

SOME STUDIES ON THE AFLATOXIN-PRODUCING ASPERGILLII IN MEAT-COLD STORES

(With 3 Tables and 3 Figures)

By

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بعض الدراسات عن عترات الأسبرجلس المفزرة لسموم الأفلاتوكسينات
فى ثلاجات حفظ اللحوم

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يعتبر تلوث اللحوم ومنتجاتها بالفطريات من أهم المشاكل التي تهدد الصحة العامة للمستهلك حيث أن لها القدرة على إفراز السموم الفطرية والتي تعد الأفلاتوكسينات من أهمها، هذا بالإضافة إلى الدور الرئيسي الذي تلعبه الفطريات في فساد اللحوم ومنتجاتها والذي يؤدي إلى خسائر اقتصادية كبيرة بسبب إعدام تلك اللحوم أو منتجاتها. ولهذا اتجهت هذه الدراسة إلى الفحص الميكولوجى لعدد 100 مسحة من أرضية وأسطح وجدران ثلاجات حفظ اللحوم وكذلك الهواء واللحوم بداخلها بمدينة الإسماعيلية. وقد أمكن عزل 12 نوع من الفطريات معظمها ينتمي إلى الأسبرجلس، الكلاوسبوريوم، البنيسليوم، بنسبة (34%)، (18.9%)، (16.2%) على التوالي. وقد تم تصنيف عدد 9 أنواع من جنس الأسبرجلس وكان معظمها ينتمي إلى أنواع الأسبرجلس فلافس نك (28%)، فلافس فاركولمنارس (25%)، النيجر (17.1%)، النيدبولنز (11%)، الباراستيكس (9.2%) . لقد وجدنا 52 (47.7%) عترة سامة من بين 109 عترة من مجموعة الأسبرجلس فلافس المعزولة من جميع العينات المختبرة وقد تم التعرف على الأفلاتوكسينات المفزرة بواسطة هذه العترات السامة وكانت بالمعدلات الآتية ب (28.8%)، ب (30.8%)، ج (28.8%) ، ج (28.8%)، ج (11.6%) . وقد تمت مناقشة هذه النتائج وتأثير الفطريات المعزولة وسمومها وأهميتها بالنسبة للصحة العامة للمستهلك .

SUMMARY

A total of 100 random swab samples were obtained from different meat- cold stores in Ismailia city, 20 from each of the floor, walls, air, roofs and stored meat. All samples were subjected for mycological examination for detection

of mould contamination and aflatoxin production. A total of 482 mould strains could be isolated and identified. The higher frequency percentages were isolated from floor 128 (26.5%), followed by walls 102 (21.2%), air 89 (18.5%) and roofs 66 (13.7%) as well as, from stored meat were 97 (20.1%). Twelve mould genera could be detected, where the predominant were *Aspergillus* 164 (34.0%) *Cladosporium* 91 (18.9%) and *Penicillium* 78 (16.2%). Furthermore nine *Aspergillus* species were identified, the most of which were *A.flavus* link 46 (28.0%), *A.flavus varcolumnaris* 41 (25.0%), *A.niger* 28 (17.1%), *A.nidulans* 18 (11.0%) and *A.parasiticus* 15 (3.1%). The isolated *Aspergillus flavus* group were screened for toxin production. Out of 109 isolates of *A.flavus* species, 52 (47.7%) strains were found to be aflatoxin producers. Aflatoxins B₁, B₂, G₁ and G₂ were secreted by 15 (28.8%), 16 (30.8%), 15 (28.8%) and 6 (11.6%) strains of *A-flavus* species, respectively. Significance of occurrence and public health importance of the isolated moulds, specially the aflatoxic strains were discussed.

Key words: Meat - Aflatoxins - Cold stores

INTRODUCTION

Because fungi are ubiquitous in nature, contamination of meat and its products may occur during handling, storage and subsequent manufacturing (Robb, 1993). Such moulds are not only implicated in the spoilage of meat leading to great economic losses but also constitute a major public health hazard due to the production of wide varieties of mycotoxins (Abdel-Rahman and El-Khatib, 1989).

Mycotoxins comprise a structurally diverse family of fungal toxins, many of which have been strongly implicated as chemical progenitors of toxicity in man and animal (Ramos *et al.* 1996).

Aflatoxins occupy the most important ingredients among mycotoxins, that is a collective term for a group of structurally similar coumarins, which produced mainly by the common mould *A.flavus* link and *A.parasiticus*. There are four major naturally occurring aflatoxins, the most hepatotoxic being aflatoxin B₁, and three similar compounds B₂, G₁, and G₂. One of the main aflatoxin B₁ biotransformation products is aflatoxin M₁ (Veldman *et al.*, 1992 and Zaky *et al.*, 1995). The principal biological effects are carcinogenicity, immunosuppression, mutagenicity and teratogenicity (Betina, 1989). Also aflatoxins may cause liver damage, altering lipid

metabolism and they can aid in depression of protein synthesis (Leibetseder, 1981 and Ostrowski, 1984).

Different mould genera were isolated and identified from swab samples obtained from frozen meat and meat-cold stores as water, air, floor and walls. The main isolated mould genera were *Aspergillus*, *Caldosporium*, *Penicillium*, *Mucor*, *Alternaria* and *Fusarium* (Abdel-Rahman *et al.*, 1985, EL-Daly *et al.*, 1988; Mousa *et al.*, 1988; Mansour *et al.*, 1990 and Refai *et al.*, 1993). Among these moulds, *Aspergillus* species had received a great attention as it predominated in producing aflatoxins which have great public health hazards (Lie and Marth, 1968 and Hamdy *et al.*, 1993).

Therefore, the present study is carried out to isolate and identify such mould growth, with a great attention to the genus *Aspergillus*, including screening test for determining the aflatoxin-producing strains.

MATERIAL and METHODS

A total of 100 swab specimens were collected from meat cold stores of Ismailia city, 20 from each of the floor, walls, air, roofs and stored meat. All samples were mycologically examined as follows:

1- Isolation of mould genera:

The samples were cultured by using malt extract agar and Czapeck Dox-agar media. The isolates were identified according to Raper and Fennel (1965), Zycha *et al.* (1969), Barnett and Hunter (1972) and Samson (1979).

2- Screening of the aflatoxin-producing aspergilli:

A total of 109 isolates of *A. flavus* species were inoculated at the centre of solidified fluorescent agar media in glass Petri dishes, then the plates were incubated at 25°C for 10 days (according to Hara *et al.*, 1974). The plates were examined under UV lamp for the detection of the fluorescent colour in the agar surrounding colonies.

3- Cultivation and extraction of aflatoxins:

The toxic strains of *A. flavus* group that were illuminated in the fluorescent agar, were inoculated in a rice liquid medium for 15 days at 25°C (Shotwell *et al.*, 1966).

At the end of incubation time extraction of aflatoxins is carried out using chloroform and then concentrated in a rotatory vacuum evaporator.

4- Identification and confirmation of aflatoxins:

The concentrated extracts of *A. flavus* cultures in rice liquid medium were analysed for identification and confirmation of aflatoxins by application

of thin -layer chromatography (TLC) according to Schuller and Egmond (1981).

RESULTS

The results are recorded in Tables (1,2 and 3) and Figures (1, 2 and 3).

DISCUSSION

Results given in Table (1) reveal that 482 mould strains were isolated from swab samples obtained from meat-cold stores and stored meat in Ismailia city. Higher frequency percentages could be isolated from floors 128(26%) followed by walls 102 (21.2%), air 89(19.5%) and roofs 66(13.6%), as well as from stored meat were 97(20.1%).

The frequency percentages of the isolated mould genera as recorded in Table (1) and Fig(1) show that *Aspergillus*, *Cladosporium* and *Penicillium* were the most predominant 164 (34.0%), 91(18.9%) and 18 (16.2%) respectively. While *Rhizopus* was 32 (6.6%), *Alternaria* 22 (4.5%), *Mucor* 22 (4.5%), *Paecilomyces* 21(4.4%), *Absidia* 15(3.1%), *Fusarium* 14 (2.9%), *Bortyis* 9(1.5%), *Geotrichum* 7(1.5%) and *Trichoderma* 7(1.5%). These findings are in agreement with that of Mousa *et al.*(1988) and Hamdy *et al.*(1993), while are lower than those reported by Hadlok (1970) and Abdel-Rahman *et al.* (1985).

The obtained results presented in Table(2) reveal that 164 *Aspergillus* species could be identified as *A.flavus* link 46(28.0%), *A.flavus varcolumnaris* 41(25.0%), *A.niger* 28 (17.1%), *A.nidulans* 18 (11.0%) and *A.parasiticus* 15(9.2%). Furthermore *A.oryzae*, *A.sydowii* and *A.repens* were isolated but in lower percentages; 7(4.3%), 5(3.0%) and 4(2.4%), respectively. These values are nearly similar to those recorded by Refai and Loot (1969), Hadlock (1970) and Abdel-Rahman *et al.*(1985).

Table (3) and Fig.(3) showed the aflatoxigenic *A.flavus* species which were isolated from meat-cold stores and stored meat. Out of 109 *Aspergillus* species, 52(47.7%) were aflatoxin-producing strains which could be identified as *A.flavus* link 24(52.2%), *Aflavus varcolumnaris* 17(41.5%), *A.parasiticus* 7(46.6%) and *A.oryzae* 4(57.1%). These toxic isolates provided aflatoxins B1, B2, G1 and G2 with values of 16(28.8%), 15(18.8%) and 6(11.6%), respectively. These findings are more or less in agreement with that of Taber and Schroeder (1969) and Hamdy *et al.*, (1993).

From the results obtained in the present study, it is achieved that frozen meat and cold-stored chambers are heavily contaminated with mould species (specially toxigenic *Aspergilli*) which are the main source of contamination either before or during transportation. Such contamination may be attributed to unsatisfactory hygienic measures adopted in cleaning and disinfecting of meat-cold stores, before and after storage of meat or its products (Abdel-Rahman *et al.*, 1985 and Mousa *et al.*, 1988).

Presence of the aflatoxin-producing fungi in meat-cold stores increases the probability of hazards arising from mould growth on meat. The direct hazard to human health from aflatoxins is achieved due to mycotoxigenic strains are still able to secrete mycotoxins in stored meat at suitable circumstances (Scott, 1973). Therefore, fungal contamination of meat should be controlled by strict hygienic precautions during production, transportation, processing and storage.

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Table (1): Isolated mould genera from meat cold – stores

Mould genera	Floor		Walls		Air		Roof		Meat		Total	
	No	%	No	%	No	%	No	%	No	%	No	%
Aspergillus	48	37.5	37	36.3	28	31.4	19	28.8	32	32.9	164	34.0
Cladosporium	26	20.3	19	18.6	10	11.2	21	31.8	15	15.5	091	18.9
Penicillium	13	10.2	15	14.7	30	33.7	00	00.0	20	20.6	078	16.2
Rhizopus	14	10.9	08	07.9	03	03.4	00	00.0	07	07.2	032	06.6
Alternaria	03	02.3	04	03.9	07	07.9	00	00.0	08	08.3	022	04.5
Mucor	09	07.0	00	00.0	05	05.6	08	12.1	00	00.0	022	04.5
Paecilomyces	06	04.7	05	04.9	00	00.0	04	06.1	06	06.2	021	04.4
Absidia	00	00.0	09	08.8	00	00.0	06	09.1	00	00.0	015	03.1
Fusarium	07	05.0	00	00.0	02	02.3	00	00.0	05	05.2	014	02.9
Botrytis	00	00.0	01	01.0	03	03.4	05	07.6	00	00.0	009	01.9
Geotrichum	00	00.0	04	03.9	00	00.0	03	04.5	00	00.0	007	01.5
Trichoderma	02	01.6	00	00.0	01	01.1	00	00.0	04	04.1	007	01.5
Total & percentage	128	26.5	102	21.2	89	18.5	66	13.7	97	20.1	482	100

Table (2) : Isolated Aspergillus species from meat – cold stores

Aspergillus spp.	Floor		Wall		Air		Roof		Meat		Total	
	No	%	No	%	No	%	No	%	No	%	No	%
A.flavus – link	13	27.1	12	32.4	07	25.0	04	21.0	10	31.3	46	28.0
A.flavus varcolumnaris	11	22.9	08	21.6	09	32.1	06	31.6	07	21.9	41	25.0
A.niger	09	18.7	07	18.9	05	17.9	03	15.8	04	12.5	28	17.1
A.nidulans	04	08.3	03	08.1	02	07.1	05	26.3	04	12.5	18	11.0
A.parasiticus	06	12.5	04	10.9	03	10.7	00	00.0	02	06.3	15	09.2
A.oryzae	03	06.3	00	00.0	01	03.6	00	00.0	03	09.2	07	04.3
A.sydowii	00	00.0	02	05.4	01	03.6	00	00.0	02	06.2	05	03.0
A.repens	02	04.2	01	02.7	00	00.0	01	05.3	00	00.0	04	02.4
Total & percentage	48	29.2	37	22.6	28	17.1	19	11.6	32	19.5	164	100

Table (3) : Isolated Aflatoxin – producing strains of A.flavus group.

A.flavus group	Total Toxic strains		AFB ₁		AFB ₂		AFG ₁		AFG ₂		
	No.	+ ve %	No.	%	No.	%	No.	%	No.	%	
A.flavus link	46	24	52.2	09	37.5	07	29.2	06	25.0	02	08.3
A.flavus varcolumnaris	41	17	41.5	04	23.5	06	35.3	04	32.5	03	17.7
A.parasiticus	15	07	46.6	02	28.6	02	28.6	03	42.8	00	00.0
A.oryzae	07	04	57.1	00	00.0	01	25.0	02	50.0	01	25.0
TOTAL	109	52	47.7	15	28.8	16	30.8	15	28.8	06	11.6

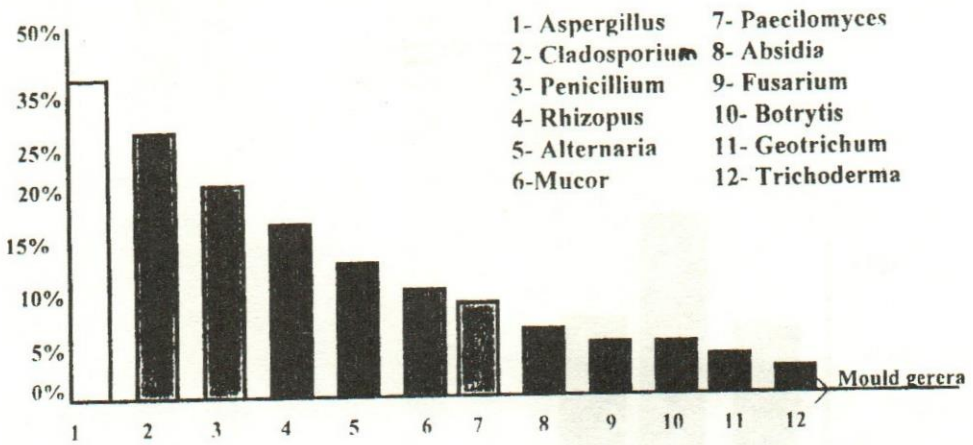


Fig.(1) Frequency of mould genera isolated from meat cold stores and stored meat

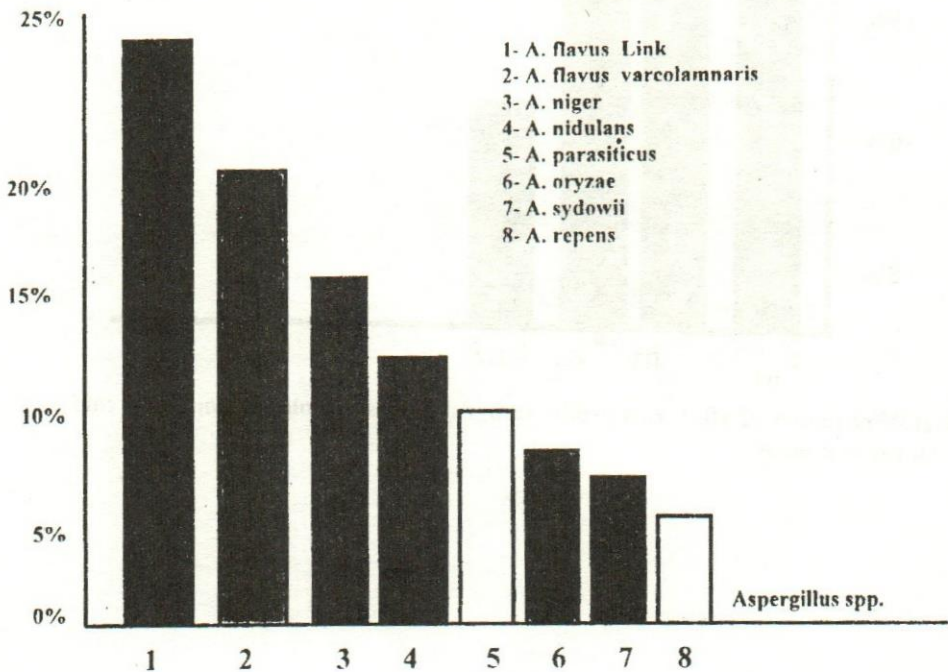
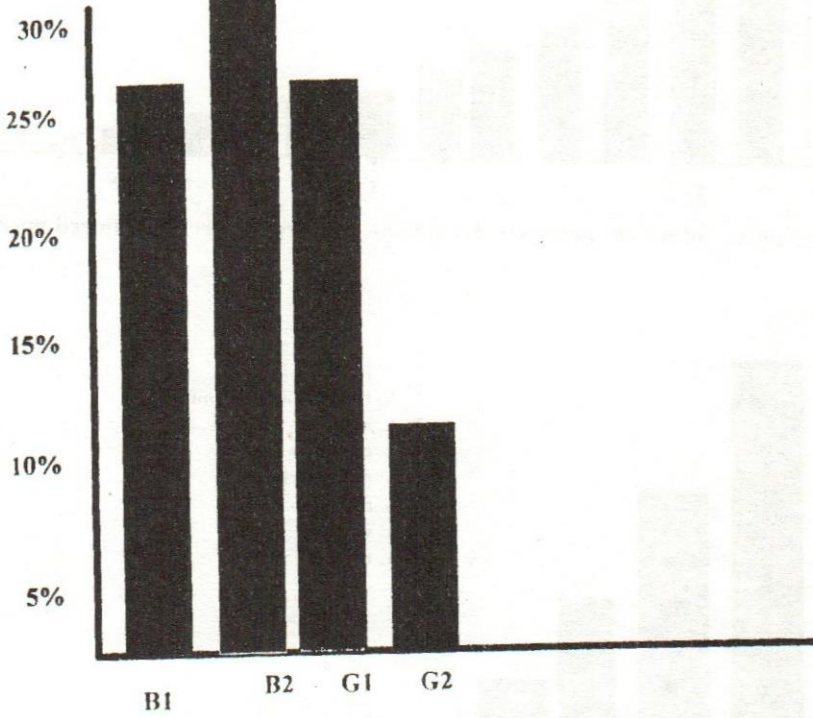


Fig.(2) Frequency of Aspergillus species isolated from meat cold stores and stored meat



Fig(3) Frequency of aflatoxins produced by *A. flavus* spp. isolated from meat cold stores and meat