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RESIDUES OF QUINOLONE GROUP OF ANTIBIOTICS IN TABLE EGGS (*)

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(With 4 Tables)

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قياس المضادات الحيوية (مجموعه الكينولون) في بعض بيض المائدة
بالاسواق

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كثير في الاونه الاخيرة استعمال المضادات الحيوية بطريقه غير سليمه قد تؤدي الى ظهور بقاياها في بيض الدواجن حيث تكمن خطورة ذلك ليس فقط في اكتساب الميكروبات الضارة مناعة ضد هذه المضادات الحيوية ولكن ايضا في ظهور بعض امراض الحساسيه بين المستهلكين لبيض هذه الدواجن خصوصا بين الاطفال . وقد تم اجراء هذا البحث لتقدير اقل تركيز يمكن تعيينه في البيض لبعض هذه المضادات الحيوية (الكينولون) والكشف عن تواجد بقايا المضادات الحيوية وهي الانروفلوكساسين- الفلبيموكوين. كما تم دراسته مدى تواجد بقايا المواد المثبطه في البيض المتداول في اسواق محافظات الاسكندريه والبحيره وكفر الشيخ وقد تبين ان اقل تركيز يمكن تقديره في البيض لكل من المستحضرين المستخدمين هو ٠,٠٢٢ ، ٠,٠٩٦ ميكروجرام لكل مللى على التوالي وقد وجد ان ميكروب الاشريشيا القولوني هو اكثر حساسيه لتحديد اقل تركيز من هذين المستحضرين من الميكروبات الاخرى وهي الزائفة المتلقه والعصويه الرقيقه والكليسيلا الصدرية . وقد تم تقدير كميته بقايا المضادات الحيوية باستخدام ٧٥ دجاجه بياضه ومقسمه الى ثلاثه مجموعات منها مجموعه تضم ٢٥ دجاجه كمجموعه ضابطه وتم اضافته كل من الانروفلوكساسين والفلبيموكوين بجرعات علاجيه الى ماء الشرب للمجموعه الاولى والثانيه على التوالي بنسبه ١٠ مللجرام لكل مللى جرام لكل كيلو جرام من وزن الطائر على التوالي لمدة ثلاثة ايام متتاليه ووجد ان الانروفلوكساسين يستمر وجوده في البيض اربعة ايام في البياض حتى اليوم الثاني من اخر يوم للعلاج اما في حالة الفلبيموكوين لوحظ ان بقايا المستحضر تستمر خمس ايام ووجد ان المواد المتبقية في البيض لمدة المتداول في الاسواق في هذه المحافظات توجد بنسب متفاوتة وان ارتفعت في البحيره وكفر الشيخ عنها في محافظه الاسكندريه وهذا وينصح بضرورة

إقرار اشتراطات خاصه عند استعمال هذه المضادات الحيويه وضرورة الكشف الدورى على البيض المتداول فى الاسواق للتأكد من خلوة من بقايا المواد الضارة لما لها من خطورة لا يستهان بها على صحة المستهلكين.

SUMMARY

This study was carried out for determination of the minimal inhibitory concentration of enrofloxacin and flumequine giving inhibition zone by using microbiological assay technique and determination of the withdrawal time of enrofloxacin and flumequine after oral application as well as the incidence of inhibitory substance in table eggs marketed in Alexandria, Behera and Kafr El-Aheikh governorates. The MICS of enrofloxacin using E.Coli, P.fluorecens ,K pneumoniae and B. subtilus as test organisms were 0.022, 0.12, 0.054 and 0.096 $\mu\text{g} / \text{ml}$ while MICS OF flumequine using the same test organismes were 0.04, 0.17, 0.096 and 0.22 $\mu\text{g} / \text{ml}$ respectively. Enrofloxacin residues could be detected at the 2nd day of oral administration and last for 2 days in albumen, and 4 days after last dose of application, while flumequine last for 3 days in albumen and 2 days in yolk after the last dose of application. The inhibitory substances in the examined egg samples purchased from Alexandria, Behera and Kafr El-Sheikh governorates were, 70, 62 and 24%; 78, 82 and 16% and 84, 64 and 48% of brown, white and Balady table eggs respectively.

Key Words: Quinolone, antibiotics table eggs.

INTRODUCTION

Antibiotics are widely used in poultry farms not only for treatment and control of infectious diseases but also as feed additives for growth promotion. The extensive use of these drugs in laying hens leads to appearance of the problem of drug residues as a result of excretion of such drugs in the eggs during and after treatment which have harmful effect on the consumers. So it is necessary to know how much and for how long a drug will continue to be secreted in eggs after hens are treated. Nowadays, flouroquinolone group (e.g. flumequine, enrofloxacin,...) is one of the most prevalent antibiotics used in poultry farms. Enrofloxacin is a new flouroquinolone chemotherapeutic agent (Brown, 1996). It has a broad spectrum activity against Gram positive and Gram negative bacteria as well as Mycoplasma and some anaerobic pathogens (Watts et al., 1993 and

Nakamura, 1995). Flumequine is a relatively new synthetic antibacterial drug especially active against Enterobacteriaceae and in veterinary field it is recommended for treatment of *E. coli*, *Salmonellae* and *Pasteurellae* infections of poultry (Giebel et al., 1982 and Duplay and Chomel, 1983).

MATERIAL and METHODS

I. Detection of the minimal inhibitory concentration:

- The reference substances: were enrofloxacin (Pt. Nr. R 177-1, Bayer and flumequine (Bremer pharma GmbH, Germany).
- The test organisms: were *E. Coli* strain, *Pseudomonas fluorescens* and *Klebsiella pneumoniae* which were obtained from Medical Research Institute, while *Bacillus subtilis* (BGA) was obtained from Animal Health Research institute.
- The Spore suspension: of the *Bacillus subtilis* (BGA) was prepared according to Pastors, (1988), then the non-spore forming organism suspension for *E. Coli* *Pseudomonas fluorescens* and *Klebsiella pneumoniae* were also prepared
- Agar diffusion test: (Bogaerts and Walf, 1980), *E. Coli* strain and standard agar II were used. Samples of egg white and egg yolk were applied by using a well technique (Coretti, 1961).
- The standard curve: of flumequine and enrofloxacin was made .
- Double fold serial dilution of the standard enrofloxacin and flumequine solutions were prepared in sterile bidistilled water, 100 μ L from each dilution were applied in a well. made in previously prepared agar plate. The plates were left for 1 hour at room temperature then incubated at 30°C for 18 hours. The inhibition zone was measured from the edge of the well to the beginning of bacterial growth. 1-2mm inhibition zone was considered negative. The test repeated at different pH 6.0, 7.2 and 8.0 for detection of the effect of pH on the efficacy of enrofloxacin and flumequine.

II. Experimental

Seventy five clinically healthy laying hens of Lohman Selected leghorn of 30 weeks age were used in this study and classified into three groups as follows:

- a) The first group: A total of 25 laying hens were supplied with 3 successive oral doses enrofloxacin (pt.Nr.R177-1, Bayer) at a

- concentration of 10 mg/kg for determination of enrofloxacin residues (withdrawal time) in eggs.
- b) The second group: A total of 25 laying hens received 3 successive oral doses of flumequine (Bremen pharma, Germany) at a concentration of 12 mg/kg for determination of flumequine residues.
- c) A total of 25 laying hens were used as a group.

III. Screening:

A survey of 450 eggs samples purchased from different localities in Alexandria, Behera and Kafr El-Sheikh markets (150 each of native and 300 of foreign breeds) and examined for possible occurrence of inhibitory substances using agar diffusion technique. *Bacillus subtilis* (BGA) was used as a test organism and standard II agar as a test medium (Pastoors, 1988).

RESULTS

Are presented in tables 1-4.

DISCUSSION

I. Minimal inhibitory concentration (MICS):

The MICs of enrofloxacin using *E. coli*, *P. fluorescens*, *K. pneumoniae* and *B. subtilis* as test organisms were 0.022, 0.12, 0.054 and 0.096 $\mu\text{g} / \text{ml}$ at pH 7.2, respectively. While the MICs of flumequine using the same test organisms were 0.04, 0.17, 0.096 and 0.22 $\mu\text{g} / \text{mL}$ at pH 6, respectively (Table 1). *E. coli* as a test organism is more sensitive for determination of enrofloxacin and flumequine residues in egg contents than the other abovementioned organisms and this selection of *E. coli* as a test organism agreed with Zehl, (1989) Prescott and Yielding, (1990) Samaha et al., (1991) and Khodary and Abd El-Latif, (1997).

II. Withdrawal time:

Enrofloxacin was firstly detected in egg white at the 2nd day of the oral administration at a level of 0.96 $\mu\text{g}/\text{ml}$ and also be detected during the period of application (3 successive oral doses of 10 mg/kg body weight) as well as 2 days after the last dose of application at a level of 0.3 $\mu\text{g}/\text{ml}$. Enrofloxacin could be detected in egg yolk at the

2nd day after the last dose of application at a level of 0.07 µg/ml and continued for 3 days (Table 2).

Flumequine residues were firstly detected in egg white at a level of 1.59 µg/ml at the 2nd-day of application and could be detected for 3 days after the last dose. While it was detected in egg yolk at the 4th day after the last dose of application and continued for 2 days (Table 3). These observations are supported by the hypothesis of the development of egg in poultry (Strukie, 1965) who stated that during the development of the egg, the period of deposition of albumin and yolk was between 12-24 hours 1-10 days, respectively before laying.

III) Incidence of inhibitory substances in table eggs:

The inhibitory substances in the examined egg samples purchased from Alexandria markets were detected in 70, 62 and 24% of brown, white and balady table eggs, respectively. The incidence of inhibitory substances in table eggs of foreign breeds (brown and white - shelled eggs) and balady breeds collected from different localities at EL-Behera and Kafr- El-Sheikh Governorates were 78,82 and 16%, respectively in El-Behera while in Kafr El-Sheikh were 84, 64, and 48% respectively (Table 4). Lower incidence of inhibitory substances in egg contents were obtained by El-Rashedy (1978), El- Bassiony (1985), Ahmed et al (1987) and Gad El Rab (1989).

The incidence of residues in the egg albumin was higher than in the yolk. These results are in accordance with those reported by Roudaut (1993). The low incidence of antibacterial agents in the examined balady eggs may due to the high resistance of these native breeds of chickens to diseases and consequently the little use of antibacterial agents and also may be due to feeding on rations free from growth promoters.

Suggestions to make assurance that eggs will be free from any antibiotic residues were discussed.

REFERENCES

- Ahmed, A. A. H.; M. S. Nagah and M. K. Moustafa (1987): Microbial Contamination of market hen eggs. *Assiut Vet Med. J.*, 18(36): 125-131.

- Bogaerts, R. and F. Wolf (1980):* A standardized method for the detection of residues of antibacterial substances in fresh meat. *Fleisch Wirtsch*, 60 (4): 672. Cited in *Waffia H. Abd Allah; A.A. Bahout and A.F. Abd-Alim (1994):* Incidence of antibacterial agents in table eggs.
- Brown, S.A. (1996):* Flouroquinolones in animals health. *J. Vet. Pharamcol. Therap.*, 19: 1-14.
- Coretti, C. (1961):* Nachweis von Antibiotika in Fleischwaren. *Fleisch Wirtsch.*, 2:119-122. Cited in *Samaha et al. (1991)*.
- Duplay, J. M. and R. Chomel (1983):* Use of flumequine as antibacterial agent in poultry production. Report from Rhone-Merienx, 17 rue Bourgelat Lyon, France.
- El-Bassiony, T.; M. K. Moustafa; A.A.H. Ahmed and S Mousa (1985):* Studies on the drugs rsidues in eggs. *Assiut Med. J.*, 9 (2): 18-29.
- El-Rashedy, A. (1978):* Studies on the identification of antibiotic products in food and food stuffs, M. V. Sci., Thesis Dept. Nutrition, High Inst. Public Health, University of Alexandria.
- Gad El Rab, H.M. (1989):* Studies on the rsidues of some antibiotics and sulphha drugs in eggs, Ph. D.Thesis, Fac. Vet. Med., Assiut University.
- Giebel, O.; M. Mazurkiewicz; A. Moroz; T. Pietrzkiez and A. Wuliezka (1982):* Evaluation of the efficacy of flumequine in the control of bacterial disease of poultry.
- Khodary, R.M. and A. E. Abd El-Latif (1997):* In vitro susceptibility of some avian pathogens to fluorquinolone compound compared with the commonly used antimicrobial agents. *Wydzial Wet, AR, Pl. Grundwaldzki, 45,350-366, Waroclaw, Poland.*
- Nakamura, S.(1995):* Venterinary use of new quinolones in Japan. *Drugs*, 49: Suppl., 2:152-158.
- Pastors, K.O. (1988):* Ein Multibakerilles screening zur qualitativen Eingenzung Antimikrobiell Wirksamer Ruckstands in Harn. Hannover, Tieraztl. Hochschs., Diss.
- Prescott, J. F.; and K.M. Yielding (1990):* In vitro susceptiblity of selected veterinary bacterial pathogens to ciprofloxacin, enrofloxacin and norfloxacin. *Can. J. Vet. Res.*, 54: 195-197.

- Roudaut, B. (1993):* Residues of sulphonamides in eggs following oral medication of laying hens. Proceedings of Euro residues II Conference, Veldhoven. The Netherlands 3-5 May.
- Samaha, I.; A.Ebrecht; L.Ellerbroek; S. Mattes and S.Wenzel (1991):* flumequine. residues in egg. Associates, A Division of Cornell University Press.
- Strukie, P.D. (1965):* Avian physiology Ed. II. Chapter 15, Ithaca, New York. Comstock Publishing Associates, A Division of Cornell University Press.
- Watts, J.L.; S. A. Salmon; J.R. Yancey; B.Nersessian and Z.V. Koumnev(1993):* Minimum inhibitory concentration of bacteria isolated from septicaemia and air sacculitis in ducks. J. Vet. Diagnostic Investigation, 5(4): 625-628.
- Zehl, U. (1989):* Beitrag zur Pharmakokinetik des neuen Gyrasehemmers, Enrofloxacin beim Pferd. Vet. Med. Diss., Hannover. Cited in Samaha *et al.* (1991): Alex. J. Vet. Sci., 13 (2): 41-42.

Table (1) : Minimal inhibitory concentrations of enrofloxacin and flumequine ($\mu\text{g} / \text{ml}$)

| Antibacterial | pH | Escherichia coli | Pseudomonas Fluorescens | Klebsiella Pneumoniae | Bacillus subtilis |
|---------------|-----|------------------|-------------------------|-----------------------|-------------------|
| Enrofloxacin | 7.2 | 0.022 | 0.12 | 0.054 | 0.096 |
| | 5.0 | 0.040 | 0.17 | 0.096 | 0.220 |
| Flumequine | | | | | |

Table (2) : Levels of enrofloxacin residues in eggs of laying hens during and after treatment :

| Days | Albumen | | | Yolk | | |
|------|----------------------|--|----------------------|--|--|--|
| | Inhibitory zone (mm) | Enrofloxacin ($\mu\text{g} / \text{ml}$) | Inhibitory zone (mm) | Enrofloxacin ($\mu\text{g} / \text{ml}$) | Enrofloxacin ($\mu\text{g} / \text{ml}$) | Enrofloxacin ($\mu\text{g} / \text{ml}$) |
| | | | | | | |
| 1 | 2 | 0.00 | 0 | 0.00 | 0.00 | 0.00 |
| 2 | 11.5 | 0.96 | 0 | 0.00 | 0.00 | 0.00 |
| 3 | 11 | 0.91 | 0 | 0.00 | 0.00 | 0.00 |
| 4 | 11 | 0.91 | 0 | 0.00 | 0.00 | 0.00 |
| 5 | 5 | 0.31 | 6 | 0.07 | 0.07 | 0.07 |
| 6 | 2 | 0.00 | 4 | 0.03 | 0.03 | 0.03 |
| 7 | 2 | 0.00 | 3 | 0.01 | 0.01 | 0.01 |
| 8 | | | 2 | 0.00 | 0.00 | 0.00 |
| 9 | | | 2 | 0.00 | 0.00 | 0.00 |

Average values

Table (3) : Levels of Flumequine residues in eggs of laying hens during and after treatment :

| Days | Albumen | | Yolk | |
|-------------------------|----------------------|--|----------------------|--|
| | Inhibitory zone (mm) | Enrofloxacin ($\mu\text{g} / \text{ml}$) | Inhibitory zone (mm) | Enrofloxacin ($\mu\text{g} / \text{ml}$) |
| | 1 | 2 | 0.00 | 0 |
| Days during application | 2 | 1.59 | 0 | 0.00 |
| | 3 | 1.43 | 0 | 0.00 |
| | 4 | 1.59 | 0 | 0.00 |
| Days after application | 5 | 1.12 | 0 | 0.00 |
| | 6 | 0.48 | 0 | 0.00 |
| | 7 | 0.00 | 6 | 0.20 |
| | 8 | 0.00 | 4 | 0.07 |
| | 9 | 0.00 | 2 | 0.00 |

* Average values

Table (4) : Incidence of antibacterial agents in table eggs :

| Type of egg | No. of examined samples | Albumin | | | | | | Yolk | | | | | |
|---------------|-------------------------|------------------|----|-----|----|-----|----|------------------|----|-----|----|-----|----|
| | | positive samples | | | | | | positive samples | | | | | |
| | | A | | B | | K | | A | | B | | K | |
| No. | % | No. | % | No. | % | No. | % | No. | % | No. | % | No. | % |
| Brown-shelled | 50 | 35 | 70 | 39 | 78 | 42 | 84 | 11 | 22 | 15 | 30 | 38 | 76 |
| White-shelled | 50 | 31 | 62 | 41 | 82 | 32 | 64 | 11 | 22 | 11 | 22 | 14 | 28 |
| Balady | 50 | 12 | 24 | 8 | 16 | 24 | 48 | 7 | 14 | 4 | 8 | 8 | 16 |

A = Alexandria governorate

B = El-Behera governorate

K = Kafr El-Sheikh governorate