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**VALIDITY OF GNRH AND MELATONIN  
INTERACTION ON SOME REPRODUCTIVE  
PERFORMANCE OF BARKI RAMS**  
(With 2 Tables and 4 Figures)

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**استخدام الهرمون الحاث لإفراز الجونادوترفين مع الميلاتونين  
لرفع الكفاءة التناسلية للكباش البرقى**

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استخدم لهذه الدراسة عدد ١٢ كبشاً برقياً بالغاً حيث تم تقسيمهم إلى أربعة مجموعات (٣ كباش في كل مجموعة) وتم حقن المجموعة الأولى بـ ١٥ جم من الميلاتونين تحت الجلد وحقنت المجموعة الثانية عضلياً بـ ١٢٥ ميكروجرام من الهرمون الحاث لإفراز الجونادوتروفين. أما المجموعة الثالثة فحقنت بالميلاتونين في اليوم الأول والثالث ثم بالهرمون الحاث لإفراز الجونادوتروفين في اليوم السابع والعاشر بنفس الجرعة والطريقة في المجموعتين السابقتين. بالنسبة للمجموعة الرابعة تم حقن الهرمون الحاث لإفراز الجونادوتروفين أولاً ثم الميلاتونين بنفس طريقة الحقن في المجموعة الثالثة. تم تجميع عينات دم وسائل منوي قبل وأثناء فترة الحقن ثم استمر التجميع حتى ٩ أسابيع. قيمت عينات السائل المنوي من حيث حجم القذفة، تركيز الحيامن نسبة الحركة الأمامية، نسبة الحيامن الحية، التشوهات الشكلية للحيوان المنوي، محيط غشاء الصفن وزمن الوثبة، كما تم قياس تركيزات الهرمون الحاث لحويصلات جراف وهرمون الأباضة وهرمون الذكورة. أسفرت النتائج على أن حقن الميلاتونين في المجموعة الأولى كان له تأثيراً معنوياً حيث زادت نسبة الحيامن الحية وتركيز الحيامن ومحيط غشاء الصفن وزمن الوثبة والتشوهات الشكلية للحيوان المنوي مع زيادة في تركيزات الهرمونات الثلاثة بالنسبة لقبل المعالجة. أما في المجموعة الثانية فقد قل حجم القذفة وزمن الوثبة وتركيز الحيوانات المنوية معنوياً أما نسبة الحيامن الحية والتشوهات الشكلية وهرمون الذكورة فقد ازدادت معنوياً عن قبل الحقن. بالنسبة للمجموعة الثالثة أظهرت النتائج انخفاضاً معنوياً في نسبة الحركة الأمامية وهرمون الذكورة بينما نتائج المجموعة الرابعة فقد أظهرت زيادة معنوية في نسبة الحيامن الحية

والهرمون الحاث لحويصلات جراف وهرمون الذكورة وقلة في هرمون الإباضة بالنسبة لقبول المعالجة وقد رجع زمن الوثبة إلى معدله الطبيعي بعد أن زاد أثناء التجربة. نستخلص من هذه الدراسة أن حقن الميلاتونين بعد الهرمون الحاث لإفراز الجونادوتروفين صالحاً لرفع الكفاءة التناسلية في الكباش البرقي.

## SUMMARY

12 Sexually, mature Barki rams were used. They were divided into four comparable groups (3 rams/each), Group (1) was injected S/C twice daily by 1.5 mg melatonin, then injections were repeated at 3<sup>rd</sup> and 7<sup>th</sup> day. Group (2) was injected by 125 µg GnRH I/M at the same regimen of group 1. Group (3) was injected by melatonin at the 1<sup>st</sup> and 3<sup>rd</sup> day than followed by GnRH injection as in the first two groups. Group (4) was injected by GnRH followed by melatonin as the third group. Blood and semen samples were collected before and all over the period of treatment and persist till 9 weeks as post treated period. Semen samples were evaluated for semen volume, sperm concentration, percentages of live spermatozoa individual motility and total sperm abnormalities. Scrotal circumference and reaction time were measured. Plasma FSH, LH and testosterone were also determined. The obtained results revealed that melatonin injection (Group 1) was significantly increased the percentages of live spermatozoa, sperm concentration, scrotal circumference, reaction time and total sperm abnormalities. Also the levels of FSH, LH and testosterone were increased. While GnRH injection (group 2) decreased significantly the semen volume, reaction time and sperm concentration but with a significant increase in percentage of line spermatozoa, total sperm abnormalities sperm concentration and testosterone secretion. Injection of melatonin followed by GnRH (group 3) showed a significant decrease in percentage of individual motility and testosterone. While injection of GnRH followed by melatonin (group 4) revealed a significant increase in percentage of live spermatozoa, FSH and testosterone with decrease in LH level. The reaction time increased then return back to the normal. It was concluded that the use of melatonin after GnRH treatment are validated for reproductive improvement of Barki rams.

*Key words: GnRH and Melatonin Interaction on Reproduction of Barki Rams.*

## INTRODUCTION

Sheep is a photoperiodic species with tendency to the short day length for stimulating their reproductive cycle (Lincoln and Kelly 1989). This tendency is created by the pineal gland which modulate photoperiodic information to the hypo thalamohypophyseal, gonadal axis (Bhagat *et al.*, 1994; Finley, *et al.*, 1995) resulting in rhythmic secretion of melatonin with affinity to darkness (Rasmussen, 1993; Malpoux *et al.*, 1996) Studies on the effect of either photoperiod (Schanbacher *et al.*, 1979; Ravault, *et al.*, 1986) or melatonin implants (Chemineau *et al.*, 1988; Fitzgerald and Stellflug, 1991) were seemed to be of practical value in sheep management. Melatonin clearly affects the activity of the reproductive axis in many diverse species, but whether melatonin acts directly on gonadal tissue to either activate or inhibit function, or at the level of the hypothalamus, is still not clear (Kennaway and Hugel, 1992). Several studies by (Valenti *et al.*, 1995) reported the inhibitory effect of Melatonin on testosterone secretion by purified rat Leydig cells. Thus, a relationship was supposed to present between melatonin and GnRH in their action on the reproductive cycle of rams (Kelly *et al.*, 1994).

So, the present study was designed to assess the effects of Melatonin and/or GnRH in a synchronizing manner on the semen characteristics, scrotal circumference, reaction time and the associated hormonal changes (FSH, LH and testosterone) in Barki rams.

## MATERIALS and METHODS

12 sexually, mature healthy Barki rams aged 3-5 years old were used. The rams were bred at the experimental farm of Animal Reproduction Research Institute (ARRI) Giza, Egypt. They were housed in an open yard under natural light condition. They were fed on farm diet which consists of concentrate mixture contains 14% crude protein and 11% crude fiber, 4 kg green corn (Darawa). Water and mineral blocks were offered *ad libitum*. At the start of the experiment (Summer season) rams were divided into four comparable groups (3 rams/each), Group (1) was injected S/C twice daily (9 am<sup>m</sup> and 9 pm<sup>e</sup>) by 1.5 mg melatonin (Memphis Co. for Pharm. & Chemical Ind. Cairo Egypt), injections were repeated at 3<sup>rd</sup> and 7<sup>th</sup> day.

Group (2) was injected I/M by 125 µg GnRH (Fertagyl) Gonadorelin, Intervet Boxmeer Holland). At the same regimen of melatonin group.

Group (3) was injected by melatonin at the 1<sup>st</sup> and 3<sup>rd</sup> day followed by GnRH injection at the 7<sup>th</sup> and 10<sup>th</sup> day by the same route and dose as in the first two groups.

Group (4) was injected by GnRH at the 1<sup>st</sup> and 3<sup>rd</sup> day followed by Melatonin at the 7<sup>th</sup> and 10<sup>th</sup> day as the third group.

Heparinized blood samples were collected from all rams 3 times before injection and served as control. After injection Heparinized samples were taken twice daily at the 1<sup>st</sup>, the 3<sup>rd</sup>, 7<sup>th</sup> day and every 15 days for 9 weeks, and in group 3 and 4 exceeded by samples taken at the 10<sup>th</sup> days. The blood samples were centrifuged for 15 minutes at 3000 rpm then plasma were separated and stored at - 20°C for hormonal assay (FSH, LH and testosterone).

FSH and LH concentrations were determined by RAI (Radioimmunoassay) following the technique described by (Zia and Coombes, 1981) Testosterone was measured by the double antibody radioimmunoassay (Yen & Jaffe, 1978) using kits obtained from DPC (Diagnostic Production Corporation, Loss Angeles CA, USA).

Semen samples were collected using artificial vagina where first and second ejaculates were pooled from each ram.

Morning semen collections from control and treated rams were done weekly in a parallel time to blood collection.

Immediately after collection semen samples were evaluated (Salisbury *et al.*, 1978) for semen volume (ml), sperm concentration ( $\times 10^6$  sperm/ml), Percentages of individual motility, live spermatozoa and abnormal spermatozoa (Bloom, 1983). Scrotal circumference (mm) and reaction time (Sec.) also was measured.

All data were statistically analyzed using Costat computer Program, version 3.03 Copy right (1986) Cottort Software.

## RESULTS

It was clear from the present results (Table 1) that melatonin injection significantly increase the percentage of live spermatozoa (treated and post treated periods), sperm concentration, scrotal

circumference, reaction time (treated period) and the percentage of total sperm abnormalities at the post treated period only. Neither semen volume nor percentage of individual motility affected by melatonin injection. Concerning the hormonal levels, melatonin injection was resulted in a significant increase in FSH, LH at the 3<sup>rd</sup> day and testosterone at the 1<sup>st</sup> day of the treated period.

At the same time, GnRH injection (Table 1) revealed a significant decrease in semen volume, reaction time (treated and post treated periods) and sperm concentration (Post treated period). While, a significant increase was observed in percentages of live spermatozoa (treated and post treated periods), total sperm abnormalities (post treated period) and sperm concentration (treated period). On the other hand, percentage of individual motility and scrotal circumference not affected by GnRH injection. Hormonal secretion under the effect of GnRH injection showed a significant increase in testosterone level at the last injection followed by a decrease in the 2<sup>nd</sup> week of treated period. No detectable change in FSH and LH concentrations.

Injection of melatonin in group 3 (Table 2) at the 1<sup>st</sup> and 3<sup>rd</sup> day showed an increased in percentage of individual motility and sperm concentration without any change in the mean values of hormonal concentrations. The following injection of GnRH at the 7<sup>th</sup> and the 10<sup>th</sup> day resulted in a significant decrease in percentages of sperm motility, live spermatozoa and testosterone level without any effect on the other parameters measured. The semen and blood samples collected after treatment showed a significant decrease in percentage of individual motility and testosterone level with a significant increase in percentage of individual motility and FSH level.

On the other hand, injection of GnRH in group 4 (Table 2) for two alternative days resulted in a significant decrease in percentage of individual motility and sperm concentration with a significant increase in FSH and testosterone concentration. The following injection of melatonin at the 7<sup>th</sup> and the 10<sup>th</sup> day revealed a significant increase in percentages of live spermatozoa, total sperm abnormalities, FSH and testosterone secretion. The post injection samples showed increase in percentage of live spermatozoa, FSH and testosterone with decrease in LH level. The reaction time increased then return back to the normal.

Fig. 1, 2, 3 and 4 showed a hormonal level of FSH, LH and testosterone under the effect of melatonin and/or GnRH injection.

## DISCUSSION

Many trials have been adopted to synchronize the reproductive cycle of sheep as photoperiodic species (Lincoln and Ebling, 1985; Lincoln and Kelly 1989 and Malpoux *et al.*, 1996). Most of these trials depended mainly on striking the hypothalamic-hypophyseal gonadal axis to evoke the sexual and testicular activities. This mechanism is controlled by some factors among them is the pineal melatonin secretion (Binkley, 1983; Bittman *et al.*, 1983 and Finley *et al.*, 1995).

The use of melatonin as a sexual promotor in sheep were reported by Morgan *et al.* (1992) and Robinson *et al.* (1993). The mechanism by which melatonin increased reproduction were through GnRH (Rasmussen, 1993), Sperm survival (Anwar *et al.*, 1996), testicular LH concentration (Amador *et al.*, 1986), testicular testosterone secretion (Boulakoud and Goldsmith, 1995) and sperm concentration, motile spermatozoa (Asher *et al.*, 1987) on the other hand, many authors reported that melatonin decrease reproductive performance through decrease LH (Christopher *et al.*, 1991), testosterone secretion (Anwar, 1987), testis width, Score count of spermatogenesis (Steinlechner Niklowitz, 1992 and Xiao *et al.*, 1994).

The data obtained showed that melatonin increased semen picture through percentage of live spermatozoa and sperm concentration as reported by Asher *et al.*, (1987). The increased effect on the percentage of live spermatozoa due to the antioxidant effect of melatonin (Poeggeler *et al.*, 1993). Also, melatonin increased FSH, LH and early testosterone which may be resulted in orgasmic reaction of testosterone on the concentration of sperm.

The activating effect of melatonin injection on hormonal level was confirmed by Morgan and Williams (1989) they proved the activating effect of melatonin on the hypothalamic receptors, enhancing the GnRH release (Rasmussen, 1993) and consequently increase gonadotropins as well as testosterone level (Amador *et al.*, 1986 and Boulakoud & Goldsmith 1995).

The use of GnRH in the reproduction was commonly accepted.

The previous reports showed an increase in testicular size (Lincoln, 1979), LH and testosterone (Jenweiz and Jhonson, 1983), Spermatogenesis (Courot and Ortavant, 1981) and improve semen production and Libido (Rao, 1990 and El-Azab *et al.*, 1996).

The present data agree with the previous results in improvement of semen picture and decreased in the reaction time. The prolongation of testosterone as a result of GnRH was previously reported with Jenweiz and Johnson (1983).

One of the most common problems in the use of GnRH in the field of reproduction is the desensitizing effect as reported by Change *et al.* (1990) and Parinaud *et al.* (1992).

This desensitizing effect of GnRH may be due to the loss of its receptors by higher or prolonged doses (Davis *et al.*, 1978 and Hertel and Perkins, 1984).

The use of melatonin pre and post GnRH treatment to decline its desensitizing effect and improve semen picture and reproductive performance were discussed in the present results.

Firstly, melatonin pre treated GnRH were resulted in decrease in percentage of individual motility and testosterone secretion with increase total sperm abnormalities accompanied by higher increase in FSH. On the other hand, melatonin post GnRH treatment resulted in improvement spermatogenesis in the form of percentage of live spermatozoa and improve the sperm concentration and semen volume as compared to GnRH injection alone. Moreover the increase in testosterone during the length of experimental periods were obtained. The third benefit is the decrease in the LH level due to its utilization by testes for sustained testosterone level. FSH also increased by combination of both treatments for spermatocytogenesis.

Finally, the use of melatonin after GnRH treatment are validated for reproductive improvement of Barki rams.

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## REFERENCES

- Amador, A.G.; Richardson, B.A.; Klemcke, H.G.; Vaughan, H.K.; Steger, R.W; Bartke, A. and Reiter, R.J. (1986):* Effect of Long-term treatment with melatonin or melatonin plus ectopic pituitary transplants on testicular LH/HCG and prolactin receptors in Juvenile Syrian hamsters (*Mesocricetus ouraatus*) *J. Reprod. Fert.* 78: 557-564.
- Anwar, M.M. (1987):* The effect of melatonin on the gonads of adult male rats. M.V.Sc. Thesis, Physiology. Fac. Med. Assiut University.
- Anwar, M.M.; Megahed, G.A.; Thorya S. El-Deeb and Shehata, H.S. (1996):* The effect of melatonin on the bull liquid semen and enzymatic release in seminal plasma. *Assiut Vet. Med. J.*, 35: 42-63.
- Asher, G.W.; Day, A.M. & Barrell, G.K. (1987):* Annual cycle of live weight and reproductive changes of farmed male fallow deer (*Dama dama*) and the effect of daily oral administration of melatonin in summer on the attainment of seasonal fertility. *J. Reprod. Fertil.*, 79: 353-362.
- Bhagat, L.; Duraiswami, S. and Muratidhar, K. (1994):* Mode of action of inhibin like pineal antigonadotropin is different from melatonin during compensatory ovarian hypertrophy. *J. Pineal Res.* 16: 193-197.
- Binkley, S.A. (1983):* Circadian rhythms of pineal function in rats. *Endocrin. Rev.*, 4: 255-270.
- Bittman, E.L.; Katnard, A.H.; Olster, D.H.; Robinson, J.E.; Yellon, S.M. & Karsch, F.J. (1983):* Pineal melatonin mediates photoperiodic control of pulsatile luteinizing hormone secretion in the ewe. *Neuroendocrin.*, 40: 409-418.
- Bloom, E. (1983):* The spermogram of bull. *Nordisk veterinar medicine*, 35: 105.
- Boulakaud, M.S. and Goldsmith, A.R. (1995):* The effect of duration of exposure to short days on the gonadal response to long days in male starlings (*Sturnus vulgaris*). *J. Reprod. and Fert.* 104: 215-217.



- Change, Ys; Kim, SH.; Shin, C.J.; Moon, Sy.; and Lee, Ju. (1990):* The efficacy of a combination administration of gonadotropin - releasing hormone against and gonadotropins for controlled ovarian hyperstimulation in IVF Program. *Asia-Oceania Journal of Obstetrics and Gynecology* 16(4): 337-345.
- Chemineau, P.; Pelletier, J.; Guerin, Y.; Colas, G.; Ravault, J.P.; Toure, G.; Thimonier, J. & Ortavant, R. (1988):* Photoperiodic and melatonin treatment for the control of seasonal reproduction in sheep and goats. *Reprod. Nutr. Develop.*, 28 (2B): 409-922.
- Christopher, S.; Max, M.; Hudson, D. and Menaker, M. (1991):* Excitatory amino acid receptors may mediated the effects of light on the reproductive system of the Golden hamster. *Biol. Reprod.* 44: 604-608.
- Costat Computer Program Copyright (1986):* Version 3:03 Copyright Software.
- Courot, M. and Ortavant, R. (1981):* Endocrine control of spermatogenesis in the ram. *J. Reprod. Fert.* (30), 47-60.
- Davies, T.F.; Dufan, M.L., and Catt, K.J. (1978):* Gonadotrophin receptors: Characterization and clinical application. *Clinics in obstetrics and gynecology* Vol. 5 (2) August.
- El-Azab, A.I.; Abdel-Ghafar, A.E.; Nasr, M.T. and Issa, E.S. (1996):* Effect of exogenous PGF<sub>2</sub>-Alph and/or GnRH on libido and semen production of aged Friesian bulls. *Vet. Med. J.*, 44(1): 529-533.
- Finley, C.M.; Gorman, M.R.; Tuthill, C.R. & Zucket, I. (1995):* Long-term reproductive effects of single long day in the Siberian hamster (*Phodopus sungorus*). *J. Biol. Rhythmus*, 10: 33-41.
- Fitzgerald, J.A. and Stellflug, J.N. (1991):* Effects of melatonin on seasonal changes in reproduction of rams. *J. Animal Sci.*, 69: 264-275.
- Hertel, C. and Perkins, J.P. (1984):* Receptor-specific mechanisms of desensitization of  $\beta$ -avenergic receptor Function. *Mol. all. Function. Mol. Cell Endocrinol.* 37, 245-256.
- Jenwe, P.E. and Johnson, B.H. (1983):* Phynotypic variation in testosterone and LH production among boars; differential response to GnRH and ACTH. *Biol. Reprod.*, 29 (2): 464-471.

- Kelly, K.K.; Goldman, B.D. and Zucker, (1994):* Gonadal growth and hormone concentrations in photoregressed Siberian hamsters: Pinealectomy versus photostimulation. *Biol. Reprod.* 51: 1046-1050.
- Kennaway, D.J. and M. Hugel H.M. (1992):* Mechanism of action of melatonin within the central nervous system. *Anim. Reprod. Sci.* 30: 45-65.
- Lincoln, G.A. (1979):* Use of pulsed infusion of luteinizing hormone releasing hormone to mimic seasonally induced endocrine changes in the ram. *J. Encor.* 83, 251-260.
- Lincoln, G.A. and Ebling, F.J. (1985):* Effect of constant release implants of melatonin on seasonal cycles in reproduction, prolactin secretion and moulting in rams. *J. Reprod. Fertil.*, 73: 241-253.
- Lincoln, G.A. and Kelly, R.W. (1989):* Test of ML23 as an antagonist to the effect of melatonin in the ram. *J. Reprod. Fertil.*, 86: 737-743.
- Malpoux, B; Viguie, C.; skinner, D.C.; thiery, J.C.; Pelletier, J. and Chemineau, P. (1996):* Seasonal breeding in sheep: Mechanism of action of melatonin. *Anim. Reprod. Sci.*, 42: 109-117.
- Morgan, P.J and Williams (1989):* Central melatonin receptors : implication for a mode of action : *Experientia* . 45: 955- 965.
- Morgan, P.J.; Barrett, Davidson, G. and Lawson, W. (1992):* Melatonin regulates the synthesis and secretion of several proteins by pars tuberalis cells of the ovine pituitary. *J. Neuro-endocr.*, 4: 557-563.
- Parinaud, J.; Oustry, P.; Bussenot, I.; Tourre, A.; Perineau, M; Plantavid, M.; Monrozies, X; Reme, J.M; Pontonieer, G. (1992):* Paradoxical ovarian stimulation in the use of LHRH analogs. *European Journal of Obstetrics, Gynecology, & Reproductive Biology* 47(2) 129-133.
- Poeggeler, B.; Reiter, R.J.; Tan, D.X.; Chen, L.D. & Manchester, L.C. (1993):* Melatonin, hydroxyl radical mediated oxidative damage and aging: A hypothesis. *J. Pineal Res.*, 14: 151-168.
- Rao, A.U. (1990):* Treatment with GnRH for correction of libido problems in Murrah bulls. *Ind. Vet. J.*, 67 (8): 760.

- Rasmussen, D.D. (1993):* Diurnal modulation of rat hypothalamic gonadotrophin releasing hormone release by melatonin in vitro. *J. Endocrin. Invest.*, 16: 1-7.
- Ravault, J.P.; Matinet, L.; Bonneford, C.; Claustrate, B. and Brun, J. (1986):* Diurnal variation of plasma melatonin concentrations. *J. Pineal Res.*, 3: 365-373.
- Robinson, J.J.; Wallace, J.M. Aitken, R.P. and McNeilly, A.S. (1993):* Effect of chronic treatment with a GnRH agonist to suppress pulsatile LH secretion on the ability of exogenous melatonin to advance estrous cyclicity in ewes. *J. Reprod. Fert.* 99, 601-608.
- Salisbury, G.W; Van Demark, N.L. and Lodge, J.R. (1978):* Physiology of reproduction and artificial insemination of cattle. W.H. Freeman Co. San Francisco, London.
- Schanbacher, D.; Bruce and Ford, J.J. (1979):* Photoperiodic regulation of ovine spermatogenesis: Relation to serum hormones. *Biol. Reprod.*, 20: 719-726.
- Steinelchner, S. and Niklowitz, P. (1992):* Impact of photoperiod and melatonin on reproduction in small mammals. *Anim. Reprod. Sci.* 30: 1-28.
- Valenti, S.; Guide, R.; Giusti, M. and Gordono, G. (1995):* In vitro acute and prolonged effects of melatonin on purified rat Leydig cell steroidogenesis and adenosine 3, 5-monophosphate production. *Endocrin.* 136: 5357-5362.
- Xiao, Y.; Jiao, S.; Tong, Y; Song, J. and Wang, G. (1994):* Seasonal changes in testicular size and serum testosterone levels and the relationship between serum testosterone concentrations and reproductive performance in male raccoon dogs. *Scienti Fur* 18: 39-43.
- Yen, S.C. and Jaffe, R.B. (1978):* Reproductive endocrinology. W.B. Saunders.
- Zia, P. and Coombes, R. (1981):* Simplified radioimmunoassay of FSH and LH in human urine. AACC Abstract.

Table (1): Effect of Melatonin / GnRH on Some Reproductive Performance (Mean  $\pm$  S.E)

Treatments	Volume		Intr. Milt %		Live Sperm %		Concentration (x10/ml)		Abnormality %		Sv Dr (mm)		Reaction time (sec)		Sv Dr (mm)		Sv Dr (mm)				
	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2			
Cow	174.0 $\pm$ 12	187.0 $\pm$ 11	85.00 $\pm$ 2.80	80.00 $\pm$ 1.60	77.00 $\pm$ 4.00	81.0 $\pm$ 2.2	23000 $\pm$ 1522	27433 $\pm$ 752	4.08 $\pm$ 1.53	4.05 $\pm$ 1.00	30.33 $\pm$ 0.32	30.33 $\pm$ 0.32	151.87 $\pm$ 10.42	151.87 $\pm$ 10.42	61.03 $\pm$ 13.40	48.96 $\pm$ 11.00	41.61 $\pm$ 8.32	38.50 $\pm$ 5.33	43.48 $\pm$ 10.17	27.3 $\pm$ 0.73	33.8 $\pm$ 0.74
Sw	150 $\pm$ 21	193 $\pm$ 20	58.33 $\pm$ 1.60	90 $\pm$ 0	72.67 $\pm$ 2.83	85.6 $\pm$ 5.89	2997 $\pm$ 756.6	34533 $\pm$ 6337	3.67 $\pm$ 2.88	3.33 $\pm$ 0.33	70.83 $\pm$ 0.53	70.83 $\pm$ 0.53	84.33 $\pm$ 3.33	30.0 $\pm$ 1.50	48.41 $\pm$ 16.22	34.48 $\pm$ 3.40	26.97 $\pm$ 6.23	18.9 $\pm$ 2.50	7.79 $\pm$ 0.64	3.88 $\pm$ 0.85	
W	133 $\pm$ 0.34	113 $\pm$ 0.12	80.67 $\pm$ 3.33	90 $\pm$ 0.00	80.33 $\pm$ 1.45	80.67 $\pm$ 1.2	45967 $\pm$ 4.1187	74533 $\pm$ 3087	3.33 $\pm$ 2.16	5.00 $\pm$ 2.08	30.00 $\pm$ 2.08	29.5 $\pm$ 1.44	48.33 $\pm$ 4.40	98.7 $\pm$ 3.77	83.19 $\pm$ 7.00	32.51 $\pm$ 8.08	80.85 $\pm$ 19.3	81.12 $\pm$ 1.95	5.80 $\pm$ 1.09	2.77 $\pm$ 0.71	
W	167 $\pm$ 0.66	140 $\pm$ 0.30	81.67 $\pm$ 3.33	88.33 $\pm$ 1.66	87.33 $\pm$ 2.49	91.07 $\pm$ 2.07	61687 $\pm$ 8.816	88133 $\pm$ 3078	4.33 $\pm$ 0.33	2.6 $\pm$ 0.33	32.27 $\pm$ 0.14	32.27 $\pm$ 0.14	116.67 $\pm$ 8.00	26.00 $\pm$ 0.63	39.97 $\pm$ 24.29	72.45 $\pm$ 15.51	40.08 $\pm$ 15.09	50.25 $\pm$ 17.53	3.44 $\pm$ 0.81	8.42 $\pm$ 1.22	
W	181 $\pm$ 0.32	173 $\pm$ 0.41	90.00 $\pm$ 2.88	80 $\pm$ 0.00	89.67 $\pm$ 2.84	80.33 $\pm$ 1.18	24633 $\pm$ 2051	24633 $\pm$ 2051	2.87 $\pm$ 0.88	4.00 $\pm$ 1.00	31.83 $\pm$ 0.92	31.83 $\pm$ 0.92	83.33 $\pm$ 4.40	64.70 $\pm$ 16.09	47.55 $\pm$ 3.86	54.49 $\pm$ 12.80	44.81 $\pm$ 24.25	71.03 $\pm$ 8.90	5.24 $\pm$ 1.12	1.10 $\pm$ 0.20	
Female	149 $\pm$ 0.14	146 $\pm$ 0.12	86.67 $\pm$ 1.59	82.67 $\pm$ 2.09	85.95 $\pm$ 1.33	85.19 $\pm$ 1.64	29667 $\pm$ 2015	19409 $\pm$ 11756	8.95 $\pm$ 3.38	7.57 $\pm$ 1.04	31.14 $\pm$ 0.40	30.52 $\pm$ 0.53	83.71 $\pm$ 0.89	37.46 $\pm$ 4.07	54.19 $\pm$ 5.40	99.74 $\pm$ 6.98	61.93 $\pm$ 5.27	44.16 $\pm$ 6.97	3.49 $\pm$ 0.59	3.31 $\pm$ 0.50	
Overall	148 $\pm$ 0.35	143 $\pm$ 0.30	80.28 $\pm$ 0.16	83.82 $\pm$ 0.27	85.03 $\pm$ 0.28	85.28 $\pm$ 0.35	32597 $\pm$ 3179	23078 $\pm$ 4608	6.89 $\pm$ 0.17	8.00 $\pm$ 0.72	31.15 $\pm$ 1.00	30.80 $\pm$ 1.38	83.33 $\pm$ 15.15	46.00 $\pm$ 16.16	52.16 $\pm$ 14.23	51.13 $\pm$ 13.80	44.11 $\pm$ 18.98	50.75 $\pm$ 15.87	4.45 $\pm$ 1.25	3.13 $\pm$ 1.38	

G1 Control  
G2 Melatonin  
G3 GnRH  
G4 GnRH+Melatonin  
P < 0.05  
.. P < 0.01  
Compared with control sample

Table (2): Effect of (Melatonin + GnRH) and (GnRH + Melatonin) on Some Reproductive Performance (Mean  $\pm$  S.E)

Treatments	Volume		Intr. Milt %		Live Sperm %		Concentration (x10/ml)		Abnormality %		Sv Dr (mm)		Reaction time (sec)		Sv Dr (mm)		Sv Dr (mm)		Sv Dr (mm)	
	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2	G1	G2
Cow	150 $\pm$ 10	150 $\pm$ 10	80.00 $\pm$ 1.00	80.00 $\pm$ 1.00	85.67 $\pm$ 0.68	80.66 $\pm$ 0.32	20333 $\pm$ 14042	17350 $\pm$ 2106	3.33 $\pm$ 1.78	6.33 $\pm$ 1.33	30.0 $\pm$ 0.57	30.0 $\pm$ 0.57	146.67 $\pm$ 15.36	160.0 $\pm$ 9.53	39.14 $\pm$ 5.43	16.03 $\pm$ 1.22	37.17 $\pm$ 19.38	79.36 $\pm$ 6.12	6.57 $\pm$ 1.73	1.31 $\pm$ 0.38
Sw	143 $\pm$ 0.48	109 $\pm$ 0.32	90.5 $\pm$ 0.6	88.33 $\pm$ 1.66	87.0 $\pm$ 2.08	83.33 $\pm$ 2.02	26460 $\pm$ 2088	21460 $\pm$ 4841	6.00 $\pm$ 1.53	8.67 $\pm$ 0.67	30.33 $\pm$ 1.81	33.10 $\pm$ 0.96	100.0 $\pm$ 3.05	34.98 $\pm$ 22.87	48.73 $\pm$ 24.89	59.17 $\pm$ 19.79	64.78 $\pm$ 9.49	26.15 $\pm$ 12.73	3.84 $\pm$ 2.07	0.19 $\pm$ 0.01
W	93 $\pm$ 0.21	104 $\pm$ 0.26	85.0 $\pm$ 0.00	85.0 $\pm$ 0.00	96.09 $\pm$ 0.31	90 $\pm$ 0.65	11133 $\pm$ 1847	23333 $\pm$ 5282	7.33 $\pm$ 1.96	7.33 $\pm$ 0.33	30.33 $\pm$ 1.32	31.0 $\pm$ 1.73	24.33 $\pm$ 11.91	83.0 $\pm$ 30.04	39.28 $\pm$ 11.87	57.86 $\pm$ 15.75	61.34 $\pm$ 19.39	43.16 $\pm$ 18.82	8.71 $\pm$ 1.37	3.44 $\pm$ 0.17
W	203 $\pm$ 0.6	183 $\pm$ 0.53	83.33 $\pm$ 0.67	88.33 $\pm$ 0.68	85.67 $\pm$ 2.26	84.66 $\pm$ 1.14	10800 $\pm$ 2003	22460 $\pm$ 4294	3.33 $\pm$ 0.33	16.00 $\pm$ 4.20	30.33 $\pm$ 1.67	31.70 $\pm$ 0.36	10.67 $\pm$ 22.25	85.33 $\pm$ 14.52	80.87 $\pm$ 9.65	15.38 $\pm$ 2.25	42.54 $\pm$ 18.51	61.38 $\pm$ 10.55	3.60 $\pm$ 2.08	3.41 $\pm$ 0.3
W	183 $\pm$ 0.49	164 $\pm$ 0.55	86.67 $\pm$ 4.44	86.66 $\pm$ 3.33	83.33 $\pm$ 3.33	83.33 $\pm$ 2.56	29550 $\pm$ 2348	26057 $\pm$ 4215	10.67 $\pm$ 5.46	16.33 $\pm$ 6.03	31.33 $\pm$ 1.26	32.0 $\pm$ 0.57	57.33 $\pm$ 1.33	88.0 $\pm$ 16.28	80.36 $\pm$ 13.83	88.20 $\pm$ 0.52	31.80 $\pm$ 8.64	36.38 $\pm$ 10.74	2.41 $\pm$ 0.36	8.50 $\pm$ 0.85
W	131 $\pm$ 0.38	131 $\pm$ 0.54	83.33 $\pm$ 0.66	90 $\pm$ 0.00	83.33 $\pm$ 0.69	83.0 $\pm$ 1.52	26950 $\pm$ 1547	35967 $\pm$ 1211	4.00 $\pm$ 1.73	7.0 $\pm$ 1.0	30.05 $\pm$ 0.50	31.0 $\pm$ 0.55	35.00 $\pm$ 12.49	130.33 $\pm$ 8.57	81.24 $\pm$ 6.68	66.21 $\pm$ 6.08	10.96 $\pm$ 3.47	17.78 $\pm$ 0.31	3.27 $\pm$ 0.50	4.29 $\pm$ 1.78
Female	145 $\pm$ 0.14	144 $\pm$ 0.15	86.67 $\pm$ 2.00	87.38 $\pm$ 0.74	85.05 $\pm$ 0.74	84.55 $\pm$ 0.74	30688 $\pm$ 1109	37294 $\pm$ 5784	8.60 $\pm$ 1.50	11.00 $\pm$ 1.19	30.08 $\pm$ 0.64	31.00 $\pm$ 0.26	64.68 $\pm$ 7.24	85.19 $\pm$ 10.37	50.89 $\pm$ 7.86	47.25 $\pm$ 10.22	57.69 $\pm$ 11.83	47.08 $\pm$ 6.54	1.99 $\pm$ 0.81	3.10 $\pm$ 0.64
Overall	148 $\pm$ 0.38	148 $\pm$ 0.38	79.11 $\pm$ 0.31	85.31 $\pm$ 0.57	87.24 $\pm$ 1.14	83.84 $\pm$ 2.45	24550 $\pm$ 1318	33525 $\pm$ 4050	9.05 $\pm$ 0.82	10.38 $\pm$ 0.82	30.38 $\pm$ 1.30	31.72 $\pm$ 0.83	84.93 $\pm$ 24.27	84.51 $\pm$ 28.10	60.82 $\pm$ 12.47	47.21 $\pm$ 17.62	47.15 $\pm$ 16.03	47.89 $\pm$ 15.02	5.04 $\pm$ 0.56	3.34 $\pm$ 0.48

G1 Control  
G2 Melatonin+GnRH  
G3 GnRH+Melatonin  
G4 GnRH+Melatonin  
P < 0.05  
.. P < 0.01  
Compared with control sample



