

## CRYPTOSPORIDIASIS IN COMMERCIAL CHICKENS

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## CRYPTOSPORIDIASIS IN COMMERCIAL CHICKENS

(With One Table and One Fig.)

By

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الإصابة بالكربتوسبورديديم فى الدجاج النامى ودجاج اللحم والبيض

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تم جمع عدد ٨٣٠ عينة من محتويات أمعاء الدجاج النامى ودجاج اللحم والبيض (٣٠٠ من الدجاج البلدى و ٥٣٠ من سلالات الدجاج الأجنبى) من محلات بيع الطيور الاجنة بمناطق مدينة الأسماعيلية وتم فحصهم لمعرفة مدى الإصابة بالكربتوسبورديديم فى الدجاج. وتبين من الفحص الميكروسكوبى لمحتويات الأمعاء: أن نسبة الإصابة الأجمالية بحويصلات الكربتوسبورديديم فى الدجاج كانت ١٠.٩٦% ونسبة إصابة دجاج اللحم (بدارى التسمين) أعمارها من ٤٥-٦٠ يوم (١٧.٦%) أعلى عما كانت فى الدجاج النامى البلدى عمر ٣-٤ شهور (٨.٥%) مع الرغم أن الاختلاف كان غير معنوى بين نسبة الإصابة بحويصلات الكربتوسبورديديم فى دجاج البيض البلدى (٤%) وسلالات البيض الأجنبى (٢%) ، وعمامة كانت نسبة الإصابة فى سلالات الدجاج الأجنبى (١٣.٢%) أعلى عما كانت فى الدجاج البلدى (٧%) وبالفحص الهستوباثولوجى للأمعاء المصابة بحويصلات الكربتوسبورديديم وجد أطوار مختلفة من طفيل الكربتوسبورديديم بالأمعاء الدقيقة للدجاج المصاب وأتضح أن الإصابة بالكربتوسبورديديم كانت أكثر إنتشارا فى مجموعات قطعان التربية والأعمار الصغيرة عما فى أعمار الطيور الكبيرة. إن الطيور المصابة بالكربتوسبورديديم ربما لها دور فى انتشار وتواجد هذا الطفيل فى البيئة وما له من علاقة بالصحة العامة.

## SUMMARY

In a survey for Cryptosporidial infection among commercial native and foreign breed chickens of different poultry markets in Ismailia city, the overall prevalence of *Cryptosporidium* sp. oocysts among the examined commercial chickens was 10.96%. The prevalence of Cryptosporidial infection in foreign fattening broilers aged 45-60 days (17.6%) was significantly higher than that reported among native semi-mature chickens aged 3-4 months (8.5%). However, there was no significant difference between Cryptosporidial positive rate of native layers (4%) and that obtained among foreign layers (2%). In general Cryptosporidial infection positive rate of commercial foreign breed chickens (13.2%) was significantly higher than that recorded among commercial native

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breed ones (7%). Histological examination of the infected chicken intestines confirmed various stages of *Cryptosporidium* sp. attached to the epithelial cells of the small intestines. Current research indicates that intestinal *Cryptosporidial* infection occurs on a flock or premises and is more likely to occur in younger birds than older birds. These findings have considerable epidemiological and public health significance for the investigated area.

**Keywords:** *Cryptosporidiasis, commercial chickens*

## INTRODUCTION

*Cryptosporidium* spp. are coccidian parasites that inhabit the microvillous border of a variety of epithelial surfaces of human beings, wild and domestic animals, poultry, fish and reptiles (LEVINE, 1984).

*Cryptosporidia* have been associated with naturally occurring respiratory and enteric diseases and have been shown experimentally to produce disease in several avian species (BERMUDEZ, *et al.* 1988). Intestinal *Cryptosporidiosis* has been associated with sever enteritis and high mortality in naturally infected bob white quail (HOERR, *et al.* 1986). *Cryptosporidial* infections of the intestinal tract have been reported in chicken (RANDALL, 1982; ITAKURA, *et al.* 1984; CURRENT, *et al.* 1986 and LINDSAY, *et al.* 1986). More recent experiments indicate that the isolates from calves, humans, deer, goats and lambs readily infect other species such as mice, rats, guinea pigs, chickens and foals without causing illness (TZIPORI, *et al.*, 1980; REESE, *et al.*, 1982 and SHERWOOD, *et al.* 1982). Mammalian isolates of this parasite readily in-

fect other hosts including humans, indicating a lack of host specificity and zoonotic potential (TZIPORI, 1983). There is little information on the prevalence of intestinal *cryptosporidiosis* for avian species whether commercial or non commercial. Therefore, this study was undertaken to determine the true prevalence of *Cryptosporidium* sp. infection in commercial chickens and also to screen for potential risk factors that may be associated with *Cryptosporidium* infection.

## MATERIAL and METHODS

### Sampling:

Between the period from October 1993 to June, 1994 a total of 830 chickens intestinal contents were collected from the slaughtered commercial native and foreign breed chickens of various purposes; 300 native chickens (200 semimature chickens aged 3-4 months and 100 layers) and 530 foreign chickens (380 fattening broilers aged 45-60 days and 150 layers). All these specimens were collected from different poultry markets in Ismailia city.

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### Examination For *Cryptosporidium* Oocysts:

The chickens intestinal contents were processed by sugar flotation within few hours of collection, in the Dept. of Avian and Aquatic medicine, Faculty of Veterinary Medicine, Suez Canal University. One drop of fluid from the levitation suspension (meniscus) was deposited on a glass slide, cover glassed and examined microscopically at 400x. The levitation suspension was also examined for the detection of *Cryptosporidium* sp. oocysts by iodine wet mount (MA and SOAVE, 1983) as confirmatory test.

### Histopathological Examination:

Fresh intestinal portions were taken from the only available 25 of 67 foreign fattening broiler intestines which had *Cryptosporidium* sp. oocysts. Fixed in 10% neutral buffered formalin and submitted for histologic processing (embedding in paraffin, staining with hematoxylin and eosin) and examined by light microscopy. Histopathologic diagnosis of intestinal *Cryptosporidiosis* was based on characteristic morphologic appearance and location of the parasites.

The analysis of data was achieved by chi-square ( $\chi^2$ ) to evaluate whether or not association existed between the risk factors and identification of *Cryptosporidium* sp. oocysts.

## RESULTS

The prevalence of *Cryptosporidium* oocysts among the examined intestinal contents of commercial chickens is summarized in table (1), the overall prevalence of *Cryptosporidium* oocysts of the commercial chickens was 10.96%. In native breed chickens, the prevalence of *Cryptosporidium* infection of semimature chickens aged 3-4 months (8.5%) was higher than that obtained among layers (4%), however, statistically there was no significant difference ( $\chi^2=2.9$ ,  $P>.05$ ). In foreign breed chickens, *Cryptosporidium* oocysts positive rate was significantly higher among fattening broilers aged 45-60 days (17.6%) than that obtained of layers (2%) ( $\chi^2=22.96$ ,  $P<.01$ ).

Although, the prevalence of *Cryptosporidial* infection of foreign fattening broilers was significantly higher than that reported among native semimature chickens ( $\chi^2=8.8$ ,  $P<.01$ ). There was no significant difference between *Cryptosporidial* positive rate among native layers and that obtained among foreign layers ( $\chi^2=0.83$ ,  $P>.05$ ).

The distribution of *Cryptosporidium* infection in the different intestinal portions was histopathologically studied on the only available 25 foreign fattening broiler intestines which had *Cryptosporidium* sp. oocysts in its contents. Twenty three (92%) out of 25 infected intestines had various stages of *Cryptosporidia* colonized

the microvillous borders in its different predilection sites. Cryptosporidia as a numerous round to oval basophilic bodies; 2 to 4µm in diameter were found in the apical portions of enterocytes of the middle and lower portion of the chickens small intestines (Fig. 1). Cryptosporidium was not observed in the duodenum, cecum and colon of the histopathologically examined intestines.

### DISCUSSION

Cryptosporidium sp. are unique among coccidia of vertebrates because of their endogenous development on the microvillous border of epithelial cells (i.e. intracellular but extracytoplasmic), lack of host specificity and their ability to infect various tissues (DHILLON, *et al.* 1981 and CURRENT, 1984). Recent attention given to Cryptosporidium sp. infection in otherwise healthy (WOLFSON, *et al.* 1885) or immunodeficient (SOAVE, *et al.* 1984) persons with diarrhea has brought about increased awareness of the probable role of Cryptosporidium as a pathogen in birds. In the present study; the overall prevalence of Cryptosporidium sp. oocysts among the examined commercial chickens (10.96%) was nearly similar to that reported by BADAWY (1989). Who found that the overall incidence of Cryptosporidium sp. oocysts of chickens was 12.5% in Egypt. Cryptosporidial infection was

more prevalent in foreign breed fattening broilers aged 45-60 days this was nearly similar with that obtained by RANDALL (1982) who found infection in 18.7% of broilers aged 4-7 weeks. On the other hand, low incidence of infection (6.6% and 6.4%) was recorded by GORHAM, *et al.* (1987) and GOODWIN and BROWN (1988) in chicken aged 21-30 days and 14-51 days respectively. A higher rate of infections was reported by SNYDER, *et al.* (1988) in chickens aged 49 (38%) and 63 days (50%). Cryptosporidium sp. oocysts positive rate recorded among the foreign breed layers (2%), in this work was a relatively similar to that recorded by LEY, *et al.* (1988).

Our data indicated that the percentage of Cryptosporidial infection among native semimature chickens aged 3-4 months and layers was in agreement with that reported by BADAWY (1989) who found Cryptosporidial infection in 9.1% in age group 3-9 months and 4.6% in age group 6-12 months.

According to the results of this work, our findings indicated that the prevalence of Cryptosporidial infection decreased with increased age of chickens. This confirms the results obtained by DHILLON *et al.* (1981); GORHAM *et al.* 1987; SNYDER *et al.* (1988) and Badawy (1989). The discrepancy between the obtained prevalence of Cryptosporidial infection among the examined chickens in the present study and

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those of other authors may be due to several factors, such as breed, age, species, management and sanitary conditions of the examined birds and the method of diagnosis used.

Most infectious organisms have ecological niches that they prefer over all others. In humans and other animals *TZIPORI, 1983*), *Cryptosporidium* sp. most commonly are found in the middle portion of the small intestine. Histological examination of the affected intestines confirmed various stages of *Cryptosporidium* sp. attached to the epithelial cells of the intestines. The severity of infection in the cases reported in the present study ranged from light to severe (*PAPADOPOULOU et al., 1988*). Concerning the predilection site of infection in the naturally infected chicken intestines, the present study emphasized that the intestinal tissues most frequently affected with the parasite were the middle and lower portions of the small intestines. This is in agreement with that by *GOODWIN, et al. 1988 and LINDSAY et al. (1986)* who stated that *Cryptosporidia* was not observed in the duodenum, and cecum in the orally inoculated chickens. Moreover, *BADAWY (1989)* found no parasites in duodenum, jejunum and cecum of the naturally infected chickens of different age groups.

The result of the present investigation disagreed with that reported by *TYZZER (1929)* who stated that,

the infection in chicken was confined to the tubular portion of the ceca. Comparison of these observations suggests that different avian *Cryptosporidium* species or strains, age and breed of the birds *BERMUDEZ et al. al., 1988 and BADAWY (1989)* may exhibit variations in tissue tropism in their hosts.

The obtained results, in this study, support the conclusion that intestinal *Cryptosporidiosis* occurs on a flock or premises basis and is more likely to occur in younger birds than older birds (*LEY et al. 1988*). In order to diagnose *Cryptosporidiosis* of the small intestine, the histology laboratory should receive clinical specimens from the middle and lower segments of the small intestine.

Cross-species infectivity studies suggest that *Cryptosporidia* are not host specific. Under experimental conditions calf isolates could infect the intestines of chickens (*TZIPORI et al., 1980*). Mammalian isolates of *Cryptosporidium* are transmissible to other species of mammals and *Cryptosporidiosis* is now recognized as a zoonoses (*TZIPORI, 1983*).

From the reported reviews and the present results, the *Cryptosporidium* sp. infections are more prevalent among chickens and it is suggested that chickens may play a role in maintenance and spread the infection in the environment. These findings have considerable epidemiological

and public health significance for this investigated area.

#### REFERENCES

- BADAWY, B.A.M. (1989): Studies on fowl Cryptosporidiosis in Egypt. Fac. of Vet Med Cairo Univ. PH.D. thesis.
- Bermudez, A.J.; Ley, D.H.; Levy, M.G.; Ficken, M.D.; Guy, J.S. and Gerig, T.M. (1988): Intestinal and Bursal Cryptosporidiosis in Turkeys following inoculation with *Cryptosporidium* sp. isolated from commercial poults. *Avian diseases*, 32: 445-450.
- Current, W.L. (1984): *Cryptosporidium* and Cryptosporidiosis of domestic animals and man. Proc. Fourth Int. symp. Neonatal Diarrhea, Saskatoon pp.293-303.
- Current, W.L.; Upton, S.J. and Haynes, T.B. (1986): The life cycle of *Cryptosporidium baileyi* sp. (Apicomplexa Cryptosporididae) infecting chickens. *J. protozoal*, 33: 289-297.
- Dhillon, A.S.; Thacker, H.L.; Dietzel, A.V. and Winterfield, R.W. (1981): Respiratory Cryptosporidiosis in broiler chickens. *Avian Dis.* 25: 747-751.
- Goodwin, M.A. and Brown, J. (1988): Histologic incidence and distribution of *Cryptosporidium* sp. infection in chickens: 68 cases in 1986. *Avian Dis.* 32:365-369.
- Goodwin, M.A.; Steffens, W.L.; Russell, I.D. and Brown, J. (1988): Diarrhea associated with intestinal Cryptosporidiosis in Turkeys. *Avian Dis.* 32:63-67.
- Gorham, S.L., Mallison, E.T., Snyder, D.B. and Odor, E.M. (1987): Cryptosporidia in the bursa of Fabricius—a correlation with mortality rates in broiler chickens. *Avian pathol.* 16: 205-211.
- Hoerr, F.J.; Current, W.L. and Haynes, T.B. (1986): Fatal Cryptosporidiosis in quail. *Avian Dis.* 30: 421-425.
- Itakura, C.; Goryo, M. and Umemura, T. (1984): Cryptosporidial infection in chickens. *Avian patho.* 13: 487-499.
- Levine, N.D. (1984): Taxonomy and review of the coccidian genus *Cryptosporidium* (protozoa, Apicomplexa). *J. protozool*; 31: 94-98.
- Lindsay, D.S.; Blagburn, B.L.; Sandermann, C.A.; Hoerr, F.J. and Ernest, T.A. (1986): Experimental *Cryptosporidium* infections in chickens: oocyst structure and tissue specificity. *Am.J.Vet. Res.* 47: 876-879.
- Ley, D.H.; Levy, M.G.; Hunter, L.; Corbett, W. and Barnes, H.J. (1988): Cryptosporidia-positive rates of avian necropsy specimens determined by examination of auramine stained fecal smears. *Avian Dis.* 32: 108-113.
- Ma, P. and Soave, R. (1983): Three-step stool examination for Cryptosporidiosis in 10 Homosexual men with protracted watery diarrhea. *J.Infect. Dis.* 147,5: 824-828.
- Papadoupoulou, C.; Xylour, F. and Zisides, N. (1988): Cryptosporidial infection in Broiler chicken in Greece. *Avian Dis* 32: 842-843.
- Randal, C.J. (1982): Cryptosporidiosis in the bursa of Fabricius and trachea in broilers. *Avian pathol.* 11: 95-102.
- Reese, N.C.; Current, W.L.; Ernst, J.R. and Bailey W.S. (1982): Cryptosporidiosis of man and calf: a case report and results of experimental infection in mice and rats. *Am. J. Trop. Med. Hyg.* 31: 226-229.
- Sherwood, D.; Angus, K.W.; Snodyrass, D.R. and Tzipori, S., (1982): Experimental Cryptosporidiosis in laboratory mice. *Infect. Immun.*, 38: 471-475.
- Soave, R.; Danner, R.L. and Honig, C.L. (1984): Cryptosporidiosis in homosexual man. *Ann. Intern. Med.* 100: 504-511.

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- Snyder, D.B.; Current, W.L.; Russek-Cohen, E.; Gorham, S.L.; Mallinson, E.T.; Marguardt, W.W. and Savage, P.K. (1988):* Serologic incidence of *Cryptosporidium* in Delmarva broiler flocks. *Poult. Sci* 67: 730-736.
- Tzipori, S.; Angus, K.W.; Campbell, I. and Gray, E.W. (1980):* *Cryptosporidium*; evidence for a single species genus. *Infect. Immun.* 30: 884-886.
- Tzipori, S. (1983):* Cryptosporidiosis in animals and humans. *Microbiol. Rev.* 47: 84-96.
- Tyzzer, E.E. (1929):* Coccidiosis in gallinaceous birds. *Am.J. Hyg.* 10: 269-284.
- Wolfson, J.S.; Richter, J.M.; Waldron, M.A.; Weber, D.J.; Mecorothy, D.M. and Hopkins, C.C. (1985):* Cryptosporidiosis in immunocompetent patients. *N. Engl. J. Med.* 312: 1278-1282.

Table (1): Prevalence of *Cryptosporidium* sp. oocysts among the examined intestinal contents of the commercial chickens

Breeds	Total No. of intestinal content specimens	Number of the chicken intestinal contents positive for <i>Cryptosporidium</i> sp. oocysts/total number of the examined chickens	
		+ve	%
Native breed			
Semimature chickens	200	17/200	8.5
Layers	100	4/100	4
Total	300	21/300	7
Foreign breed			
Fattening broilers	380	67/380	17.6
Layers	150	3/150	2
Total	530	70/530	13.2
Grand total	830	91/830	10.96



Fig. (1): Light photomicrograph of *Cryptosporidium* (arrow) parasitizing small intestinal absorptive epithelial cells, the lamina propria is infiltrated by mixed population of inflammatory cells (H & E x 400)