

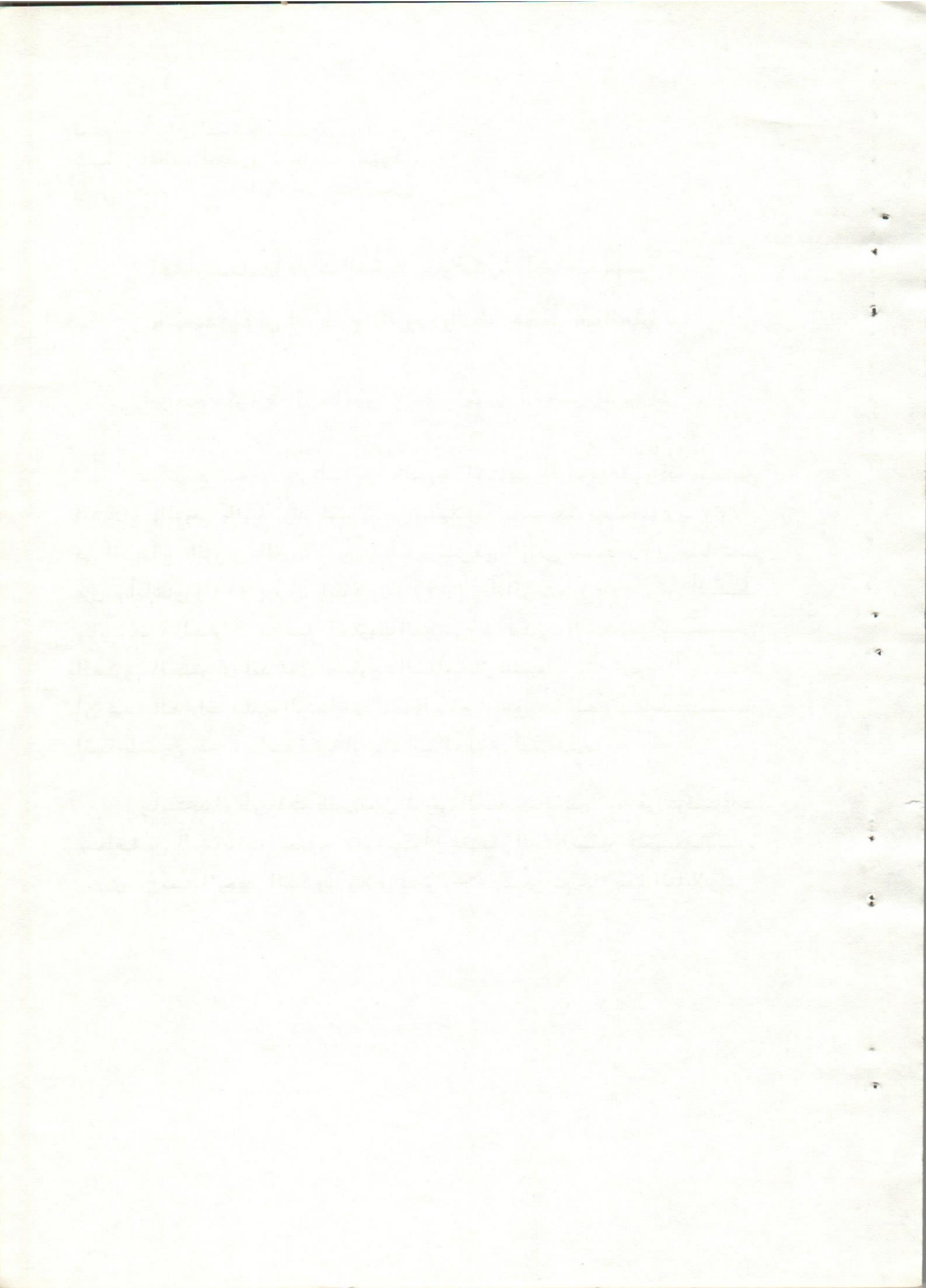
قسم : أمراض الد واجسن .
كلية : الطب البيطري - جامعة أسيوط .
رئيس القسم : أ.د / ابراهيم حسن سكر .

الختبار حساسية عترات الميكوبلازما والبكتريا المصاحبة لها والمعتمولة من الدجاج والرومي والبط بصعيد مصر العليا

ابراهيم سكر ، عادل سليمان ، صلاح موسى ، محسن الد مرداش

تم تجميع مسحات من الخنجرة والجيوب الأنفية والاكياس الهوائية من
الدجاج والرومي والبط وقد ثبت وجود ميكوبلازما جاليسبتكم بنسبة ٢٤ - ٣٢ %
في الدجاج والرومي بالترتيب وم . ميليا جريس في الرومي بنسبة ٣٠ % بينما تم
عزل م. أناثس (١٤ %) ، أ . ليدلاوى (٢٨ %) ، أ. أكزايثم (٣٢ %) من البط
وكانت هذه العترات مصاحبة للميكروب القولوني ، ميكروب السيد ومانس
والميكروب العنقودي الذهبي وميكروب السالمونيلا بللورم . وقد أوضح البحث
أن هذه العترات عالية الحساسية للجوراميسين ومتوسطة الحساسية
لليנקومايسين وغير حساسة لبقية المضادات الحيوية المستخدمة .

وباستخدام طريقة تغطيس بيض الرومي المصاب بالميكوبلازما في تركيزات
مختلفة من المضادات الحيوية فقد ثبت أن عقنار الجوراميسين تسبب في
خفض نسبة وجود الميكروب وتلاه الد وكسي سيكليين ثم جاء بعد التالان .



Dept. of Poultry Diseases,
Faculty of Vet. Med., Assiut University,
Head of Dept. Prof. Dr. I.M. Sokkar.

**IN-VITRO SENSITIVITY OF MYCOPLASMAS AND ASSOCIATED BACTERIA
ISOLATED FROM CHICKENS & TURKEYS AND DUCKS
AT THE AREA OF UPER EGYPT**

(With 3 Tables)

By
I.M. SOKKAR; A.M. SOLIMAN; S. MOUSA and M.Z. EL-DEMERDASH
(Received at 21/3/1985)

SUMMARY

Swabs from trachea, sinuses and air-sacs taken from chickens, turkeys and ducks of different ages revealed the presence of M. gallisepticum (24 and 32%) in chickens and turkeys respectively, M. meleagridis (30%) were detected in infected turkeys, while M. anatis (16%), M. gallinarum (24%), M. iners (14%), A. laidlawi (28%) and A. axanthum (32%) were recovered from ducks. These isolates were found in association with E. coli, Pseud. auroginosa, Staph. aureus and S. gallinarum pullorum. All isolates showed high sensitivity to josamycine, moderate sensitivity to lincomycine and low variable sensitivity to other antibiotics. Incubated turkey-eggs from infected farm showed good elimination of mycoplasma after dipping in josamycine solution, while inferior results were obtained with doxycycline and tylosin-tartrate treatment.

INTRODUCTION

Mycoplasmas have been isolated from man, animals and birds, most of these mycoplasmas are pathogenic and cause specific diseases. Mycoplasmas were first isolated from poultry (NELSON, 1935), Several disease conditions were reported due to mycoplasma species. CRD in chickens and sinusitis in turkeys caused by M. gallisepticum was described by AMIRA (1976) and SOLIMAN (1982). Air-sacculitis in turkey poults, late incubation mortality and poor growth rate due to M. meleagridis were reported by GHAZIKHANI and YAMAMOYO (1974). Air-sacculitis and ascitis in ducks due to M. anatis, M. gallinarum, M. iners, A. laidlawii and A. axanthum were described by ROBERTS, (1964), KARPAS, (1969), EL-EBEEDY, (1976), FAWZIA, (1976) and SOLIMAN, (1985). Complicated air-sacculitis in turkey- poults infected with M. meleagridis and E. coli were recorded by MOHAMED et al. (1970). Also E. coli, Staph. aureus, Pseud. auroginosa and S. gallinerum pullorum were recovered from upper and lower respiratory organs by BERGMAN, et al. (1980) and KIBENGE and WILCOX, (1983). MURATA, et al. (1981) and SOLIMAN, (1982, 1985) tested the effect of several antibiotics on mycoplasma isolates, and reported greater drug resistance to tylosin-tartrate. Also RAO, et al. (1976) showed that 98.8, 93.3, 46.6 and 76% of 345 E. coli strains were resistant to erythromycin, streptomycin, oxytetracycline and chlorotetracycline respectively. YOON, et al. (1981) and EL-BAKRY (1983) applied the in-vitro sensitivity test on E. coli, S. gallinarum pullorum Staph. aureus and Pseud. auroginosa, they reported the greater resistance of various E. coli strains and Pseud. auroginosa to erythromycin, neomycin, sulphonamides and Nitrofurantion. An increased rate of hatchability

and low recovery rate of mycoplasmas in treated turkey-eggs by egg-injection or egg-dipping by using various antibiotics were reported by (YAMAMOTO and BIGLAND, 1966 & ELFTERESCU, *et al.* 1972 & MACAPES, *et al.* 1976, 1977 & ELMAHI and HOFSTAD, 1978).

The present work was planned to cover the following items:

- Isolation and identification of mycoplasmas and the associated bacteria from chicken, turkey and duck flocks in the area of Upper Egypt.
- Application of the in-vitro sensitivity test on the recovered strains and associated bacteria against the available drugs to choose the effective ones.
- Trials for treating infected turkey eggs by antibiotic dipping to control mycoplasma infection in turkeys.

MATERIAL and METHODS

I- Isolation and identification of mycoplasmas and associated bacteria:

1) **Mycoplasmas:** tracheal, sinuses and air-sac swabs were collected on Brain-Heart-infusion broth, from living and dead chickens, turkeys and ducks of different ages. Samples were cultured as described by SABRY (1968). Inoculated broth and agar media were incubated at 37°C, however agar plates were incubated in moist candle jar under reduced oxygen tension. After 3-days plates were examined microscopically for appearance of characteristic colonies. The suspected colonies were subjected to further identification: digitonine-sensitivity test (FREUNDT, *et al.* 1979), biochemically (SABRY, 1968 & ERNO and STIPKOVITIS, 1973) and Serologically (CLYDE, 1964 and KROGSGARD-JENSEN, 1972).

2) **Associated bacteria:** Corresponding samples to that for mycoplasmas were inoculated on nutrient-broth and incubated at 37°C. This was followed by subculturing on blood-agar, MacConky (agar, S-S agar and Crystal-Violte-blood agar. Subcultures were incubated at 37°C for 24 hours. Suspected colonies were picked up and subjected to further biochemical and serological identification (CRUICKSHANK, *et al.* 1975).

II- In-Vitro sensitivity of mycoplasma and associated bacteria against antimicrobial agents:

Sensitivity discs produced by Oxoid-laboratories, England and Ugjhon Company, U.S.A. were used, while sensitivity discs for josamycin (0.05 mg/ml) were prepared from "alplucine 20%" produced by Virbac-laboratories, France, and "Doxycycline" discs were prepared from "Vibravet" produced by Pfizer, New York (0.1 mg/ml). The test was carried out on mycoplasmas after CLYDE (1964), and on associated bacteria after KOLMER, *et al.* (1951).

III- Antibiotic treatment of naturally infected turkey-eggs with mycoplasmas:

A total of 200 turkey-eggs were obtained from El-Wadi El-Gadid turkey farm the had the history of mycoplasma infection. Eggs were divided into four group of 50 eggs each. Eggs were dipped in antibiotic solutions at 2-5°C for one minute. Antibiotic solutions were prepared from "alplucine, 50 p.p.m.", Vibravet 50 p.p.m." and "Tylosine-tartarate 200 p.p.m." The fourth group of eggs served as non treated control. At the end of incubation dead-embryos and hatched poult after two weeks old, were subjected to Post-mortum and mycoplasma examination.

RESULTS

Results of isolation and identification of mycoplasmas and associated bacteria and the in-vitro sensitivity of these isolates as well as antibiotic turkey-egg treatment are illustrated in tables 1,2,3 respectively.

IN-VITRO SENSITIVITY OF MYCO. AND ASSOCIATED BACT.

DISCUSSION

It is clear from the results of isolation and identification that mycoplasmosis in poultry constitute an important economic problem causing high losses in flocks at the area of Upper Egypt. Results of recovery of *M. meleagridis* from turkey air-sac of different ages agreed with FREY, et al. (1968) and YAMAMOTO (1978). As well as results of *M. gallisepticum* recovery from turkey sinuses exudate were agreed with SOLIMAN (1982) while its recovery from trachea, air-sacs and sinuses of chickens were agreed with AHMED (1980). The recovery of *M. anatis* & *M. gallinarum* & *M. iners* & *A. laidlawii* and *A. axanthum* from duck samples were parallel with observations of KARPAS and FABRICANT (1969), FAWZIA (1976), EL-EBEEDY (1976) and SOLIMAN (1985).

Results of recovery of associated bacteria were agreed with BERGMAN, et al. (1983).

Results of the in-vitro sensitivity indicates that all mycoplasma strains and associated bacteria were highly sensitive to josamycin and varied from sensitive to weak sensitivity to other antibiotics, while drug resistance was recorded in Tylosin-tartrate, Nitrofurantion, Neomycin and oxytetracycline. These results were agreed with KLEVEN, et al. (1971), ELMAHI, et al. (1978), HAMDY, et al. (1980), YOON, et al. (1981), SOLIMAN (1982), EL-BAKRY (1983), and SOLIMAN (1985).

Results of egg-dipping in antibiotics revealed the higher sensitivity of josamycin and greater resistance to Tylosintartrate and this agreed with GHAZIKHANIAN and YAMAMOTO (1969), MURATA, et al. (1981).

Results of sensitivity test points to the need of periodical testing of bacterial isolates to detect their sensitivity to the available antimicrobial agents in order to choose the more effective drugs for using in the prophylactic and therapeutic programs.

REFERENCES

- Ahmed, S.Z. (1981): Studies on mycoplasmosis in poultry. M.V.Sc., Faculty of Vet. Med., Cairo Univ.
- Amira, H.M. (1974): Haematological and Histopathological changes in turkeys naturally infected with *M. meleagridis*. M.V.Sc. Thesis, Fac. Vet. Med., Cairo Univ.
- Bergman, V.; Kohler, B. and Vogel, K. (1980) *Saphylococcus aureus* infection of fowls on industrialized Poultry Units. I type of infection. Archiv Fur experimentelle Veterinarmedizin 34 (6): 891.
- Bigland, C.H. (1971): Experimental control of *M. meleagridis* in turkeys, by the dipping of eggs in tylosin and spiramycin. Can. J. Comp. Med. 34: 26-30.
- Clyde, W.A. (1964): Mycoplasma species identification based upon growth inhibition test by specific antisera. J. Immunology 92: 958-965.
- Cruickshank, R.; Duguid, J.P. and Swain, R.H. (1975): Medical Microbiology 12 15 ed. E.S. Livingstone Limited Edinburgh and London.
- El-Bakry, M.S. (1983): Some studies on *Pseudomonas* infection in chickens. M.V.Sc. Thesis (Poultry Dis.). Fac. of Vet. Med., Assiut Univ.
- El-Ebeedy, A.A. (1977): Studies on mycoplasma infection of turkeys, geese and ducks. Candidate D. Thesis, Hungarian Academy of Science, Budapest.
- Elmahi, M.M. and Hofstad, M.S. (1978): Prevention of egg transmission of *M. meleagridis* by antibiotic treatment of naturally and experimentally infected turkey eggs. Avian Dis. 23 (1): 88-94.

- Ern, H. and Stipkovitis, L. (1973): Bovine Mycoplasmas: Culture and biochemical studies. *Acta. Vet. Scand.*, 14: 436-449.
- Fawzia Mohamed Mostafa (1976): Studies on isolation and characterization of mycoplasma and acholeoplasma of ducks. M.V.Sc., Dept. of Microb.; Fac. of Vet. Med., Cairo University.
- Freundt, E.A.; Erno, H. and Lemcke, R.M. (1979): Identification of mycoplasmas in: *Methods in microbiology*. Vol. 13. T. Bergan, and J.R. Norris, eds. Academic press, London. pp. 391-401.
- Frey, M.L.; Hanson, R.P. and Anderson, D.P. (1968): A medium for the isolation of avian mycoplasma. *Am. J. Vet. Res.* 24: 2163-2171.
- Ghazikhanian, G. and Yamamoto, R. (1969): Tylosin resistant strains of *M. meleagridis*. *Proc. 3rd Calif. Poultry Symp. Univ. Calif. Davis.* pp. 36-37.
- McCapes, R.H.; Yamamoto, R.; Ghazikhanian, G.; Dungan, W.M. and Ortmyer, H.B. (1977): Antibiotic egg injection to eliminate *M. meleagridis* infection. *Avian Dis.*, 21(1): 57-68.
- Mohamed, Y.S.; Moorhead, P.D. and E.H. Bohl (1970): *M. meleagridis* and *E. coli* infections in germ free and S.P.F. turkey poults: production of complicated air-sacculitis. *Amer. J., Vet. Res.*; 31: 1637-1643.
- Murata, M.; Kajikawa, M.; Kuniyasu, C. and Yoshida, Y. (1981): Sensitivity in vitro of *M. gallisepticum* to antibiotics. *J. of Fac. App. Biological Sci. Hiroshima Univ.*, 19 (1): 55-68.
- Nelson, J.B. (1935): Cocco-bacilliform bodies associated with an infectious fowl coryza. *Science*, 82: 43-44.
- Nelson, R.C.; Dungan, W.M. and Larsen, C.T. (1974): Comparison of the performance of *M. meleagridis* free and infected poults. *Proc. 23rd West Poul. Dis. Conf. and 8th Poul. Health Symp. Univ. Calif. Davis.* pp. 66-69.
- Rao, M.V.; Kulshreshtha, S.B. and Kumar, S. (1976): Drug resistance in *E. coli* isolated from the intestinal tract of poultry. *Ind. J. of animal Sci.* 44 (6): 345.
- Ghazikhanian, G. and Yamamoto, R. (1974): Characterization of pathogenic and non-pathogenic strains of *M. meleagridis*: Manifestation of disease in turkey embryos and Poults. *Am. J. Vet. Res.* 35: 417-424.
- Hamdy, A.H.; Said, Y.M.; Kleven, S.H.; Yamamoto, R.; Newman, J.A. and Kartzner, D.D. (1980): Linco-spectin medication of *M. meleagridis* air-sacculitis in turkey-poults. *Avian Dis.*, 23 (3): 670-681.
- Karpas, D. and Fabricant, J. (1969): Recovery of *M. anatis* from ducks. *Proc. Annu. Meet. Northeast. Conf. Avian Dis. 41st Univ. Maine P.* 37.
- Kleven, S.H.; Pomeroy, B.S. and Nelson, R.C. (1971): Ineffectiveness of antibiotics treatment of semen in the prevention of egg-transmission of *M. meleagridis* in turkeys. *Poultry Sci.* 4 (5): 1522-1523.
- Kolmer, J.A.; Spulding, E.H. and Robinson, H.W. (1951): *Approved laboratory technique* 5th Ed. Appleton century-Corfts, Inc., New-York.
- Krogsgard-Jensen, A. (1972): Mycoplasma growth-precipitation as a serodiagnostic method. *Appl. Microbiol.*, 23: 553-558.
- McCapes, R.H.; Yamamoto, R.; and Ortmyer, H.B. (1976): Injecting antibiotics into turkey hatching eggs to eliminate *M. meleagridis* infection. *Avian Dis.*, 19 (3): 506-514.
- Roberts, D.H. (1964): The isolation of an influenza A-virus and amycoplasma associated with duck sinusitis. *Vet. Rec.*, 76: 470-473.
- Rzedzicki, M.A. (1975): In-Vitro antibiotics sensitivity of some strains of *Sal. pullorum* and *Sal. gallinarum*. *British Poultry Sci.* 8.

IN-VITRO SENSITIVITY OF MYCO. AND ASSOCIATED BACT.

- Sabry, M.Z. (1968): Characterization and classification of avian mycoplasmas. Ph.D. Thesis, Cornell Univ., USA, pp. 257.
- Sokkar, I.; Nafei, E.; A.A. Ibrahim; M. Shahata; S. Mousa; Hashim, S. and A.El-Tamawi (1983): The role played by microbial infections on hatchability rate of duck-embryos. Assiut Vet. Med. Journal (In press).
- Soliman, A.M. (1982): Some studies on the incidence of Mycoplasmosis in turkeys in Upper Egypt. M.V.Sc. Thesis (Poultry Dis.). Fac. of Vet. Med. Assiut Univ. (1982).
- Soliman, A.M. (1985): Further investigations on duck-mycoplasmosis in Upper Egypt. Ph.D. Thesis (Poultry Dis.). Fac. of Vet. Med. Assiut Univ. (1985).
- Yamamoto, R. (1978): M. meleagridis in genital tract of turkey hens and its role in egg-transmission. In "Diseases of poultry" (M.S. Hofstad) 7th Ed. pp. 250-260. Iowa State Univ. Press. Ames.
- Yamamoto, R. and Adler, H.E. (1958): Characterization of pleuropneumonia-like organisms of avian origin. II-Culture; biochemical; morphological and further serological studies. J. infectious Dis., 102: 243-245.
- Yamamoto, R. and Ortmyer, H.B. (1967): Hatcher and intraflock transmission of M. meleagridis. Avian Dis., 11: 481-512.
- Yoon, Y.D.; Kim, I.M. and Kim, B.H. (1981): Antimicrobial drug susceptibility of salmonella isolated from various animals. Korean. J. of Vet. Public Health 5: (1): 19.

Table (1)
Recovery Rate of Mycoplasmas and associated bacteria from chicken and Turkey and duck samples

Isolates	Chicken			Turkey			Duck		
	No. Examined	No. Positive	Percentage %	No. Examined	No. Positive	Percentage %	No. Examined	No. Positive	Percentage %
<u>M. gallisepticum</u>	50	12	24	50	16	32	50	--	--
<u>M. meleagridis</u>	50	--	--	50	15	30	50	--	--
<u>M. anatis</u>	50	--	--	50	--	--	50	8	16
<u>M. gallinarum</u>	50	--	--	50	--	--	50	12	24
<u>M. iners</u>	50	--	--	50	5	10	50	7	14
<u>A. laidlawii</u>	50	--	--	50	6	12	50	14	28
<u>A. awanthum</u>	50	--	--	50	--	--	50	16	32
<u>E. coli</u>	50	20	40	50	12	24	50	8	16
<u>S. gallinarum pullorum</u>	50	7	14	50	3	6	50	3	6
<u>Staph. aureus</u>	50	5	10	50	3	6	50	2	4
<u>Pseud. aeruginosa</u>	50	12	24	50	15	30	50	10	20

IN-VITRO SENSITIVITY OF MYCO. AND ASSOCIATED BACT.

Table (2)
Results of in-vitro sensitivity of Mycoplasmas and associated Bacteria

Isolate strains	Josamycine 0.05 mg/ml.	Doxycycline 0.1 mg/ml.	Tylosine- Tartarate 0.1 mg/ml.	Spectinomycin 100 ug	Lincomycin 20 ug	trythromycin Erythromycin 15 ug	Nitrofurantion 30 ug	Oxytetracycline 30 ug	Neomycin 30 ug	Chloramphenocol 30 ug	Ampicillin 10 ug	Tetracycline 30 ug
<u>M. gallisepticum</u>	+++	++	-	+	++	+	-	+	-	-	+	+
<u>M. meleagridis</u>	+++	+	-	+	++	+	-	+	-	-	+	+
<u>M. anatis</u>	+++	+	+	+	++	+	+	+	-	+	+	+
<u>M. gallinarum</u>	+++	+	+	+	++	+	+	+	+	+	+	+
<u>M. iners</u>	+++	+	+	+	++	+	+	+	+	+	+	+
<u>A. laidlamii</u>	+++	+	+	+	++	+	+	+	+	+	+	+
<u>A. axanthum</u>	+++	+	+	+	+	+	-	+	+	+	+	+
<u>E. coli</u>	+++	++	-	-	++	++	+	+	+	+	+	+
<u>S. gallinarumpollorum</u>	+++	+	-	-	++	+	-	+	-	+	+	+
<u>Staph. gurus</u>	+++	+	-	-	++	+	+	+	+	+	+	+
<u>Pseud. aeruginosa</u>	+++	-	-	-	++	+	+	+	+	+	+	+

++++ highly sensitive & +++ sensitive & ++ Moderately sensitive & + weakly sensitive and - Resistant.

Table (3)
Results of Hatching Rate and Recovery Rate of Mycoplasmas from Antibiotic treated turkey eggs

Antibiotic drugs	Number of Eggs:				Hatched Poults Recovery Rate of Mycoplasma from							
	Inoc- ulated	Infer- tile	Alive at day:			Number	Hatch- ability Perce- ntage %	Dead Embryos		Hatched Poults after two- week age		
			7	15	21			No.exam.	No.Pos.	No.exam.	No.with air-sac	No.Pos.
Josamycin*	50	9	41	35	33	30	73	11	2	30	5	8
Doxycycline**	50	9	35	28	18	15	36	26	14	15	8	12
Tylosine-tartrate	50	8	32	25	15	12	28	30	28	12	9	11
Control	50	10	30	25	12	9	22	31	28	9	7	9

* Josamycin = alphucine-20% Verbac Lab., France.

** Doxycycline = Vibravet Pfizer lab., U.S.A.