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HALOPHILIC BACTERIA IN SOME DAIRY PRODUCTS SOLD IN ASSIUT CITY

(With 6 Tables)

By

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البكتريا المحبة للملح فى بعض منتجات الالبان المباعة فى مدينة اسيوط

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تم فحص عدد ٢٠٠ عينة عشوائية من بعض منتجات الالبان المخزنة بالتعليق وتشمل الجبن الدمايطى والجبن الكاريش المخزن بالتعليق و المش وكذلك الكشك وذلك للتعرف على مدى تلوثها بالميكروبات المحبة للملحة وعدد هذه البكتيريا وأنواعها وخطورتها الصحية. وقد جمعت العينات بطريقة عشوائية من أسواق مدينة اسيوط المختلفة ومحلات البقالة حيث تم جمع ٥٠ عينة لكل منتج فى أوعية زجاجية معقمة وتم نقلها بدون تأخير الى المعمل حيث تم فحصها. أولا تم تقدير الرقم الهيدروجيني (pH) وكذلك نسبة الملح فى كل العينات المفحوصة ووجد أن متوسطهما كان ٤,٦٤ , ٥,١٧ , ٥,٣٢ , ٤,١١ بالنسبة للرقم الهيدروجيني و ٩,٨٨% , ١٠,٥٥% , ١١,٨١% , ٧,٣٢% بالنسبة لكمية الملح فى عينات الجبن الدمايطى , الجبن الكاريش , المش والكشك على التوالى. أما بالنسبة للفحص البكتريولوجى فقد وجد أن جميع العينات (١٠٠%) كانت تحتوى على البكتريا المحبة للملحة وذلك باستنباتها على مستبتين مختلفين وهما Halophilic agar المحتوى على ٣% ملح , Halophilic agar المحتوى على ١٠% ملح. بالنسبة للمستبت الأول كان متوسط العدد الكلى للبكتريا المحبة للملحة فى العينات المفحوصة على الترتيب كالاتى ٩٩× ١٠ , ١٧× ١٠ , ٢٩× ١٠ , ٩× ١٠ / جرام بينما باستنابات العينات على المستبت الثانى كان العدد الكلى لهذه البكتريا كالاتى ٣١× ١٠ , ٦٢× ١٠ , ٢٥× ١٠ , ١٣× ١٠ لكل جرام على التوالى. وقد تم عزل وتصنيف ٧٠٠ عترة من العينات المختلفة بنسب متفاوتة وكانت كالتالى:

Staph.aureus, *Staph. epidermidis*, Micrococci, *B. cereus*, *B. licheniformis*, *B.coagulence*, *B.subtilis*, *B.mycoids*, *E. coil* and *Proteus species*.

S.aureus, *Staph. epidermedis*, Micrococci, *E.coli* and عزل بينما لم يتم *Proteus species*. من عينات الكشك. كذلك تمت دراسة الخاصية السمية لميكروبات ال *Staph. aureus* المعزول من جميع العينات المفحوصة (٩٧ عترة) وقد وجد أن ٢٤ (٢٤,٨%) من هذه العترات مفرزة للسموم المعوية وهى: A, B, Enterotoxins AB, AC, ABC وذلك بالاعداد التالية ٩, ٦, ٤, ٣, عترة على الترتيب. كذلك تمت دراسة الخاصية السمية لميكروب ال *B.cereus* المعزول من هذه العينات (١٠٠ عترة) وقد وجد أن ال diarrheal enterotoxin يفرز من حوالى ٣٨ عترة (٣٨%) من ال *B.cereus* وذلك بالاعداد التالية ١٨, ٩, ١٨, ٥, من ٤٣, ٢٢, ١٩, ١٦ عترة معزولة من عينات الجبن الدمياطى, الجبن الكاريش, المش والكشك على التوالى. كذلك تمت مناقشة خطوره هذه الميكروبات على الصحة العامة بالاضافة الى الشروط والاحتياطات الواجب اتخاذها نحو تقليل تلوث هذه المنتجات بهذه الانواع من البكتريا .

SUMMARY

A total of 200 random samples of pickled Damietta, pickled kareish cheese, mish and kishk (50 each) were collected from different markets and shops in Assiut City. The average pH values of the examined samples were 4.64, 5.17, 5.32 and 4.11, while the average values of NaCl% were 9.88, 10.55, 11.81 and 7.32, of the examined samples, respectively. The samples were examined for numbers and types of halophilic bacteria. It was found that all of the examined samples contained (100%) halophilic bacteria using halophilic agar either containing 3% or 10% salt. The average numbers of halophilic bacteria recovered on halophilic agar containing 3% salt were 99×10^4 , 17×10^4 , 29×10^5 , 9×10^4 /g of the examined pickled Damietta, kareish cheese, mish and kishk samples respectively. Corresponding counts on halophilic agar containing 10% NaCl were 31×10^4 , 62×10^3 , 25×10^4 and 13×10^3 /g of the examined samples, respectively. Seven hundred (700) isolates were recovered from the examined samples in different percentages. These isolates were identified biochemically as *S.aureus*, *S.epidermidis*, Micrococci, *B.cereus*, *B.licheniformis*, *B. coagulence*, *B. subtilis*, *B. mycoids*, *E. coli* and *Proteus species*. However, *S.aureus*, *Staph.epidermedis*, micrococci, *E. coli* and *Proteus species* failed to recover from the examined kishk samples. Enterotoxigenicity of *S.aureus* isolates reveal that 24 (24.8%)

out of 97 strains recovered from the examined samples were found to produce enterotoxins A, B, AB, BC and ABC in numbers of 9, 6, 4, 3 and 2 strains, respectively. *B.cereus* could be isolated from pickled Damietta, pickled kareish, mish and kishk samples in different percentages. Diarrheal enterotoxin was produced by 38% of the tested *B.cereus* strains including 18, 9, 6, 5 out of 43, 22, 19 and 16 *B.cereus* isolates recovered from the examined samples, respectively. The public health hazard of these pathogens and preventive measures were discussed.

Key words: Halophilic, Dairy, Sold.

INTRODUCTION

Halophilic microorganisms usually require certain minimal concentrations of sodium chloride (NaCl) for their growth. Their requirement for salt in general, is not an exclusive need, since many halophiles require low levels of K^+ , Mg^{++} , and other cations and anions in addition to NaCl (Macleod, 1965; Dundas, 1977 and Kushner, 1978). Furthermore, the apparent requirement for NaCl is not specific for some bacteria, and other salts and sugars can be substituted. The level of salt required by microorganisms varies greatly. Therefore, the types of microorganisms associated with a particular salt food depend on the concentration and type of salt, as well as, type of food.

The most practical classification of halophilic bacteria is based on the level of salt required (Eimhjellen, 1965; Gibbons, 1969 and Kushner, 1978). Slight halophilic bacteria grow optimally in media containing 0.5 to 3% salt and the moderate halophiles grow in media containing 3 to 15% salt, while extreme halophiles can grow in media containing 15-30% salt. Moreover, another group of bacteria capable of growing in salt concentration 5% and frequently higher up to 12% salt or more, as well as, in media containing no salt are called halotolerant. They are gram-positive bacteria and are species of *Micrococcaceae*, the *Bacillaceae* and some of *Coryne bacterium species*. However, the species *Bacillaceae* and *Micrococcus* are considered moderately halophilic and are involved in spoilage of salted food.

Some human pathogens, such as *Staph.aureus* and *Cl.perfringens* and some strains of *Cl.botulinum* can be responsible for food poisoning (Riemann, 1969 and Reimann *et al.* 1972). Some salted dairy products including pickled Damietta, pickled kareish cheese, mish and kishk are

considered among the moderately salted food (1-15% salt by weight). These products are susceptible to microbial spoilage, and also are more likely to contain viable human pathogens. Therefore, this study was planned to secure the numbers and types of halophilic bacteria in such products as well as their public health hazard.

MATERIAL and METHODS

Collection of samples:

A total of two hundred (200) random samples of pickled Damietta, kariesh cheese, mish and kishk (50 each) were collected from different markets and shops in Assiut City. The samples were dispatched to the laboratory in sterile glass wide mouth bottles with a minimum of delay, where they were prepared for microbiological examination according to A.P. H.A. (1985).

1- Determination of pH value:

The pH value of the examined samples was determined using a pH meter (an Orion Model 701) equipped with standard electrode.

2- Determination of salt content:

Salt content of the examined samples was determined according to Atherton and Nev Lander (1977).

3- Enumeration of halophilic bacteria:

Ten fold serial dilutions were prepared from each prepared sample using halophilic broth (Gibbons, 1969). Halophilic counts were made using halophilic agar (Gibbons, 1969) according to Baross and Matches (1984). Representative colonies of different shapes and color were picked up on agar slants for further identification.

4- Identification of the isolated bacteria:

The isolated bacteria in pure cultures were identified on the basis of Gram stain, catalase and oxidase tests, as well as other biochemical reactions according to Finegold and Martin (1982).

5- Enterotoxigenicity of the isolated *Staph.aureus* strains:

The identified strains of *Staph.aureus* were tested for production of different enterotoxins using Reserved Passive Latex Agglutination Test (Unipath/Oxoid TD 900) as described by Shingaki, et al. (1981).

6- Enterotoxigenicity of isolated strains of *B.cereus*:

The identified strains of *B.cereus* were tested for their enterotoxigenicity using Reserved Passive Latex Agglutination Test (Unipath/Oxoid TD 930) as reported by Shingaki, et al. (1981). The test

was developed for detecting diarrheal enterotoxin only using microtiter plate.

RESULTS

The obtained results are recorded in Tables, 1,2,3,4,5 and 6

DISCUSSION

The results summarized in Table 1 showed that the average values of pH and salt contents of the examined pickled Damietta and kareish cheese, mish and kishk samples were, respectively 4.64, 5.17, 5.32 and 4.11 for pH and 9.88, 10.55, 11.81 and 7.32% for salt contents. It has been proved that pH value of these products decreased a long the days of storage, which may be due to accumulation of lactic acid produced from lactose fermentation. On the contrary, the salt contents of such pickled products increase gradually during storage period accompanied by decrease in moisture content (water phase). This was comparable to the results obtained by Amer *et al.* (1979) and Ahmed *et al.* (1983). As kishk is a dried fermented milk product, its pH value and salt content do not change greatly during long storage period, and they are nearly similar to what they were in the finished product after its preparation.

Evaluation of the products for slightly halophilic bacteria (Table 2), reveals that all of the examined samples contained these bacteria that recovered on a medium containing 3% NaCl. The average counts of these types were 99×10^4 , 17×10^4 , 29×10^5 and 9×10^4 /g of the examined Damietta and kareish cheese, mish and kishk samples, respectively. Also, the data of the Table 3 point out that using halophilic agar containing 10% salt favored the recovery of moderately halophilic bacteria in all of the examined samples in varying numbers, but comparatively lower than slight halophiles. The average counts of moderately halophilic bacteria were 31×10^4 , 62×10^3 , 25×10^4 and 13×10^3 /g of the examined samples, respectively.

Bacteriological identification of the different isolates recovered from the products on both types of media reveals that 97, 80 and 104 out of 700 isolates recovered from the examined products were identified as *Staph.aureus*, *Staph.epidermidis* and *Micrococcus* spp, respectively. The examined pickled Damietta and kareish cheese and mish yielded 40, 32

and 25 strains of *Staph.aureus*; 34, 27 and 19 strains of *Staph. epidermedis* and 46, 31 and 27 strains of *Micrococcus spp.*, respectively. The kishk samples yielded no of these isolates.

Furthermore, the data of Table 4 show that 100, 22, 57, 79 and 95 out of 700 isolates recovered from the examined products were identified as *B.cereus*, *B.licheniformis*, *B. coagulase*, *B. subtilis* and *B.mycoids*. The *B. cereus* could be isolated from pickled Damietta (43 strains), pickled kareish (22 strains), mish (19 strains) and kishk samples (16 strains). Out of 22 isolates identified as *B.licheniformis*, 12, 4, 6 and 0 strains were isolated from the examined samples, respectively. Moreover, the examined products yielded 18, 15, 13 and 11 strains of *B. coagulase*; 28, 22 19 and 10 *B. subtilis* strains and 25, 30, 22 and 18 strains of *B.mycoids*, respectively. On the other hand, out of the total isolates (700) recovered from the examined samples, 36 and 30 strains could be identified as *E. coli* and *Proteus spp.*, and they recovered from the examined samples cultured on medium containing 3% salt. *E. coli* could be isolated from pickled Damietta (21 strains), pickled kareish (6 strains) and mish (9 strains). Out of 30 *Proteus spp.*, 16, 8 and 6 strains could be isolated from the examined samples, respectively, while they failed to recover from kishk samples. *E.coli* and *Proteus spp.* have been isolated from Damietta and kareish cheese and other dairy products examined by Ahmed et al. (1988 a); Ahmed and Sallam (1991) and Abdel-Hady et al. (1995).

It is apparent from the previous data that kishk samples yielded only *Bacillus spp.* and failed to yield the other types of isolates, this could be attributed to the low moisture content of the product as well as the ability of *Bacillus spp.* to withstand the different stress factors. This was confirmed previously by Ahmed (1980). However, *Staph.aureus*, *Staph.epidermedis*, *Micrococcus spp.* and *Bacillus spp.* are considered moderately halophilic bacteria, they could be identified among the slight halophilic on medium containing 3% NaCl. This could be attributed to the fact that such bacteria are halotolerant and capable of growing in medium containing 5% salt and frequently higher up to 12% or more, as well as, they are able to grow on medium containing no salt.

Regarding the results recorded in Table 4, similar findings were achieved by Ahmed (1978 and 1980), who could isolate *Staph.aureus*, *Staph. epidermeidis* and *Micrococcus* from the examined Damietta and kareish cheese samples, and failed to detect them in any of the examined kishk samples. Furthermore, *S.aureus* organisms were isolated from

Damietta and kareish cheese samples examined by Ahmed et al. (1988 b) and Tawfek et al. (1988). Presence of *Staph.aureus* in such types of cheese made from unpasteurized milk is not surprizing in view of fact, that the milk itself is an endogenous source of *Staph.aureus* in dairy products. Mishandling during processing and distribution can serve as an additional sources of contamination.

Occurrence of *B.cereus* in the examined Damietta and Kareish cheese has been confirmed by El-Naway et al. (1982) and Saad (1985). The high frequency distribution of *B.cereus* among the isolates recovered from the examined products could be attributed to the contamination from surroundings, careless during processing, storage and distribution of such products, besides the ability of *Bacillus spp.* to survive pasteurization temperature as well as other stress factors.

Studying the enterotoxins production by the isolated *Staph.aureus* as recorded in Table 5, it was found that out of 97 *Staph.aureus* strains, 24 strains (24.8%) were found to be enterotoxigenic. They produce enterotoxins A (9 strains); B (6 strains); AB (4 strains); BC (3 strains) and ABC (2 strains). Out of 40 *Staph.aureus* recovered from Damietta cheese 11 strains (27.5%) were enterotoxins producers, 4 and 3 strains produced enterotoxins A and B, respectively, while one strain produced 2 enterotoxins A and B, and 2 strains produced B and C. Only one strain secreted 3 enterotoxins A, B and C. Kareish cheese samples had 7 enterotoxigenic strains (21.9%) out of 32 *Staph.aureus* recovered from the examined samples. They secreted enterotoxins A, B, AB and BC by 3, 1, 2 and 1 strain, respectively. Entertoxins A, B, AB and ABC were produced by 6 (24%) out of 10 *Staph. aureus* strains isolated from mish samples. Enterotoxins A and B were produced by 2 strains each, while one strain produced two enterotoxins, A and B. One strain secreted 3 enterotoxins AB and C. Abdel-Hakiem (1992) recorded lower occurrence of enterotoxigenic strains that 15 (18.7%) out of 80 *Staph.aureus* recovered from cheese samples produced enterotoxins. Enterotoxins A and C were produced by 2 and 10 strains, respectively. One strain produced D and 2 strains secreted AC. Different data were obtained by Ahmed (1980) who found lower findings that 8 out of 89 *Staph.aureus* strains recovered from Damietta cheese were enterotoxigenic and they produced A, B and E (one strain each), AE and BD were produced by 2 strains each, while one strain secreted 3 enterotoxins ABE. From kareish cheese one out of 13 *Staph.aureus* strains produced enterotoxin D, however, Ahmed et al. (1988 c) reported

that *Staph.aureus* recovered from Damietta and kareish cheese samples failed to produce enterotoxins A or C.

The public health hazard of enterotoxigenic *Staph.aureus* has been well documented and incriminated in several cases of food poisoning out breaks due to consumption of contaminated dairy products (Casman and Bennet 1965; Zehren and Zehren, 1968; Bryant et al., 1988 and Bone et al., 1989). In addition to food poisoning, *Staph.aureus* has been reported to cause clinical infection in the skin and mucous membranes due to epidermolytic toxin. It causes chronic purulent inflammations of all organs, wound infections, abscesses, pyoderma, osteomyelitis and can lead to septicemia (Johnson, 1981).

Testing the isolated *B.cereus* strains recovered from the examined samples for diarrheal enterotoxins proved that 38 (38%) out of 100 *B.cereus* strains were diarrheal enterotoxin producers (Table 6). Out of 43 *B. cereus* strains isolated from Damietta cheese samples 18 (41.9%) were enterotoxigenic while 9 (40.9%), 6 (31.6%) and 5 (31.3%) out of 22, 19, 16 *B.cereus* strains recovered from kareish, mish and kishk samples, respectively, were found to be diarrheal enterotoxins producers (Table 6). Higher incidence of enterotoxigenic *B.cereus* was detected by Abdel-Hakeim (1992) who stated that 34% of the tested *B.cereus* recovered from cheese samples were diarrheal enterotoxin producer. Also, Granum et al. (1993) recorded a higher percentages (59%) of enterotoxigenic *B.cereus* strains. It has been well established that *B.cereus* was found to be the cause of several food poisoning outbreaks (Hauge, 1950 & 1955); Nikodemusez et al. (1962); Midura et al. (1970); Public Health Laboratory Service (1972); Taylor and Gilbert (1975); Mossel, (1982); Midura et al. (1970) and Gianella and Brasile (1979).

Considering the public health hazard of *E.coli* organisms, they have been associated with severe diarrhea in infants and young children. Also, they are involved in food poisoning gastroenteritis (Riley et al., 1983; Anon, 1987; Hitchins et al., 1992 and Anon, 1994). Also, the presence of *Proteus spp.* in the examined samples is of public health importance as they have been encountered in some cases of summer diarrhea in infants and were also incriminated in some cases of urinary tract affections (Smith and Canant, 1960 and Mackie and Maccartney, 1962).

Furthermore, it has been well reported that these bacteria (*Bacillus spp.*, *E.coli* and *Proteus*, are capable to decarboxylate one or more amino acids with production of biogenic amines during the long

storage period of these products. These bacteria decarboxylate the amino acid tyrosine and histidine liberating tyramine and histamine (Rice *et al.*, 1976 and Beutling, 1993). Presence of these biogenic amines in dairy products could be of public health hazard associated with their consumption. It has been reported that tyramine and histamine are responsible for cutaneous symptoms (itching and flushing), gastrointestinal symptoms, hemodynamic symptoms (hypo or hypertension), as well as, neurologic symptoms (Murray *et al.*, 1982 and Varnam and Sutherland, 1995).

It is clearly evident from this study that, some dairy products are preserved by salting and pickling in brine solution. However, they are still harbouring certain types of bacteria which are called halophilic or halotolerant bacteria. Some of these halophiles, including *Staph. aureus*, *B. cereus*, *E. coli* and proteus are of public health hazard through implication in several food poisoning outbreaks. Furthermore, some of these bacteria have the ability to carboxylate amino acid during the long period of storage producing biogenic amines (tyramine and histamine) which are of public health hazard. Pasteurization of milk is an essential step followed by strict hygiene during processing, storage and handling of such products to destroy or even minimize the existing bacteria to negligible numbers that represent no health hazard.

REFERENCES

- A.P.H. A. (1985): Standard Methods for the Examination of Dairy Products. 15th Ed. Am. P. H. A., Washington, D. C.
- Abd El-Hady, H.M.; Halawa, M.A. and El-Shinawy, S. H. (1995): Surveillance of Enterohemorrhagic *Escherichia coli* (*E. coli* O157: H7) in milks and Kareish cheese. *Assiut Vet. Med. J.* Vol. 33, No. 66.
- Abdel-Hakiem, E.H. (1992): Incidence and public health importance of some food poisoning organisms in milk and some dairy products. Ph. D. thesis, Faculty of Vet. Med., Suiz Canal University.
- Ahmed, A. A-H. (1978): A survey of the incidence of coagulase positive staphylococci in milk and dairy products. M.V. Sc. Thesis, Fac. Vet. Med. Assiut Univ.

- Ahmed, A. A-H. (1980): Studies on the enterotoxigenicity of *Staph. aureus* isolated from milk and milk products. Ph. D. Thesis, Fac. Vet. Med. Assiut Univ.
- Ahmed, A. A-H.; Abdel-Rahman, A. H. and Moustafa, M. K. (1988 a): Incidence of Enterobacteriaceae in some selected food stuffs. Assiut Vet. Med. J. 20, 4 (104-109)
- Ahmed, A. A-H.; Ahmed, S.H. and Moustafa, M.K. (1988 b): Occurrence of Fecal coliforms and enteropathogenic E.coli (EEC) in Egyptian soft cheese. J. Food Prot., 51 (6) 442-444.
- Ahmed, A. A-H.; Moustafa, M.K.; Saad, N.M. and Ahmed, S.H. (1988 c): occurrence of staphylococci in milk and some dairy products. Assiut Vet. Med. J. 19: 86-89.
- Ahmed, A. A-H.; Moustafa, M.K. and Marth, E.H. (1983): Growth and survival of *Staph. aureus* in Egyptian Domiati cheese. J. Food Prot. 46: 412-415.
- Ahmed, A. M. and Sallam, S.S. (1991): Public Health significance of coliforms in raw milk and Domiati cheese. Assiut, Vet. Med. J. Vol. 25, No. 50, 89-92.
- Amer, S. N.; Fahmi, A. H. and El-Batwy, M. A. (1979): A comparison between some properties of Domiti cheese made with adult bovine and Calves rennet as affected by different salt concentration. Egyptian J. Dairy Sci. 7: 55-62.
- Anon (1987): Outbreak of gastrointestinal disease. Ontario, J. Food Prot., 50, 436-439.
- Anon (1994): Outbreak of *Escherichia coil* O157:H7 associated with contaminated milk. International Food safety News, 3 (7), 74.
- Atherton, H.V. and Newlander, J.A. (1977): A chemistry and testing of dairy products. 4th ed. AVI Publishing company. Inc. Westport-Connecticut.
- Baross, J.A. and Matches, J.R. (1984): In The compendium of Methods for Microbiological examination of Foods. American Public Health Association. Washington, D. C. USA.
- Beutling, D.V. (1993): Studies on the formation of tyramine by microbes with food hygienic relevance. Arch. Lebensmittelhyg. 44. 83.
- Bone, F.J.; Bogie, D. and Morgan. Jones, S.C. (1989): Staphylococcal food poisoning from sheep's milk cheese. Epid. Inf., 103, 249-258.

- Bryant, R.G.; Jarvis, J. and Guibert, T. (1988): Selective enterotoxin production by *Staph.aureus* strains implicated in a foodborne outbreak. J. Food Prot. 51: 130-131.
- Casman, E.P. and Bennete, R.W. (1965): Detection of staphylococcal enterotoxin A in foods. Appl. Microbiol. 13: 181-189.
- Dundas, I.E.D. (1977): Physiology of halobacteriaceae. In H. Rose and D. W. Tempest, Vol. 15. Advances in Microbiology and physiology. Academic Press. London.
- El-Naway, M.A.; El-Mansy, H.A. and Khalafalla, S.M. (1982): *Bacillus cereus* in some Egyptian dairy products. Zbl. fur Bakt., Mikrobiol. und Hyg. 4: 541-547.
- Eimhjellen, K. (1965): Isolation of extremely halophilic bacteria. pp. 126-138. In M. Schlegel (ed) Anreicherungskultur und Mutantenauslese. Gustav Fisher Verlag. Stuttgart, Germany.
- Finegold, S.H. and Martin, W.J. (1982): Bailey and Scott. Diagnostic Microbiology. 6th Ed. C.V. Mosby Co. St. Louis, London
- Giannella, R.A. and Brasile, L. (1979): A hospital foodborne outbreak of diarrhoea caused by *Bacillus cereus*. J. Infec. Dis. 139: 366-370.
- Gibbons, N.E. (1969): Isolation, growth and requirements of halophilic bacteria. pp. 169-183. In J. R. Norris and D. W. Ribbons (ed). Methods in Microbiology. Academic Press. New York, N. Y. Vol. 3B.
- Gramum, P.E.; Brynestad, S. and Kramer, J.M. (1993): Analysis of enterotoxin production by *Bacillus cereus* from dairy products food poisoning incidents and non-gastrointestinal infections. Int. J. Fd. Mic., 17,4,269-279.
- Hauge, S. (1950): *Bacillus cereus* as a cause of food poisoning. Nordisk. Hyg. Tidsk. 31: 184-206.
- Hauge, S. (1955): Food poisoning caused by aerobic sporeforming bacilli. J. Appl. Bact. 18: 591-595.
- Hitchins, A.D.; Hartman, P.A. and Todd, E. C.D. (1992): Coliforms *Eschericia coil* and its toxins. Chapter 24, Compenium of methods for the microbiological examination of foods. 3rd. Ed. Am. Public Health Assoc. Washington, D. C.
- Johnson, A.D. (1981): Production of biochemically different types of exfoliatin from two strains of *Staph. aureus*. Staphylococci and Staphylococcal Infection. Ed. Gustav Fischer Verlag, Stuttgart and New York.

- Kushner, D.J. (1978):* Life in high salt and solate concentration: Halophilic bacteria. pp. 317-368. In D. J. Kushner (ed) Microbial life in extreme Environments. Academic Press. London, England.
- Mackie, T.J. and Maccartney, J.E. (1962):* Hand book of practical bacteriology, 10th. Ed., E. and S. Livingstone Ltd., London.
- Macleod, R.A. (1965):* The question of the existence of specific marine bactria Bacteriol. Rev. 29: 9-23.
- Midura, T.M.; Wood, R. and Leonard, A.R. (1970):* Outbreak of food poisoning caused by *Bacillus cereus*. Health Rep. 85:879-881.
- Mossel, D.A. (1982):* Microbiology of foods. 3rd Ed. The Univ. of Uterecht, The Netherlands.
- Murray, C.K.; Hobbs, G. and Gilbert, R.G. (1982):* Scombrototoxin and Scombrototoxin-Like Poisoning from canned fish. J. Hyg. Camb. 88: 215
- Nikodemusz, I.; Bondar, S.; Bojan, M.; Kiss, M.; Kiss, P.; Loczko, M. and Papay, D. (1962):* Aerobic sporeformer as food poisoning. Zbl. Bakt. I. Orig. 184: 462-470.
- Public Health Laboratory Service (1972):* Bacillus cereus food poisoning Brit. Med. J. 1: 189-192.
- Rice, S.L.; Eitenmiller, R.R. and Koehler, P.E. (1976):* Biologically active amines in food: A review. J. Milk food Technol. 39: 353.
- Riemann, H. (1969):* Food processing and preservation effects. PP. 489-541. In H. Riemann (ed) Food borne infection and Intoxication. Academic Press. New. York. Chapter XII.
- Riemann, H.; Lee, W.H. and Genigeorgis, C. (1972):* Control of *Clostridium botulinum* and *Staphylococcus aureau* in semi-preserved meat products. J. Milk food Technol. 35. 514-523.
- Riley, L.W.; Remis, R.S.; Helgerson, S.D. McGee, H.B.; Wells, J.G.; Davis, B.R.; Herbert, R. J.; Olcott, E.S.; Johnson, L.M.; Hargrett, N.T.; Blake, P.A. and Cohen, M.L. (1983):* Hemorrhagic colitis associated with a rare *Escherichia coli* serotype. N. Eng. J. Med., 308: 681-685.
- Saad, Nagah. M. (1985):* Occurrence of *Bacillus cereus* in milk and milk products in Assiut City. Ph. D. Thesis. Fac. Vet. Med. Assiut Univ.

- Shingaki, M.; Igarashi, H.; Fujikawa, H.; Ushida, H.; Terayama, T. and Sakai, S. (1981): Study on Reversed Passive Latex Agglutination for the detection of staphylococcal enterotoxins. Ann. Rep. Tokyo Metr. Res. 32: 128-131.*
- Smith, D.T. and Conant, N.F. (1960): Zinser Microbiology, 12th Ed., Appleton Century Craft. INC. New York.*
- Tawfek, N.H.; Sharaf, O.M. and Hewedy, M.M. (1988): Incidence of pathogens and staphylococcal enterotoxins in Kareish cheese. Egyptian J. Dairy Sci. 16:295-300.*
- Taylor, A.J. and Gilbert, R. J. (1975): Bacillus cereus food poisoning a provisional serotyping scheme. J. Med. Microbiol. 8. 543-550.*
- Varnam, A.H. and Sutherland, J.P. (1995): Meat and meat products .1st Ed. Chpman and Hall, 2-6 Boundary Row, London SE 18 HN, UK.*
- Zehren, V.L. and Zehren, V.E. (1968): Examination of large quantities of cheese for staphylococcal enterotoxins A. J. Dairy Sci. 51:635-644.*

Table 1. Average pH values and NaCl% of the examined samples

Product	No. of samples	average pH value	average NaCl %
Pickeled Damietta	50	4.64	9.88
Pickeled kareish	50	5.17	10.55
Mish	50	5.32	11.81
Kishk	50	4.11	7.32

Table 2. Incidence and counts of halophilic bacteria in the examined samples (using halophilic agar containing 3% NaCl)

Samples	Positive samples		Counts/g		
	No/50	%	Min.	Max.	Average
Pickeled Damietta	50	100	12×10^3	42×10^5	99×10^4
Pickeled kareish	50	100	85×10^2	90×10^4	17×10^4
Mish	50	100	29×10^3	24×10^6	29×10^5
Kishk	50	100	11×10^2	22×10^4	9×10^4

Table 3. Incidence and counts of halophilic bacteria in the examined samples (using halophilic agar containing 10% NaCl)

Samples	Positive samples		Counts/g		
	No/50	%	Min.	Max.	Average
Pickeled Damietta	50	100	4×10^2	22×10^5	31×10^4
Pickeled kareish	50	100	5×10^2	40×10^4	62×10^3
Mish	50	100	6×10^2	35×10^6	25×10^4
Kishk	50	100	6×10^2	90×10^3	13×10^3

Table 4. Frequency distribution of the isolated halophilic bacteria recovered from the examined samples using two different halophilic agar media

Isolates	Total	Pickeled Damietta		Pickeled Kareish		Mish		Kishk	
		No.	%	No.	%	No.	%	No.	%
<i>S.aureus</i>	97	40	41.2	32	33.0	25	25.8	0	0.0
<i>S.epidermidis</i>	80	34	42.5	27	33.8	19	23.7	0	0.0
<i>Micrococci</i>	104	46	44.2	31	29.8	27	26.0	0	0.0
<i>B.cereus</i>	100	43	43.0	22	22.0	19	19.0	16	16.0
<i>B.licheniformis</i>	22	12	54.5	4	18.2	6	27.3	0	0.0
<i>B.coagulans</i>	57	18	31.6	15	26.3	13	22.8	11	19.3
<i>B.subtilis</i>	79	28	35.4	22	27.8	19	24.1	10	12.7
<i>B.mycoids</i>	95	25	26.3	30	31.6	22	23.2	18	18.9
<i>E.coli</i>	36	21	58.3	6	16.7	9	25.0	0	0.0
<i>Proteus spp.</i>	30	16	53.3	8	26.7	6	20.0	0	0.0
Total	700	283	40.4	197	28.1	165	23.6	55	7.9

Table 5. Coagulase, Tnase and enterotoxins produced by the isolated *Staph. aureus* strains

The products examines	No. of strains	Coagulase +ve strains		Tnase +ve strains		Enterogenic strains		Types of enterotoxins																					
								A			B			AB			BC			ABC									
		No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%								
Pickeled Damietta	40	33	82.5	26	65.0	11	27.5	4	36.4	3	27.3	1	9.1	2	18.2	1	9.1	2	18.2	1	9.1	0	0.0	1	2.5	0	0.0	1	2.5
Pickeled Karcish	32	24	75	18	56.3	7	21.9	3	42.9	1	14.3	2	28.6	1	14.3	0	0.0	1	28.6	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
Mish	25	10	76	13	52.0	6	24	2	33.3	2	33.3	1	16.7	0	0.0	0	0.0	1	16.7	0	0.0	0	0.0	0	0.0	0	0.0	1	16.7
Kishk	0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0	0	0.0
Total	97	76	78.4	57	58.8	24	24.8	9	9.3	6	9.3	4	4.1	3	3.1	2	2.1	3	3.1	2	2.1	0	0.0	2	2.1	0	0.0	2	2.1

Table 6. Diarrheal enterotoxin produced by the isolated *B.cereus* strains

Products examined	No. of strains tested		Toxicogenic strains	
	No.	%	No.	%
Pickeled Damietta	43	41.9	18	41.9
Pickeled karcish	22	40.9	9	40.9
Mish	19	31.6	6	31.6
Kishk	16	31.3	5	31.3
Total	100	38.0	38	38.0