

**THE EFFECT OF HIGH CONCENTRATE DIET
ON THE HISTOLOGICAL STRUCTURE OF THE
SMALL INTESTINE OF EARLY WEANED LAMBS**
(With 2 Tables and 19 Figures)

By

MONA ABD EL-FATAH ALI

(Received at 27/12/1999)

**تأثير تغذية الحملان حديثة الفطام على عثيقة عالية المركزات
على التركيب النسيجي للأمعاء الدقيقة**

منى عبد الفتاح على

اجري هذا العمل لدراسة تأثير تغذية الحملان حديثة الفطام بعلائق عالية المركزات على النمو وعلى التركيب النسيجي للأمعاء الدقيقة. أجريت هذه التجربة على ١٨ حمل قسمت على ثلاث مجموعات. المجموعة الأولى واعتبرت المجموعة الضابطة تكونت من عدد ٦ حملان عمر ١٢٠ يوم وغذيت على ٨٠% مركزات و ٢٠% سبلاخ الذرة. المجموعة الثانية تكونت من ٦ حملان حديثي الفطام عمر ٦٠ يوم وغذيت على ٩٠% مركزات و ١٠% سبلاخ الذرة والمجموعة الثالثة تكونت من ٦ حملان حديثي الفطام عمر ٦٠ يوم وغذيت على ١٠٠% مركزات فقط. تركت الحملان لتتغذى دون قيد على المركزات من خلال عذايات. وقد أظهرت الدراسات ان مخاطية الامعاء الدقيقة للمجموعة الضابطة تتكون من خملات ضيقة وقصيرة مع عدد معوية عديدة وطويلة. ووجد ان الظهارة المبطننة للأمعاء تكون من خلاية كاسية تكثر في الغدد المعوية عن الخملات وخلايا صمادية وخلاية محبة للفضة. وجد ايضا عقد ليمفاوية موجودة في الطبقة تحت المخاطية في الجزء العلوي من الامعاء (Duodenum) اما الخلايا المعوية داخلية الافراز فكانت عديدة في الجزء العلوي من الامعاء الدقيقة عنها في الجزء السفلي وكذلك كانت عديدة في الغدد المعوية عنها في الخملات. ظهرت الخلايا المعوية داخلية الافراز في غدد Brunner كانت الخلايا المعوية داخلية الافراز متعددة الاشكال فمنها المثلث والمخروطي والكاسي. وقد اظهرت الدراسات الهستولوجية انه هناك تحلل في الطبقة الطلائية المبطننة لخملات الامعاء الدقيقة في الحملان التي تغذيت على مركزات عالية. وكانت التغيرات الهستولوجية في امعاء الحملان التي تغذيت على ٩٠% مركزات عبارة عن اتساع واحتقان شديد في الاوعية الدموية مع علامات تحلل في الطبقة الطلائية المبطننة للخملات وقد تطورت مظاهر التحلل في امعاء الحملان التي تغذيت على ١٠٠% مركزات حيث ازداد عدد خلايا البلازما والخلايا الليمفاوية. ومن هذه النتائج ننصح المربي الذي يريد فطام الحملان الصغيرة بعد ٦٠ يوم

فقط من تاريخ الولادة واستبدال اللبن بالعلائق عالية التركيز أن يتم ذبح الخراف عندما تصل إلى وزن ٤٣ كجم حتى لا يزداد الضرر بالحيوان.

SUMMARY

The mucosa of the small intestine of the lambs was formed of short, narrow finger like villi and numerous long intestinal glands. The goblet cells were more abundant in the crypts than on the sides of villi. Lymphatic nodules were seen between the submucosal glands in the duodenum. The enteroendocrine cells showed a descending pattern of distribution from the duodenum to the ileum and more abundant in the crypts and base of the villi than in the upper parts of the villi. The enteroendocrine cells may occasionally found scattered between the cells lining the submucosal glands. (Brunner's glands). The enteroendocrine cells were mainly triangular, spindle or flask-shaped. The dark brown granules of the enteroendocrine cells were basally located and occasionally forming a crescent-like appearance around the unstained nucleus. The lining epithelium of the intestinal villi of early-weaned lambs fed 90% concentrate diets showed various degrees of desquamation and degeneration. The signs of degeneration increased in the small intestine of early-weaned lambs fed on 100% concentrate diets.

Key words: Diet, Histological Structure, Small Intestine

INTRODUCTION

Sheep numbers have declined markedly for several reasons, including lack of profitability compared to beef cattle production. New production techniques could reverse this pattern. Lambs can be early-weaned to dry lot and finished on high concentration diets (Bakr, and Hegazi, 1997). High-energy diets with a low roughage contents (0-10%) are efficient to promote high growth rate, (Thérize, 1975).

The small and large intestines form together a long tube adapted for the digestion, absorption of nutrients and absorption of salt and water (Smith *et al.*, 1988).

The small intestine has two principal functions (1) to complete the digestion of food by the action of appropriate enzymes, (2) to absorb selectively the final products of digestion into the blood and lymph vessels. (Turk 1982, Bank 1993 and Matthew and Robert 1996).

The aim of the present study is to illustrate the changes in the histological structure of small intestine of early-weaned lambs finished on 100% or 90% concentrate diets or feeder lambs finished on 80% concentrate diet.

MATERIALS and METHODS

Eighteen lambs were purchased from the grazing flock in Kafr El-Sheikh Governorate. These lambs were divided into 3 groups: (each one contains 6 lambs) (Table 2).

Group I: (Considered as a control group) feeder lambs, 120 days old, fed on 80% concentrate and 20% corn silage.

Group II: Early-weaned lambs, 60 days old, fed on 90% concentrate and 10% corn silage.

Group III: Early-weaned lambs, 60 days old, fed on 100% concentrate.

Ingredient compositions of concentrate finishing diets were shown in Table (1). 14-days adaptation period was used to adjust lambs to their respective high-concentrate diets. At day 15 lambs were weighed (initial weight) and switched to be self-fed the high concentrate diets. The lambs were individually weighed in early morning every 21 days throughout study. Immediately after weighing at the final conclusion of the feed trial, three lambs from each group were slaughtered.

Specimens from the small intestine of lambs were taken just after slaughtering. The specimens were fixed either in Bouin's fluid or in 10% neutral buffered formalin. After a proper fixation, the specimens were dehydrated, cleared and embedded in paraffin wax. Sections of 5 microns thick were obtained and stained with Haematoxylin and Eosin, Crossman's trichrome stain, periodic acid Schiff's technique and Grimelius silver nitrate impregnation method (Grimelius 1968). All stain techniques were adapted to that reported by Bancroft and Stevens (1990).

Table 1: Ingredients composition of high concentrate finishing diets.

Item	100% concentrate	90% concentrate	80% concentrate
Ingredient, % of DM			
Whole white corn	73.30	65.40	55.00
Soybean meal	21.00	21.00	21.00
Corn silage	-	10.00	20.00
Ground limestone	2.10	2.00	2.00
Trace-mineralized salt	1.00	1.00	1.00
Ammonium sulfate	0.50	0.50	0.50
Neomycin sulfate	0.10	0.10	0.10
Vitamin mix	0.002	0.0004	0.0002

DM = Dry matter

RESULTS

The average values of the initial and final body weights and daily gain are presented in Table (2). Early-weaned lambs fed 100% concentrate diet were reached to 43.4 kg more earlier than those fed 90% concentrate diet. With 100% concentrate diet the final body weight (43.4 kg) of early-weaned lambs was increased nearly 3 folds the initial body weight (15.40 kg) while with feeder lambs fed 80% concentrate diet (41.5 kg) was nearly 2 folds the initial body weight (23.7 kg) in the same period 83 days reflecting a faster average daily gain over(83 day) feeding period.

Table 2: Average values of performance parameters of early-weaned and feeder lambs fed high concentrate diets.

Item	Group I	Group II	Group III
No. of lambs	6	6	6
% of concentrate	80%	90%	100%
Initial body wt. Kg	23.70	15.00	15.40
Final body wt. kg	41.50	43.40	43.40
Experimental period (days)	83	125	83
Average daily gain/kg	0.21	0.23	0.34

Histological findings:

The wall of the small intestine of control group was formed of mucosa, submucosa, muscular layer and serosa.

The mucosa of the small intestine was formed of short, narrow finger like villi and numerous long intestinal glands filled nearly all lamina propria to reach muscularis mucosa (Fig. 1).

The epithelium lining the intestinal villi and intestinal crypts consisted of tall columnar cells, goblet cells and argyrophil cells.

The tall columnar cells contained vacuolated eosinophilic cytoplasm with rounded basally located nucleus. The apical surface of these cells showed a pronounced striated border (Fig. 2).

The goblet cells were scattered among the tall columnar cells. They were more abundant in the crypts than on the sides of the villi (Fig. 3).

The central core of the intestinal villi and the subepithelial lamina propria contained blood capillaries, smooth muscle fibres, fibroblasts, lymphocytes, plasma cells, eosinophils and central lacteals (Fig. 4). The central lacteals were numerous, dilated and occupied nearly the length of the villi (Fig. 3). They were lined with one layer of endothelial cells. (Fig. 5).

The muscularis mucosa consisted of thin layer of longitudinally arranged smooth muscle fibres (Fig. 6).

The submucosa of the duodenum contained clusters of branched, coiled tubular glands (Brunner's glands) that open into the intestinal crypts (Fig. 6). These glands were lined with cuboidal to low columnar cells contained vacuolated cytoplasm with basally located oval or flat nuclei (Fig. 7). Few argyrophilic cells were demonstrated scattered among the mucus cells of Brunner's glands (Fig. 8a, b).

Secondary lymphatic nodules with germinal centers were seen between the submucosal glands in the duodenum (Fig. 9).

The muscular layer was formed of inner circular and outer longitudinal layers of smooth muscle fibres. The inner layer was thicker than outer layer (Fig. 9).

In sections impregnated with silver, the enteroendocrine cells granules were stained dark brown. The distribution and shape of the enteroendocrine cells were variable in different regions of the intestine. Moreover, these cells showed a descending pattern of distribution from the duodenum to the ileum and more abundant in crypts and base of villi than the upper parts of villi (Fig. 10a, b, c).

The enteroendocrine cells were mainly triangular, spindle or flask-shaped. The cytoplasm of these cells demonstrated fine dark brown granules which were located in the basal parts of cells and

occasionally forming a crescent-like appearance around the unstained nucleus (Fig. 11a & b & c & d & e). All enteroendocrine cells rest on the basement membrane of the crypts and villi but, some cells occupied the whole length of the epithelium and showed a luminal contact, while other cells did not reach the lumen.

The lining epithelium of the intestinal villi of early-weaned lambs fed 90% concentrate diets showed various degrees of desquamation and degeneration. (Fig. 12a & b). The cytoplasm of the lining epithelium of the villi was more acidophilic than that in control group and the apical surface of the columnar cells appeared strongly acidophilic (Fig. 13), but the lining epithelium of the intestinal glands was still contact and their cytoplasm was less acidophilic than that of the villi and the apical surface of the columnar cells gave a marked positive reaction with acid fuchsin in trichrome stain (Fig. 14 & 15). At this group there was more lymphocytic infiltration and caseated acidophilic materials surrounded by inflammatory cells in the lamina propria (Fig. 16), also, the blood vessels were enlarged and engorged with blood.

The signs of degeneration increased in the small intestine of early-weaned lambs fed on 100% concentrate diets (Fig. 17). These changes were associated with an increase in the number of plasma cells and lymphocytes (Figs. 18 a & b). The cells lining the intestinal glands were also acidophilic (Fig. 19).

DISCUSSION

Our investigation revealed that the epithelium lining the intestinal villi and crypts consisted of tall columnar, goblet and argyrophil cells. The apical surface of columnar cells of control group showed a pronounced striated border, where the apical surface of the columnar cells of early-weaned lambs fed 90% or 100% concentrate were strongly acidophilic and gave a strong reaction with acid fuchsin stain. Humphrey and Turk (1974) and Telford and Bridgman (1990), revealed that the striated border is composed of parallel rows of microvilli of even height. These microvilli increase the luminal surface of each enterocyte, (Bayer *et al.*, 1975), from which extend a network of hair-like structures termed the glycocalyx. The glycocalyx and the microvilli known as the epithelial brush border, contain many of enzymes responsible for the terminal digestion of carbohydrate and

protein while the special carrier protein necessary for the absorption of hexose sugars, amino acids, vitamins, minerals and electrolytes situated on the surface of microvilli (Smith *et al.*, 1988).

The present study revealed that the goblet cells were scattered among the tall columnar cells. Telford and Bridgman (1990), reported that these cells release copious amount of mucus to protect the lining cells from abrasion and to lubricate the passage of material along the digestive tract. They may also serve to assist in absorption (Turk, 1982). It has been emphasized that, the layer of mucus secreted by the goblet cells constitutes the first line of defence for the intestine by its tenacious adherence to the underlying tissue and through its general impermeability to the destructive chemical agents (Holander, 1954, Cecil, 1990 and El-Gengchy, 1990).

The present investigation showed that the Brunner's glands were lined with cuboidal to low columnar cells contained vacuolated slightly basophilic cytoplasm. Lesson *et al.* (1985) and Telford and Bridgman (1990), reported that the secretion of Brunner's glands is strongly alkaline which neutralize the acidic chyme of the stomach and thus protects the duodenal mucosa from autodigestion. Also, to bring the intestinal contents to the optimum pH for pancreatic enzyme action (Junqueira *et al.*, 1989). Banks (1993), suggested that these glands may be the aboral continuations of pyloric glands displaced to the submucosa.

Grimelius silver impregnation, in our study, showed few argyrophilic cells scattered among the mucus cells lining the Brunner's gland. Recent immunofluorescence studies suggest that Brunner's glands contains urogastrone, a peptide hormone, that inhibits HCl production in the stomach (Lesson *et al.*, 1985, and Telford and Bridgman, 1990) and stimulate the epithelial cell proliferation (Junqueira *et al.*, 1989).

The muscular layer of the small intestine of lambs was formed of inner circular and outer longitudinal smooth muscle fibres. The inner circular layer serves to constrict and dilate different regions of the intestinal length and acting to mix the chyme with the digestive enzymes while the outer longitudinal layer is responsible for the peristaltic waves that slowly drive the chyme forward (Shecler, 1996).

Some of the enteroendocrine cells stimulate, and others inhibit certain alimentary processes. Their overlapping activities collectively regulator and integrate the activities of the GI tract, (Telford and Bridgman 1990).

The present studies revealed some variabilities in the distribution and shape of the enteroendocrine cells in different the regions of the intestine. They showed a descending pattern of distribution from the duodenum to the ileum. Banks (1993), reported that the argentaffin cells secrete serotonin, motilin, pancreatic glucagon, cholecystokinin, vasoactive intestinal peptide, somatostatin, gastrin, neurotensin, secretin and enteroglucagon. Also, Matthew and Robert (1996), added that these cells synthesize and store histamine and release it when stimulated by acetylcholine or gastrin. Histamine diffuses to nearby parietal cells to stimulate HCl secretion. On other hand, Kirkegaard *et al.* (1982), reported that glucagon and other related peptides secreted by the endocrine cells act as powerful inhibitors of gastric acid secretion. This fact could explain the abundance of endocrine cells in the proximal part of intestinal tract as observed in the present study.

Our results revealed that the enteroendocrine cells were more concentrated in the crypts and base of villi than the upper part of villi. This location could be attributed to the slow rate of migration of the newly formed enteroendocrine cells from the bases of the crypts to their superficial parts (Ferreira, 1971).

In the present study, different shapes of enteroendocrine cells could be demonstrated. Their cytoplasm showed fine dark brown granules, mainly basally located and occasionally forming a crescent like appearance around the unstained nucleus. In agreement to that reported by Schofield and Silva (1968), Nossier (1992) and Kassab (1996), in which the argyrophilic granules either occupied basal position, or sparsely distributed in some cells and densely packed in others.

The basal accumulation of the argyrophilic granules indicates the possibility of secretion by exocytosis across the basal membrane to the lamina propria (Forssman *et al.* 1969, Nichols *et al.*, 1974, Cristina *et al.*, 1978, Sjolund *et al.*, 1983 and Luis and Thompson, 1988).

The present study revealed that the lining epithelium of the intestinal villi of early-weaned lambs fed 90% concentrate diets showed various degrees of desquamation and degeneration. The signs of degeneration increased in the small intestine of early weaned lambs fed on 100% concentrate diets. McGee *et al.* (1992), stated that the dietary fibre is an important but not essential constituent of the diet. Fibre passes along the gastrointestinal tract acting as a sponge, and modulates function by delaying gastric emptying, slowing the absorption of

nutrients from the jejunum, possibly modifying bile acid absorption from the ileum, providing nutrition to caecal bacteria and increasing weight.

In our study, the number of plasma cells was increased. This increase is a hallmark of subacute and chronic inflammation (Telford and Bridgman, 1990). This inflammation may be due to that the whole corn, consumed by sheep, is crushed during eating and ruminating and an amount may escape degradation in the rumen and pass intact or coarse crushed to the small intestine (Qrskov, 1986).

Mona (1997) found a similar results in the liver and kidney of lambs fed on 90% or 100% concentrate where the histological studies of liver and kidney revealed signs of degenerative changes.

In conclusion, finishing of early-weaned lambs on a diet based on 100% concentrate diet resulting in greater daily gain compared with those finishing on 90% concentrate diet, and 10% corn silage or feeder lambs finished on 80% concentrate diet and 20% corn silage, but the histological studies of small intestine of early weaned lambs finished on 100% or 90% concentrate diet showed signs of inflammation and degeneration in the mucosa, so, we advice that these lambs must be slaughter immediately after this period of fattening (when reach to 43 kg).

ACKNOWLEDGEMENT

The author would like to acknowledge Dr.Bakr and Dr.Hegazi, staff member of animal feeding Fac. Vet. Med. Kafr El sheikh, for carrying out the experiment and supplying the specimens of this work.

REFERENCES

- Bakr, A.A. and Hegazi, E.M. (1997):* Finishing of early-weaned lambs on high concentrate diets. *Zagazig Vet. J. Vol. 25 No. 1":* pp. 26-37.
- Bancroft, J.D. and Stevens, A. (1990):* Theory and Practice of Histological techniques. Churchill Livingstone, Edinberg, London.
- Banks, W.J. (1993):* Applied Veterinary Histology. 3rd ed. Mosby Year Book, St. Louis, Baltimore, Boston, London, Toronto.

- Bayer, R.C.; Chawan, C.B.; Bird, F.H. and Musgrave (1975):* Characteristics of the absorptive surface of the small intestine of the chicken from 1 day to 14 weeks of age. *Poultry Science*, 54: 155-169.
- Cecil, T. (1990):* Cecil Essential of Medicine. 2nd ed. p. 385, W.B. Saunders Company, Philadelphia, London.
- Cristina, M.L.; Lehy, T.; Zeitoun, P. and Dufougeray, F. (1978):* Fine structural classification and comparative distribution of endocrine cells in normal human large intestine. *Gastroenterology*, 75: 20-28.
- El-Gengehy, T. (1990):* Study of goblet cells in different parts of the duodenum of adult rat. *Egypt. J. Histol.*, 13(2): 351-356.
- Ferreira, M.N. (1971):* Argentaffin and other endocrine cells of the small intestine in the adult mouse. I. Ultrastructure and classification. *Am. J. Anat.*, 131, 315-330.
- Forssman, W.G.; Orzi, L.; Pictet, R.; Renold, A.E. and Rouiller, C. (1969):* The endocrine cells in the epithelium of the gastrointestinal mucosa of the rat. *J. Cell Biol.*, 40: 692-715.
- Grimelius, L. (1968):* The argyrophil reaction in islet cells of adult human pancreas studied with a new silver nitrate procedure. *Acta Soc. Med. Upsal.* 73: 271-294.
- Holander, F. (1954):* The two components mucus barrier. Its activity in protecting the gastro-intestinal mucosa against ulceration. *Arch. Int. Med.*, 93, 107.
- Humphrey, C.D. and Turk, D.E. (1974):* The ultrastructure of normal chick intestinal epithelium. *Poultry Science*, 53: 990-1000.
- Junqueira, C.L.; Carneiro, J. and Kelley, O.R. (1989):* Basic Histology. 6th ed. a LANGE Medical Book.
- Kassab, A.M. (1996):* Some histological and histochemical studies on the development of the mucosa of the small intestine in goat. M.V.Sc. Thesis, Fac. Vet. Med. Tanta University.
- Kirkegaard, P.; Moody, A.J.; Holst, J.J.; Loud, F.B.; Skov, O.P. and Christiansen, J. (1982):* Glicentin inhibits gastric acid secretion in the rat, *Nature*, 297: 156-157.
- Lesson, R.C.; Lesson, S.T. and Paparo, A.A. (1985):* Text Book of Histology. 5th ed. W.B. Saunders Company. Philadelphia, London, Toronto, Mexico City, Tokyo. pp. 343-357.
- Luis, F. and Thompson J.C. (1988):* Neuroendocrine potential of the colon and rectum. *Gastroenterology*, 94, 832-844.

- Matthew, N.L. and Robert, M.B. (1996):* Principles of Physiology, 2nd ed. Mosby, Year Book, Inc.
- McGee, D.; James, O.; Isaacson, G. and Wright A. Nicholas (1992):* Oxford Textbook Of Pathology. Vol. 1. Principles of pathology 1st ed. Oxford University Press. New York.
- Mona, A.E. Ali (1997):* The effect of high concentrate diet on the histological structure of the liver and kidney of early weaned lambs. Benha Vet. Med. J. Vol. 8 No. (1): 119-135.
- Nichols, D.B.; Cheng, S.H. and Leblond, C.P. (1974):* Variability of the shape and argentaffinity of the granules in the enteroendocrine cells of the mouse duodenum. J. Histoch. Cytochem., 22(10): 929-944.
- Nosseir, D.A. (1992):* Postnatal study of the enterochromaffin cells in the large intestine of New Zealand rabbits. Egypt. J. Histol., 15(2): 339-355.
- Qrskov, E.R. (1986):* Starch digestion and utilization in Ruminants. J. Anim. Sci. 63: 1634.
- Schofield, G.C. and Silva, D.G. (1968):* The fine structure of enterochromaffine cells in the mouse colon. J. Anat. 103, 1-13.
- Sheeler, Ph. (1996):* Essentials of Human Physiology 2nd ed. WCB. Wm. C. Brown Publishers. Boston, Chicago, London, Mexico City.
- Sjolund, K.; Sanden, G.; Hakanson, R. and Sundler, F. (1983):* Endocrine cells in human intestine: An immunocytochemical study. Gastroenterology, 85, 1120-1130.
- Smith, E.D.; Paterson, R.C.; Scratchered, T. and Read, W.N. (1988):* Textbook of Physiology 11th ed. ELBS. Churchill Livingstone.
- Telford, R.I. and Bridgman (1990):* Introduction to Functional Histology. 7th ed. Harper & Row, Publisher.
- Thériez, M. (1975):* In: Livestock feeds and feeding. p. 331. Church, D.C. (1991), Prentice-Hall International, Inc., Englewood Cliffs, New Jersey.
- Turk, D.E. (1982):* The anatomy of the avian digestive tract as related to feed utilization. Poultry Science, 61: 1225-1244.

FIGURES

- Fig. 1 & 2:** Small intestine of control lambs showing: mucosa (A), submucosa (E), muscular layer (O) muscularis mucosa (K) intestinal villi (V), crypts (J), central lacteal (N), goblet cell (T), columnar cells (R), striated border (arrow) and lymphocytes (S) (H & E stain x: (1) 40, (2) 1000).
- Fig. 3:** Small intestine of control lambs showing: the distribution of goblet cells (T), crypts (J) and central lacteal (N) (PAS-H tech., x: 100).
- Fig. 4 & 5:** Small intestine of control lambs showing: plasma cells (S), eosinophil (x), blood capillary (W) lymphocyte (Z) and central lacteal (N) (H & E stain x: 1000).
- Fig. 6 & 7:** The duodenum of control lambs showing: Brunner's glands (U), muscularis mucosa (K). (H & E stain, x (6): 100, (7): 1000).
- Fig. 8a & b:** Small intestine of control lambs showing: The enteroendocrine cells (two arrows) in Brunner's gland (U) (Grimelius stain x: (a): 100, (b) 1000).
- Fig. 9:** Small intestine of control lambs showing: Lymphoid follicles (Q) with germinal center (m) embedded in Brunner's glands (U) (H & E stain, x: 100).
- Fig. 10a & b & c:** Sections in duodenum, Jejunum and ileum respectively showing distribution of enteroendocrine cells (arrows). (Grimelius stain x 100).
- Fig. 11a & b & c & d & e:** Sections in intestinal tract of lambs showing different shapes of enteroendocrine cells (Grimelius stain).
a, b: In crypts of small intestine (x: 400).
c, d: In villi of small intestine (x: 1000).
e: In Brunner's glands (x: 400).
- Fig. 12a & b:** Small intestine of lambs fed 90% concentrate diet (H & E stain x: (a) 100, (b) 400).
- Fig. 13:** Small intestine of lambs fed 90% concentrate diet showing acidophilia in the apical border of columnar cells (arrows) (H & E stain x: 400).
- Fig. 14:** Small intestine of lambs fed 90% concentrate diet showing acid fuchsin positive apical surface (arrow) (Crossmon's trichrome stain x 1000).

- Fig. 15:** Small intestine of lambs fed 90% concentrate diet showing intestinal glands arrows (H & E stain x 400).
- Fig. 16:** Small intestine of lambs fed 90% concentrate diet showing lymphocytic infiltration (arrow) and caseated acidophilic material (two arrows) (H & E stain x 100).
- Fig. 17:** Small intestine of lambs fed 100% concentrate diets (H & E x 100).
- Figs. 18a & b:** Small intestine of lambs fed 100% concentrate diets showing plasma cell (arrow) intraepithelial and in the lamina propria, respectively (H & E stain x 1000).
- Fig. 19:** Small intestine of lambs fed 100% concentrate diet showing acidophitic apical border in the crypts and interepithelial plasma cell (arrow) (H & E stain x 400).











