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SERO-PREVALENCE OF *TOXOPLASMA GONDII* IN SMALL RUMINANTS

(With One Table)

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المسح المصلى للتوكسوبلازما جوندى فى المجترات الصغيرة

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هدفت هذه الدراسة إلى إجراء مسح مصلى ليبان مدى الإصابة (الأجسام المضادة) بعدوى التكسوبلاز ما جوندى من أربع مدن في المملكة العربية السعودية وهى الرياض ، المزاحمية المجمعة وعسير تم جمع 449 عينة مصل من أغذام وماعز تعانى من الإجهاض المتأخر وتتراوح أعمار ها بين 3 - 6 سنوات في الفترة من أكتوبر 2008 إلى أبريل 2010. تم إختبار هذه العينات باستخدام اختبار تفاعل الإنزيم المناعي المرتبط الغير المباشرة (الإيليزا). عدد ثلاثمائة وأربعين عينة كانت إيجابية لوجود الأجسام المناعية المصادة للتوكسوبلازما وبنسبة (76.84 ٪) لمجمل العينات. تم تسجيل أعلى نسب تواجد للأجسام المناعية: 100 (33.56%) في مدينة الرياض العينات. تم تسجيل أعلى نسب تواجد للأجسام المناعية: 100 (33.56%) في مدينة الرياض المزاحميه و 60 (2005%) في مدينة المجمعة على التوالي . تعد هذه الدراسة الأولى لتسجيل تواجد طفيل توكسوبلازما جوندى بين قطعان المجترات الصغيرة التى تعاني الإجهاض المتأخر بالمملكة العربية المعودية الأمر الذي يتطعان المجترات المعنيرة التي تعاني الإجليل تواجد طفيل توكسوبلازما جوندى بين قطعان المجترات الصغيرة التى تعاني الإجهاض المتأخر بالمملكة العربية السعودية الأمر الذي يتطلب مزيدا من الدراسة عن مين الموليل

SUMMARY

The current study aimed to determine the sero-prevalence of infection from four cities in kingdom of Saudi Arabia; Al Riyadh, Al Muzahmiah, Al Magmah and Asir. Samples were collected from October 2008 to April 2010. A total of 449 serum samples were collected from 3-6 years old sheep and goat suffered from late abortion. The obtained samples tested using an indirect enzyme linked immunosorbant assay (ELISA). Three hundred forty five (76.84%) of the all investigated samples were found to be positive, of which the heights ratios were 110 in Al Riyadh (33.56%) and Asir 130 (49.51%), while the lowest ratio was 45 (5.91%),

60 (10.95%) in Al Muzahmiah and Al Magmah, respectively. To the author knowledge, this is the first report indicated the existence of *Toxoplasma gondii* among small ruminants herds suffered from late abortion in KSA; so further investigations among other species to explore the potential for spread and diagnosis of *Toxoplasma gondii* are recommended.

Key words: ELISA, Small ruminants, Sero-prevalence, Toxoplasma gondii.

INTRODUCTION

Toxoplasmosis is a true zoonotic disease caused by infection with an obligate intracellular protozoan parasite; Toxoplasma gondii (Dubey and Beattie, 1988 and Buxton, 1990) which has a world wide distribution (Buxton, 1991; Akca and Neriman, 2010). While; a wide range of warmblooded animals including human being act as intermediate hosts for the parasite; the only known final hosts are cats and other felids. Cats excrete environmentally- resistant oocysts in their feces, while the organism can infect a wide range of animal species (Dubey and Beattie, 1988; Dubey, 1994). Toxoplasmosis is a world problem which indicated that T.gondi oocysts contaminate the environment due to the presence of large number of infected stray cats (Dubey and Beattie, 1988; Buxton, 1990). Oocysts shed continuously in the cat's faeces from 4 until 14 days after infection, with an expected peak output of tens millions of 6-8 days, thus; fifty grams of infected cat faeces may contain as many as million of oocysts (Dubey and Frenkel, 1972), which can remain sporulate so for over a year (Dubey, 1977).

Toxoplasmosis is a common infection of sheep and goat (Dubey and Jones, 2008), bovine (Bekele and Kasali, 1989; Akca and Neriman, 2010), equine (Riemann *et al.*, 1975) and Camiledae (Elamina *et al.*, 1992 and Sadrebazzaz *et al.*, 1998) world wid. Toxoplasmosis is a principal cause of abortion and neonatal death (Dubey and Jones, 2008). Hosts become infected by ingestion of food or milk contaminated with oocysts (Meireles *et al.*, 2003 and Skinner *et al.*, 1990) or by ingesting under cooked meat from infected animals with *Toxoplasma gondii* (Dubey *et al.*, 2005).

The presence of antibodies against *T.gondii* in the serum of the hostes with either recent or past *Toxoplasma* infection has been demonstrated by different techniques (Handman *et al.*, 1980; Kasper *et al.*, 1983 and Sharma *et al.*, 1983).

The sero-prevalence of Toxoplasmosis in goat and sheep varies among different countries and regions within a country, and up to 77% prevalence has been reported (Tenter *et al.*, 2000). Sheep and goats are very economic food animals; act as an important source of both meat and milk for humans in many countries (Dubey, 1990 and Huong *et al.*, 1998). Infection with *Toxoplasma gondii* in sheep and goat not only resulted in significant reproduction disorders as abortion or neonatal mortalities; but also has implication for public health since consumption of infected sheep and goat meat or milk can facilitate zoonotic transmission (Waldeland and Loken, 1991; Tenter and Johnson, 1997).

The purpose of this study was to investigate the sero-prevalence of *T. gondii* among sheep and goat herds from different regions in KSA.

MATERIALS and METHODS

1. Serum samples:

Blood samples were obtained from 449 sheep and goats showed late abortion, from 4 cities of KSA (Al Riyadh- Al Muzahmiah – Al Magmah – and Asir). Blood samples were collected by jugular vein puncture. Serum was removed from the clotted blood samples by centrifugation at 3000 rpm for 10 minutes then aspirate and stored at -20°C until tested.

2. Enzyme linked Immunosorbant assay (ELISA):

The ELISA kits were supplied by Institute of Pourquier, France and the test was carried out according to the manufacture's instructions. Each sample; individually tested in single well. The results were expressed as the percentage of the mean absorbance value of the samples (S) to the mean absorbance value of the positive (P) control sample provided with the diagnostic kit. The resultant S/P ratio was expressed as a percentage (S/P%) according to manufacture's recommendation where; sera with S/P% less than 40% were regarded as negative, between 40% - 50% as suspicious, and more than 50% as positive.

RESULTS

Out of 449 tested small ruminant sera, 345 (76.84%) were found to be positive to anti-toxoplasma IgG antibodies. The total positives seroprevalence were 345out of 449 representative 76.84%, of which 110 (33.56%), 45 (5.91%), 60 (10.95%) and 130 (49.51%) in Al Riyadh, Al Muzahmiah, Al Magmah, and Asir respectively (Table, 1).

Table 1: The sero-prevalence (antibodies) of *Toxoplasma gondii* infection from four cities in Saudi Arabia.

Examined		species		+ve		-ve			
animal	No. of animals	sheep	goat	Sheep	goat	sheep	goat	Total positive	positive percentage
City									
Al Riyadh	137	41	96	30	80	11	16	110	33.56
Al									
Muzahmiah	59	20	39	13	32	7	7	45	5.91
Al Magmah	82	28	54	20	40	8	14	60	10.95
Asir	171	51	120	35	95	16	25	130	49.51
Total	449	140	309	98	247	42	62	345	76.84

DISCUSSION

Toxoplasma gondii is an obligate intracellular zoonotic protozoan with a worldwide distribution. It infects humans as well as a broad spectrum of vertebrate hosts (Kim *et al.*, 2008).

Goat is one of the most important animals for meat and milk production (Pita Gondim *et al.*, 1999) which could be turned into potential sources of human Toxoplasmosis.

Toxoplasmosis in small ruminante have been widely studied due to its importance to public health, since the dissemination of the parasite for man can occur through the direct contact with domestic animal. The high prevalence of *T. gondii* infection indicated continues exposure of goats to infection due to heavy environmental contamination with oocysts shed from the observed stray cats in the farms. Similar observation was also reported in Italy (Masala *et al.*, 2003).

A plethora of commercially available products has been released in the past few years which use enzyme immunoassay (EIA) technology for detection of proteins and parasitic antigens. ELISA is a more effective screening test for *T. gondii* infection (Kim *et al.*, 2008). Most of the early methods were qualitative assessments of the presence or absence of antibodies by a color change of liquid in a microtiter plates. ELISA methods were developed to meet the demands for more rapid test results. The techniques were reported to be simple enough to be performed by nontechnical employees or sophisticated setting (Facklam, 1987).

The world sero-prevalence of Toxoplasmosis in small ruminants has been shown to have great variability, Tenter *et al.* (2000) indicated value from 0% to 77% in Ethiopia, and 81.6% in Nigeria for goats (Chantal

et al., 1994 and Arene, 1984), this agreed with the results in the present study, the sero-prevalence was (76.8 %) this indicated that Toxoplasmosis is one of the most important caustic agent of abortion and infertility in sheep and goat in Saudi Arabia.

In our present study, the high sero-prevalence of *Toxoplasma* gondii antibodies in sheep and goat may be due to the fact that; cats were extensively distributed throughout the investigated regions. Previous reports on the prevalence of *T. gondii* in Korea, 37.0% (17/46) of cats raised on Jeju island were sero-positive by ELISA (Kim and kim, 1989), 13.1% (26/198) of stray cats in a rural area near Chinju-si were positive (Sohn and Nam, 1999), in addition; a high prevalence of Toxoplasmosis within hot and humid environments is attributed to longer viability of *T.gondii* oocysts of cat under humid conditions (Fleck, 1972 and Fayer, 1981).

To the author knowledge, this is the first report indicated the existence of *Toxoplasma gondii* among small ruminant herds suffered from late abortion in KSA. Although; our focus here is on goat and sheep; further investigations are recommended among other species taking in consider that the prevalence of *toxoplasma* antibodies and infection were recorded world widely in bovine, equide in addition; it is urgently needed to control urban stray cat population and to reduce the risk of zoonotic transmission of toxoplasmosis to other animal hosts and humans. Therefore; high ratio of *T. gondii* antibodies represents an essential advance for the development of new reagents for the diagnosis of this parasitic disease.

REFERENCES

- Akca, A. and Neriman Mor (2010): Seroprevalence of Toxoplasma gondii in cattle in the Province of Kars, Turkey as Determined by ELISA. Journal of animal and Veterinary Advances 9 (5): 876-878.
- Arene, F.O.I. (1984): The prevalence and public health significance of Toxoplasma gondii in indigenous meat animals in the Niger delta. Tropical Medicine and Parasitology, 35: 133-135.
- Bekele, T. and Kasali, O.B. (1989): Toxoplasmosis in sheep, goats and cattle in central Ethiopia. Veterinary Research Communications. 13 (5): 371-375.
- Buxton, D. (1990): Ovine Toxoplasmosis: a review Journal of the Royal Society of Medicine. 83: 509-511.

- Buxton, D. (1991): Toxoplasmosis. In. Diseases of Sheep. Edited by Martin W.B., Aitken I.D., Blackwell Scientific Publications, Oxford, pp. 49-58.
- Chantal, J.; Dorchies, P.H. and Legueno, B. (1994): Study on some Zoonoses in Djibouti Rrpublic. Revue. de Medecin Veterinaire, 145: 633-640.
- Dubey, J.P. (1977): Toxoplasma, Hammondia, Besnoitia, Sarcocystis, and other tissue cyst_forming coccidian of man and animals. In Kreier J.P(Ed). Parasitic protozoa III. New York. Academic Press, pp. 101-237.
- Dubey, J.P. (1990): Status of toxoplasmosis in sheep and goats in the united state. J. Am. Vet. Med. Assoc. 196: 259-262.
- Dubey, J.P. (1994): Toxoplasmosis. J. Am. Vet. Assoc. 205: 1593-1598.
- Dubey, J.P. and Beattie, C.P. (1988): Toxoplasmosis of Animals and Man 1st Ed., CRC Press Inc., Raton, Florida, pp. 61-80.
- Dubey, J.P. and Frenkel, J.K. (1972): Cyst induced Toxoplasmosis in cats. Journal of Parasitology.19: 155-177.
- Dubey, J.P. and Jones, J.L. (2008): Toxoplasma gondii Infection in humans and animals in the United States. Int. J. Parasitol., 38: 1257-01278.
- Dubey, J.P.; Hill, D.E. and Sreekumar, C. (2005): Toxoplasmopsis. In The Merck Veterinary Manual, Kahn, (Eds). Merck and Co., New Jersey, pp. 549.
- Elamina, E.A.; Eliasa, S.; Daugschies, A. and Rommelb, M. (1992): Prevalence of *Toxoplasma gondii* antibodies in pastoral camels (Camelus dromedarius) in the Butana plains, mid-Eastern Sudan. Veterinary Parasitology. 439 (3-4): 171-175.
- *Facklam, R.R. (1987):* Specificity study of kits for detection of group A streptococci directly from throat swabs. J. Clin. Microbiol. 25: 504-508.
- *Fayer, R. (1981):* Toxoplasmosis update and public health implications. Can. Vet. J. 22: 344-352.
- Fleck, D.G. (1972): The sero-epidemiology of *Toxoplasma* infection in man. Proc. Royal. Soc. Med., 65: 50.
- Handman, E.; Goding, J.W. and Remington, J.S. (1980): Detection and characterization of membrane antigens of *Toxoplasma gondii*. J. Immunol. 124: 2578.
- Huong, L.T.; Jungsltom, B.L.L.; Uggla, A. and Bjokman, C. (1988): Prevalence of antibodies to Neospora caninum and Toxoplasma

gondii in Cattle and water buffaloes in Southern Vietnam, Vet. Parasitol, 75: 53-57.

- Kasper, L.H.; Crabb, J.H. and Pfefferkorn, E.R. (1983): Purification of a major membrane protein of *Toxoplasma gondii* by imunoabsorption with a monoclonal antibody. J. Immunol. 130: 2407.
- *Kim, S.H. and Kim, Y.J. (1989):* On the distribution of *Toxoplasma* antibodies in Cheju-do. Distribution of *Toxoplasma* antibodies in swine, cats and butchers. Korean J. Vet. Res. 29: 333-342
- Kim Hye-Youn; Yun-Ah Kim, S.; Kang, H.S.; Lee, H.G.; Rhie, Hye-Jin Ahn, Ho-Woo Nam and Sang-Eun Lee (2008): Prevalence of *Toxoplasma gondii* in Stray Cats of Gyeonggi-do, Korea. Korean J. Parasitol. Vol. 46, No. 3: 199-201.
- Masala, G.; Porcu, R.; Tanda, A.; Ibba, B.; Satta, G. and Tola, S. (2003): Survey of ovine and caprine Toxoplasmosis by IFAT and PCR assay in Sardina, Italy. Veterinary Parasitology. 117: 15-21.
- Meireles, L.R.; Galisteo, J.R.A.J. and Andrade, Jr.H.F. (2003): Serological survey of antibodies to *Toxoplasma gondii* in food animals from Sao Paulo state, Brazil. Brazilian Journal of Veterinary Research and Animal Science. 40: 267-271.
- Pita Gondim, L.F.; Barbosa, H.V.; Riberio Filho, C.H. and Saeki, H. (1999): Serological Survey of antibodies of *Toxoplasma gondii* in goats, sheep, Cattle and water buffaloes in Bahia State ,Brazil. Veterinary Parasitology. 82: 273-276.
- Riemann, HP.; Smith, AT.; Stormont, C.; Ruppanner, R.; Behymer, DE.; Suzuki, Y.; Franti, CE. and Verma, BB. (1975): Equine toxoplasmosis: a survey for antibodies to Toxoplasma gondii in horses. Am. J.Vet. Res. 36(12): 1797-1800.
- Sadrebazzaz, A.; Haddadzadeh, H. and Shayan, P. (1998): Seroprevalence of *Neospora caninum* and *Toxoplasma gondii* in camels (Camelus dromedarius) in Mashhad, Iran. Parasitology. 6: 600-601.
- Sharma, S.D.; Mullenax, J.; Araujo, F.G.; Erlich, M.H.A. and Remington, J.S. (1983): Western blot analysis of the antigens of *Toxoplasma* gondii recognized by human IgM and IgG antibodies. J. Immunol. 131: 977.
- Skinner, L.J.; Timperley, A.C.; Wightman, D.; Chatterton, J.M. and Ho_Yen, D.O. (1990): Simultanus diagnosis of Toxoplasmosis in goats and goat owner's family. Scandinavian Journal of Infectious Diseases 22: 359-361.
- Sohn, W.M. and Nam, H.W. (1999): Western blot analysis of stray cat sera against *Toxoplasma gondii* and the diagnostic availability of

monoclonal antibodies in sandwich-ELISA. Korean J Parasitol. 37: 249-256.

- Tenter, A.M. and Johnson, A.M. (1997): Phylogeny of the tissue cystforming coccidia. Adv. Parasitol., 39: 69-139.
- Tenter, A.M.; Heckeroth, A.R. and Weiss, L.M. (2000): Toxoplasma gondii from animals to human. Int. J. Parasitol., 30: 1217-1258.
- Waldeland, H. and Loken, T. (1991): Reproduction failure in goats in Norway: An investigation in 24 herds, Acta Vet. Scand., 32: 535-541.