# POTENTIALS PROTECTIVE EFFECT OF RUTA GRAVEOLENS AS ANTIOXIDANT ON CISPLATIN INDUCED TESTICULAR DAMAGE IN ALBINO RATS

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#### ABSTRACT

The present study was established to investigate that Ruta graveolens' ability to alleviate cisplatin-induced testicular dysfunction in albino rats. Thirty rats were randomly divided into six groups (A-F) of 5 rats each: Group A which attended as control (-) received water; Group B was intraperitoneally (i.p) injected 10 mg kg<sup>-1</sup> bw. cisplatin on 6 weeks as control (+). Groups C and D were assumed 50 and 100 mg of R. graveolens via oral administration, respectively, for 7 weeks and then exposed to i.p administration of 10 mg kg<sup>-1</sup>bw cisplatin on the 6<sup>st</sup> week ; Groups E and F received 10 mg kg<sup>-1</sup> bw of cisplatin intraperitoneally and then recieved 50 and 100 mg kg<sup>-1</sup> bw of R. graveolens for 6 weeks orally. Cisplatin-exposed Group B rats had decreased sperm characteristics and higher sperm morphological abnormalities, as well as altered histological architecture of seminiferous tubules and lower glutathione (GSH) levels in the testes. These parameters in R. graveolens alone treated Groups C and D were not markedly different compared with negative control group ;such as sperm count recorded 102.54±3.91 compared with 146.25±3.12 in the control (-) and 56.41±4.10 in the control (+). The rats with the combined treatment in Groups E and F showed significantly improved sperm parameters, testicular histology architecture and antioxidant enzymatic activities. Conclusively, R. graveolens has protective potential against cisplatin damage

Keywords : cisplatin, Ruta graveolens, testis, male albino rats

# **INTRODUCTION**

Cisplatin (CP) is a DNA alkylating anti-neoplastic drug created from platinum that is commonly utilised as a front-line adjuvant therapy for malignancies such as testicular, gut, stomach, head, neck, ovarian, cervical, germ cell tumours, and non-small cell lung carcinoma (Yildirim et al., 2011). Although the use of CP in the treatment of testicular cancer, even at an advanced stage of the disease, has encouraging results, it is also associated with undesired side effects such as testicular weight loss, azoospermia, and transient or permanent loss of male reproductive capability (Türk et al., 2008). Spermatogenesis is a complex process that is influenced extensively by hormonal levels and environmental factors, particularly temperature. Sertoli cells, Leydig cells, and peritubular cells are among the testicular cells involved in this occurrence (Huleihe et al., 2004 and Yamaguchi et al., 2008). Because of its alkylating properties, CP therapy inhibits nucleic acid synthesis in germ cells, particularly in spermatogonia (Seaman et al., 2003). Damage to Leydig cells caused by CP resulted in testosterone secretion suppression (Lirdi et al., 2008). The pathogenesis of reproductive abnormalities caused by CP exposure is commonly related to oxidative stress, which causes the antioxidant immune system to deteriorate.Within the human body protection against oxidative stress is achieved by selfdefense enzymes that catalytically remove the free radicals and other reactive species. Superoxide dismutase, catalase, and glutathione peroxidase are examples of these enzymes (Farias-Silva et al., 2007), CP, on the other hand, inhibits the activity of these enzymes, causing damage in testicular tissue. Chemotherapeutic oxidative treatments are currently non-selective in their potency and kill rapidly dividing cells, including cancer, normal cells, and stem cells. Such harmful substances also affect spermatogenic cells and cause infertility in men (Chirino and Pedraza-Chaverri, 2009). Antioxidant supplementation may be effective in reducing cisplatin-induced toxicity, according to a growing body of research. (Ali et al., 2006 and Yagmurca et al., 2007). As a result, developing combinatorial medicines that minimise CP resistance while minimising its negative effects is critical. Plants have a variety of elements that can help avoid oxidative stress-related illnesses, such as male reproductive diseases (*Shah et al.,2014*), As a result, it's regularly included in recipes with chemotherapy medications to help safeguard against their negative effects (*HemaIswarya and Doble, 2006*). Flavonoids are potent antioxidants that prevent lipid peroxidation and platelet aggregation (*Lirdi et al., 2008*), protect the tissue from free radicals by direct scavenging ROS, reactive nitrogen species (RNS), and activating antioxidant enzymes.

Ruta graveolens (R. graveolens) has long been used in folk medicine to cure a variety of ailments, including eye difficulties, rheumatism, dermatitis, pain, and a variety of inflammatory disorders (Ratheesh and Helen, 2007). A variety of chemical compounds have been identified from various portions of the plant, including alkaloids, coumarins, volatile compounds, terpenoids, flavonoids, and furoquinolines (Kuzovkina et al.,2004). Additional pharmacological applications of R. graveolens have been discovered in contemporary pharmaceutical trials, including antioxidant and antiinflammatory properties (Ratheesh and Helen, 2007), antinociceptive and antipyretic (Loonat, Amabeoku, 2014) antiulcerogenic (Tarique et al., 2016), antidiabetic, antibacterial (Toserkani et al., 2011) and antifungal (Meepagala et al., 2005) as well as antiandrogenic effects (Khouri et al.,2005).

The goal of this study was to see if ethanolic extracts of S. officinalis and R. graveolens could protect albino rats from testicular and cardiac toxicity caused by two regularly used insecticides, chlorpyrifos and methomyl. With the protective benefits of *R. graveolens* in mind, this study was conducted to assess *R. graveolens'* ability to protect against CP-induced histamine release.

*The present study aimed to* estimate the protective role of *R*. *graveolens* on the improvement of reproductive function in cisplatin induced toxicity of testes in male albino rats.

### **Materials and Methods**

#### <u>1-chemicals:-</u>

CP injection (Sigma-Aldrich, St. Louis, MO, United States of America) was dissolved in saline and a dose of 10 mg/kg body weight of CP was chosen based on previous research to produce testicular toxicity (*Afsar et al.,2015*). *R. graveolens* were obtained from Experimental Station of

*R. graveolens* were obtained from Experimental Station of Medical Plants (ESMP), Faculty of Pharmacy, Cairo University.

### Preparation of Ethanolic Extracts of R. graveolens

The leaves of and *R. graveolens* were washed and dried in good aerated shaded area. The dried leaves were powdered by an electric grinder. The powder of *R. graveolens* was soaked in absolute ethyl alcohol for 72 hours and the mixture was filtered. The solvent in the filtrate was evaporated by rotatory evaporator at 50°C and high pressure. The obtained viscous extract was kept in deep freezer until used.

## 2- Experimental animals:-

Male adult albino rats (30) (60 days- old, weighing 130:150g) were collected from the National Research Center's animal house in Cairo, Egypt and fed on a basal diet for eight days. The basal diet consisted of corn starch 70%, casein 10% corn oil 10%, salt mixture 4%, vitamin mixture 1% and cellulose 5% according to (AOAC, 2005). Rats were raised and kept in an air-conditioned animal housing under particular pathogen-free conditions, with a 12:12 h daylight/dark cycle and unlimited access to food and water. Animal facilities followed and supervised all ethical guidelines for animal treatment. All animal experiments received approval from the animal care committee, National Research Center.

## 3- Experimental design:-

30 male albino rats were divided into 6 groups as follows: -

A. Control: rats orally received water (5 ml kg<sup>-1</sup> bw) for 6 weeks.

B. Cisplatin (CIS): rats were given (10 ml kg<sup>-1</sup> bw) single dose of CIS intraperitoneally on the  $6^{st}$  weeks of the experiment as described by *Amin et al.* (2012).

C. 50 mg *R. graveolens*: rats received (50 ml kg<sup>-1</sup> bw) of *R. graveolens* for 6 weeks orally and then administered 10 ml kg<sup>-1</sup> bw of cisplatin intraperitoneally on the  $6^{\text{st}}$  week.

D. 100 mg *R. graveolens*: rats received 50 ml kg<sup>-1</sup> bw of *R. graveolens* for 6 weeks orally and then administered 10 ml kg<sup>-1</sup> bw of cisplatin intraperitoneally on the 6st week.

E. 50 mg *R. graveolens*: rats received 10 ml kg<sup>-1</sup> bw of cisplatin intraperitoneally and then recieved 50 ml kg<sup>-1</sup> bw of *R. graveolens* for 6 weeks orally.

F. 50 mg *R. graveolens*: rats received 10 ml kg<sup>-1</sup> bw of cisplatin intraperitoneally and then recieved 100 ml kg<sup>-1</sup> bw of *R. graveolens* for 6 weeks orally.

Five days after the administration of cisplatin, the rats were sacrificed for the evaluation of biochemical and histological variations in the testicular tissue. The mean body weights were noted once a week during the experimental period.

#### <u>Sperm count</u>

Five pairs of caudal epididymis were minced in distilled water and filtered through a nylon mesh to assess epididymal sperm count. As detailed by Pant & Srivastava (2003), the spermatozoa were counted using a haemocytometer and the enhanced Neubauer (Deep 1/ 10 m; LABART, Munich, Germany) chamber.

#### **Biochemical** assays

*Hormone analysis* Concentration of LH and FSH in the serum of different treatment groups were estimated through GenWay Biotech, Inc. Immunoassay Test Kits

Five right testes were homogenised in 50 mm Tris-HCl buffer (pH 7.4) containing 1.15% potassium chloride, and the homogenate was centrifuged at 10 000 g for 15 min at 4 °C. The supernatant was collected for the evaluation of Lipid peroxidation was quantified as MDA following the method defined by al. (2000)and expressed Farombi et as lm MDA g<sup>-1</sup> tissue. CAT activity was resolute as defined by Aebi (1974) and expressed as units mg<sup>-1</sup> of protein. GSH was estimated at 412 nm using the method designated by Jollow et al. (1974).

## Statistical analysis

Statistical analysis was passed out using one-way analysis of variance, followed by Tukey's post hoc test for multiple comparison. The results were expressed as group mean standard error of mean, while the level of significance was P < 0.05.

## **Results**& **Discussion**

In the present research, measured several hormonal, biochemicals, and histological parameters correlated to testicular toxicity and oxidative stress in the testis tissue to assess the protective effect of *R. graveolens* against Cis common effective antineoplastic agents quite used in the treatment of numerous types of cancers including urogenital system carcinoma. However, Cis has side effects such as nephrotoxicity, ototoxicity, cardiotoxicity, hepatotoxicity, and gonadal toxicity (*Hassen et al.,2021*).

#### Body weight:

Data are given in table 1, proved that the average body weights of Cis group recorded a very highly significant decrease

(p < 0.05) as compared to the control group. At the same time the average body weights of Cis rats treated with either R(LD)+Cis or R(HD)+Cis revealed a highly significant increase (p < 0.05) as compared to Cis group. Moreover, data showed a very highly significant increase in the body weight gain of Cis group treated with both Cis+R(LD)and Cis+R (HD) in relation to Cis group.

The present study revealed a significant decrease in body weight of male albino rats given Cis. Similar observation was recorded by *Hassen et al.,2021* who used Cis intraperitoneal in rats and observed a decrease in body weight of rats.

**Table 1:** Effect of cisplatin and different treatments of *R. graveolens* on body weight.

Groups Parameters	Control A	CIS B	R(LD)+CIS C	R(HD)+CIS D	CIS+R (LD) E	CIS+R (HD) F
Body weight gain (g)	244.84 ± 8.80	130.79 ± 8.42	198.79 ± 8.42*	201.77 ± 21.55*	155.11 ± 19.19 *	176.77 ± 21.55 *

(LD: low dose, HD: high dose)

Values are presented as mean ±SEM

\*: statistically significant compared to corresponding value in control group

(P<0.05)

## Changes in Sperm Parameters

The data shown in Table 2 exhibited a significant (P < 0.05) reduction in live/dead ratio (L-D), sperm count, and motility with accompanied increased percentage of morphologically after treatment with Cis (B group) relative to the control group. On the other hand *R. graveolens* pretreatment at doses of 50 and 100 mg kg/ bw significantly (P < 0.05; P < 0.01, respectively) ameliorated these altered sperm parameters in a dose-dependant manner in

Groups C and D rats, respectively, relative to Group B. Interestingly, Groups E and F rats that did not receive pretreatment with graded doses of *R. graveolens* presented no significant difference (P < 0.05) in the values of sperm parameters compared with Group B. (table 1).

In the current investigation, Cis administration contributed to a reduction in sperm count as a result of Cis toxic side-effects, resulting in a diminished testicular activity. These findings agreed with several documentations on the adverse effects of Cis on sperm characteristics. The formation of free radicals and diminished Cis-induced antioxidant enzymes result in a fast loss of intracellular adenosine triphosphate (ATP), impairing sperm motility and viability (*Türk et al.,2008*). Also, reducing sperm motility and increasing abnormal sperm rate in rats treated with cisplatin could be caused by lipid peroxidation of unsaturated fatty acids in the sperm plasma membrane, resulting in loss of fluidity and function (*Ateşşahin et al.,2006*).

Graded doses of *R. graveolens* pre-treatment offered marked protection against this Cis toxicity dose dependently.

Table 2:	Effect	of	cisplatin	and	different	treatments	of	<i>R</i> .	graveolens on
sperm cou	ınt.								

groups	Control A	CIS B	R(LD)+C IS C	R(HD)+C IS D	CIS+R (LD) E	CIS+R (HD) F
Sperm count (10 <sup>6</sup> mm <sup>3</sup> /cell)	146.25±3. 12	56.41±4.1 0	97.56±3.5 4*	102.54±3. 91*	77.14±2.15	82.14±2.65 *

(LD: low dose, HD: high dose)

Values are presented as mean ±SEM

\*: statistically significant compared to corresponding value in control group

(P<0.05)

## Hormone Results

The results of T, LH and FSH in the control and other studied groups are illustrated in Table (3). Indicating that Cis administration altered the secretion of reproductive and pituitary hormones. Cis inoculation considerably (p0.01) lowered T, LH, and FSH concentrations, which were significantly improved (P0.05; P0.01, respectively) following *R. graveolens* Cis inoculation considerably (p0.01) lowered T, LH, and FSH concentrations, which were significantly improved (P0.05; P0.01, respectively) following R. graveolens, however the hormonal concentrations did not restore to normalized control values in Groups E and F rats did not receive pretreatment with graded doses of *R. graveolens*.(table3)

Serum gonadotropin releasing hormone (GnRH) including LH, FSH levels aid in making conclusions regarding reproductive pathologies. The decrease in serum T concentration in drugtreated animals indicates either a direct effect of the chemical on Leydig cells or an indirect effect via changes in hormonal regulation of the hypothalamic-pituitary axis (HPA) owing to oxidative stress. (Latif et al., 2008). It has also been found that abnormal intratesticular testosterone concentrations hinder spermatogenesis. (Azu et al., 2010). LH stimulates the generation of testosterone in Leydig cells by activating FSH to interact with Sertoli cells and stimulate spermatogenesis (Guideline, 2001). Cis displayed a notable decrease of T, LH, and FSH concentrations in serum, according to the results of the current experiment. T has already been shown to be suppressed by Cis. Administration of R. graveolens results in potential intensification of lowered levels of T, LH and FSH and might result in regulation of HPA. Pretreatment with R. graveolens in Groups C and D rats, more significantly maintained serum hormone levels compared to Cis plus R. graveolens, indicating the preventive potential of R. graveolens.

**Table 3:** Effect of Cisplatin and different treatments of *R. graveolens* on reproductive hormone.

groups	Control A	CIS B	R(LD)+C IS	R(HD)+CI S	CIS+R (LD)	CIS+R (HD)
parameters	/		C	D	E	F
LH (ng/ml)	2.61±.01	$1.23 \pm .005$	$1.98 \pm .004$	2.01±.09*	$1.46 \pm .03$	1.84±.07*
FSH (ng/ml)	2.11±.06	.97±.03	1.86±.06*	1.99±.007*	1.51±.09	1.65±.05
Testesteron e (ng/ml)	4.65±.07	2.10±.09	3.05±.06*	3.45±.04*	2.56±.08	2.98±.008

(LD: low dose, HD: high dose)

Values are presented as mean ±SEM

\*: statistically significant compared to corresponding value in control group

(P<0.05)

#### **Biochemical analysis**

Table 4 shows the data the of *R*. on outcomes graveolens pretreatment against Cis-induced damage of antioxidant systems and lipid peroxidation in testicular tissues. There was a significant (P < 0.01) decrease in the testicular catalase (CAT) and glutathione (GSH) enzymes of Group B compared with control rats. However, testicular levels of CAT and GSH enzymes of rats in Groups C and D increased significantly (P < 0.05) when compared to the cisplatin-treated Group B rats. Besides, lipid peroxidation raised significantly (P < 0.01) in Group B relative to another group. R. graveolens pre-treatment in Groups C and D significantly (P < 0.01) ameliorated the altered MDA levels when compared to Group B. The levels of the antioxidant enzymes, the CAT, GSH activity, and MDA in the R. graveolens treated Groups E and F rats were not significantly (P <0.05) different from the Group B. (table 4)

Oxidative stress is described by an excessive reactive oxygen species (ROS) and remains a notable cause of male infertility (*Iwasaki & Gagnon, 1992*). Reactive oxygen formations and reactions with biological structures are usually attended by antioxidant systems (*Riley, 1994*). In the current study, Cis recorded that the rat's antioxidant profile developed as testicular

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MDA increased and TAC reduced, with GSH showing that and non-enzymatic antioxidant enzymatic molecules were insufficient scavenge free radicals generated to from cisplatin (Adejuwon et al., 2015). The increase in the amount of MDA may be attributed to the excess output of ROS attacking the cell membrane due to a deficiency of antioxidant enzymes (Nita et al.,2016). The decrease in their activity could be explained by the consumption of these enzymes during the transformation of free radicals into less damaging or harmless metabolites, or by the direct inhibitory effect of cisplatin on enzyme function. The testis is one of the most sensitive organs to oxidative stress and peroxidative damage, as well as low antioxidant concentration efficiency, which leads to decreased sperm vitality, motility, and infertility. (Beytur et al., 2012 and Hassen et al., 2021).

In Groups C and D rats, pretreatment with R. graveolens significantly improved the damaged antioxidant defence systems in a dose-dependent manner, enhancing the profiles of cisplatin-depleted antioxidant enzymes (CAT) and intracellular GSH, as well as the level of MDA produced in the testes. These observed ameliorations are suggested to be connected to the phyto-antioxidants components particularly flavonoids in *R. graveolens* leaves. These findings are similar to earlier reports on optimizing Phyto-antioxidants benefits in alleviating cisplatin-induced testicular injury as documented by *Jahan et al.*(2018)

It is worth discussing that the flavonoid, a group of polyphenolic compounds found mainly in plants of the Rutaceae family, have been shown to exhibit a series of biological effects among which stand out the inhibition of LPO and platelet aggregation, due to their antioxidant properties and their ability to remove free radicals and chelating divalent cations (*Ashour et al.2017*). *R. graveolens* was known for its free radical scavenging activity, which was revealed in the current study to be related to its phenolic and flavonoid components, which greatly increased antioxidant status and lowered lipid peroxidation.

**Table 4:** Effect of cisplatin and different treatments of *R. graveolens* on antioxidant and oxidative stress markers.

	Control	CIS	R(LD)+C	R(HD)+C	CIS+R	CIS+R
groups	Α	В	IS	IS	( <b>LD</b> )	(HD)
			С	D	E	F
parameters						
MDA	.62±.11	2.51±.05	1.25±.10*	1.04±.16*	2.10±.15	$2.06 \pm .18$
(n. mol/mg)						
GSH		.98±.05	2.45±.15*	3.6±.26*	1.65±.14	1.98±.23*
( <b>m.</b>						
mol/mg)	4.65±.16					
CAT	19.02±0.1	8.36±0.17	$12.02 \pm .09$	$16.25 \pm .18$	9.15±.15	9.99±0.24
(U/min)	4		*	*		

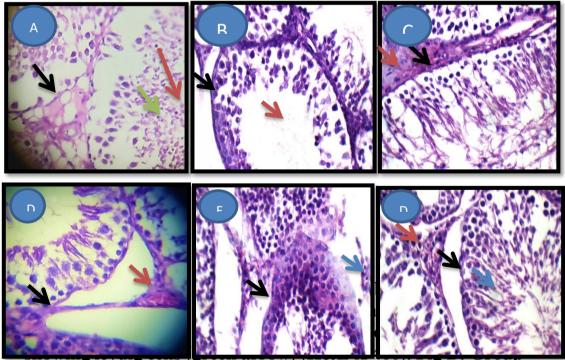
(LD: low dose, HD: high dose)

Values are presented as mean  $\pm$ SEM

\*: statistically significant compared to corresponding value in control group

(P < 0.05)

#### Histopathological Analysis



B; rue (LD): showing tubules with mildly thick BM (black arrow), average germinal lining up to full spermatogenesis, and interstitium showing increased number of Leydig cells (red arrow) (H&E X 400). Fig C: Group C; rue (HD) : higher

power view showing tubules with mildly thick BM (black arrow), average germinal lining up to full spermatogenesis, and average interstitium with average Levdig cells (red arrow) (H&E X 400). Fig D: Group D; Cisplatin: higher power view showing tubules with thick detached BM (black arrow), marked reduction of germinal lining with few sperms (red arrow), and interstitium showing increased number of leydig cells (H&E X 400). Fig E: Group E; Cis + rue (LD): showing tubules with mildly thick BM (black arrow), mild reduction of germinal lining with moderate number of sperms, and interstitium showing increased number of levdig cells (blue arrow) (H&E X 400). Fig F: Group F; Cis + rue (HD): : higher power view showing tubules with average BM (black arrow), average germinal lining up to full spermatogenesis (blue arrow), and average interstitium with average Leydig cells (red arrow) (H&E X 400).

#### Conclusion

The current study substantiates that reproductive toxicity induced by *Cis* is associated with enhanced oxidative stress. *R. graveolens* as a possible antioxidant might be employed in combination with or before chemotherapeutic drug treatment to evade the related side effects. Results increase improved sperm parameters, testicular histology architecture and antioxidant enzymatic activities. Conclusively, *R. graveolens* has protective potential against cisplatin damag.

## References

- Aebi H (1974) .Catalase. In: Methods of Enzymatic Analysis. Bergmeyer HU (ed.). Academic Press, New York, pp. 673– 677.
- Adejuwon, S.A.; Femi-Akinlosotu, O.M.; Omirinde, J.O (2015). Cisplatin-induced testicular dysfunction and its amelioration by *Launaea taraxacifolia* leaf extract. *Andrologia*, 47, 553–559, doi:10.1111/and.12302.
- 3. Ali BH, Al Moundhri MS (2006). Agents ameliorating or augmenting the nephrotoxicity of cisplatin and other platinum

compounds: a review of some recent research. Food Chem Toxicol.;44:1173-83.

- 4. Amin A, Mahmoud-Ghoneim D, Syam MI, Daoud S (2012). Neural network assessment of herbal protection against chemotherapeutic induced reproductive toxicity. Theor Biol Med Model 9:1–14.
- 5. AOAC Official Methods of Analysis of the Association of Official Analytical Chemists, 18th ed., Washington, D.C.; 2005.
- Ashour M B., Ahmed O M., Asran A M., Ali M A (2017). Assessment of the Preventive Effects of *Salvia officinalis* and *Ruta graveolens* Ethanolic Leaf Extracts on Chlorpyrifos- and Methomyl-induced Renal Toxicity and Oxidative Stress in Albino Rats. International Journal of Prevention and Treatment, 6: 34-44.
- 7. Afsar T, Khan MR, Razak S, Ullah S, Mirza B (2015). Antipyretic, anti-inflammatory and analgesic activity of Acacia Hydaspica R. Parker and its phytochemical analysis. BMC Complement Altern Med.;15:1.
- Ateşşahin, A.; Sahna, E.; Türk, G.; Ceribaşi, A.O.; Yilmaz, S.; Yüce, A.; Bulmuş, O (2006). Chemoprotective effect of melatonin against cisplatin-induced testicular toxicity in rats. *J. Pineal Res.*, 41, 21–27, doi:10.1111/j.1600-079X.2006.00327.x.
- 9. Azu OO, Duru FI, Osinubi AA, Noronha CC, Elesha SO, Okanlawon AO (2010). Protective agent, Kigelia Africana fruit extract, against cisplatin-induced kidney oxidant injury in Sprague–Dawley rats. Asian J Pharma Clin Res.;3:84–8.
- 10.Beytur, A.; Ciftci, O.; Oguz, F.; Oguzturk, H.; Yilmaz, F (2012). Montelukast attenuates side effects of cisplatin including testicular, spermatological, and hormonal damage in male rats. *Cancer Chemother. Pharmacol.*, *69*, 207–213, doi:10.1007/s00280-011-1692-y.
- 11.Chirino YI, Pedraza-Chaverri J (2009). Role of oxidative and nitrosative stress in cisplatin-induced nephrotoxicity. Exp Toxicol Pathol.;61:223–42.
- 12. Farias-Silva E, Cola M, Calvo TR, Barbastefano V, Ferreira AL, Michelatto DDP, de Almeida ACA, Hiruma-Lima CA,

Vilegas W, Brito ARS (2007). Antioxidant activity of indigo and its preventive effect against ethanol-induced DNA damage in rat gastric mucosa. Planta Med.;73:1241–6.

- 13.Farombi EO, Tahnteng JG, Agboola AO, Nwankwo JO, Emerole GO (2000) Chemoprevention of 2acetylaminofluorene induced hepatotoxicity and lipid peroxidation in rats by kolaviron a Garcinia kola seed extract. Food Chem Toxicol :535–541.
- 14.Guideline OO (2001). 425: acute oral toxicity—up-and-down procedure. Paris: OECD Guidelines for the Testing of Chemicals Organization for Economic Cooperation and Development.
- 15.Hassen, M.T.; Mohamed, H.K.; Montaser, M.M.; El-Sharnouby, M.E.; Awad, N.; Ebiya, R.A (2021).
  Molecular,Immunomodulatory, andHistopathological Role of Mesenchymal Stem Cells and Beetroot Extracton Cisplatin Induced Testicular Damage in Albino Rats. *Animals*, *11*, 1142.https://doi.org/10.3390/ani11041142
- 16.Huleihel M, Lunenfeld E (2004). Regulation of spermatogenesis by paracrine/ autocrine testicular factors. Asian J Androl.;6:259–68.
- 17.HemaIswarya S, Doble M (2006). Potential synergism of natural products in the treatment of cancer. Phytother Res.;20:239–49.
- Iwasaki A, Gagnon C (1992) Formation of reactive oxygen species in spermatozoa of infertile patients. Fertil Steril 57:409–416.
- 19.Jahan S., Munawar A., Razak S., Anam S., Qurat Ul Ain1, Ullah H., Afsar T., Abulmeaty M. and Almajwal A (2018). Ameliorative effects of rutin against cisplatin-induced reproductive toxicity in male rats. BMC Urology 18:107.
- 20.Jollow DJ, Mitchell JR, Zamppaglione Z, Gillette JR (1974) Bromobenzene induced liver necrosis: protective role of glutathione and evidence for 3,4-bromobenzene oxide as the hepatotoxic metabolites. Pharmacology 11:151–157.
- 21.Kuzovkina I, Al'terma, I, Schneider B (2004). Specific accumulation and revised structures of acridone alkaloid

glucosides in the tips of transformed roots of *Ruta graveolens*. Phytochemistry, 65: 1095-1100.

- 22.Khouri NA, EL-Akawi Z (2005). Antiandrogenic activity of *Ruta graveolens L* in male albino rats with emphasis on sexual and aggressive behavior. Neuro Endocrinol. Lett., 26: 823-829.
- 23.Latif R, Lodhi GM, Aslam M (2008). Effects of amlodipine on serum testosterone, testicular weight and gonado-somatic index in adult rats. J Ayub Med Coll Abbottabad.;20(4):8–10.
- 24.Lirdi LC, Stumpp T, Sasso-Cerri E, Miraglia SM (2008). Amifostine protective effect on cisplatin-treated rat testis. Anat Rec.;291:797–808.
- 25.Loonat F, Amabeoku GJ (2014). Antinociceptive, antiinflammatory and antipyretic activities of the leaf methanol extract of *Ruta graveolens L*. (*Rutaceae*) in mice and rats. Afr J Tradit Complement Altern Med., , 11:173-81.
- 26.Meepagala KM, Schrader KK, Wedge DE, Duke SO (2005). Algicidal and antifungal compounds from the roots of *Ruta graveolens* and synthesis of their analogs. Phytochemistry, , 66: 2689-2695.
- 27.Nita, M.; Grzybowski, A (2016). The Role of the Reactive Oxygen Species and Oxidative Stress in the Pathomechanism of the Age- Related Ocular Diseases and Other Pathologies of the Anterior and Posterior Eye Segments in Adults. *Oxid. Med. Cell. Longev.* 3164734, doi:10.1155/2016/3164734.
- 28.Pant N, Srivastava SP (2003). Testicular and spermatotoxic effects of quinalphos in rats. J Appl Toxicol 23:271–274.
- 29.Riley PA (1994). Free radicals in biology: oxidative stress and the effects of ionizing radiation. Int J Radiat Biol 65:27–33.
- 30.Ratheesh M, Helen A (2007). Anti-inflammatory activity of *Ruta graveolens* Linn on carrageenan induced paw edema in Wistar male rats. Afr. J. Biotechnol., 6: 1209-1211.
- 31.Seaman F, Sawhney P, Giammona C, Richburg J (2003). Cisplatin-induced pulse of germ cell apoptosis precedes longterm elevated apoptotic rates in C57/BL/ 6 mouse testis. Apoptosis.;8:101–8.
- 32.Shah NA, Khan MR, Naz K, khan MA (2014). antioxidant potential, DNA protection, and HPLC-DAD analysis of

neglected medicinal Jurinea dolomiaea roots.Biomed Res Int.;2014

- 33. Tarique M, Siddiqui HH, Khushtar M, Rahman MA (2016). Protective effect of hydro-alcoholic extract of *Ruta graveolens* Linn. leaves on indomethacin and pylorus ligation-induced gastric ulcer in rats. J Ayurveda Integr Med., 7:38-43.
- 34. Toserkani A, Jalali MR, Najafzaheh H (2011). Changes of lipid profiles, glucose, and hemogram after administration of *Ruta graveolens* extract in diabetic rats. Comp. Clin. Pathol., 10: 1331-1333.
- 35.–Türk G, Ateşşahin A, Sönmez M, Çeribaşi AO, Yüce A (2008). Improvement of cisplatin-induced injuries to sperm quality, the oxidant-antioxidant system, and the histologic structure of the rat testis by ellagic acid. Fertil Steril.;89:1474–81.
- 36.Yamaguchi K, Ishikawa T, Kondo Y, Fujisawa M (2008). Cisplatin regulates Sertoli cell expression of transferrin and interleukins. Mol Cell Endocrinol.; 283:68–75.
- 37.Yagmurca M, Bas O, Mollaoglu H, Sahin O, Nacar A, Karaman O, Songur A (2007). Protective effects of erdosteine on doxorubicin-induced hepatotoxicity in rats. Arch Med Res.;38:380–5.
- 38. Yildirim NC, Kandemir FM, Benzer F (2011). Beneficial effects of grape seed extract against cisplatin-induced testiculer damage in rabbits. Dig J Nanomater Biostruct.;6:155–9.

مجلة البحوث في مجالات التربية النوعية

لعشبة الروتا جرافولينز كمضاد للأكسدة علي تلف الخصيتين في التأثير الوقائي فئران الألبينو في المحدث من السيسبلاتين د. صفاء مصطفى عبد الفتاح فايد قسم الاقتصاد المنزلي –كلية التربية النوعية –جامعة عين شمس – القاهرة – مصر

الملخص العربى

تم اجراء هذه الدراسة لمعرفة قدرة عشبة روتا جبرولين على ضعف الخصية المحدث من السسبلاتين في فئران التجارب. استخدم في هذه الدراسة عدد ٣٠ فأرا تم تقسيمهم عشوائيا الي ست مجموعات (A-F) "كل مجموعة تحتوى على ٥ فئران" . المجموعة A تم تغذيتها على غذاء اساسي وتم امدادها بالمياه. تم حقن المجموعة B بجرعة ١٠ملجم سسبلاتين /كجم وزن داخل الغشاء البروتوني واستخدمت هذه المجموعة كمجموعة ضابطة موجبة "مصابة". تم اعطاء المجموعتين C & D (٥٠ و ١٠٠ ملجم من مستخلص الروتا لمدة ٦ أسابيع ثم تم حقن فئران هذه المجموعات بالسسبلاتين بجرعة ١٠ملجم سسبلاتين /كجم وزن داخل الغشاء البروتوني. تم حقن فئران المجموعتين E & F بالسسبلاتين بجرعة ١٠ملجم سسبلاتين /كجم وزن داخل الغشاء البروتوني ثم تم بعد ذلك اعطائهم (٥٠ و ١٠٠ ملجم من مستخلص الروتا لمدة ٦ أسابيع). أظهرت النتائج أن فئران المجموعة B التي تم حقنها بالسسبلاتين انخفاضا في خصائص الحيوانات المنوبة ، وزبادة في التشوهات المورفولوجية للحيوانات المنوبة، والبنيةالنسيجية المشوهة للأنابيب المنوبة، وإنخفاض نشاط مستوبات الجلوتاثيون في الخصيتين. في حين أظهرت نتائج المجموعتين C & D والتي تلقت مستخلص عشبة روتا جيرولين قبل حقنها بمادة المسبلاتين عدم وجود اختلاف ملحوظ بينهم وبين المجموعة الضابطة السالبة. بينما أظهرت نتائج الفئران في المجموعتين F & F تحسنا ملحوظا في تقديرات الحيوانات المنوبة، وبنية الخصية وانشطة الانزبمات المضادة للأكسدة.

الكلمات المفتاحية: سسبلاتين ، روتا جبرولينز ، الخصية ، فئران