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INCIDENCE OF BACILLUS CEREUS IN SOME FOOD STUFFS WITH THE SPECIAL REFERENCE TO ITS PRODUCTION OF THERMONUCLEASE ENZYME IN ASSIUT CITY (With 3 Tables)

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مدى تواجد ميكروب الباسيللس سيريس فى بعض أنواع الأغذية بمدينة أسيوط مع الإشارة إلى إنتاجه لأنزيم السيرمونيوكليز

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تضمن البحث فحص ٢٠٠ عينة من بعض أنواع الأغذية فى مدينة أسيوط بواقع ٤٠ عينة لكل من اللانشون والسجق والجبن القريش والاييس كريم والزبادى وذلك لمعرفة مدى تواجد ميكروب الباسيللس سيريس بها باستخدام طريقة الفرد السطحى على مستتبت KG. وأثبتت النتائج عن تواجد الميكروب فى عينات اللانشون والسجق والجبن القريش والاييس كريم بنسب ٥٠٪، ٥٧٪، ٣٧,٥٪، ٨٥٪، على التوالى وكان متوسط العدد الكلى للباسيللس سيريس $3,8 \times 10^5$ ، $4,5 \times 10^4$ ، $1,52 \times 10^4$ ، لم تتمكن من عزل الميكروب من أى من عينات الزبادى. وقد شمل الجزء الثانى من البحث تعيين أنزيم السيرمونيوكليز الثابت والمحطم بالحرارة. وشمل الجزء الثالث من البحث تأثير المضادات الحيوية على العترات المعزولة من الميكروب وأوضحت النتائج أن جميع العترات أظهرت حساسية بنسبة ١٠٠٪ لكل من باستراسين والكلور مفينكول والارثرومايسين والجنناميسين والكاناميسين ونيومايسين والاستربتومايسين والنتراسيكلين. وعلى عكس ذلك جميع العترات غير حساسة للكولستين والبنسلين. هذا وقد نوقشت الأهمية الصحية للميكروب والشروط الواجب مراعاتها لإنتاج أغذية سليمة.

SUMMARY

A total of 200 random samples of meat and milk products (40 samples of luncheon, sausage, kareish cheese, ice cream and yoghurt) were collected from Assiut city. The samples were examined

bacteriologically to enumerate and isolate *Bacillus cereus* using surface plating technique on KG agar plates. *Bacillus cereus* was found in 20 (50%); 23(57%); 15(37.5%) and 34(85%) samples of luncheon, sausage, kareish cheese and ice cream with a mean values of 3.8×10^5 , 5×10^4 , 1.52×10^3 and 4.5×10^4 cfu/g or ml, respectively. *Bacillus cereus* could not be isolated from yoghurt. 55.43% of isolated *Bacillus cereus* produce thermolabile Deoxyribonuclease at PH (9) while 14.13% produce it at pH (6.7). On the other hand thermostable Deoxyribonuclease activated only at pH(9) with incidence of 3.26%. All *Bacillus cereus* strains were highly sensitive to bacitracin, chlormephenicol, erythromycin, gentamycin, kanamycin, neomycin, streptomycin and tetracycline, whereas all isolates were resistant to colistin and penicillin. The public health importance and sanitary measures to minimize contamination of food stuffs with *Bacillus cereus* are discussed.

Key words: Food stuffs - Bacillus cereus - Thermomuclease enzyme

INTRODUCTION

Bacillus cereus is a large Gram positive bacillus resembling *Bacillus anthracis*, except that it is motile and lacks the glutamic acid capsule. Also, *B.cereus* is one of the most important heat resistance aerobic sporeforming bacteria, which belong to the genus *Bacillus* (Griffith, 1992).

B.cereus is widely distributed in the environment, the organism can be introduced into food from soil, air, water, equipment and workers.

The unique combination of both thermotolerant and psychrotrophic properties of *B.cereus* represent recurrent problems of meat and milk industries. The organism associated with defects such as off-flavours, sweet curdling and spoilage of both meat and milk products due to protease, lipase and phospholipase produced by *B.cereus* and this led to economic losses (Giffel et al., 1995).

The importance of *B.cereus* which may be present in meat and milk products (especially luncheon, sausage, kareish cheese, ice cream and yoghurt) is its harmful effect on human when consumed, therefore *B.cereus* is recognized as one of the potential organisms for human food poisoning [Christiansson, (1992), Baker and Griffiths (1993) and Granum et al. (1993)].

Two different forms of food poisoning have been attributed to consumption of contaminated food with *B.cereus*, a rapid onset emetic form

which is characterized by acute attack of nausea, vomiting and abdominal cramps, which occurs within 30 mins. to 5 hours, heat-stable enterotoxin is responsible for emetic form while the second diarrheal form occurs within 8 - 16 hours after ingestion of contaminated food, this is due to heat-labile enterotoxin produced during multiplication of *B.cereus* in situ. This form is characterized by diarrhea, nausea and abdominal cramps (Patrick *et al.*, 1994). The number of such organisms required to produce disease in man is generally of order $10^6 - 10^8$, however in compromised consumers a much smaller dose (1.2×10^3) may cause illness (Giannella and Brasile, 1979).

The purpose of this investigation is to study the incidence and enumeration of *B.cereus* in luncheon, sausage, kareish cheese, ice-cream and yoghurt sold in Assiut city.

Testing the isolated strains of *B.cereus* for production of thermolabile and thermostable Deoxyribo nucleas. Study the antibiotic sensitivity of isolated *B.cereus* strains. Discuss the public health importance of *B.cereus*.

MATERIAL and METHODS

Two hundred random samples including Luncheon, sausage, kareish cheese, ice-cream and yoghurt (each 40 samples) were collected from super markets, markets, dairy shops and groceries in Assiut city. The samples were transferred in sample cases and delivered promptly to the laboratory.

Preparation of samples:

In a sterile mortar, to 10 grams of thoroughly mashed sample (luncheon, sausage and kareish cheese) 90 ml of peptone water 1% were added and thoroughly mixed to make a dilution 1:10, on the other hand ice-cream samples were melted in a thermostatically controlled water bath ($40\text{C}^{\circ} \pm 1 \text{C}^{\circ}$) for 15 minutes and well mixed. For yoghurt samples each was thoroughly mixed and 10ml or g were added to 90ml peptone water 1% to make a dilution 1 :10, from the supernatant fluid 1ml was taken by using sterile pipette to make 10-fold serial dilutions technique.

From the prepared samples 10-fold serial dilutions up to 10^{-4} were made, using sterile peptone water.

Enumeration and isolation of *B.cereus*:

One tenth ml amount of each of the prepared dilutions was carefully transferred and evenly spread over a dry surface of KG agar plates as recommended by Kim and Goepfert, 1971. Following

incubation at 37 C° for 24h, KG plates were examined for typical colonies, which were dry flat and surrounded by wide cloudy zones.

Colonies presumed to be *B.cereus* were transferred to nutrient agar slants. Gram's stain and motility test were performed on each isolate. Confirmatory tests were based on carbohydrate utilization, nitrate reduction, and production of acetymethyl- carbinyol according to Cowan and Steel (1974). Numbers of confirmed *B.cereus* / g or ml were counted and recorded.

Surveying the isolated strains of *B.cereus* for production of thermolabile and thermostable Deoxyribonuclease at pH (6.7) and pH (9):

Toluidine blue -O- DNA agar (TDA).was prepared and 15ml. quantities were pipetted into plates. After solidification, wells of 2mm. diamter were cut in the agar, and agar cuts were removed with a metal canula. Over night brain-heart infusion broth cultures (before and after heating, in a boiling waterbath, for 15 minutes) were added to the wells using micropipette and incubated at 37C°. positive results were indicated by development of bright pink zones arround wells after 3 and 24 h. of incubation (Lachia, *et al* 1971).

Antibiotic sensitivity of isolated *B.cereus* strains:

The diffusion method described by Cruickshank *et al* (1975) was performed using different types of sensitivity discs obtained from Bio Merieux France.

RESULTS

The results are presented in Tables (1-3).

DISCUSSION

The results given in table (1) showed that out of 40 samples of Lunchoen, *B.cereus* could be isolated from 20 (50%) with a maximum count of 3×10^6 /g, while the minimum count was 2×10^3 /g with a mean value of 3.8×10^5 . Similar observation were reported by Hefnawy *et al.* (1984) and Lotfi *et al.* (1988), on the other hand Ashmowee (1994) could detecte the organism in Lunchoen samples with high incidence 92%. The mean count /g was nearly similar count.

Out of 40 samples of susage *B.cereus* could be isolated from 23 (57%) with a count of 5×10^5 /g as a maximum, 7×10^2 /g as a minimum

and a mean value of 5×10^4 Table (1). These results were agreed with that reported by El-said (1992) and Lotfi *et al.* (1988). High incidence (92%) was reported by Ashmawee (1994) while low one (28%) was reported by Hefnawy *et al.* (1984).

From this study the incidence of *B.cereus* in meat products pointed that the meat products contained high *B.cereus* count and this may be attributed to contamination of flesh used for manufacture of such products. Mincing machines, grinders, equipments and knives considered a main source of contamination of meat during processing. Moreover addition of spices, which often contain a large number of spore formers, to meat led to marked increase in bacterial population. Unsatisfactory hygienic measures during preparation, handling and distribution, may play a role in the contamination (Hefnawy *et al.* (1984). Lotfi *et al.* (1988); El-said (1992) and Ashmawee (1994).

In case of kareish cheese *B.cereus* was isolated from 15(37.5%) of examined samples, with a maximum count of $3 \times 10^4/g$ and a minimum count of $2 \times 10^2/g$. The mean value was $1.52 \times 10^3/g$. (Table 1). This is somewhat agree with the incidence of (45%) reported by Ell-Boudy (1985), while low incidence (18% and 9.09%) were reported by Saad (1985) and Aman and Ahmed (1997) respectively.

Regarding results of ice-cream samples *B.cereus* could be isolated from 34(85%) of examined samples, in counts ranging from 10 to $9 \times 10^4/ml$ with a mean value of $4.5 \times 10^4/ml$. The results obtained are nearly similar to that reported by Abdel Haleem (1995), while incidence of 38% and 40% were reported by Al-ashmawy *et al.* (1996) and Rangasamy *et al.* (1993) respectively. Low results (11.36% and 16%) were reported by Ahmed (1980) and El-Bagoury (1992).

B.cereus could not be isolated from examined samples of yoghurt. These results are similar to those obtained by Ahmed *et al.* (1983) and Saad (1985). The failure of detecting these organisms in yoghurt may be due heating of milk during manufacturing and, or from the inhibitory effect of lactic acid bacteria on *B.cereus* (Saad, 1985).

Generally, *B.cereus* can gain access to dairy products from different sources during its manufacturing, handling and directly from contaminated ingredients of animal (milk). *B.cereus* gain entry into milk by variety of routes. Contamination via the udder can occur either by infection with *B.cereus* causing acute mastitis (Jones and Turnbull 1981). or by sticking direct from soil, bedding, dung, fodder, pasture and water (Becker and Terplan 1989). Other possible sources may be milk handling equipment,

transport vessels and processing equipment (Heddehem and Vlaemynck 1992).

The incidence of deoxyribonuclease (DNase) producing *B.cereus* recovered from 92 samples of meat and milk products was shown in (Table 2). The incidence of thermolabile DNase positive *B.cereus* at pH (9.0) was greatest in lunchoen 13(65%), followed by ice-cream 20(58.8%), susage 12(52.2%) and kareish cheese 6(40%) while thermolabile DNase positive *B.cereus* at pH(6.7) was 20%, 14.71%, 13.04% and 10% in case of kareish cheese, ice-cream, susage and lunchoen, respectively.

Goepfert et al. (1972) mentioned that the pathogenic character of *B.cereus* is due to a toxic protein like component, secreted by the organism during growth, this exotoxin was described to be thermolabile. Moreover thermostable DNase was recovered only on 2(8.7%) susage and 1(6.7%) kareish cheese at pH (9).

The production of thermolabile and thermostable DNase by *B.cereus* from milk and meat products were reported by Saad (1985) and Lotfi et al. (1988).

Results in this study indicate that some strains of *B.cereus* are capable to produce thermostable DNase. Moreover, *B.cereus* thermostable DNase production was activated at pH 9 than pH 6.7 and this was similar to that of staphylococcal thermostable DNase. So the production of the enzyme by *B.cereus* should be viewed with concern.

It is evident from (Table 3) that all isolates of *B.cereus* were highly sensitive to bacitracin, chlormephenical, erythromycine, gentamycin, kanamycin, neomycin, streptomycin and tetracycline. On the other hand, all isolates were resistant to colistin and pencillin. These results were in agree with that reported by Al-Ashmay et al. (1996) and Chopra et al. (1980).

It is apparent from the results obtained in this study that meat and milk products sometimes could contain *B.cereus* strains with various levels of contamination. Furthermore if the conditions are suitable, multiply rapidly and produce sufficient toxin to induce symptoms of food poisoning. Therefore, the effective measures should be applied to reduce the number of *B.cereus* in the ingredient through; proper heating (temperature/time) during preparation and filling of the product, adequate refrigeration temperature of the product after the preparation, during distribution and in the house hold and finally prevention of the contamination of raw product by the equipment in relation to scale of production.

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Table (1) Incidence and count of *Bacillus cereus* in examined samples of meat and milk products.

| Products | No. of examined samples | Postive samples | | Count / g or ml | | |
|----------------|-------------------------|-----------------|------|-----------------|-----------------|--------------------|
| | | No. | % | Minimum | maximum | Mean |
| Lunchoen | 40 | 20 | 50 | 2×10^3 | 3×10^6 | 3.8×10^5 |
| Susage | 40 | 23 | 57 | 7×10^2 | 5×10^5 | 5×10^4 |
| kareish cheese | 40 | 15 | 37.5 | 2×10^2 | 3×10^4 | 1.52×10^3 |
| Ice-cream | 40 | 34 | 85 | 10 | 9×10^4 | 4.5×10^4 |
| Yoghurt | 40 | 0 | 0 | 0 | 0 | 0 |

Table (2) The incidence of isolated strains of *Bacillus cereus* producing deoxyribonuclease from meat and milk products.

| Products | No. of strains tested | Deoxyribonuclease producing <i>Bacillus cereus</i> | | | | | |
|----------------|-----------------------|--|-------|---------|-------|--------------|------|
| | | Thermolabile | | | | Thermostable | |
| | | pH(9.0) | | pH(6.7) | | pH(9.0) | |
| | | No. | % | No. | % | No. | % |
| Lunchoen | 20 | 13 | 65 | 2 | 10 | 0 | 0 |
| Susage | 23 | 12 | 52.2 | 3 | 13.04 | 2 | 8.7 |
| Kareish cheese | 15 | 6 | 40 | 3 | 20 | 1 | 6.7 |
| Ice-cream | 34 | 20 | 58.8 | 5 | 14.71 | 0 | 0 |
| Total | 92 | 51 | 55.43 | 13 | 14.13 | 3 | 3.26 |

Table (3): Antibiotic susceptibility of 92 strains of *Bacillus cereus* isolated from meat and milk products.

| Antimicrobial agent | Disk Content | Resistant | | Intermediate | | Sensive | |
|---------------------|--------------|-----------|-------|--------------|------|---------|-------|
| | | No. | % | No. | % | No. | % |
| Ampicillin | 10 mcg | 15 | 16.30 | 2 | 2.17 | 75 | 81.52 |
| Bacitracin | 10 iu | 0 | 0 | 0 | 0 | 92 | 100 |
| Chlormephenicol | 30 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Colistin sulphate | 10 mcg | 92 | 100 | 0 | 0 | 0 | 0 |
| Erythromycin | 15 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Gentamycin | 10 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Kanamycin | 30 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Naladixic acid | 30 mcg | 14 | 15.55 | 2 | 2.17 | 76 | 82.6 |
| Neomycin | 30 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Pencillin | 10 iu | 92 | 100 | 0 | 0 | 0 | 0 |
| Streptomycin | 10 mcg | 0 | 0 | 0 | 0 | 92 | 100 |
| Tetracycline | 30 mcg | 0 | 0 | 5 | 5.43 | 87 | 94.56 |

| Year | 1900 | 1901 | 1902 | 1903 | 1904 | 1905 | 1906 | 1907 | 1908 | 1909 | 1910 |
|--------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Population | 1,000,000 | 1,050,000 | 1,100,000 | 1,150,000 | 1,200,000 | 1,250,000 | 1,300,000 | 1,350,000 | 1,400,000 | 1,450,000 | 1,500,000 |
| Area (sq. miles) | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Population Density | 10,000 | 10,500 | 11,000 | 11,500 | 12,000 | 12,500 | 13,000 | 13,500 | 14,000 | 14,500 | 15,000 |

| Year | 1911 | 1912 | 1913 | 1914 | 1915 | 1916 | 1917 | 1918 | 1919 | 1920 | 1921 |
|--------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Population | 1,550,000 | 1,600,000 | 1,650,000 | 1,700,000 | 1,750,000 | 1,800,000 | 1,850,000 | 1,900,000 | 1,950,000 | 2,000,000 | 2,050,000 |
| Area (sq. miles) | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Population Density | 15,500 | 16,000 | 16,500 | 17,000 | 17,500 | 18,000 | 18,500 | 19,000 | 19,500 | 20,000 | 20,500 |

| Year | 1922 | 1923 | 1924 | 1925 | 1926 | 1927 | 1928 | 1929 | 1930 | 1931 | 1932 |
|--------------------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|-----------|
| Population | 2,100,000 | 2,150,000 | 2,200,000 | 2,250,000 | 2,300,000 | 2,350,000 | 2,400,000 | 2,450,000 | 2,500,000 | 2,550,000 | 2,600,000 |
| Area (sq. miles) | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 | 100 |
| Population Density | 21,000 | 21,500 | 22,000 | 22,500 | 23,000 | 23,500 | 24,000 | 24,500 | 25,000 | 25,500 | 26,000 |