidaclopride, a Potent Inhibitor of Array of Digestive Enzymes of *Helicoverpa armigera* (Hubner) (Lepidoptera: Noctuidae). Adv Biotech & Micro., 4(1):555626-555629.
- Bradford, M.M. (1976). A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Proteins Utilizing the Principle of Protein-dye Binding. Anal. Biochem., 72:248-254.
- Brown, E.S. and Dewhurst, C.F. (1975). The Genus *Spodoptera* (Lepipodptera: Noctuidae) in África and the Near East. Bull. Res., 65:221-262.
- Campion, D.G.; Bettany, B.W.; Mcginnigle, J.B, and Tailor, L.R. (1977). The Distribution and Migration of *Spodoptera littoralis* (Boisduval) (Lepidoptera: Noctuidae), in Relation to Meteorology on Cyprus, Interpreted from Maps of pheromone trap samples. Bulletin of the Entomolgy Research, 67(3):501-522.
- Costat, (2007). Costat Software Package, CoHort Software Inc., Berkeley CA, USA, Release 5.5.
- Crompton, M, and Birt, L.M. (1967). Changes in the Amounts of Carbohydrates, Phosphagen, and related Compounds During the Metamorphosis of the Blowfly, *Lucilia cuprina*. J. Insect Physiol., 13:1575-1595.
- Desneux, N.; Decourtye, A, and Delpuech, J. M. (2007). The Sublethal Effects of Pesticides on Beneficial Arthropods. Annu. Rev. Entomol., 52:81-106.
- Domínguez, F. (1993). Plagas y Enfermedades de las Plantas Cultivadas. Ediciones Mundi-Prensa. Madrid. pp. 821, essay.
- Dubios, M.; Gilles, K.A.; Hamilton, J.K.; Rebers, P.A, and Smith, F. (1956). Colorimetric Method for Determination

of Sugars and Related Substances. Analyt. Chem., 28(3):350-356.

- El-Defrawi, M.E.; Toppozada, A.; Mansour, N, and Zeid, M. (1964). Toxicological Studies on the Egyptian Cotton Leafworm, *Prodenia litura* I. Susceptibility of Different Larval Instars to Insecticides. J. Econ. Entomol., 57(4):591-593.
- Emmanouil, R.; Christina, S, and Staurakaki, M. (2013). Toxicity of Insecticides to Populations of Tomato Borer *Tuta absoluta* (Meyrick) from Greece. Pest Management Science, 69 (7):834-840.
- Feng-Juan, M.A.; Yong-Dan, L.I, and Xi-Wu, G.A.O. (2012). Sublethal effects of Metaflumizone on the Development and Reproduction of the Beet Armyworm, *Spodoptera exigua*. Chinese Journal of Applied Entomology, 2012-02.
- Finney, D.J. (1971). Probit analysis, 3rd Ed. Cambridge University Press, Cambridge, United Kingdom, pp. 318.
- Ge, W.; Yan, S.; Wang, J.; Zhu, L.; Chen, A, and Wang, J. (2015). Oxidative Stress and DNA Damage Induced by Imidacloprid in Zebrafish (*Danio rerio*). J Agric Food Chem., 63(6):1856-1862.
- Gómez-Clemente, F, and Del Rivero, J. M. (1951). La "rosquilla negra" (*Prodenia litura* F.). Bol. Pat. Veg. Ent. Agr., 19:221-278.
- Habig, W.H.; Pabst, M.J, and Jakoby, W. B. (1974). Glutathione S-transferase the First Enzymatic Step in Mercapturic Acid Formation. J. Biol. chem., 249(22):7130-7139.
- Han, W.S.; Ren, C.C.; Yan, H.Y.; Zhang, S.F.; Shen, F.Y, and Gao, X.W. (2012).
  Sublethal Effects of Metaflumizone on Abamectin-Resistant and Susceptible Strains of the Diamondback Moth, *Plutella xylostella* (Lepidoptera: Plutellidae). Acta Entomologica Sinica, 55(6):694-702.
- Haynes, K.F. (1988). Sublethal Effects of Neurotoxic Insecticides on Insect Behavior. Annual Review of Entomology, 33:149-168.
- Hatem, A.E.; Shawer, D.M, and Vargas-Osuna, E. (2016). Parasitism and Optimization of *Hyposoter didymator* (Hymenoptera: Ichneumonidae) Rearing on *Spodoptera littoralis* and *Helicoverpa armigera* (Lepidoptera:

Noctuidae). Journal of Economic Entomology, 109 (3): 1058-1063.

- Hatem, A.E.; Laila, E.S, and Hassan, A.T. (2015). Effects of Host Plants as Larval Feeding on the Biology *Spodoptera littoralis* (Boisd.) (Lepidoptera: Noctuidae). Egyptian Journal of Applied Sciences, 30(3):81-95.
- Jemec, A.; Tisler, T.; Drobne, D.; Sepcic, K.; Fournier, D, and Trebse, P. (2007). Comparative Toxicity of Imidacloprid, of its Commercial Liquid Formulation and of Diazinon to a Non-Target Athropod, the Microcrustacean *Daphnia magna*. Chemosphere, 68(8): 1408-1418.
- Karaagac, S. (2012). The Current Status of *Tuta absoluta* (Meyrick) (Lepidoptera: Gelechiiidae) in Turkey and Baseline Toxicity of some Insecticides. Bulletin OEPP/EPPO Bulletin, 42(2):333-336.
- Khakame, S. K.; Wang, X. L. and Wu, Y. D. (2013). Baseline Toxicity of Metaflumizone and Lack of Cross Resistance between Indoxacarb and Metaflumizone in Diamondback Moth (Lepidoptera: Plutellidae). Journal of Economic Entomology, 106(3):1423-1429.
- Kristopher, S.S.; Weizhong, S.; Yoshiko, N.;
  Vincent, L.S, and Ke, D. (2010).
  Mechanism of Action of Sodium Channel Blocker Insecticides (SCBIs) on Insect Sodium Channels. Pestic Biochem Physiol., 97(2):87-92.
- Kullik, S. A.; Sears, M. K, and Schaafsma,
  A. W. (2011). Sublethal Effects of Cry 1F Bt Corn and Clothianidin on Black Cutworm (Lepidoptera: Noctuidae) Larval Develop. J. Eco. Entom., 104(2):484-493.
- Lee, S.H, and Clark, J.M. (1998). Permethrin Carboxylesterase Functions as Nonspecific Seque-stration Proteins in the Hemolymph of Colorado Potato Beetle, Pestic. Biochem. Physiol., 62(1):51-63.
- Lidia, C, and Justin, C. (2007). Metaflumizone-A new Ectoparasiticide for Dogs and Cats. Medicamentul veterinar/Veterinary drug, 1(2):40-43.
- Lorenzo, F, and David, K. (2015). Alternatives to Neonicotinoid Insecticides for Pest Control: case Studies in Agriculture and Forestry.

Environ. Sci. Pollut. Res. Int., 22(1): 135-147.

- Mahmoud, D.M.; Hamed, R.K.A.; Seufi, A.M.; Salama, M.S.; Diwan, N.M.L, and El Shafei, A. M. (2009). Toxicity and Biological Effects of Three Egyptian Isolates of Baculovirus on the Cotton leafworm, *Spodoptera littoralis* (Boisd.). Egyptian Academy J. Biolog. Sci., 2(1): 219-226.
- Moussa, M. A.; Zaher, M. A. and Kotby, F. (1960). Abundance of the Cotton Leafworm, *Prodenia litura* (F), in Relation to Host Plants. I. Host Plants and Their Effect on Biology (Lepidoptera: Agrotidae, Zenobiinae). Bull. Soc. Ent. Egypte, 44:241-245.
- Nasr, E.A.; Tucker, M.R. and Campion, D.G. (1984). Distribution of Moths of the Egyptian Cotton Leafworm, *Spodoptera littoralis* (Boisduval) (Lepidoptera: Noctuidae), in the Nile Delta Interpreted from Catches in a Pheromone Trap Network in Relation to Meteorological Factors. Bull. Ent. Res., 74:487-494.
- Nikhil, S. and Verma, K. (2015). Evaluation of New Chemistry Against *Agrotis segetum* (Denis & Schiffermuller). Journal of Insect Science (Ludhiana), 28(2):264-268.
- Powell, M. E. A. and Smith, M. J. H. (1954). The Determination of Serum Acid and Alkaline Phosphatase Activity with 4aminoantipyrine. J. Clin. Pathol., 7 (3):245-248.
- Radwan, M.A, and Mohamed, M.S. (2013). Imidacloprid Induced Alterations in Enzyme Activities and Energy Reserves of the Land Snail, *Helix aspersa*. Ecotoxicol Environ Saf., 95(9):91-97.
- Russell, R.M.; Robertson, J.L, and Savin, N.E, 1977. POLO: A New Computer Program for Probit Analysis. Bull. Entomol. Soc. Am., 23(3):209-213.
- Sandor, A.; Sarospataki, M, and Farkas, S. (2015). The Mode of Action of Neonicotinoids on Insects. [Hungarian]. Novenyvedelem, 51(1): 14-24.
- Sayyed, A. H. and Wright, D. J. (2006). Genetics and Evidence for an Esterase-Associated Mechanism of Resistance to Indoxacarb in a Field Population of Diamondback Moth (Lepidoptera: Plutellidae), Pest Manag. Sci., 62(11):1045-1051.

- Shao, X.; Xia, S.; Durkin, K.A, and Casida, J. E. (2013). Insect Nicotinic Receptor Interactions in Vivo with Neonicotinoid, Organophosphorus, and Methylcarbamate Insecticides and a Synergist. Proceedings of National Academy of Sciences of USA, 110(43):17273-17277.
- Simpson, D.R.; Bulland, D.L, and Linquist, D. A. (1964). A Semimicrotechnique for Estimation of Cholinesterase Activity in Boll Weevils. Ann. Ent. Soc. Amer., 57:367-371.
- Smirle, M.J.; Zurowski, C.L.; Lowery, D.T, and Foottit, R.G. (2010). Relationship of Insecticide Tolerance to Esterase Enzyme Activity *in Aphis pomi* and *Aphis spiraecola* (Hemiptera: Aphididae). J. Econ. Entomol., 103(2):374-378.
- Stein, R.T, and Soderlund, D.M. (2012). Role of the Local Anesthetic Receptor in the state-dependent Inhibition of Voltage-gated Sodium Channels by the Insecticide Metaflumizone. Mole. Pharmacol., 81(3): 366-374.
- Thany, S. H. (2009). Agonist Actions of Clothianidin on Synaptic and Extrasynaptic Nicotinic Acetylcholine Receptors Expressed on Cockroach Sixth Abdominal Ganglion. Neurotoxicology. 30(6):1045-1052.
- Tian, X.; Rui, S.; Xing, X, and Su, J.Y. (2014). Biochemical Mechanisms for Metaflumizone Resistance in Beet Armyworm, Spodoptera exigua. Pesticide Biochemistry and Physiology, 113(7):8-14.
- Tofangsazi, N.; Cherry, R.H.; Beeson, R.C, and Jr, A.S.P. (2015). Concentration-Response and Residual Activity of Insecticides to Control *Herpetogramma phaeopteralis* (Lepidoptera: Crambidae) in St. Augustinegrass. Journal of Economic Entomology, 108(2):730-735.
- Van, A. K. (1962). A Study of House Fly Esterase by Means of Sensitive Colourimetric Method. J. Insect Physiol., 8:401-416.
- Vargas-Osuna, E. (1985). La Reproducción de *Spodoptera littoralis* (Lepidoptera: Noctuidae) y sus Alteraciones por el Virus de la Poliedrosis Nuclear (VPN) (Baculoviridae: Baculovirus). Tesis

Doctoral. Universidad de Córdoba, pp. 175.

Yalcin, M.; Mermr, S.; Kozac, L.D, and Turgut, C. (2015). Insecticide Resistance in Two Populations of *Tuta absoluta* (Meyrick, 1917) (Lepidoptera: Gelechiidae) from Turkey. Türk. Entomol. Derg., 39 (2):137-145.

Zhang, Z.; Hong, L.J, and Gao, X.W. (2012). Sublethal Effects of Metaflumizone on *Plutella xylostella* (Lepidoptera: Plutellidae). J. Int. Agricult., 11(7):1145-1150.

# **ARABIC SUMMERY**

دراسات بيوكيميائية وسمية وتطوريه و تكاثرية لاثنين من المبيدات الحشرية الجديدة على دودة ورق القطن الكبرى

> **عادل السيد حاتم - هشام أحمد سرور - عبد الناصر توفيق حسن** مركز البحوث الزراعيه - معهد بحوث وقايه النباتات - الدقى- جيزة - مصر a\_hattem@yahoo.com, hamsor98@hotmail.com

تم دراسة إستجابة دودة ورق القطن الكبرى للتركيزات القاتلة وللتركيز تحت المميت لكل من مبيد الكلوثيانيدين و مبيد الميتافلوميزون بالمعاملة ليرقات العمر الرابع عن طريق الغمر لورق الخروع لوحظ أنة قد إز دادت نسبة الموت بزيادة التركيز لكلا المبيدين وقد أظهرت النتائج أن الجرعة النصفية القاتلة LC50 ليرقات العمر الرابع هي ٢٤,٧٤ و ٢٠,٤١ جزء في المليون لمبيد الكلوثياندين ومبيد الميتافلوميزون على التوالي. وتمت دراسة التأثيرات تحت المميتة عن طريق معاملة يرقات العمر الرابع بالتركيز النصفي القاتل لكلا المبيدين. وقد أظهرت النتائج زيادة فترة تطور البرقات بصورة معنوية مقارنة بالكنترول لكلا المبيدين، و هي الفترة من المعاملة بالمبيد و حتى التعذير، أيضا لوحظ إنخفاض فترة طور العذراء بشكل ملحوظ لكلا الجنسين ولكن بصورة أقل و بشكل ملحوظ في عذراء الذكور المعاملة بالكلوثيانيدين، وإنخفض وزن العذراء بشكل كبير لكلا المبيدين مقارنة بالكنترول. ومَّع ذلك لم يتم ملاحظة أي فروق معنوية في فترة وضع البيض بينما في فترة وضع البيض المخصب تم ملاحظة فروق معنوية في فترة ما قبل وضّع البيض عند معاملة اليرقات مع ميتافلوميزون، وإنخفضت نسبة الخصوبة لكلا المبيدين مَّقارنة بالكنترول وطول فترة حياة الفراش لكلا الجنسين لكلا المبيدين. وقد أظهر التحليل البيوكيميائي أن معاملة يرقات العمر الرابع بالجرعة النصفية القاتلة LC<sub>50</sub> لكلا المبيدين لم يكن لهما تأثير معنوي على نشاط إنزيم أستيل كولين إستريز في تجانس اليرقات، فقد إنخفضت معنويا محتويات اليرقات من البروتين الكلي، وإنخفاض الألكلاين فوسفاتيز، وبيتا إستريز، وزيادة نشاط جلوتاثيون-s-ترانسفيرز. وبالإضافة إلى ذلك، أثرمبيد ميتافلوميزون بشكل ملحوظ على محتوى اليرقات من الكربو هيدرات الكلية، في حين أن مبيد كلوثيانيدين أدى إلى إنخفاض كبير في نشاط إنزيم ألفا إستيريز. كلمات المفتاحية: تركبزُ اتحت ممبت ،الحبوبة ، خصوبة البيض، طول عمرُ الفراش، القباسات البيوكيميائية ِ