

Bacteriological Profile of Bacteria isolated from Skin and Soft Tissue Infections

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Abstract

Background: Skin & soft tissue infections (SSTIs) are common type of infections either community or hospital aquired with a significant morbidity.

Aims: To identify the microbiological profile of Gram negative bacteria causing skin and soft tissue infections with detection of antibiotic susceptibility pattern of isolates.

Material and methods: A cross-sectional study of skin and soft tissue infections was conducted from April 2019 till January 2020. A total of 200 pus samples from SSTIs obtained from patients at the

general surgery outpatient clinic of Fayoum University Hospital. Bacterial isolation and identification was performed according to conventional microbiological methods. Antimicrobial susebtipilty test was performed by Kirby-Bauer Disc diffusion method.

Results: A total of 197 bacteria were isolated from 200 pus samples. There were 129/197 (65.4%) Gram positive cocci and 68 (34.5%) Gram negative bacilli. Among the Gram negative bacilli, *Pseudomonas aeruginosa* was the most common isolated organism (28/197 (14.2%) followed by *Klebsiella* species (27/197 (13.7%). About

11/68 (16.2%) of the Gram - negative bacteria isolates was identified as Extended spectrum beta lactamase (ESBL) while 23/68 (33.8%) were carbapenem resistant. Gram - negative bacteria were most susceptible to piperacillin/Tazobactam and meropenam (45 /68 (66.2%) and 44/68 (64.4%) respectively).

Conclusion: An alarming increase in antibiotic resistant pattern of Gram negative bacilli causing SSTIs was reported. Wise use of antibiotics and implementation of antimicrobial stewardship is a mandatory need nowadays.

Key Words – Skin & soft tissue infections , Extended spectrum beta lactamase , Antibiotic susceptibility pattern.

1) Introduction

Skin and soft tissue infections (SSTIs) are characterized by inflammatory microbial invasions of the epidermis, dermis and subcutaneous tissue, which can result in prolonged hospitalization, disability, and antibiotic therapy all over the world [1]. Several factors can contribute in SSTI classification , such as site of infection,

causative agent, , depth of infection, clinical presentation, prognosis and severity of infection [2]. The most common organisms that are involved in community acquired SSTIs include gram-positive cocci-like *Staphylococcus aureus*, *Staphylococcus pyogenes* infection [2]. Recently Gram-negative bacteria, such as *Escherichia coli* and *Pseudomonas aeruginosa* started to be an emerging pathogen that affect morbidity and mortality in skin and soft tissues [3]. Gram-negative bacteria as *P. aeruginosa* and *Klebsiella* species are resistant to some (multidrug resistant) or all (Pandrug resistant) of the antibiotic classes commonly used to treat Gram-negative bacteria: penicillins, cephalosporins, carbapenems, monobactams, quinolones, aminoglycosides, tetracyclins and polymyxins [3]. Efficient treatment of these infection depend on recognition of microbiological characteristics and antimicrobial resistance [4]. Since limited data are available concerning soft tissue infection, mortality rate and antibiotic susceptibility of Gram-negative bacteria in our hospital settings , this study aimed to evaluate the clinical and bacteriological profiles of bacterial isolates in skin and soft tissue infection including, the number and

type of potential bacteriological pathogens along with antibiotic susceptibility and pattern of isolates.

2) Material and Methods

A cross-sectional study of skin and soft tissue infection was conducted from April 2019 till January 2020, clinical samples were obtained from the patients at general surgery outpatient clinic of Fayoum University Hospital, the collected samples were processed at the Department of Medical Microbiology and Immunology, Faculty of Medicine, Fayoum University. The study was approved by Fayoum Faculty of Medicine Ethical Committee (no.58)

2.1) Collection of samples

The lesions were cleaned with sterile normal saline. Special care was taken to avoid contamination by normal flora of skin or mucus surface, where possible pus was aspirated or exudates collected, and serial number were given to code each specimen with careful labeling. The specimens were transported within 2 hours to the laboratory.

2.2) Bacterial identification

Specimens were inoculated onto different culture media including (nutrient agar, MacConkey agar, blood agar) (Oxoid Ltd., Basingstoke, UK) under aerobic and anaerobic conditions (using candle jar method) at 37°C overnight. The isolates were identified by Gram staining, colony morphology and standard biochemical tests: catalase, slide and tube coagulase, oxidase, esculin hydrolysis, bacitracin sensitivity test, indole production, citrate utilization, H₂S production, urease, and triple sugar iron fermentation tests [5].

2.3) Antimicrobial susceptibility testing

Antibiotic susceptibility tests were performed using Kirby-Bauer Disc diffusion method on Mueller-Hinton agar (MHA) (bioMérieux, Marcy L'Etoile, France) according to Clinical and Laboratory Standard Institute (CLSI) guidelines. The following antibiotics were tested (Oxoid Ltd., Basingstoke, UK): Amoxicillin (25 µg), Amoxicillin / Clavulanic acid (20/10 µg), oxacillin (1µg), piperacillin-tazobactam (100/10µg), erythromycin (15µg), azithromycin (15µg) ceftriaxone (30µg), Cefepime (30µg), ceftiofloxacin (30µg), Meropenam (10 µg) [6].

2.4) Extended spectrum beta lactamase and carbapenem resistant detection

Double- disc synergy test: The test inoculum (0.5 McFarland turbidity) was spread onto Mueller-Hinton agar (MHA) (bioMérieux, Marcy L'Etoile, France) using a sterile cotton swab. A disc of ((Oxoid Ltd., Basingstoke, UK) Amoxicillin / Clavulanic acid (AMC) (20/10 µg) was placed on the surface of MHA; then discs of cefpodoxime

(30 µg), ceftriaxone (30µg), Cefepime (30µg), ceftiofloxacin (30µg), Meropenam (10 µg) were kept around it in such a way that each disc was at distance ranging between 16 and 20 mm from (AMC) disc (centre to centre). The plate was incubated at 37 °C overnight. The organisms were considered to be producing ESBL when the zone of inhibition around any of the expanded-spectrum cephalosporin discs showed a clear-cut increase towards the augmentin disc. Meropenam resistance was used as the indication for carbapenemase production [7].

3) Results

Of the 200 samples 197 culture showed bacterial growth. All positive samples carried single pathogen. Higher distribution of infection was demonstrated among male patients (57.9%). Range of age was (1.5y to 70 y) with mean value of age was 35.5. Data are expressed in the form of mean \pm standard deviation (SD) (Table.1).

Table 1: Demographic characteristics of patients included in the study

Ages (years)	Mean	SD
	35.5	17.9
Sex (N=197)	N	%
Male	114	57.9%
Female	83	42.1%

By studying Skin and soft tissue infections distribution among different body site, extremities were the most affected site (150/197, 76.1%) while head and neck were the least affected site (3/197, 1.5%). The most prevalent type of infection was abscess formation which represented 175 /197 (88.8%) out of the total cases (**Table. 2**).

Table 2: Sites and types of skin and soft tissue infections

Site of infection	(N)	(%)
Extremities	150	76.1%
Trunk	44	22.3%
Head and Neck	3	1.6%
Type of infection		
Abscesses	175	88.8%
Surgical Site Infections	22	11.2%

Out of the 197 isolates 129 (65.4%) were Gram positive cocci and 68 (34.5%) were Gram negative bacilli.

Pseudomonas aeruginosa had the highest prevalence 28/197 (14.2%), while *Proteus* species was the least frequent isolated bacteria 5/197 (2.5%) (**Table. 3**).

Table 3: Type of isolated bacteria

Type of isolated bacteria	(N) (Total=197)	(%)
<i>E- coli</i>	8	4.06%
<i>Klebsiella species</i>	27	13.7%
<i>Proteus species</i>	5	2.5%
<i>Pseudomonas aeruginosa</i>	28	14.2%
<i>Staphylococcus aureus</i>	129	65.4%

Antibiotic susceptibility pattern of isolated Gram-negative bacteria is given in (Table. 4).

Table 4: Antimicrobial susceptibility among Gram-negative bacteria

Antibiotic	Gram- negative bacteria (Total=68)					
	Sensitive		Intermediate		Resistant	
	N	%	N	%	N	%
Erythromycin	22	32.4%	0	0.0%	46	67.6%
Azithromycin	21	30.9%	8	11.8%	39	57.4%
Amoxicillin, Clavulanic acid	29	42.6%	0	0.0%	39	57.4%
Cefoxitin	35	51.5%	0	0.0%	33	48.5%
piperacillin/tazobactam	45	66.2%	0	0.0%	23	33.8%
Ceftriaxone	25	36.8%	0	0.0%	43	63.2%
Meropenam	44	64.7%	0	0.0%	24	35.3%
Cefepime	22	32.4%	0	0.0%	46	67.6%

Our results showed that, 23/68 isolates of Gram-negative bacteria (32.2%) were carbapenem resistant and 11/68 isolates (15.4%) were extended spectrum beta lactamase (ESBL) producing isolates. *Klebsiella spp* isolates were the most resistant isolates, among *Klebsiella spp*. 13 /27 (48.1%) were carbapenem resistant and 7/27 (25.9%) were ESBL (Resistance to penicillin, cephalosporins and monobactam aztreonam) (Table. 5), (Figure. 1).

Table 5: Frequency of Extended spectrum beta lactamase producers and carbapenem resistant among Gram - negative isolates

Type of resistance	Gram - negative bacteria (N=68)							
	<i>E coli</i> (N=8)		<i>Klebsiella spp</i> (N=27)		<i>Pseudomonas aeruginosa</i> (N=28)		<i>Proteus spp</i> (N=5)	
	N	%	N	%	N	%	N	%
Carbapenem resistant (N=23)	0	0.0%	13	48.1%	10	35.7%	0	0%
Extended spectrum beta lactamase producers (N=11)	0	0.0%	7	25.9%	4	14.3%	0	0%

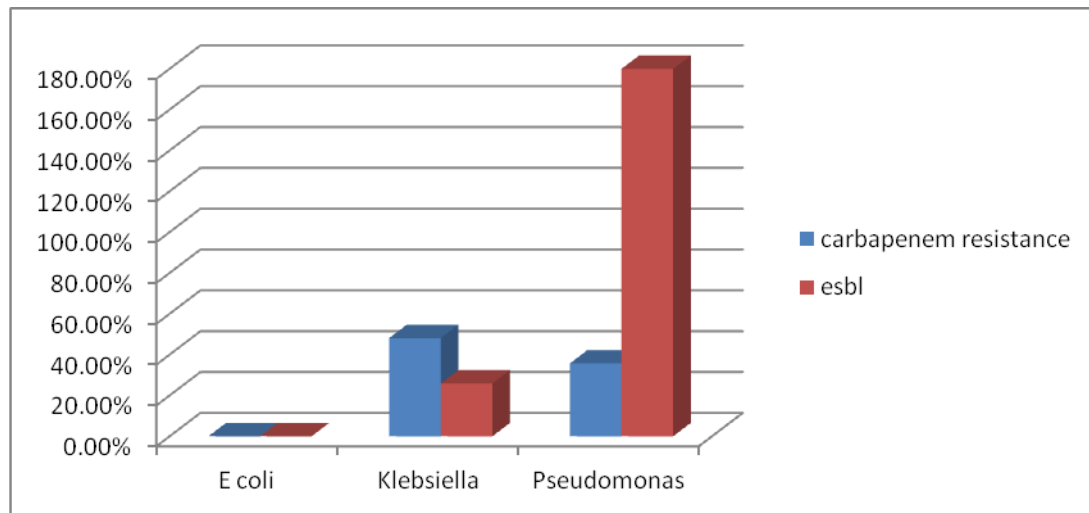


Figure. 1: Percentage of Extended spectrum beta lactamase producers and carbapenem resistant among Gram -negative bacteria

(4) Discussion

Antimicrobial resistant Gram-negative bacteria are one of the most important current threats to public health. The spread of resistant bacteria into the community is a crucial development, and is associated with increased morbidity, mortality, healthcare costs and antibiotic use. Prompt identification

of the causative pathogen requires that patients with SSTI be risk stratified according to the likelihood of resistance to enable early recognition and swift initiation of appropriate therapy. Our study aimed to assess the expanding role of Gram-negative bacteria in skin and soft tissue infections and their antibiotic susceptibility pattern. Higher distribution of infection was

demonstrated among male patients (57.9%) with mean value of age 35.5 which come in agreement with **Malhotra et al, (2012)** where the percentage of male was (67.21%) and the major age group was 31-40 years [8]. *Pseudomonas aeruginosa* was the most predominant organisms among the Gram-negative bacteria, similar finding was reported in a study conducted by **Mohanty et al, (2004)**, *Pseudomonas aeruginosa* was the most frequently isolated organism among the predominant five organisms isolated from skin and soft tissue infections in patient admitted to the hospital [9]. Maximum resistance of Gram-negative bacteria was seen against azithromycin 47(69%), erythromycin 46 (67.6%) and cephalosporins like cefepime 46 (67.6%) and ceftriaxone 43(63.2%) followed by amoxicillin, clavulanic acid 39 (57%) which come in partial agreement with **Afroz et al, (2015)** where the percentage of cephalosporins resistant GNB were (90.32%) [10]. Resistance of Gram-negative organisms was minimum against piperacillin/tazobactam 23(33.8%)

followed by meropenem 24 (35.4%) and Cefoxitin (57.4%) which is like other studies 11,12 13. Our results showed that, 23/68 isolates of gram-negative bacteria (32.2%) were carbapenem resistant and 11/68 isolates (15.4%) were ESBL producing isolates. *Klebsiella spp* was the most resistant isolated Gram-negative bacteria, 13/27 (48.1%) were carbapenem resistant and 7/23(30.4%) were ESBL producers. This rate of resistance is lower than other studies conducted in India on soft tissue infections which documented 72.37% ESBL producers among *E. coli*, 68.79% among *Acinetobacter spp* and 58.9% among *Pseudomonas spp* [9]. Use of antibiotics, inappropriate dose and duration of antibiotics, lack of compliance to the hospital antibiotic policy, lack of awareness among the prescribing clinicians and patients about antibiotic resistance are the main underlying factors of exaggeration of antimicrobial resistance [14].

5) Conclusion

In this present study the most common isolate in skin and soft tissue infection among Gram-negative bacteria is *Pseudomonas aeruginosa* followed by *Klebsiella spp.* The alarming rise in multi drug resistant Gram-negative bacteria infections implement the great need to identifying the epidemiology of the bacteria causing skin and soft tissue infections and their sensitivity pattern. Following antibiotic stewardship will help in avoiding unnecessary medication with ineffective antibiotics and prevent development drug resistance.

6) References

- [1]- Deleo FR, Otto M, Kreiswirth BN and Chambers HF (2010): Community associated meticillin resistant *Staphylococcus aureus*. *Lancet*; 375: 9725-1557.
- [2]- Esposito S, Noviello S and Leone S (2016). Epidemiology and microbiology of skin and soft tissue infections: *Curr Opin Infect Dis*; 29:109–15.
- [3]- Buhl M, Peter S and Willmann M (2015): Prevalence and risk factors associated with colonization and infection of extensively drug-resistant *Pseudomonas aeruginosa*. *Anti-Infect. Ther* ; 13:1159–1170.
- [4]- Jääskeläinen IH, Hagberg L, Forsblom E and Järvinen A (2017): Microbiological Etiology and Treatment of Complicated Skin and Skin Structure Infections in Diabetic and Nondiabetic Patients in a Population-Based Study. *Open Forum Infect Dis*; 4 : 0-44.
- [5]- Collee JG, Marr W. Specimen collection,; Collee JG, Fraser AG, Marmion BP, Simmons A, editors. Mackie and McCartney's (2012): culture containers & media. In *Practical Medical Microbiology*; 95-111.
- [6]- CLSI (2012). Performance Standards for Antimicrobial Disc Susceptibility Tests; Approved standard 11th edition. CLSI document M02-A11. Wayne, PA: Clinical and Laboratory Standards Institute.
- [7]- Jarlier V, Nicolas M, Fournier G and Philippon A (1988). Extended spectrum β -lactamases conferring transferable resistance to newer β -lactam agents in

- Enterobacteriaceae: Hospital prevalence and susceptibility patterns. *Rev Infect Dis*;10:867-78.
- [8]- Malhotra SK, Malhotra S, Dhaliwal GS and Thakur A (2012): Bacteriological Study of Pyodermas in a tertiary Care Dermatological Center. *Indian J Dermatol* ; 57 : 358-61
- [9]- Mohanty S, Kapil A, Dhawan B and Das BK (2004): Bacteriological and antimicrobial susceptibility profile of soft tissue infections from northern India. *Indian J Med Sci*;58:10-15.
- [10]- Afroz, Z. , Metri C and Jyothi P. (2015). Bacteriological profile and antimicrobial susceptibility pattern of skin and soft tissue infections among gram negative bacilli in a tertiary care hospital of south India. *Journal of Pharmaceutical Sciences and Research*; 7: 397-400.
- [11]- Sah P, Khanal R and Upadhaya S (2013): Skin and soft tissue infections: Bacteriological profile and antibiotic resistance pattern of the isolates. *J Universal College of Medical Science* ;18-21.
- [12]- Soumya K and Jaya S (2014): Prevalence and antimicrobial susceptibility patterns of bacteria isolated from skin and wound infections. *J Microbiol. Biotech. Res*; 4 : 39-45.
- [13]- Matthew S. Dryden (2010): Complicated skin and soft tissue infection. *J Antimicrob Chemother*; 65 : 35–44.
- [14]- Seni J, Najjuka CF, Kateete DP, Makobore P, Joloba ML, Kajumbula H, et al (2013): Antimicrobial resistance in hospitalized surgical patients: a silently emerging public health concern in Uganda. *BMC Res Notes*; 27(6):298.