# Systematic review of opportunistic parasites among Egyptian immunocompromised individuals from 2010 to 2020

Review Article

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### ABSTRACT

Opportunistic parasites are commonly linked with immunocompromised individuals due to weakness in their immune system. Alteration in their cellular and humoral responses leads to hindrance of T and B lymphocytes from efficiently acting against opportunistic pathogens. Accordingly, immunocompromised patients present increased susceptibility to different microorganisms including viral, bacterial, fungal, and parasitic infections. Several conditions are commonly associated with host immune system impairment. Among them enrolled in the present review were malignancy, chronic liver diseases, diabetes mellitus, renal failure, organ transplantation, and inflammatory bowel disease. The most common reported opportunistic parasites include species of *Cryptosporidium*, *Blastocystis*, and *Microsporidium*, as well as *T. gondii*, *C. cayetanensis*, *I. belli*, and *S. stercoralis*. The objective of the present systematic review is to increase awareness concerning opportunistic parasitosis among Egyptian immunocompromised individuals from 2010 to 2020 with particular reference to their relative detection rates and risk factors of infection.

Keywords: Blastocystis, Cryptosporidium, Microsporidium, opportunistic, T. gondii

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#### **INTRODUCTION**

The immune system is composed of B and T lymphocytes, phagocytic cells and complement system. Immunodeficiency occurs when one part of the immune system is not efficient or has lost its function, and it may be either primary (congenital) or secondary (acquired). Primary immunodeficiency diseases (PID) are hereditary disorders, caused by mutations of specific genes. They result in increased susceptibility to infections and a predisposition to autoimmune diseases and malignancies. While secondary immunodeficiency diseases are caused by infectious agents such as human immunodeficiency virus (HIV), or corticosteroid chemotherapy, organ transplants, metabolic diseases, irradiation, malnutrition, and environmental conditions<sup>[1-3]</sup>. According to the component of the immune system, PIDs are classified as adaptive or innate immunity disorders<sup>[4]</sup>. More than 150 different types of PID were recorded to date<sup>[1]</sup>. Disorders of adaptive immunity include cellular and humoral immunodeficiency disorders due to defects in both T and B lymphocytes development, differentiation, and maturation. Since B cell mediated-antibody production requires intact T lymphocyte function, so any defects in the T lymphocyte will lead to combined immunodeficiency disorders. Disorders of innate immunity are due to failure of the innate system resulting in delay in the induction of the immune response and may worsen outcomes of infections<sup>[5,6]</sup>.

On the other hand, several studies revealed that interleukins (IL) 12, and 18, and interferon gamma  $\,$ 

(IFN- $\gamma$ ) may play a role in improvement of the protective immunity against cryptosporidiosis<sup>[7-9]</sup>. The major source of IL-12 is the dendritic cells that are involved in mediating the immune responses in the host<sup>[10]</sup>. They have a potentially important role in protection against infection, and they are also involved in degradation and transport of antigens to the lymph nodes, and release chemokines in response to cryptosporidiosis<sup>[11]</sup>. Therefore, opportunistic parasites are common in immunocompromised individuals when the CD4<sup>+</sup> T lymphocyte counts fall below 200 cells/ $\mu$ l<sup>[12-14]</sup>.

Childhood and elderly populations are susceptible to opportunistic infections due to depression of their cellular immunity that leads to increase in disease morbidity<sup>[15]</sup>. The relationship between protein-energy malnutrition and immunitv increases vulnerability to infectious diseases causing immunological impairments<sup>[16]</sup>. Alcohol slows down the functions of the phagocytes as well as changes the production of cytokines; hence alcohol is considered an immunosuppressant factor<sup>[17,18]</sup>. Besides, alcohol affects the humoral and cellular system resulting in dysregulation of the immune system. That renders the patient susceptible to infectious pathogens, resulting in increased risk of opportunistic infections in the lungs; infections after surgery and liver diseases<sup>[17,19]</sup>.

Viral infections, in general, induce the production of IFN- $\alpha$ , that inhibits G1 phase of the cell cycle leading to temporary immunosuppression. Besides, HIV can suppress the immune system by destruction of the

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CD4<sup>+</sup> T lymphocytes<sup>[20]</sup>. Meanwhile, cytotoxic drugs impair the immune system by destroying all cells, not only cancer and proliferating cells, thus causing nonspecific immunosuppression<sup>[20,21]</sup>.

Opportunistic parasitosis may not cause severe pathological disorders in immunocompetent individuals, because of their normally functioning immune system<sup>[22]</sup>. In contrast, in immunocompromised patients, impairment of host immune system alters cellular and humoral responses and hinders T and B lymphocytes from acting efficiently against the infection<sup>[23]</sup>. Hence, immunocompromised patients are a vulnerable group to microbial infections especially opportunistic parasites<sup>[24,25]</sup>. Immunosuppression establishes favorable conditions for opportunistic parasites to flourish against the host system causing clinical diseases<sup>[26]</sup>. Impaired host immune systems are observed in different disease conditions such as malignancy, acquired immunodeficiency diseases (AIDS), organ transplants, corticosteroid chemotherapy, autoimmune and metabolic diseases, irradiation, malnutrition, environmental conditions, as well as in the elderly, and young children<sup>[22]</sup>.

In developing countries, intestinal parasitosis represents a major public health problem due to the low standard of personal hygiene and inadequate sanitation. Opportunistic parasites play an important role in eliciting diseases especially among immunosuppressed patients and children<sup>[27]</sup>. The WHO reported that about three billion people are globally infected with intestinal parasites because all opportunistic parasites inhabit the gastrointestinal tract except T. gondii and *Microsporidium* spp. on some occasions<sup>[28,29]</sup>. A major cause of morbidity and mortality among severely immunocompromised patients is partially attributed to opportunistic intestinal infections due to watery diarrhea and/or disseminated pathological manifestations<sup>[30]</sup>. Detection rates of enteric protozoa among immunocompromised patients ranged from 26.5% to 100% in different governorates of Egypt<sup>[31,32]</sup>.

The aim of the present systematic review is to determine the extent of opportunistic parasitosis detected among immunocompromised patients in Egypt throughout the last decade.

# Detection rates of opportunistic parasites in Egyptian immunocompromised hosts

Egypt is in the north eastern part of Africa and in 2020 it was recorded to have the highest population density of ~ 104 million inhabitants<sup>[33]</sup>. It is comprised of 27 Governorates. and over 90% of the population live in 10% of the whole area along the River Nile and Nile Delta in the northern part of the country.

Table (1) shows the prevalence of opportunistic parasites in Egyptian Governorates. In Cairo, a study conducted in 2010 reported a high rate of opportunistic

parasites among immunocompromised patients as compared to healthy controls (30% and 10%, respectively). The highest rate was detected among patients suffering from malignancy (18%), followed by equal rates (6%) among chronic liver failure (CLF) and diabetes mellitus (DM)<sup>[34]</sup>. Four years later, El-Mahallawy *et al.*<sup>[35]</sup> revealed that the overall parasitic infection was 50.6% among children with cancer compared to diarrheic immunocompetent cases (41%) attending the National Cancer Institute (NCI), Cairo University. Another study conducted by Wassef et al.<sup>[24]</sup> recorded a higher rate of opportunistic parasites in cancer patients (57%) than in control group (43%). Higher rates were observed among patients suffering from solid tumor as compared with those having hematological malignancies (63% vs 46%, respectively). In addition, those under radiotherapy showed higher infection rate than those under chemotherapy (68% vs 52%, respectively). The investigators attributed their results to reduction in local and cell-mediated responses in immunosuppressed patients<sup>[24]</sup>.

In Alexandria Governorate, Hassanein *et al.*<sup>[32,36]</sup> conducted two studies among patients suffering from inflammatory bowel disease (IBD) and acute lymphocytic leukemia (ALL) in children. Both studies detected the highest rates of opportunistic parasitosis (100% and 90.6%, respectively) as compared to their controls (72.5% and 58.1%, respectively), and the investigators attributed their results to immunosuppression, susceptibility to infection with opportunistic pathogens and diagnosis using a combination of different techniques<sup>[32,36]</sup>.

In addition, two other studies were conducted among immunocompromised patients. The first in 2019 in which Shehata *et al.*<sup>[37]</sup> reported that the prevalence rate of intestinal parasitosis among hemodialysis (HD) patients was significantly high as compared to apparently healthy individuals (52.5% vs. 12.0%, respectively) and no helminths infection was found. In the second more recent report, Elsayad *et al.*<sup>[38]</sup> revealed high parasitic infections in 79% of all samples obtained from patients with renal disorders; 40% in patients undergoing HD and 39% in those with chronic renal diseases (CRD), as compared to healthy control (10%).

In Sohag Governorate, two studies were conducted. In the first study, a high rate was detected among diabetic patients (25%). Type I-DM showed a higher rate of infection as compared to type II-DM (52% and 16% respectively)<sup>[39]</sup>. The second study was conducted in children and revealed a high rate of intestinal parasitosis in those on chemotherapy as compared to the control group (94% vs 35%, respectively)<sup>[40]</sup>.

In Minya, a study conducted among immunocompromised children showed high prevalence of opportunistic parasites as compared

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Governorate	Immunocompromised hosts	No.#	%	Reference		
	Malignancy	150	57.0			
	Control	50	43.0			
	Solid tumor	100	63.0	[24]		
	Hematological malignancies	50	46.0	[24]		
	Solid tumor under radiotherapy	50	68.0			
	Solid tumor under chemotherapy	50	52.0			
Cairo	Immunocompromised	100	30.0			
	Malignancy	40	18.0			
	CLF	30	6.0	[34]		
	DM	30	6.0			
	Control	20	10.0			
	Malignancy and diarrhea	89	50.6	[2]]		
	Diarrheic immunocompetent	100	41.0	[35]		
	IBD	40	100.0	[22]		
	Control	40	72.5	[32]		
	ALL	117	90.6	[0,6]		
	Control	117	58.1	[36]		
Alevandria	HD	120	52.5	[07]		
Alexanuria	Control	100	12.0	[37]		
	Patients with renal disorders	100	79.0			
	HD	50	40.0	[20]		
	CRD	50	39.0	[38]		
	Control	50	10.0			
	DM	100	25.0			
	Type-I DM	25	52.0	[0.0]		
	Type-II DM	75	16.0	[39]		
Sohag	Control	100	10.0			
	Children on chemotherapy	100	94.0			
	Control	100	35.0	[40]		
	Immunosuppressed	200	94.0	5445		
Minya	Immunocompetent	250	60.0	[41]		
Menoufia	Liver transplantation	50	16.0	[42]		

**#:** Total number examined; **ALL:** Acute lymphocytic leukemia; **CRD:** Chronic renal disease; **DM:** Diabetes mellitus; **HD:** Hemodialysis; IBD: Inflammatory bowel disease.

with immunocompetent children (94% vs 60%, respectively)<sup>[41]</sup>. In Menoufia Governorate, Saad *et al.*<sup>[42]</sup> reported opportunistic parasitic infections (16%) among patients suffering from hepatic diseases. One year later, a study conducted in Dakahlia Governorate showed a high rate of intestinal protozoa (85.5%) that included *G. lamblia* (36.6%), *C. parvum* (30.3%), and *E. histolytica/E. dispar* (27.6%) among hematological malignancy patients with diarrhea<sup>[43]</sup>.

## Commonly reported opportunistic parasites *Blastocystis* spp.

*Blastocystis* spp. are anerobic protozoa inhabiting the gastrointestinal tract (GIT) in four different distinct forms: cyst, ameboid, granular and vacuolar forms. Blastocystosis can be acquired by fecal-oral route, although the role of cyst forms is unknown<sup>[44]</sup>. The main symptoms include acute, chronic, and intermittent gastroenteritis, abdominal pain, abdominal distension, or constipation. A study reported that *Blastocystis* spp. are associated with colonic inflammation<sup>[45]</sup>, and another two studies conveyed that they are not pathogenic<sup>[46,47]</sup>. *Blastocystis* spp. can be detected in wet mount preparations of fresh stools and cultured stool samples in Jones' media<sup>[48]</sup>. Iodoquinol and metronidazole are the most common drugs prescribed for treatment<sup>[49]</sup>.

Table (2) shows the reported prevalence of *Blastocystis* spp. in some Egyptian Governorates. In Cairo; El-Shazly *et al.*<sup>[50]</sup> recorded an infection of 14% among chronic liver diseases (CLD) and that was attributed to possible poor environmental hygiene. A higher rate (28.5%) was reported among neoplastic patients in 2016<sup>[24]</sup>. Similarly, Ismail and Fadl<sup>[51]</sup> in 2019 diagnosed blastocystosis in 30% of patients with renal transplantation attending in Nephrology Unit of Kasr Al-Aini, Faculty of Medicine, Cairo University.

In Alexandria, Hassanein *et al.*<sup>[32]</sup> reported that blastocystosis was significantly high among patients with IBD as compared to the control (65% vs 17.5%, respectively). More or less similar infection rates were detected in 54.5% of immunocompromised and 67.4%

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<b>Table 2.</b> Prevalence of <i>Blasocystis</i> spp. among immunocompromised patients in five Egyptian Governorates.							
Governorate	Immunocompromised hosts	No.#	%	Reference			
Cairo	Neoplastic patients CLD Renal transplantation	150 50 50	28.5 14.0 30.0	[24] [50] [51]			
	IBD Control	40 40	65,0 17.5	[32]			
Alexandria	HD Control	120 100	24,2 13.0	[37]			
	ALL Control	55 46	54,5 67.4	[52]			
Minya	Immunosuppressed	200	12.1	[41]			
Menoufia	Liver transplantation	50	12.0	[42]			
Dakahlia	Hematological malignancy Malignancy with radiotherapy Malignancy with chemotherapy	145	21.4 38.5 18.5	[43]			

**#:** Total number examined; **ALL:** Acute lymphocytic leukemia; **CLD:** Chronic liver disease; **HD:** Hemodialysis; **IBD:** Inflammatory bowel disease.

of immunocompetent ones<sup>[52]</sup> and that was attributed to pharmacologic effect of cytotoxic drugs on the parasite among the immunocompromised<sup>[53]</sup>. Patients undergoing HD showed higher infection rates as compared to their controls (24.2% vs 13%, respectively)<sup>[37]</sup>.

In Minya Governorate, the parasite was detected in 12.1% of immunocompromised children<sup>[41]</sup>, which conformed with the recorded prevalence among liver transplantation patients in Menoufia Governorate<sup>[42]</sup>. Higher rates were detected in Dakahlia Governorate among patients who had hematological malignancy (21.4%), while those treated with radiotherapy had higher infection rate than those receiving chemotherapy (38.1% vs 18.5%, respectively)<sup>[43]</sup>. The recorded high prevalence may be due to restriction of the study to diarrheic cases besides other contributing factors such as environmental, socio-economic factors, residence, water source, and food supply.

#### Cryptosporidium spp.

Cryptosporidium spp. are world wild intracellular zoonotic protozoa and are recognized globally as a major cause of chronic diarrhea in immunocompromised patients resulting in significant morbidity and mortality<sup>[54-56]</sup>. Sporulated oocysts containing four sporozoites are infective on excretion and therefore transmissible by the fecal-oral route. Complicated AIDS patients with neoplasm and acute leukemia, as well as dairy or cattle farm workers, children in day care and owners of infected dogs or cats are at greatest risk<sup>[57]</sup>. Cryptosporidiosis is self-limited in immunocompetent individuals meanwhile it may cause severe acute diarrhea in immunocompromised patients, weight loss, nausea, and vomiting<sup>[54]</sup> leading to malnutrition and cognitive function impairment as well as growth retardation in infants<sup>[58]</sup>. Cryptosporidiosis is diagnosed using acid fast stain (AF), rapid immunochromatographic test, immunofluorescent microscopy, PCR-restriction fragment length polymorphism, multiplex allele-specific-PCR, and quantitative real-time PCR<sup>[59,60]</sup>.

In Cairo (Table 3), cryptosporidiosis was detected in 23.5% among neoplastic patients<sup>[24]</sup> and a low rate of 7% was observed among immunocompromised ones<sup>[34]</sup>. Patients with renal transplantation attending the nephrology unit of Kasr Al-Aini showed 10% prevalence, and the same was found among children suffering from CLD<sup>[50,51]</sup>. Another study conducted on liver cirrhosis patients revealed only 3.3% C. parvum infection by n-PCR and no infection was detected among the control group<sup>[61]</sup>. Recently, Amin *et al.*<sup>[62]</sup> detected cryptosporidiosis microscopically among elderly individuals attending outpatient clinics of Internal Medicine Hospital, Cairo University using AF, immunochromatographic test (ICT), ELISA and nested-PCR (3.7%, 6.3%, 6.7% and 3.7%, respectively), and genotype 1 and 2 were reported after using fragment length polymorphism on n-PCR assays.

In Alexandria (Table 3), the high rate of cryptosporidiosis was detected among patients suffering from IBD as compared to control (77.5% vs 20%, respectively)<sup>[32]</sup>. Meanwhile, lower rates were detected among children having ALL (42.7%)[36], followed by 32.5% vs 11% among HD patients and their counterparts respectively with statistically significant differences<sup>[37]</sup>.

In Dakahlia Governorate (Table 3), infection rates were 36%, 32%, 30% and 22% among patients suffering from liver cirrhosis with ascites, hepatocellular carcinoma, CLD and liver cirrhosis without ascites respectively<sup>[63]</sup>. A similar rate was observed among patients with hematological malignancy attending the Oncology and Radiotherapy Department, Mansoura University Hospital (30.3%)<sup>[43]</sup>. Liver transplant recipients showed also 20% infection in Menoufia Governorate<sup>[42]</sup>.

Cryptosporidiosis showed a high infection rate among immunocompromised children as compared to immunocompetent children in Minya Governorate (60.2% vs 42.2% respectively)<sup>[41]</sup>. This report was followed by a study by ElNadi *et al.*<sup>[39]</sup> in Sohag Governorate among diabetic patients (5%). Meanwhile, children on chemotherapy in Sohag Governorate<sup>[40]</sup> showed a high rate of 45%. Later, Mohamed *et al.*<sup>[60]</sup> detected *Cryptosporidium* oocysts in 45% of diarrheic immunocompromised patients attending the outpatient clinics of pediatric, oncology and internal medicine departments in Sohag University Hospitals. The recorded prevalence rate was ascribed to continued poor personal hygiene, inadequate supply of drinking water, low standard environmental conditions and inadequate waste disposal system<sup>[60]</sup> (Table 3).

#### Cyclospora cayetanensis

*Cyclospora cayetanensis* is an emerging coccidian parasite. The infective disporocystic disporozoic oocyst is unsporulated when passed and therefore is not transmitted by the feco-oral route. Sporulated stages are transmitted by contaminated water and food<sup>[64]</sup>. The overall number of cases reported in US increased from 2017 to 2019 to attain 2,408 cases possibly due to updating of diagnostic testing procedures, e.g., multiplex molecular test<sup>[65]</sup>. The infection may be asymptomatic or associated with self-limited or severe diarrhea<sup>[66]</sup>. Immunocompromised individuals suffer from prolonged diarrhea that lasts up to several months with remission and relapse. Although, the extra-intestinal infection is rare, complications such as Guillain-Barré and Reiter's syndromes were reported<sup>[67]</sup>. For diagnosis, flow cytometry is more sensitive than AF stain<sup>[68]</sup>. Renocal-sucrose gradient sedimentation proved to be effective in processing oocysts concentration and purification for flow cytometry<sup>[69]</sup>. Interestingly, PCR is increasingly used in research and outbreak investigations<sup>[70,71]</sup>. Trimethoprim-sulfamethoxazole (TMP-SMZ) is the drug of choice used as prophylaxis to prevent recurrent episodes among infected immunocompromised patients<sup>[72,73]</sup>.

Table (3) shows low rates of *Cyclospora* oocysts recorded as 2.5% and 3% among neoplastic and immunocompromised patients, respectively in Cairo<sup>[24,34]</sup>, and 10% among patients with renal transplantation attending the nephrology unit of Kasr Al-Aini, Faculty of Medicine, Cairo University<sup>[51]</sup>. That was in line with a study done among patients suffering from IBD in Alexandria (10%)<sup>[32]</sup>. The highest rate of *C. cayetanensis* was detected among children suffering from ALL in Alexandria (22.2%) <sup>[36]</sup>. In Minya Governorate 7.8% was recorded among immunocompromised children<sup>[41]</sup>, and 6% were positive among liver transplant recipients in Menoufia Governorate<sup>[42]</sup>.

#### CystoIsospora belli

Previously known as *Isospora belli*, it is known to cause uncommon diarrheal illness termed cystoisoporiasis. It is an opportunistic protozoan in immunosuppressed human hosts<sup>[74,75]</sup>. After *in vitro* 

Covernerate	Immuno compromized hosts	No #	Pre	Doforonco			
Governorate	minunocompromised nosts	NO."	Cryptosporidium spp.	C. cayetanensis	C. belli	Reference	
	Neoplastic patients	150	23.5	2.5	0.5	[24]	
	Immunocompromised	100	7.0	3.0		[34]	
	CLD	50	10.0			[50]	
	Renal transplantation	50	10.0	10.0		[51]	
Cairo	Liver cirrhosis	60	3.3			[61]	
			3.7 (AF)*				
	Fldorly individuals	270	6.3 (ICT)*			[62]	
		270	6.7 (ELISA)*			[02]	
			3.7 (nPCR)*				
	IBD	40	77.5	10.0		[32]	
Alexandria	ALL	117	42.7	22.2	1.7	[36]	
	HD	120	32.5			[37]	
	Diabetic patients	100	5.00			[39]	
Sohag	Children on chemotherapy	100	45.0			[40]	
	Diarrheic immunocompromised	40	45.0			[60]	
Minya	Immunosuppressed	200	60.2	7.8	9.7	[41]	
Menoufia	Liver transplantation	50	20.0	6.0		[42]	
	Hematological malignancy	145	30.3			[43]	
	Liver cirrhosis with ascites	50	36.0				
Dakahlia	Hepatocellular carcinoma	50	32.0			[(2]	
	CLD	150	30.0			႞ၒၣ႞	
	Liver cirrhosis without ascites	50	22.0				

Table 3. Prevalence of intestinal coccidia among immunocompromised patients in six Egyptian Governorates.

**#:** Total number examined; <sup>\*</sup>: Methods used for diagnosis; **ALL:** Acute lymphocytic leukemia; **CLD:** Chronic liver disease; **CRL:** Chronic renal failure; **DM:** Diabetes mellitus; **HD:** Hemodialysis; **IBD:** Inflammatory bowel disease.

sporulation, *C. belli* disporocystic tetrasporozoic oocysts are transmitted through contaminated food or water, so it is not a zoonosis<sup>[76]</sup>. It may cause mild diarrhea, abdominal discomfort, and low-grade fever in immunocompetent individuals. In contrast infected immunocompromised patients have extreme diarrhea. anorexia, weight loss, abdominal pain, cramps, loss of appetite, nausea, vomiting, and fever, that can last from weeks to months<sup>[77]</sup>. Molecular tools are more sensitive than microscopic examination for oocyst detection. The extended-range PCR approach, that offers a promising test for diagnosis of parasitic diseases that elude diagnosis using conventional methods, may be applied<sup>[78]</sup>. For this infection, TMP-SMZ combination are the drugs of choice and are better than antibiotics for treating diarrhea. In patients with AIDS, a single TMP-SMZ double-strength tablet, 3 times a week, was commonly used for long-term suppression of *C. belli*<sup>[79]</sup>. Table (3) shows that *C. belli* was detected in a small sample (0.5%) among neoplastic patients in  $Cairo^{[24]}$ ; and higher rates were recorded among ALL children attending El-Shatby Hospital in Alexandria and immunocompromised children in Minya Governorate (1.7% and 9.7%, respectively)<sup>[36,41]</sup>.

#### Microsporidium spp.

*Microsporidium* spp. previously considered as obligate unicellular spore forming eukaryotic parasites was recently phylogenetically classified as fungus<sup>[80]</sup>. It was elicited as an opportunistic infection associated with diarrhea in immunocompromised patients, patients with neoplasms, transplant recipients, diabetics, and in elderly individuals<sup>[81-83]</sup>. Humans acquire infection through ingestion of contaminated food or water, direct contact with broken skin or eyes, trauma, sexual transmission and trans-placentally<sup>[84]</sup>. Microsporidiosis in immunocompromised patients is characterized by severe and chronic diarrhea with massive weight loss especially in HIV/AIDS patients, as well as nausea, vomiting, malabsorption, and dyspepsia<sup>[85-87]</sup>. In 2017, Kazemi *et al.*<sup>[88]</sup> reported that multiplex nested PCR targeting internal transcribed spacer (ITS), small subunits (SSU) and large subunits (LSU) of ribosomal DNA (rDNA) identified intestinal *Microsporidium* with high sensitivity as compared to traditional techniques such as modified trichrome staining. Albendazole is effective against *Encephalitozoon* (*Enc.*) spp., meanwhile Fumagilin is more broadly effective against *Enc.* spp. and *Enterocytozoon* (*Ent.*) *bieneusi*<sup>[89]</sup>. In HIV patients, microsporidiosis is treated by anti-retroviral therapy through restoration of immune competence<sup>[90]</sup>.

The highest records of microsporidiosis were recorded in Alexandria Governorate. Table (4) shows four studies conducted in Alexandria among patients suffering from IBD, non-HIV immunocompromised patients, children suffering from ALL and HD patients (90%, 77.3%, 60.7%, and 11.7%, respectively)<sup>[32,36,37,91]</sup>. In contrast, lower rates were reported in Cairo (9.5%) among neoplastic patients<sup>[24]</sup>, and 2% each among immunocompromised patients and patients suffering from CLD<sup>[34,50]</sup>. In Cairo, stool samples were collected from cancer patients suffering from leukemia and lung, liver, breast, and colon cancer. The samples were examined by different stains (modified trichrome blue, acridine orange, and calcofluor white), together with regular PCR. The latter showed the highest sensitivity rate and diagnosed *Ent. intestinalis* in 17% of the sample examined<sup>[92]</sup>. In 2016, approximately 5% was recorded among hematological malignancy patients undergoing radiotherapy and chemotherapy in Dakahlia Governorate<sup>[43]</sup> and 3% among diabetic patients in Sohag Governorate<sup>[39]</sup>. In the Faculty of Medicine, Ain Shams University, spores were microscopically detected in 13.9% of all individuals examined and only one case was missed by Nested and RFLP-PCR<sup>[93]</sup>. The infection rate was higher among immunocompromised as compared to immunocompetent (14.5% and 13.3%, respectively) cases. Additionally, a detailed genotyping study showed equal number of immunocompromised

Table 4. Prevalence of Microsporidia spp. among immunocompromised patients in four Egyptian Governorates.

Governorate	Immunocompromised hosts	No.#	%	Reference
	Neoplastic patients	150	9.5	[24]
	Malignancy, DM and CRF	100	2.0	[34]
	CLD	50	2.0	[50]
Cairo	Cancer patients	100	17.0	[92]
	All samples microscopically	323	13.9	
	Immunocompromised patients	173	14.5	[93]
	Immunocompetent	150	13.3	
	IBD	40	90	[32]
Alexan duia	ALL	117	60.7	[36]
Alexandria	HD	120	11.7	[37]
	Non-HIV immunocompromised patients	44	77.3	[91]
Sohag	Diabetic patients	100	3.0	[39]
Dakahlia	Hematological malignancy Malignancy with radiotherapy Malignancy with chemotherapy	145	5.0	[43]

#: Total number examined; ALL: Acute lymphocytic leukemia; CLD: Chronic liver disease; CRL: Chronic renal failure; DM: Diabetes mellitus; IBD: Inflammatory bowel disease; HD: Hemodialysis.

individuals, five cases, with each of *Ent. bieneusi, Enc.* spp., and mixed infection among 15 cases. Meanwhile, six immunocompetent individuals had *Ent. bieneusi*, 3 had *Enc.* spp. and 10 had mixed infection<sup>[93]</sup>.

#### Strongyloides stercoralis

This nematode worm causes strongyloidiasis and is endemic in 25% of tropical and subtropical regions. In the USA, the highest rates of infection were observed among residents of the southeastern states, immigrants, refugees, travelers, and military personnel<sup>[94]</sup>. Complications were associated with persistent infection, high worm burden and high mortality<sup>[95]</sup>. Its life cycle begins by skin penetration with infective filariform larvae, found in soil contaminated with human feces. This is followed by migration to the lungs and penetration into the alveolar air sacs, then ascent to the tracheobronchial tree and swallowing<sup>[96]</sup>. After that, they mature into adult worms that burrow into the mucosa of duodenum and jejunum. In the GIT lumen fertilized females produce eggs from which hatch noninfectious rhabditiform larvae that pass in the feces<sup>[97]</sup>. In autoinfection, rhabditiform larvae mature into the filariform form within the GIT and penetrate perianal skin or colonic mucosa leading to increased burden of infection<sup>[98]</sup>. Among immunocompromised patients, fatal hyperinfection with disseminated disease was attributed to autoinfection<sup>[99]</sup>. In a case report of angioimmunoblastic T-cell lymphoma, Abdelrahman *et al.*<sup>[100]</sup> documented, the association of immunosuppressive therapy and steroids as a primary cause of fatal strongyloidiasis hyper infection. Recorded prevalence in table (5) showed high rate of 13.6% strongyloidosis among immunosuppressed children in Minya Governorate<sup>[41]</sup>, as compared to 2.6% estimated by Hassanein *et al.*<sup>[36]</sup> among leukemic children in Alexandria city, and no infection was found among the control groups.

#### Toxoplasma gondii

This obligate intracellular parasite affects about onethird of the world population<sup>[101]</sup>. It can be transmitted by ingestion of tissue cysts in meat or ingestion of mature disporocystic tetrasporozoic oocysts in contaminated fruits, vegetables and drinking water; besides congenital transmission, organ transplantation, blood transfusion, and accidental inoculation in the laboratory<sup>[102]</sup>. Retinochoroiditis is the most frequent clinical manifestation of congenital and acquired *Toxoplasma* infections<sup>[103,104]</sup>. Chronic toxoplasmosis is associated with various autoimmune disorders, including rheumatoid arthritis, systemic sclerosis, inflammatory bowel syndrome and autoimmune thyroid disorders<sup>[105]</sup>; and causes impairment of the innate immune state that is responsible for proliferation in autoantibodies production<sup>[106]</sup>. Mori *et al.*<sup>[107]</sup> reported that loop-mediated isothermal amplification (LAMP) has increasingly appeared as an alternative molecular method to PCR for diagnosis of toxoplasmosis. Currently, recommended drugs act against *T. gondii* tachyzoites, and do not eradicate the bradyzoite. The most effective drug is pyrimethamine because it blocks dihvdrofolate reductase that is involved in parasite reproduction<sup>[108-110]</sup>.

Table (6) shows that a low rate of toxoplasmosis was detected among immunocompromised patients in Cairo Governorate (6%)<sup>[34]</sup>. In Dakahlia Governorate, *Toxoplasma* IgG-antibody showed higher rate of infection (92.6%) as compared to *Toxoplasma* IgM (13.6%) among patients suffering from late stage of liver cirrhosis. Besides, it was higher than in those with chronic non cirrhotic HCV (76.9% and 12.8%, respectively) as compared to the controls (15% and 7.5%, respectively)<sup>[111]</sup>. In Menoufia Governorate, *Toxoplsama* IgG-antibody showed higher rate of infection as compared to *Toxoplasma* IgM-antibody

Table 5. Preva	alence of S ster	<i>coralis</i> in two	Egyntian (	Governorates
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Governorate	Immunocompromised hosts	No.#	%	Reference
Alexandria	ALL children Immunocompetent	117 117	2.6 0.0	[36]
Minya	Immunosuppressed children Immunocompetent	200 250	13.6 0.0	[41]

#: Total number examined; ALL: Acute lymphocytic leukemia.

Tal	οle	e 6	. Preva	lence of	Τ.	gondii	among	immunocompron	nised	l patients i	n four	Egyptian	i Go	vernorates
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Governorate	Immunocompromised hosts	No.#	Toxoplasmosis%*	IgG%	IgM%	Reference			
Cairo	Malignancy, DM, CRF	100	6.0			[34]			
Alexandria	Liver cirrhosis Chronic HCV non cirrhotic	81 39		92.6 76.9	13.6 12.8	[111]			
Menoufia	Liver transplant recipients	50		28.0	18.0	[42]			
	DM-type I DM-type II		86.37 66.69			[113]			
Qalyoubia	Rheumatic arthritis patients	25	54.0			[112]			
#: Total number	: Total number examined; *: Overall prevalence of toxoplasmosis.								

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among liver transplant recipients (28% and 18%, respectively)<sup>[42]</sup>.

In Qalyoubia Governorate, the overall prevalence of toxoplasmosis showed a high infection rate in rheumatic arthritis patients as compared to the controls (54% and 32.0%, respectively)<sup>[112]</sup>, while in Menoufia Governorate, the rate was higher among DM-type I patients than those of DM-type II (86.37% and 66.69%, respectively)<sup>[113]</sup>.

In conclusion, the highest rates of opportunistic infections were recorded from each of Alexandria, Sohag and Minya Governorates. In Alexandria City, IBD patients presented high rates of *Blastocystis* spp. and *Cryptosporidium* spp., meanwhile children with ALL showed highest rates of C. cavetanensis. Both C. belli and S. stercoralis were largely detected in immunocompromised patients in Minya Governorate. *Microsporidium* spp. was highly demonstrated among non-HIV immunocompromised patients in Alexandria City. Finally, toxoplasmosis was mostly detected among DM-I in Menoufia Governorate, and IgG and IgM antibodies were highly detected among patients with liver cirrhosis in Dakahlia Governorate. Reasons for variations in patterns in parasite distribution are unclear but reflects that parasite colonization in the intestinal tract might be affected by immunosuppression.

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#### REFERENCES

- Geha RS, Notarangelo L, Casanova JL, Chapel H, Fischer A, Hammarstrom L *et al.* Primary immunodeficiency diseases: an update the International Union of Immunological Societies Primary Immunodeficiency Diseases Classification Committee. J Allergy Clin Immunol 2007; 120(4):776–794.
- 2. Ochs HD, Smith CIE, Puck JM. Primary immunodeficiency diseases. A molecular and genetic approach, 2<sup>nd</sup> edition, Oxford University Press, New York, 2006.
- Stiehm ER, Ochs HD, Winkelstein JA. Immunodeficiency disorders in infants and children, 5<sup>th</sup> edition, Elsevier Saunders, Philadelphia, 2004.
- 4. Notarangelo LD, Fischer A, Geha RS, Casanova JL, Chapel H, Conley ME *et al.* Primary immunodeficiencies: 2009 update. J Allergy Clin Immunol 2009; 124(6):1161-1178.
- 5. Notarangelo LD. Primary immunodeficiencies. J Allergy Clin Immunol 2010; 125(2):S182-194.
- 6. Bonilla FA, Bernstein IL, Khan DA, Ballas ZK, Chinen J, Frank MM *et al.* Practice parameter for the diagnosis

and management of primary immunodeficiency. Ann Allergy Asthma Immunol 2005, 94(5):S1-63.

- Ehigiator HN, McNair N, Mead JR. *Cryptosporidium* parvum: the contribution of Th1-inducing pathways to the resolution of infection in mice. Exp Parasitol 2007; 115:107–113.
- Ehigiator HN, Romagnoli P, Borgelt K, Fernandez M, McNair N, Secor WE *et al*. Mucosal cytokine and antigen-specific responses to *Cryptosporidium parvum* in IL-12p40 KO mice. Parasite Immunol 2005; 27(1-2):17–28.
- Tessema TS, Schwamb B, Lochner M, Forster I, Jakobi V, Petry F. Dynamics of gut mucosal and systemic Th1/ Th2 cytokine responses in interferon-gamma and interleukin-12p40 knock out mice during primary and challenge *Cryptosporidium parvum* infection. Immunobiology 2009; 214(6):454–466.
- 10. Trinchieri G. Interleukin-12 and the regulation of innate resistance and adaptive immunity. Nat Rev Immunol 2003; 3(2):133–146.
- 11. Ponnuraj EM, Hayward AR. Intact intestinal mRNAs and intestinal epithelial cell esterase, but not *Cryptosporidium parvum*, reach mesenteric lymph nodes of infected mice. J Immunol 2001;167(9):5321–5328.
- Shah UV, Purohit BC, Chandralekha D, Mapara MH. Co-infection with *Cryptosporidium, Isospora* and *S. stercoralis* in a patient with AIDS: A case report. Ind J Med Microbiol 2005; 21(2):137–138.
- Assefa S, Erko B, Medhin G, Assefa Z, Shimelis T. Intestinal parasitic infections in relation to HIV/AIDS status, diarrhea and CD4 T-cell count. BMC Infect Dis 2009; 9:155.
- 14. Teklemariam Z, Abate D, Mitiku H, Dessie Y. Prevalence of intestinal parasitic infection among HIV positive persons who are naive and on antiretroviral treatment in Hiwot Fana Sepecialized University Hospital, Eastern Ethiopia. ISRN AIDS 2013; 2013(2): 324329.
- 15. Dey AB, Chatterjee P, Das PC. Immune Status in the Elderly. Medicine Update. 2012; 22:721-724.
- 16. Keusch GT. The history of nutrition: malnutrition, infection, and immunity. J Nutr 2003; 133(1): 336S-340S.
- 17. Molina PE, Happel KI, Zhang P, Kolls JK, Nelson S. Focus on: Alcohol and the immune system. Alcohol Res Health 2010; 33(1-2):97-108.
- Szabo G. Alcohol's contribution to compromised immunity. Alcohol Health Res World 1997; 21(1):30-41.
- 19. Greenfield TK, Ye Y, Bond J, Kerr WC, Nayak MB, Kaskutas LA *et al.* Risks of alcohol use disorders related to drinking patterns in the U.S. general population. J Stud Alcohol Drugs 2014; 75(2):319-327.
- 20. Playfair JHL, Chain BM. Immunology at a Glance. Blackwell Publishing, USA, 7<sup>th</sup> edition; 2001; 92 pages
- 21. Gorczynski R, Stanley J. Clinical Immunology. Landes Bioscience, Texas. 1999.
- 22. Zope A, Pai A, De A, Baveja SM. Opportunistic intestinal parasites in HIV infected individuals and its correlation with the CD4 counts. RRJMHS 2014; 3(3):55-59.

- 23. Romagnani S. Biology of human TH1 and TH2 cells. J Clin Immunol 1995; 15(3):121-129.
- 24. Wassef R, Rizk E, Abdel-Malek R. Prevalence of enteric opportunistic parasites in immunocompromised cancer patients. OFID 2016; 3(1):582.
- 25. Mikati T, Griffin K, Lane D, Matasar M, Shah MK. International travel patterns and travel risks for stem cell transplant recipients. J Travel Med 2015; 22(1):39–47.
- 26. Tekle B. Guidelines for management of opportunistic infections and anti-retroviral treatment in adolescents and adults in Ethiopia. Addis Ababa, Ethiopia. Federal MOH 2008: 1-109.
- 27. Ferreira MS. Infections by protozoa in immunocompromised hosts. Mem Inst Oswaldo Cruz 200; 95(1):159-162.
- 28. WHO Department of control of neglected tropical diseases. Sustaining the drive to overcome the global impact of neglected tropical diseases: Second WHO report on neglected diseases. WHO 2013:140.
- 29. Botero JH, Castaño A, Montoyal MN, Ocampo NE, Hurtado MI, Lopera MM. A preliminary study of the prevalence of intestinal parasites in immunocompromised patients with and without gastrointestinal manifestations. Rev Inst Med trop S Paulo 2003; 45(4):197-200.
- Jose RJ, Brown JS. Opportunistic bacterial, viral and fungal infections of the lung. Medicine (Abingdon) 2016; 44(6):378–383.
- 31. Monib MEM, Hassan AA, Attia RAH, Khalifa MM. Prevalence of intestinal parasites among children attending Assiut University Children's Hospital, Assiut, Egypt. J Adv Parasitol 2016; 3(4):125-131.
- Hassanein F, El-Masry SA, Hassan A. Gastrointestinal disorders and intestinal parasitic infections. Lambert Academic Publishing 2012: Ps146; ISBN 978-3-659-21248-2.
- 33. Egypt Demographics. Available form: https:// www.worldometers.info/world-population/egyptpopulation/. [Accessed on: May 2021]
- Baiomy AM, Mohamed KA, Ghannam MA, Shahat SA, Al-Saadawy AS. Opportunistic parasitic infections among immunocompromised Egyptian patients. J Egypt Soc Parasitol 2010; 40(3):797-808.
- 35. El-Mahallawy H, El Basha NR, Zaki MM, El-Arousy M, Elswaifi SF, Abo-hashem EM. A comparative study on enteric parasitic infections in immunocompetent and immunosuppressed children in Egypt. Comp Clin Pathol 2014; 23(5);1509–1514.
- Hassanein F, Shaibat-El-Hamd Z, El-Masry SA. Acute lymphocytic leukemia and ecto- and endo-parasites. Lambert Academic Publishing 2013: Ps145. ISBN 978-3-659-21270-3.
- Shehata AI, Hassanein F, Abdul-Ghani R. Opportunistic parasitoses among Egyptian hemodialysis patients in relation to CD4<sup>+</sup> T cell counts: A comparative study. BMC Infect Dis 2019; 19(480):1-9.
- Elsayad MHM, Maharem DA, Ali FAS, Abd El-latif NF. Detection of intestinal protozoan infections with stress on *Blastocystis*, *Microsporidia* in Egyptian chronic

kidney disease patients. J Egypt Soc Parasitol 2020; 50(3): 513-521.

- ElNadi NA, Hassanien HA, Ahmed AM, Abd Allah AK. Intestinal parasites in diabetic patients in Sohag University Hospitals, Egypt. J Egypt Soc Parasitol 2015; 45(2):443-449.
- El-Hady HA, Ahmed NS, Taha MAA, Abd El-Kareem NM, Bakheet RA. Intestinal parasites in children receiving chemotherapy. J Egypt Soc Parasitol 2017; 47(2):375-380.
- 41. Abdel-Hafeez EH, Ahmad AK, Ali BA, Moslam FA. Opportunistic parasites among immunosuppressed children in Minya district, Egypt. Korean J Parasitol 2012; 50(1):57-62.
- 42. Saad AE, Elkersh WM, Afifi AF, Hawash Y, Shendi SS. Coccidian parasites in liver-transplant recipients. Menoufia Med J 2015; 28(3):635-641.
- Abdel-Magied AA, El-Ghanam WA, El-Nemr HI, El-Henawy AA. Prevalence of intestinal parasites in cancer therapy recipients with concurrent diarrhea. IJTDH 2016; 15(1):1-7.
- 44. Stenzel DJ, Lee MG, Boreham PF. Morphological differences in *Blastocystis* cysts an indication of different species? Parasitol Res 1997; 83:452-457.
- 45. Carrascosa M, Martinez J, Perez-Castrillon JL. Hemorrhagic proctosigmoiditis and *Blastocystis hominis* infection. Ann Intern Med 1996; 124(2):278-279.
- 46. Shlim DR, Hoge CW, Rajah R, Rabold JG, Echeverria P. Is *Blastocystis hominis* a cause of diarrhea in travelers? A prospective controlled study in Nepal. Clinic Infect Dis 1995; 21(1):97-101.
- 47. Al-Tawil YS, Gilger MA, Gopalakrishna GS, Langston C, Bommer KE. Invasive *Blastocystis hominis* infection in a child. Arch Pediatr Adolesc Med 1994; 148(8): 882-885.
- 48. Jones WR. The experimental infection of rats with *Entameba histolytica*; with a method for evaluating the anti-amebic properties of new compounds. Annals of Trop Med and Parasitol 1946; 40:130-140.
- 49. Qadri SM, Al-Okaili GA, Al-Dayel F. Clinical significance of *Blastocystis hominis*. J Clinic Micro 1989; 27:2407-2409.
- El-Shazly LBE, El-Faramawy AAM, El-Sayed NM, Ismail KA, Fouad SM. Intestinal parasitic infection among Egyptian children with chronic liver diseases. J Parasit Dis 2015; 39(1):7–12.
- 51. Ismail MAM, Fadl HO. *Cyclospora* infection in renal transplant recipient. J Egypt Soc Parasitol 2019; 49(3):727-730.
- 52. Eassa SM, Ali HS, El Masry SA, Abd El-Fattah AH. *Blastocystis hominis* among immunocompromised and immunocompetent children in Alexandria, Egypt. Ann Clin Lab Res 2016; 4(2):1-6
- Gharavi MJ, Ashraf F, Vosough P, Rokni MB. Survey of intestinal parasitic infection in leukemic children and evaluation of their serum immunoglobulins. Iranian J Publ Health 2003; 32:19-21.
- Ryan U, Zahedi A, Paparini A. *Cryptosporidium* in humans and animals: A one health approach to prophylaxis. Parasite Immunol 2016; 38(9):535-547.
- 55. Zahedi A, Paparini A, Jian F, Robertson I, Ryan U. Public health significance of zoonotic *Cryptosporidium* species

in wildlife: critical insights into better drinking water management. Int J Parasitol Parasites Wildl 2016; 5(1): 88-109.

- Thompson RC, Ash A. Molecular epidemiology of *Giardia* and *Cryptosporidium* infections. Infect Genet Evol 2016; 40:315–323.
- Desai NT, Sarkar R, Kang G. Cryptosporidiosis: An underrecognized public health problem. Trop Parasitol 2012; 2(2):91-98.
- 58. Shrivastava AK, Kumar S, Smith WA, Sahu PS. Revisiting the global problem of cryptosporidiosis and recommendations. Trop Parasitol 2017; 7(1):8-17.
- 59. Khurana S, Chaudhary P. Laboratory diagnosis of cryptosporidiosis. Trop Parasitol 2018; 8(1):2–7.
- 60. Mohamed SR, El-Hady AH, Ahmed AM. Evaluation of immunochromatographic assay for diagnosis of cryptosporidiosis. J Egypt Soc Parasitol 2020; 50(3):477-482.
- 61. Shahat SA, El-Badry AA, El-Sheety AG, El Faramawy MS, Ismael NF, Abo-Mandil ME. Genotypic prevalence of *Cryptosporidium* in Egyptian patients with liver cirrhosis. AIMJ 2020; 1(2):225-231.
- 62. Amin NM, Raafat A, Morsy SM. Detection rate and genotyping of *Cryptosporidium* spp. and its relation to copro TNF- $\alpha$  in elderly Egyptians attending outpatient clinics of Cairo University Hospitals. PUJ 2021; 14(1):77-85.
- 63. Ibrahim MA, Abdel-Ghany AE, Abdel-Latef GK, Abdel-Aziz SA, Aboelhadid SM. Epidemiology and public health significance of *Cryptosporidium* isolated from cattle, buffaloes, and humans in Egypt. Parasitol Res 2016; 115(6):2439–2448.
- 64. Ortega YR, Sanchez R. Update on *Cyclospora cayetanensis*: A food-borne and waterborne parasite. Clin Microbiol Rev 2010; 23(1):218–234.
- 65. CDC. Domestically Acquired Cases of Cyclosporiasis: United States, May–August 2019. https://www.cdc.gov/ parasites/cyclosporiasis/outbreaks/2019/a-050119/ index.html. [Accessed on May 2021].
- Doller PC, Dietrich K, Filipp N, Brockmann S, Dreweck C, Vonthein *et al.* Cyclosporiasis outbreak in Germany associated with the consumption of salad. Emerg Infect Dis 2002; 8(9):992–994.
- 67. Zar FA, El-Bayoumi E, Yungbluth MM. Histologic proof of acalculous cholecystitis due to *Cyclospora cayetanensis*. Clin Infect Dis 2001; 33(12: E140–E141
- Dixon BR, Bussey JM, Parrington LJ, Parenteau M. Detection of *Cyclospora cayetanensis* oocysts in human fecal specimens by flow cytometry. J Clin Microbiol 2005; 43(5):2375–2379.
- Riner DK, Mullin AS, Lucas SY, Cross JH, Lindquist HAD. Enhanced concentration and isolation of *Cyclospora cayetanensis* oocysts from human fecal samples. J Microbiol Methods 2007; 71(1):75–77.
- Soldan OCP, Vásquez FV, Varas AG, Cordón GP, Soto JRV, Sánchez-Moreno M *et al.* Intestinal parasitism in Peruvian children and molecular characterization of *Cryptosporidium* species. Parasitol Res 2006; 98(6):576– 581.

- 71. Hussein EM. Molecular identification of *Cyclospora* spp. using multiplex PCR from diarrheic children compared to others conventional methods. J Egypt Soc Parasitol 2007; 37(2):585–598.
- 72. Verdier RI, Fitzgerald DW, Johnson WD Jr, Pape JW. Trimethoprim-sulfamethoxazole compared with ciprofloxacin for treatment and prophylaxis of *Isospora belli* and *Cyclospora cayetanensis* infection in HIVinfected patients: A randomized, controlled trial. Ann Intern Med 2000; 132(11):885–888.
- Pape JW, Verdier RI, Boncy M, Boncy J, Johnson WD Jr. *Cyclospora* infection in adults infected with HIV: Clinical manifestations, treatment, and prophylaxis. Ann Intern Med 1994; 121(9):654–657.
- 74. Minnaganti VR. Cystoisoporiasis. Medscape Updated: Apr 17, 2018. Available from: https://emedicine. medscape.com/article/219776-overview
- Gutierrez Y. In: Diagnostic Pathology of Parasitic Infections with Clinical Correlations (2<sup>nd</sup> edn). Oxford University Press, New York, 2000; pp: 769
- Neira PO, Barthel EM, Wilson GL, Muñoz NS. *Isospora belli* infection in HIV positive patients. Report of two cases and literature review. Rev Chil Infect 2010; 27(3):219-227.
- 77. Néstor VJ, Germán AO, Cecilia DR, Cristina E, Víctor CA, Elisabet PG, *et al.* Molecular characterization of Cysto*lsospora belli* and unizoite tissue cyst in patients with acquired immunodeficiency syndrome. Parasitology 2010; 138(3):279-286.
- Murphy SC, Hoogestraat DR, SenGupta DJ, Prentice J, Chakrapani A, Cookson BT. Consultations in molecular diagnostics, molecular diagnosis of cystoisosporiasis using extended-range PCR screening. JMD 2011; 13(3):359-362.
- 79. Malik S, Samantaray JC, Bagga A, Das A. Refractory isosporiasis. Indian J Pediatr 2005; 72(5):437-439.
- Hibbett DS, Binder M, Bischoff JF, Blackwell M, Cannon PF, Eriksson OE, *et al.* A higher-level phylogenetic classification of the Fungi. Mycol Res 2007;111(5):509-547.
- 81. Didier ES, Weiss LM. Microsporidiosis: Not just in AIDS patients. Curr Opin Infect Dis 2011; 24(5):490-495.
- 82. Antonios SN, Tolba OA, Othman AA, Saad MA. A preliminary study on the prevalence of parasitic infections in immunocompromised children. J Egypt Soc Parasitol 2010; 40(3):617–630.
- 83. Sak B, Brady D, Pelikanova M, Květoňová D, Rost M, Kostka M *et al.* Unapparent microsporidial infection among immunocompetent humans in the Czech Republic. J Clin Microbiol 2011; 49(3):1064–1070.
- Anane S, Attouchi H. Microsporidiosis: epidemiology, clinical data and therapy. Gastroenterol Clin Biol 2010; 34(8-9):450-464.
- Lallo MA, Hirschfeld MMP. Encephalitozoonosis in pharmacologically immunosuppressed mice. Exp Parasitol 2012; 131(3):339-343.
- Santín M, Fayer R. Microsporidiosis: *Enterocytozoon bieneusi* in domesticated and wild animals. Res Vet Sci 2011; 90(3):363-371.

- Endeshaw T, Kebede A, Verweij JJ, Wolday D, Zewide A, Tsige K, *et al.* Intestinal microsporidiosis in diarrheal patients infected with human immunodeficiency virus-1 in Addis Ababa, Ethiopia. Jpn J Infect Dis 2006; 59(5): 306–310.
- Kazemi E, Tavalla M, Maraghi S, Yad MJ, Latifi M. Frequency of microsporidial infection in immunocompromised patients with staining and molecular methods based on internal transcribed spacer region gene in two cities of Southwest Iran during 2013-2014. AJPRHC 2017; 9(1):7-16.
- 89. Didier ES. Microsporidiosis: an emerging and opportunistic infection in humans and animals. Acta Trop 2005; 94(1): 61–76.
- 90. Pozio E, Morales MA. The impact of HIV-protease inhibitors on opportunistic parasites. Trends Parasitol 2005; 21(2):58–63.
- 91. Abu-Akkada SS, El Kerdany EDH, Mady RF, Diab RG, Khedr GAE, Ashmawy KI *et al. Encephalitozoon cuniculi* infection among immunocompromised and immunocompetent humans in Egypt. IJP 2015; 10(4):561-570.
- 92. El Sobky MM, El Nahas NS. Detection and differentiation between Enterocytozoon bieneusi and Encephalitozoon intestinalis species in cancer patient's stools using PCR compared with different staining methods. PUJ 2012; 5(1):19-26.
- 93. Abd-Elbaki MH, Arafa MA, Abd-El Hameed DM, Habib KS, Abdel Rahman AA, Anwar MA. Intestinal microsporidiosis: prevalence and genetic study of Egyptian isolates. PUJ 2020; 13(1):35-45.
- 94. Posey DL, Blackburn BG, Weinberg M, Flagg EW, Ortega L, Wilson M, *et al.* High prevalence and presumptive treatment of schistosomiasis and strongyloidiasis among African refugees. Clin Infect Dis 2007; 45(10):1310-1315.
- 95. Segarra-Newnham M. Manifestations, diagnosis, and treatment of *Strongyloides stercoralis* infection. Ann Pharmacother 2007; 41(12):1992-2001.
- Schupf, N, Ortiz, M, Kapell, D, Rudelli RD. Prevalence of intestinal parasite infections among individuals with mental retardation in New York State. Ment Retard 1995; 33(2):84-90.
- 97. Lindo, JF, Robinson, R, Terry, SI, Vogel P, Gam AA, Neva FA *et al.* Age-prevalence and household clustering of *Strongyloides stercoralis* infection in Jamaica. Parasitology 1995; 110(1):97-102.
- Siddiqui AA, Genta RM, Berk SL. Life cycle of *Strongyloides* stercoralis. In: Guerrant RL, Walker DH, Weller PF, (Eds). Infectious Diseases: Principles, Practices and Pathogens. Churchill-Livingstone Elsevier, Philadelphia; 2006. p. 1274.
- 99. Keiser PB, Nutman TB. *Strongyloides stercoralis* in the immunocompromised population. Clin Microbiol Rev 2004; 17(1):208-211.
- 100. Abdelrahman, MZ, Mohammad, Z, Rahmah, N, Norsyahida A, Madihah B, Azlan H *et al.* Fatal septicemic

shock associated with *Strongyloides stercoralis* infection in a patient with angioimmunoblastic T-cell lymphoma: A case report and literature review. Parasitol Int 2012; 61(3):508-511.

- 101. Peng HJ, Chen XC, Lindsay DS. A review: competence, compromise, and concomitance-reaction of the host cell to *Toxoplasma* gondii infection and development. J Parasitol 2011; 94 (4):e620-e628.
- 102. Montoya JG, Liesenfeld O. Toxoplasmosis. Lancet 2004; 363(9425): e1965-e1976.
- 103. Delair E, Latkany P, Noble AG, Rabiah P, McLeod R, Br'ezin A. Clinical manifestations of ocular toxoplasmosis. Ocul Immunol Inflamm 2011; 19(2):91–102.
- 104. Olariu TR, Remington JS, McLeod R, Alam A, Montoya JG. Severe congenital toxoplasmosis in the United States: clinical and serologic findings in untreated infants. Pediatr Infect Dis J 2011; 30(12):1056–1061.
- 105. Carter CJ. Toxoplasmosis and polygenic disease susceptibility genes: extensive *Toxoplasma gondii* host/ pathogen interactome enrichment in nine psychiatric or neurological disorders. J Pathog 2013; 2013:965046.
- 106. Prandota J. *T. gondii* infection acquired during pregnancy and/or after birth may be responsible for development of both type 1 and 2 diabetes mellitus. J Diabetes Metabol 2013; 4(2):55.
- 107. Mori Y, Kanda H, Notomi T. Loop mediated isothermal amplification (LAMP): recent progress in research and development. J Infect Chemother 2013; 19(3):401–411.
- 108. Sobrin L, Kump LI, Foster CS. Intravitreal clindamycin for toxoplasmic retinochoroiditis. Retina 2007; 27(7):952-957.
- 109. Soheilian M, Ramezani A, Azimzadeh A, Sadoughi MM, Dehghan MH, Shahghadami R *et al.* Randomized trial of intravitreal clindamycin and dexamethasone versus pyrimethamine, sulfadiazine, and prednisolone in treatment of ocular toxoplasmosis. Ophthalmology 2011; 118(1):134-141.
- 110. Soheilian M, Sadoughi MM, Ghajarnia M, Dehghan MH, Yazdani S, Behboudi H *et al.* Prospective randomized trial of trimethoprim/sulfamethoxazole versus pyrimethamine and sulfadiazine in the treatment of ocular toxoplasmosis. Ophthalmology 2005; 112(11):1876-1882.
- 111. El-Nahas HA, El-Tantawy NL, Farag RE, Alsalem AMA. *Toxoplasma gondii* infection among chronic hepatitis C patients: A case control study. Asian Pac J Trop Med 2014; 7(8):589-593.
- 112. El-Sayed NM, Kishik SM, Fawzy RM. The current status of *Toxoplasma gondii* infection among Egyptian rheumatoid arthritis patients. Asian Pac J Trop Dis 2016; 6(10):797-801.
- 113. Beshay EVN, El-Refai SA, Helwa MA, Atia AF, Dawoud MM. *Toxoplasma gondii* as a possible causative pathogen of type-1 diabetes mellitus: Evidence from case-control and experimental studies. Exp Parasitol 2018; 188:93e101.