

Chemical and functional properties of free-gluten biscuit making from corn, quinoa and millet flours

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ABSTRACT

The present study was carried out to evaluate the chemical and functional properties of gluten-free biscuit formulation and their relation to final product quality making from different ratios of corn, quinoa and millet flours. The result showed that quinoa seeds contained acceptable range of saponin. Washing and soaking processes succeeded to reduce saponin content from 0.035% to 0.022%. Corn flour had higher percentage of moisture (6.86%) than quinoa (5.41%) and millet (4.87%). The highest percentage of fat was 9.72% in corn followed by millet 7.9% and quinoa 6.55%. Quinoa and millet are worthy of consideration as an important grain source of protein being 15.10 and 12.50% in dry matter, respectively, while it was 9.20% in corn. Millet and quinoa had high percentage of crude fibers (4.28% and 3.94%, respectively) as compared to corn (2.76%). Starch content as an important part in carbohydrate was 41.29% in corn, 46.97% in quinoa, and 43.85% in millet.

Millet generally contains significant amounts of essential amino acids particularly the sulphur containing amino acids methionine and cysteine (2.87 and 3.60, respectively) compared to quinoa and corn. Quinoa is a good source of minerals iron (4.47), calcium (82.78), magnesium (169.55) and potassium (1508.64 mg/100g). Vitamins soluble in fat (Vit. A and Vit. E) were found to be the highest ratio in corn followed by quinoa then millet.

Results also revealed that water holding capacity (WHC) was increased in (quinoa+corn) followed by millet. Corn with millet and quinoa increased wettability actions while, quinoa + millet recorded the lowest values. Also, increasing the level of corn flour increased sensory scores of biscuits for over all acceptability as seen in blended (25%Q +75% corn) followed by (75%Q +25%C) and it was 74.3±9.6 and 71.4±8.6 respectively. Whereas, control biscuit (100% corn) sample had the highest value in all parameters and over all acceptability was (87.2±9.8) compared to other tested samples. In conclusion, addition of corn flour by each ratio to quinoa or millet recorded good values and satisfied acceptable about blended the three samples with each other.

Keywords: Corn, Quinoa, Millet, Biscuit, Free Gluten, Composite Flours, Functional Properties.

INTRODUCTION

Corn is considered as one of the major cultivated crops in Egypt. Its production is increasing steadily. However, the majority of the crop production is directed for animal and poultry feeding, in spite of the shortage in the cereal-based food

stuffs. Therefore, it would be beneficial to introduce new manufactured corn products to the Egyptian food market such as Tortillas. In the last decade the volatility of corn prices, consequence of a continuous increase in biofuels production as well as oil price rise and speculation (Ajanovic, 2011),

has caused the increase in the production. The future of this crop is bright because it is environmentally more flexible than other cereals and shows better tolerance to diseases, drought, and pests than its parental species costs of nixtamalized corn flour (Darvey *et al.*, 2000).

Quinoa (*Chenopodium quinoa* Willd) has added popularity worldwide appreciations to the attractive nutritional profile. Starch is the major component of quinoa grain and makes up to 70% of the dry matter. The starch acting a vital role in functional properties of quinoa and associated food products (Zhu and Li, 2018). The flours obtained from quinoa seeds, can be used for elaborated bread or biscuits. Nowadays, in the shop, diverse products with a 20% content of quinoa are commercially available i.e., backed products, infant foods, and gluten free products (Pellegrini and Agostoni, 2015; Wang and Zhu, 2016). Furthermore, the gluten-free kind of quinoa seeds makes to this pseudocereal a valued dietary source of digestible protein for persons with gluten sensitivity and coeliac disease (Tang *et al.*, 2015). This wide range of use quinoa seeds due to its versatility as food component, representing an motivating field of research due to the high content of different macromolecules and phyto-chemicals content in their seeds (Gordillo-Bastidas *et al.*, 2016). This pseudocereal holds more biological value proteins and bioavailable essential amino acids, dietary fiber, unsaturated lipids, complex carbohydrates and other beneficial bioactive compounds such as polyphenolic compounds resulting in enormous helpful health properties to customers (Wu, 2015; Fischer *et al.*, 2017). These substances have already presented diverse *in vitro* biological potentials (Gawlik-Dziki *et al.*, 2013) and *in vivo* activities against various diseases and

metabolic conditions (Graf *et al.*, 2015; Gordillo-Bastidas *et al.*, 2016).

Millets are one of the cereals besides the other major cereal crops in Egypt such as wheat, rice, and maize. Millets are major food sources for millions of people, especially those who live in hot, dry areas of the world because of their ability to grow under hard weather conditions like restricted rainfall. (Adekunle *et al.*, 2012). It is the major source of energy, protein and still part of the major diet in most African countries because it has many nutritious, health benefits and medical functions, and its uses in food industry sector. (Amadou *et al.*, 2011). Millets are classified with sorghum, maize, and Coix (Job's tears) in the grass sub-family *Panicoideae* (Yang *et al.*, 2012). Millet is gluten-free, thus a premium option for people inmate from celiac diseases regularly irritated by the gluten content of wheat and other more common cereal grains. It is also beneficial for people who are suffering from atherosclerosis and diabetic heart disease (Gélinas *et al.*, 2008).

Composed Flour is types of flour from grains other than wheat, legumes, carrot and tubers can be a mixture of flours other than wheat flour. (Okpala and Okoli, 2011). Composite flours are recently manufactured not only to improve the desired functional properties of end product based on them but also to improve nutritional composition (Ubbor and Akobundu, 2009). Good nutritional value of cereals concerns with their proteins, carbohydrates and fiber contents and appreciable quantities of vitamins and minerals (Hill and Path, 1998).

Looking to supply a gluten-free product with improved acceptance and that promote a possible increase in the absorption of vitamins and minerals in individuals with celiac disease, gluten-free product (da Silva and Conti-Silva, 2018). Studies performed on teenagers and young

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adults have exposed that biscuits are a popular foodstuff consumed by a varied range of population due to their varied taste, long shelf life and comparatively low cost. Millet is gluten-free, therefore a best option for persons inmate from celiac diseases regularly irritated by the gluten contented of wheat and other more common cereal grains. It is also helpful for persons who are suffering from atherosclerosis and diabetic heart disease (Lubna and Vidhu, 2012).

In that sense, the aim of this research study was to evaluate the chemical and functional properties of gluten-free biscuit formulation and their relation to final product quality making from different ratios of corn, quinoa and millet flours.

MATERIALS AND METHODS

Materials

Corn (*Zea mays*) seeds were obtained from Cereals and Pulses Maize Department, Field Crops Research Institute- Agricultural Research Center, Egypt. It was cleaned manually to remove stones, grit, chaff and other impurities then milled.

Quinoa (*Chenopodium quinoa Willd.*) seeds were obtained from Egyptian Company for Oils and Natural Products, Egypt. It was washed with water at 60°C (with agitation) during one hour 1:10 (w/v). Seeds were dried at 60°C using a convective dryer according to Margarita *et al.* (2010) then milled.

Millet (Pearl millet) seeds were obtained from local market. It was cleaned manually then milled.

Guar gum and Ssl (Sodium stearoyl-2-lactylate) obtained from Chemitec International Technology Center- 6th of October City, Egypt. Other ingredients (fat, sugar, egg, baking powder, and salt) were purchased from local market.

Preparation of whole flour sample: For compositional and nutritional analysis, all the samples under study were milled using a Hammer mill laboratory type [DCFH-48-Germany], to obtain flour through sieve 0.1mm.

Methods:

Proximate analysis

Moisture, protein, fat, crude fiber, starch and ash content of the investigated samples (corn, quinoa and millet) were carried out according to the AOAC (2007). Total carbohydrate contents were tested quantitatively according to Kostas *et al.* (2016). The absorbance was measured at a wavelength of 490 nm using UV-Vis Shimadzu Spectrophotometer (UV-1601 PC).

Saponin was determined following the described method in Mastebroek *et al.* (2000)

Minerals (Fe, Ca, K, and Mg) contents were measured using Atomic Absorption (GBC 932/933-England) according to procedure outlined by AOAC (2007).

Vitamins namely (A), (C), (E) and vitamin (B complex) content were determined according to J. of Chromatography B 830:41-46 (2006), A 935:71-76 (2001), B 816:67-72 (2005) and B 816:67-72 (2005), respectively.

Amino acids of the investigated samples were carried out as described by the method of the Association of Official Analytical Chemists (AOAC, No.994.12, 2012) using Amino acid analyser biochrom 30 U.K.

Physical properties

Bulk density: Loose and tapped bulk densities were calculated by the equation given by Baysal *et al.* (2003):

$$P_b \text{ g/ml} = M/(V^* \text{ or } V_1)$$

Where: M = Amount of the sample poured into the cylinder

V^* = First scale reading

V_1 = Volume measured after 20 tapings

Water holding capacity (WHC) was determined according to Jongaroontaprangsee *et al.* (2007). It was calculated as the amount of water retained by the sample (g/g DMB) as follows:

$$\text{WHC (g/g)} = \frac{\{\text{Residue fresh weight (g)} - \text{Residue dried weight (g)}\}}{\text{Residue fresh weight (g)}}$$

Sample	Mixtures of flour sample	Sample	Mixtures of flour sample
Control	100% corn	7	75% quinoa +25% corn
1	25% millet+25% quinoa +50% corn	8	50% millet+25% quinoa +25% corn
2	25% millet+50% quinoa +25% corn	9	50% millet+50% quinoa
3	25% millet+75% quinoa	10	50% millet+50% corn
4	25% millet+75% corn	11	75% millet+25% quinoa
5	25% quinoa +75% corn	12	75% millet+25% corn
6	50% quinoa +50% corn	7	75% quinoa +25% corn

The eggs were initially homogenized with a hand blender (Braun, Kronberg, Germany) for a few seconds. Then the sugar, shortening, and syrup were mixed for 20 s with eggs in a moulinx mixer (LM240-France). Half of the flour and all other ingredients were mixed for 20 s. The remaining flour was added and mixed for 140 s to give a total mixing time of 3 min. Following a rest time of 20 min the dough was sheeted to a final thickness of 3 mm using a pastry break. Dough pieces with a diameter of 70 mm were cut and placed on a non-stick baking tray and baked for 8 min in a deck oven (More -Turkey) at 230°C top heat and 200°C bottom heat. After 40 min cooling at room temperature, the biscuits were placed in polyethylene bags (Tilman *et al.*, 2003).

Sensory evaluation of biscuits

Sensory evaluation of the biscuit samples was performed by 10 panelists of the Home Economic Dept., Fac. of Specific

Wettability was performed according to the method of Pearson (1976).

Process of Biscuit samples

The straight dough procedure of biscuit processing was done using the following ingredients: flour 100g, sugar 35g, fat 30g, egg 27.5g, syrup 5g, guar gum 2.5g, salt 0.5g, Ssl 0.5g and baking powder 0.5g. Flour of (corn, millet and quinoa) was supplemented with tested samples at different percentage of 25, 50, 75 % as follows:

Education, Ain Shams univ. according to (Tilman *et al.*, 2003). Palatability tests were considered in terms of color (20), break& shred (20), crumb color (20), surface character (20), mouth feel (20) and overall acceptability (100), (Padma and Prabhasankar, 2013)

Statistical analysis

Analysis of Variance and Duncan's multiple range tests at 5% level of significance was used to compare mean values of the tested factors. The analysis was carried out using the PROC ANOVA procedure of Statistical Analysis System (SAS, 1996).

RESULTS AND DISCUSSION

Technical aspects

Saponins are plant glycosides that impart a bitter taste and tend to foam in water solutions. Until recently, saponins have been considered to be highly toxic, nevertheless, those present in foodstuffs are

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non-toxic and it has been recommended that they may be even beneficial in human food (Vilche *et al.*, 2003).

The results showed that quinoa seeds contain saponin with (0.035%). After washing and soaking saponin content was reduced to 0.022%. So, it is in the acceptable range according to Koziol (1992) who reported that, quinoa was categorized as “sweet” being saponin free on with <0.11% saponin on a fresh weight basis or “bitter” with >0.11% saponins.

Saponins were reported by Valencia-Chamorro (2003) as the main anti-nutritional factor found in quinoa grain. Most of the saponins were found concentrated in the outer husk of the grain (perianth, pericarp, seed coat, and a cuticle-like layer) which facilitated their subtraction industrially by abrasive dehulling or traditionally by soaking and washing the grains with water. The amount of saponins present depends on the variety of quinoa. It is higher in bitter-flavor varieties than in sweet, or low-saponins, varieties. Quinoa contains saponins in the amount from 0.1% to 5%

Chemical composition of corn, quinoa and millet flours

The obtained results of corn, quinoa and millet grain are shown in Table (1). Corn flour had the highest percentage of moisture (6.86%) compared to quinoa (5.41%) and millet (4.87%). On the other hands, millet flour has the highest percentage of ash (4.86%) followed by corn (4.26%) and quinoa (2.97%). The highest percentage of fat was found in corn (9.72%) followed by millet (7.90%) and quinoa (6.55%). Comai *et al.* (2007) demonstrated that, quinoa lipids appear to be a high-quality edible vegetable oil, same in the fatty-acid composition to soybean oil. Tang *et al.* (2015) recently revealed that the fatty acid composition of quinoa is

9.9_12.3% saturated fat with palmitic acid predominant. Mono-unsaturated fat is 25.0_28.7% total fat and mainly oleic acid. Polyunsaturated fat is 56.20_58.3% total fat and is predominantly two essential fatty acids, linoleic acid (18:2n-6, an omega-6 fatty acid) and α -linolenic acid (an omega-3 fatty acid). The unsaturated fatty acids are well protected from oxidation by a high level of naturally occurring vitamin E present in the forms of tocopherols and tocotrienols. The omega-6/omega-3 ratio is approximately 6/1, which is more favorable than other plant oils regarding potential health benefits.

Quinoa and millet are worthy of consideration as important grain sources of protein being 15.10 and 12.50% in dry matter, respectively while corn is (9.20%). Matiacevich *et al.* (2006) reported that, quinoa seeds have a high nutritional value in comparison to most cereals. The protein content of quinoa seeds showed from 8% to 22%, which is higher on average than that in common cereals such as corn. In quinoa, maximum of the protein is positioned in the embryo. Albumins and globulins are the major protein fraction (44–77% of total protein), which is greater than that of prolamins (0.5–7.0%). Quinoa is considered to be a gluten-free grain for the reason that it content very little or no prolamin. Quinoa provides a nutritional, economical, easy-to-prepare, flavorful food source which is of particular relevance for people with gluten intolerance, such as those with celiac disease (Valencia-Chamorro, 2003).

Millet is unique among the cereals because of their richness in protein especially significant amounts of essential amino acids, energy value, fat and minerals (Devi *et al.*, 2011).

Also, results indicated that, millet and quinoa had high percentage of crude fiber (4.28%) and (3.94%), respectively as compared to corn (2.76%). These data were

in agreement with Lamothe *et al.* (2015), who reported that quinoa contains 10% total dietary fiber. Fiber is the carbohydrate fraction which is resistant to enzymatic digestion and absorption in the small intestine, and which usually undergoes full or partial fermentation in the large intestine. Dietary fiber is considered essential for optimal digestive health, and it also adds functional benefits (Brownawell *et al.*, 2012).

Results also indicated that no significant differences ($p > 0.05$) among the corn, quinoa and millet samples in carbohydrates (69.94, 69.95 and 69.84%) respectively. Starch is an important part in carbohydrate, results showed that corn was (41.29%), quinoa (46.97%) and millet was (43.85%). Valencia-Chamorro, (2003)

reported that the main component in quinoa contains of carbohydrates, and varies from 67% to 74% of the dry matter. Starch makes about 52–60%. The starch composite is located in the perisperm of the seeds; starch can be present as simple units or as spherical aggregates. Other, such as monosaccharides (2%) and disaccharides (2.3%), crude fiber (2.5–3.9%), and pentosans (2.9–3.6%). While carbohydrates are found in small amounts Devi *et al.* (2011) mention that millets show relatively higher than other cereals carbohydrate (72%) comprises of starch as the main constituent and the non-starchy polysaccharides which amounts to 15–20% of the seed matter as an unavailable carbohydrate dietary fiber content and complements which are the health benefits of the millet.

Table (1): Major chemical constituents (g\100g dry matter) of Corn, Quinoa and Millet.

Constituents (%)	Mean \pm SDM of Samples		
	Corn	Quinoa	Millet
Moisture	6.86 \pm 1.13 ^a	5.41 \pm 0.50 ^{ab}	4.87 \pm 0.61 ^b
Ash	4.26 \pm 0.13 ^b	2.97 \pm 0.03 ^c	4.86 \pm 0.00 ^a
Protein	9.20 \pm 1.10 ^c	15.10 \pm 1.40 ^a	12.50 \pm 2.90 ^b
Fat	9.72 \pm 0.13 ^a	6.55 \pm 0.20 ^b	7.9 \pm 0.20 ^{ab}
Carbohydrates	69.94 \pm 1.84 ^a	69.95 \pm 2.22 ^a	69.84 \pm 3.80 ^a
Starch	41.29 \pm 0.70 ^b	46.97 \pm 0.52 ^a	43.85 \pm 0.90 ^b
Crude Fiber	2.76 \pm 2.30 ^b	3.94 \pm 2.60 ^a	4.28 \pm 4.70 ^a

Data are presented as means \pm SDM ($n=3$).

Means within a row with different letters are significantly different at $P \leq 0.05$.

Amino acids

The 17 amino acids and their compositions identified of corn, quinoa and millet showed in Table (2). The highest amount of essential amino acids was Leucine which had a value of 9.89% dry matter in corn followed by millet (9.76%) then quinoa 8.42%. While the lowest one was Methionine which represent 2.85, 2.24 and 2.17% in quinoa, millet and corn respectively. Quinoa recorded the highest value of Lysine (5.30%) comparable to corn and millet. Millet generally contains

significant amounts of essential amino acids particularly the sulphur containing amino acids methionine and cysteine (2.87 and 3.60), respectively comparing with quinoa and corn. These results agree with Villa *et al.* (2014) who reported that seeds of quinoa content high protein average 12-18%. Moreover, this protein is of an exceptionally high quality and is particularly rich in balanced composition of essential amino acids, such as sulfur amino acids, lysine and aromatic amino acids, higher than those recommended by FAO/WHO (2011) and

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which are deficient in most grain crops but necessary for proper nutrition in humans. This fact results in protein content comparable to casein that of whole dry milk (Villa *et al.*, 2014) and this is in the line with results that showed millet and quinoa had the superior percentage of protein (12.50 and 15.10%) respectively while corn had (9.20%). In general, cereal proteins including millets are limited in lysine and tryptophan content and vary with cultivar (Devi *et al.*, 2011).

Out of the 17 amino acids observed, 7 were classified as essential amino acids, and 2 were semi-essential (histidine and Arginine) is essential for children. Quinoa recorded the highest values of histidine and Arginine 3.38 and 9.34% respectively.

While, millet recorded the highest values of non-essential amino acids such as Alanine, Aspartic, ½Cysteine and Glutamic acids (8, 8.08, 3.60 and 19.44%) respectively. Also in total non-essential amino acids millet was the highest value 54.8% followed by corn was 52.85% then quinoa (50.57%). While, quinoa recorded the highest value in total essential and semi essential amino acids 32.93 and 12.72% respectively.

Essential amino acids must be consumed each day to replace the amino acids lost during normal metabolism, and to rebuild and repair the body (FAO, 2013). Vega-Galvez *et al.* (2010) reported that quinoa protein can supply over 180% of the daily recommended intake of essential amino acids for adult nutrition.

Table (2). Essential, non essential and semi essential amino acids (%) of corn, quinoa and Millet.

Amino acids (A.A) %	Corn	Quinoa	Millet
Essential Amino Acid (EAA)			
Isoleucine (ILE)	3.15	3.77	4.16
Leucine (LEU)	9.89	8.42	9.76
Lysine (LYS)	2.63	5.30	2.80
Phenylalanine (PHE)	4.24	4.57	4.92
Threonine (THR)	3.37	3.38	3.12
Valine (VAL)	4.57	4.64	5.76
Methionine (MET)	2.17	2.85	2.24
Total EAA	30.02	32.93	32.76
Non-essential amino acids (NEAA)			
Alanine (ALA)	6.85	7.17	8.00
Aspartic (ASP)	6.87	8.08	8.08
½Cysteine (CYS)	2.39	2.12	3.60
Glutamic (GLU)	16.96	14.37	19.44
Glycine (GLY)	3.48	5.17	3.04
Proline (PRO)	8.37	6.44	6.00
Serine (SER)	4.13	3.91	3.12
Tyrosine (TYR)	3.80	3.31	3.52
Total NEAA	52.85	50.57	54.80
Semi essential amino acids (SEAA)			
Histidine (HIS)	2.93	3.38	2.56
Arginine (ARG)	4.13	9.34	4.64
Total SEAA	7.06	12.72	7.20
Total AA	89.93	96.22	94.76

Minerals (Macro and micro elements)

The results presented in Table (3) summarize the mineral composition of corn, quinoa and millet. Potassium (K) had the highest value of the three samples followed by magnesium (Mg) and calcium (Ca) had the least value of samples among the macro-elements.

Quinoa recorded the highest results of minerals, iron (4.47), calcium (82.78), magnesium (169.55) and potassium (1508.64 mg/100g). This data agrees with Valencia-Chamorro (2003) who reported that Quinoa is a good source of minerals. It contains more iron, calcium, zinc and magnesium, than common cereals. USDA (2015) reported that, many minerals in quinoa are present in greater quantities than other grains, including phosphorus, magnesium, potassium, calcium, iron, zinc,

and copper. The process of saponin removal decreases vitamin and mineral contents to an extent.

Millet takes the second level of minerals with 3.99, 72.59 and 157.69 mg/100g of iron, calcium and magnesium respectively. This result is close to Vijayakumari *et al.* (2003) who mentioned that millet is a rich source of iron and calcium. Calcium shortage leading to bone and teeth disorder, iron deficiency leading to anemia can be overwhelmed by presenting finger millet in our daily diet. Singh and Srivastava (2006) reported that, the iron content of 16 millet varieties ranged from 3.61 mg/100g to 5.42 mg% with a mean value of 4.40 mg/100g and this data agree with the result in Table (3) where millet contain 3.99 mg/100g.

Table (3). Mineral composition (mg/100g dry wt) of corn, quinoa and Millet.

Minerals (mg/100g)	Corn	Quinoa	Millet
Macro- elements			
Calcium (Ca)	55.03±0.1 ^c	82.78±±0.4 ^a	72.59±±0.1 ^b
Magnesium (Mg)	157.11±±0.1 ^b	169.55±±0.3 ^a	157.69±±0.2 ^{ab}
Potassium (K)	1284.62±±0.3 ^b	1508.64±±0.1 ^a	1202.18±±0.2 ^c
Micro-elements			
Iron (Fe)	2.99±±0.2 ^c	4.47±±0.2 ^a	3.99±±0.4 ^b

While the results indicated that corn is less than quinoa in potassium 1284.62mg/100g and iron 2.99 mg/100g as seen in Fig (1). All these minerals are necessary for physiological development

and general well being of human being and animals. The deficiency of one or more of these mineral elements may constitute nutritional disorder in human (Abiose and Ikujenlola, 2014).

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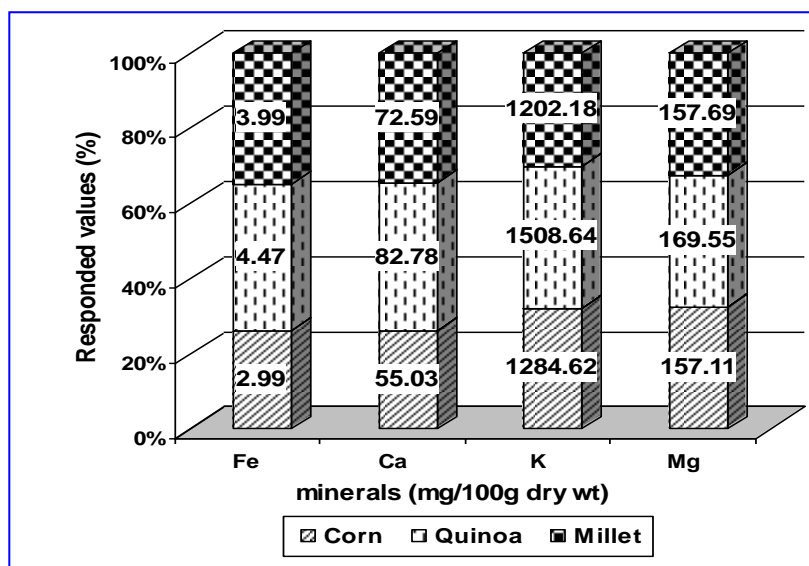


Fig. (1). Macro and micro minerals (mg/100g) of quinoa, millet and corn seeds.

Vitamins

Table (4) and Figure (2) show that quinoa seeds have more vitamin C, Nicotinic and Pyridoxin (67.34, 224.27 and 97.33ppm respectively) followed by corn then millet (8.01, 103.86 and 38.97 ppm) and (7.64, 66.17 and 35.42ppm) respectively. These results agree with USDA, (2015) mentioned that quinoa seeds are a rich source of vitamins, counting vitamin A precursor β -carotene, thiamin/vitamin B1, riboflavin/vitamin B2,

niacin/vitamin B-3, ascorbic acid/vitamin C, folic acid/vitamin B9 and vitamin E B6, and pantothenic acid. Also, Fitzpatrick *et al.* (2012) mentioned that quinoa seeds are a good and rich source of vitamins, which are required in the human diet to act as enzymatic cofactors in metabolism, regulate cell growth and development, protect against oxidative damage, improve vision, and play beneficial roles in various other physiological processes.

Table (4). Vitamins analyses (ppm) of corn, quinoa and millet seeds.

Vitamins	Corn	Quinoa	Millet
Vitamin A ($\mu\text{g}/100\text{g}$)	391.79 ^a	294.42 ^b	115.68 ^c
Vitamin E ($\mu\text{g}/100\text{g}$)	40.48 ^a	11.71 ^b	8.33 ^c
Vitamin C (ppm)	8.01 ^b	67.34 ^a	7.64 ^c
Vitamin B complex (ppm)			
Nicotinic acid (B3)	103.86 ^b	224.27 ^a	66.17 ^c
Thiamin (B1)	3.07 ^a	2.788 ^b	2.65 ^b
Pyridoxin (B6)	38.97 ^b	97.33 ^a	35.42 ^c
Folic acid	15.23 ^a	7.60 ^b	7.88 ^b
Ribiflavin (B2)	28.06 ^b	13.94 ^c	38.92 ^a
B12	29.07 ^c	71.95 ^b	116.01 ^a

*Values are means of triplicate readings.

Means within a raw with different letters are significantly different at $P \leq 0.05$.

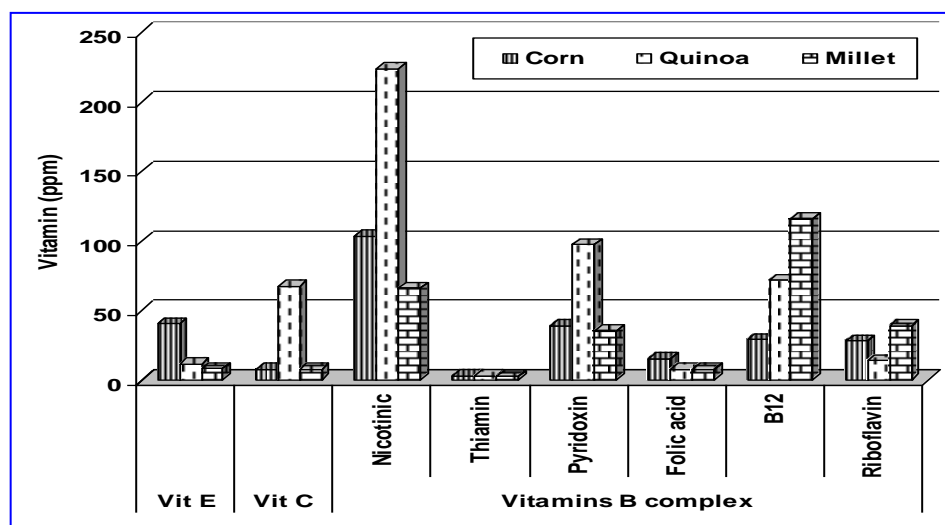


Fig. (2). Comparison between corn, quinoa and millet on vitamins E, C and B complex.

The vitamins soluble in fat (Vit. A and Vit.E) are found to be the highest ratio in corn followed by quinoa then millet (391.7885 and 40.48049 $\mu\text{g}/100\text{g}$), (294.4231 and 11.71329 $\mu\text{g}/100\text{g}$) and (115.6823 and 8.333006 $\mu\text{g}/100\text{g}$), respectively.

Water holding capacity (WHC)

Functional properties are controlled by the composition and structure of proteins and the interactions of proteins with one another and with other substances. Water-holding capacity (WHC) is an important protein–water interaction that occurs in various food systems. WHC represents the ability of a protein matrix to absorb and retain bound, hydrodynamic, capillary, and physically entrapped water against gravity (Damodaran and Paraf, 1997). The obtained results show that water holding capacity WHC is increased in (25% quinoa +75% corn) was 0.87 g/gDMB followed by(50% millet + 25% quinoa + 25% corn) 0.72 g/g as seen in Fig (3). While the samples (75%

millet + 25% quinoa) and (25 % millet+ 25% quinoa+50% corn) were recorded the least values 0.66g/g DMB comparing with the control 100% corn 0.71g/gDMB as seen in Table (5). On the other hand, these results suggested that dietary fibers from all samples containing quinoa with corn in different ratios (70 g/g) could aid gel formation and enhance texture stability of food products such as bread and other baked products. In contrast, low WHC of samples containing quinoa with millet ranged between 66-69 g/gDMB may be due to the damage of fiber matrix and the collapse of the pore during grinding. Jideani (2011) reported that water holding capacity of highly protein content is very significant as it affects the texture, juiciness, and taste of food formulations and in particular the shelf-life of bakery products. Water plays an important role in the main changes that occur thru baking, which include starch gelatinization, protein denaturation, yeast- and enzyme-inactivation, flavor and color formation (Pomeranz, 1985).

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Table (5). Loss bulk density, tapped bulk density, WHC and Wettability of tasted flower samples.

Blended samples	Bulk density (g/cm ³)			WHC (g/g DMB)	Wettability (S)
	Loss	Tapped	Difference		
25%M+75%Q	0.406±0.01 ^c	0.631±0.02 ^e	0.225	0.69±0.02 ^e	2.88±0.47 ^e
25%M+75%C	0.402±0.004 ^c	0.672±0.01 ^{de}	0.270	0.69±0.20 ^e	3.81±0.39 ^{abc}
50%M+50%Q	0.409±0.01 ^c	0.737±0.05 ^{bcd}	0.328	0.68±0.03 ^f	3.50±0.06 ^{bcde}
50%M+50%C	0.400±0.00 ^c	0.715±0.02 ^{bcd}	0.315	0.67±0.02 ^g	3.39±0.08 ^{bcde}
75%M+25%Q	0.407±0.01 ^c	0.696±0.03 ^{bcde}	0.289	0.66±0.02 ^h	3.23±0.06 ^{cde}
75%M+25%C	0.426±0.03 ^{bc}	0.770±0.10 ^{ab}	0.344	0.67±0.03 ^g	3.06±0.61 ^{de}
25%Q+75%C	0.409±0.01 ^c	0.744±0.03 ^{bcd}	0.335	0.87±0.03 ^a	3.42±0.17 ^{bcde}
50%Q+50%C	0.428±0.01 ^b	0.669±0.01 ^{de}	0.241	0.70±0.02 ^d	3.22±0.05 ^{cde}
75%Q+25%C	0.423±0.01 ^{bc}	0.632±0.03 ^e	0.209	0.70±0.03 ^d	3.92±0.17 ^{ab}
25%M+25%Q+50%C	0.416±0.02 ^c	0.745±0.02 ^{bcd}	0.329	0.66±0.01 ^h	3.92±0.32 ^{ab}
25%M+50%Q+25%C	0.483±0.01 ^a	0.763±0.03 ^{bc}	0.280	0.69±0.014 ^e	3.72±0.24 ^{bcd}
50%M+25%Q+25%C	0.400±0.00 ^c	0.685±0.03 ^{cde}	0.285	0.72±0.02 ^b	4.46±0.08 ^a
100%C	0.455±0.03 ^{ab}	0.830±0.04 ^a	0.375	0.71±0.02 ^c	4.01±0.48 ^{ab}

*Values are means of triplicate readings. DMB= Dry matter basis Q = quinoa M= millet C = corn Means within a column with different letters are significantly different at P≤ 0.05.

To establish the techno-functional properties of quinoa flours, water-holding capacity (WHC) was assessed. WHC allow assessing the flour aptitude to retain water under a centrifugal gravity force, considering physically entrapped, capillary, bound and hydrodynamic water. Ogungbenle *et al.* (2009) who mentioned

that quinoa flours were capable of retaining 147% of its weight in water.

WHC is important indicator to evaluate the functions of dietary fiber because it is always closely related to the cholesterol-lowering ability of dietary fiber (Li *et al.*, 2013). WHC increased gradually with adsorption time increasing in the first 2 h, and then WHC kept stable.

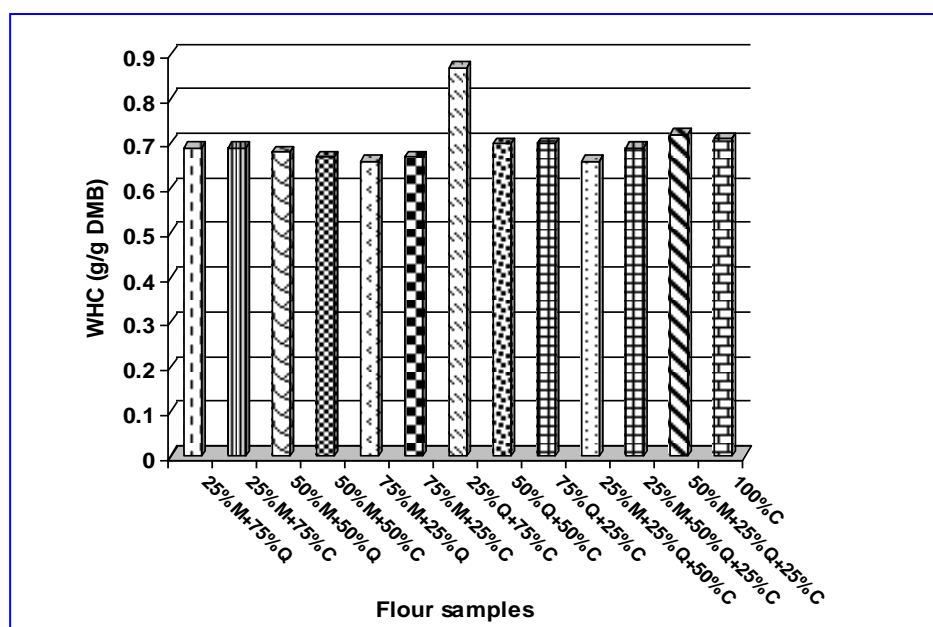


Fig. (3). Comparison between blended flour samples of corn, quinoa and millet on WHC.

Bulk density

The loose and tapped bulk density of the investigated samples is given in Table (5). Bulk density of the prepared powder was completely differed within the kind of the samples. The highest loose porosity values were found for 25% Millet+50% Quinoa+25% Corn samples were 0.483 g/cm^3 . While the corn sample was the highest value in the tapped density of 0.830 g/cm^3 followed by 75% Millet + 25% Corn (0.770 g/cm^3) and the difference was 0.375 and 0.344 g/cm^3 respectively. This trend may be related to the particle size of the granules which being of lowest diameter or indicated that degree of fineness was absolutely inherent within the fat content which acting as adhering substances and preventing the molecules to be oriented in a homogenous matter. Other possible explanation is that, the presences of higher level of fat (as see in Table 1) usually minimize the bulk density. These results are close to the opinion of Peleg and Bagley (1983) who stated that, it is still evident by that the properties of any low-density powders cannot be explained by gemotric consideration only, but also to their physical as well as their chemical properties. The lowest differences between loss and tapped bulk density were found in 75%Q+25%C (0.209 g/cm^3) followed by 25% M+75% Q (0.225 g/cm^3). Krokida and Maroulis (1999) stated that the significant different bulk density values of a product can be instigated by difference in particle size or dry matter content. It is of interest to mention that most food powders are known to be cohesive, which means that their particle attraction forces are significantly higher in relation to the particles own weight, (Dobbs *et al.*, 1982). Another notable exception to this trend is the case of fine powders that very

cohesive even in their dry form. (Baysal *et al.*, 2003).

Wettability

Wettability actions of the different samples were also measured as seen in the Table (5). The following results were obtained, the highest wettability was (50% Millet+25% Quinoa+25% Corn) 4.46 S and on the contrary 25% Millet+75% Quinoa recorded the lowest wettability 2.88 S. Typically, a increase wettability time for sample was associated with an increase in wettability in all samples from 3: 4 S in all tested sample.

Wettability is one of the properties that may influence the general reconstitution and/or mixing characteristics. Many of conventional dry sample powders need long time to wet reflecting little wettability, because of low specific surface area and particle's texture/microstructure, and chemical composition Liapis and Bruttini (1995). Wettability is a measure of the ability of powder to absorb water. Thus, the more the wetting time, the lower the wettability Singh and Rai (1998). These results are mainly correlated with the structural configurations and the chemical constituents of the tested samples. Such pattern of results is in parallel with Liapis and Bruttini (1995) who also demonstrated that in some cases, complete structural rigidity may hamper or contradicted rehydration due to the absence of pathways for the entrance of water.

Sensory characteristics of biscuits

The Sensory evaluation of biscuits baked from suggested powder samples from corn + Millet + Quinoa at ratios 25, 50 and 75 % are shown in Table (6).

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Table (6). Mean score values of the sensory attributes of biscuits mad from tested flour samples.

Treatments	Mean of scoring tested parameters out of (20)					Over all acceptability (100)
	Color	Break & shred	Crumb color	Surface character	Mouth feel	
25% M+25% Q+50% C	13.4±2.1 ^{bc}	12.7±2.6 ^{bc}	12.0±2.2 ^{cd}	12.6±3.1 ^b	11.3±1.5 ^d	58.3±11.5 ^e
25% M+50% Q+25% C	12.4±2.0 ^c	13.1±2.3 ^{bc}	11.4±1.7 ^d	13.1±2.8 ^b	11.1±1.5 ^d	59.3±8.2 ^d
25% M+75% Q	12.4±1.7 ^c	12.1±1.7 ^c	11.9±2.3 ^{cd}	12.3±2.1 ^b	12.9±1.3 ^{dc}	58.8±8.3 ^e
25% M+75% C	13.6±3.5 ^{bc}	13.5±3.0 ^{bc}	14.0±3.5 ^{bc}	13.4±3.8 ^b	13.2±2.9 ^{bc}	67.7±12.9 ^{bc}
25% Q+75% C	15.0±2.1 ^b	14.9±2.1 ^b	15.5±1.9 ^b	15.0±3.0 ^b	14.9±1.9 ^b	74.3±9.6 ^b
50% Q+50% C	14.4±3.6 ^{bc}	13.1±2.6 ^{bc}	14.6±2.8 ^b	14.1±2.4 ^b	13.2±3.1 ^{cd}	69.4±9.9 ^{bc}
75% Q+25% C	14.2±2.7 ^{bc}	14.5±3.3 ^{ab}	13.5±2.1 ^{bc}	15.1±2.2 ^{ab}	14.1±1.7 ^{bc}	71.4±8.6 ^{bc}
50% M+25% Q+25% C	13.4±2.1 ^{bc}	14.2±2.0 ^{ab}	14.6±0.9 ^b	14.0±2.8 ^b	12.4±1.8 ^d	68.6±6.6 ^c
50% M+50% Q	13.7±2.0 ^{bc}	13.1±2.5 ^{bc}	13.6±2.3 ^c	12.3±1.6 ^b	12.7±2.2 ^{cd}	65.4±7.0 ^{bc}
50% M+50% C	14.6±2.9 ^{bc}	14.4±2.1 ^{ab}	13.3±2.7 ^{bc}	13.9±1.8 ^b	13.7±1.5 ^{bc}	69.9±8.2 ^{bc}
75% M+25% Q	12.6±1.2 ^{bc}	12.5±1.2 ^{bc}	13.3±2.0 ^{bc}	13.1±1.8 ^b	13.0±2.1 ^c	64.5±4.7 ^{cd}
75% M+25% C	13.4±2.0 ^{bc}	14.1±2.4 ^{ab}	13.2±1.6 ^{bc}	15.0±2.8 ^{ab}	13.9±2.4 ^{bc}	69.6±6.1 ^{bc}
100% C	19.2±1.6 ^a	16.5±3.5 ^a	17.6±2.9 ^a	17.4±3.6 ^a	16.5±3.4 ^a	87.2±9.8 ^a

*Values are means of triplicate readings. Q = quinoa M= millet C = corn

Means within a column with different letters are significantly different at $P \leq 0.05$.

The obtained results indicated that, increasing the level of corn flour increased sensory scores of biscuits for color, Break and shred, crumb color, surface character, mouth feel and over all acceptability as seen in blended 25%Q +75% corn followed by 75%Q +25%C over all acceptability was 74.3±9.6 and 71.4±8.6 respectively as seen in Figure (4). Whereas control biscuit (100% corn) sample had the highest value in all parameters and over all acceptability was (87.2±9.8) compared to other tested samples. While there were no significant differences in samples (25% Millet+25% Quinoa+50% Corn) and (25% Millet+50%

Quinoa+25% Corn) in surface character and mouth feel were and they recorded the lowest values, the overall capacity of them was 58.3±11.5 and 59.3±8.2 respectively. In conclusion, addition corn flour by each ratio to quinoa or millet record good values and satisfied acceptable about blended the three samples with each other. These results agree with Handa *et al.*, (2012) who reported that, corn flour is related to cultural or social preferences and some of the products are more suitable for commercial trade because they require further processing or provide convenience and extended shelf life.

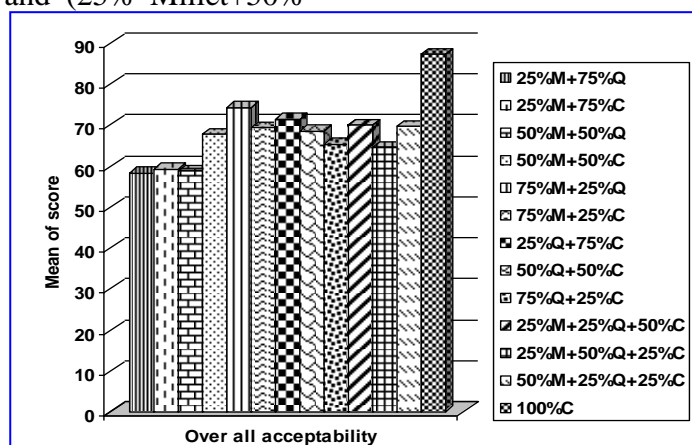


Fig. (4). Over all acceptability of biscuits made from blended flour samples of corn, quinoa and millet.

REFERENCES

- A.O.A.C. (2007). Official Methods of Analysis of the Association of Official Analytical Chemists. 18th Ed. Published by Association of Official Analytical Chemists, Gaithersburg, MD, USA.
- A.O.A.C. (2012). Official Methods of Analysis (19thed.). Arlington, VA, USA: Association of Official Analytical Chemists.
- Abiose, S.H. and Ikujenlola, A.V. (2014). Comparison of chemical composition, functional properties and amino acids composition of quality protein maize and common maize (*Zea mays* L). *Afr. J. Food Sci. Technol.*, 5(3):81-89.
- Adekunle A.A., Ellis-Jones J., Ajibefun I., Nyikal R.A., Bangali S., Fatunbi O., Angé A. (2012). Agricultural innovation in Sub-Saharan Africa : experiences from multiple-stakeholder approaches. ISBN 978-9988-8373-2-4.
- Ajanovic, A. (2011). Biofuels versus food production: does biofuels production increase food prices? *Energy*, 36 (4): 2070-2076. At: <https://doi.org/10.1016/j.energy.2010.05.019>
- Amadou, I.; Gbadamosi, O.S. and Guo-Wei, L. (2011). Millet-based traditional processed foods and beverages-A review. *Cereal Food World*, 56(3):115–121.
- Baysal, T.; Icier, F.; Ersus, S., and Yildiz, H. (2003). Effects of microwave and infrared drying on the quality of carrot and garlic. *Eur. Food Res. Technol.*, 218: 68-73.
- Brownawell, A.M.; Caers, W.; Gibson, G.R.; Kendall, C.W.C. , Lewis, K.D. Ringel Y and Slavin JL. (2012). Prebiotics and the health benefits of fiber: current regulatory status, future research, and goals. *J Nutr.*; 142:962–974.
- Comai, S.; Bertazzo, A.; Bailoni, L.; Zancato, M.; Costa, C.V.L. and Allegri G. (2007). The content of proteic and nonproteic (free and protein bound) tryptophan in quinoa and cereal flours. *Food Chem.*, 100: 1350–1355.
- da Silva, T.F. and Conti-Silva, A.C. (2018). Potentiality of gluten-free chocolate cookies with added inulin/oligofructose: Chemical, physical and sensory characterization. *LWT - Food Sci. Technol.*, 90:172-179
- Damodaran, S. and Paraf, A. (1997). Food proteins and their applications. Marcel Dekker, New York, 696.
- Darvey, N.L.; Naeem, H., and Gustafson, J.P. (2000). Triticale: production and utilization. Chapter 9 in: *Handbook of Cereal Science and Technology*, 2nd ed. (eds. K. Kulp, J. Ponte). Marcel Dekker, New York, pp: 257-274.
- Devi, P.B.; Vijayabharathi, R.; Sathyabama, S.; Malleshi, N.G. and Priyadarisini, V.B. (2011). Health benefits of finger millet (*Eleusine coracana* L.) polyphenols and dietary fiber: a review. *J. Food Sci. Technol.*, 51(6): 1021–1040. DOI: 10.1007/s13197-011-0584-9
- Dobbs, A.J.; Peleg, M.; Mudgett, R.E. and Rufner, R. (1982). Some physical characteristics of active dry yeast. *Powder Technol.*, 32-75.
- FAO/WHO (2011). Quinoa: An ancient crop to contribute to world food security. Regional Office for Latin America and the Caribbean.
- FAO (2013). Dietary protein quality evaluation in human nutrition. Report of an FAO Expert

Chemical and functional properties of free-gluten biscuit making from corn, quinoa and millet flours

- Consultation March 31-April 2, 2011. Food and Nutrition Paper 92.
- Fischer, S.; Wilckens, R.; Jara, J.; Aranda, M.; Valdivia, W.; Bustamante, L.; Grafa, F. and Obala, I. (2017). Protein and antioxidant composition of quinoa (*Chenopodium quinoa* Willd) sprout from seeds submitted to water stress, salinity and light condition. *Ind. Crops Prod.*, 107: 558–564.
- Fitzpatrick, D.; Scanlon, E.; Krüse, J.; Vos, B.; Evans-Hurson, R.; Fitzpatrick, E. and McSweeney, S. (2012). Blend uniformity analysis of pharmaceutical products by Broadband Acoustic Resonance Dissolution Spectroscopy (BARDS), *Int. J. Pharm.*, 438(1–2): 134-139
- Gawlik-Dziki, U.; Świeca, M.; Sułkowski, M.; Dziki, D.; Baraniak, B., and Czyz, J. (2013). Antioxidant and anticancer activities of *Chenopodium quinoa* leaves extracts – In vitro study. *Food Chem. Toxicol.*, 57: 154–160.
- Gélinas, P.; McKinnon, C. M.; Mena, M. C., and Méndez, E. (2008). Gluten contamination of cereal foods in Canada. *Int. J. Food Sci. Technol.*, 43(7):1245–1252.
- Gordillo-Bastidas, E.; Díaz-Rizzolo, D.A.; Roura, E.; Massanés, T. and Gomis, R., (2016). Quinoa (*Chenopodium quinoa* Willd), from nutritional value to potential health benefits: an integrative review. *J. Nutr. Food Sci.*, 63 (3): 1–10.
- Graf, B.L.; Rojas-Silva, P.; Rojo, L.E.; Delatorre-Herrera, J.; Baldeón, M.E. and Raskin, I. (2015). Innovations in health value and functional food development of quinoa (*Chenopodium quinoa* Willd). *Comp. Rev. Food Sci. Food Saf.*, 14(4): 431–445.
- Handa, C.; Goomer, S. and Siddhu, A. (2012). Physicochemical properties and sensory evaluation of fructooligosaccharide enriched cookies. *J. Food Sci. Technol.*, 49(2): 192–199.
- Hill, M.J. and Path, F.R. (1998). Cereals, dietary fiber and cancer. *Nutr. Res.*, 18(4): 563-659.
- Jideani, V.A. (2011). Functional properties of soybean food ingredients in food systems. In Ng PT-B editor; Soybean - Biochemistry, Chemistry and Physiology: InTech, 2011.
- Jongaroontaprangsee, S.; Tritrong, W. and Chokanaporn, W., (2007). Effects of drying temperature and particle size on hydration properties of dietary fiber powder from lime and cabbage by-products. *Int. J. Food Proper*, 10: 887–897.
- Kostas, T.E.; Wilkinson, J.S.; White, A.D. and Cook, J.D. (2016). Optimization of a total acid hydrolysis based protocol for the quantification of carbohydrate in macro algae. *J. Algal Biomass Utin.* 7(1):21-36.
- Koziol M.J. (1992). Chemical composition and nutritional evaluation of quinoa (*Chenopodium quinoa* Willd). *J. Food Composition Anal.*, 5: 35–68.
- Krokida, M.K. and Maroulis, Z.B. (1999). Effect of microwave drying on some quality properties of dehydrated products *Drying Technol.*, 17(3): 449–466.
- Lamothe, L.M.; Srichuwong, S.; Reuhs, B.L. and Hamaker, B.R. (2015). Quinoa (*Chenopodium quinoa* W.) and amaranth (*Amaranthus caudatus* L.) provide dietary fibres high in pectic substances and xyloglucans. *Food Chem.*, 167: 490–496.
- Li, T.; Zhong, J.Z.; Wan, J.; Liu, C.M.; Le, B.Y.; Liu, W. and Fu, G.M., (2013). Effects of micronized okara dietary

- fiber on cecal microbiota, serum cholesterol and lipid levels in BALB/c mice. *Int. J. Food Sci. Nutr.*, 64: 968e973.
- Liapis, A. and Bruttini, R. (1995). Freeze Drying, In: MUJUMDAR, A.S. *Handbook of Industrial Drying*. Marcel Dekker. New York.
- Lubna, M. and Vidhu, A.B. (2012). Fortification of biscuit with flaxseed: biscuit production and quality evaluation. *J. Environ. Sci. Toxicol. Food Technol. (IOSR-JESTFT)*, 1(5): 6-9.
- Margarerita, M.; Vega-Galvez, A.; Lopez, J.; Parada, G.; Sanders, M.; Aranda, M.; Elsa, V. and Karina, D. (2010). Impact of air-drying temperature on nutritional properties, total phenolic content and antioxidant capacity of quinoa seeds (*Chenopodium quinoa* Willd). *Industrial Crops Products*, 32: 258-263.
- Mastebroek, H.D.; Limburg, H.; Gilles, T. and Marvin, H.J.P. (2000). Occurrence of saponin in leaves and seeds of Quinoa (*Chenopodium quinoa* Willd). *J. Sci. Food Agric.*, 80:152–156
- Matiacevich, S.B.; Castellion, M.L.; Maldonado, S.B. and Buera, M.P. (2006). Water-dependent thermal transitions in quinoa embryos. *Thermochimica Acta*, 448: 117–122.
- Ogunbenle, H.N.; Oshodi, A.A. and Oladimeji, M.O. (2009). The proximate and effect of salt applications on some functional properties of quinoa (*Chenopodium quinoa*) flour. *Pak. J. Nutr.* 8 (1): 49–52.
- Okpala, L.C. and Okoli, E.C. (2011). Nutritional evaluation of cookies produced from pigeon pea, cocoyam and sorghum flour blends. *Afr. J. Biotechnol.*, 10(3): 433-438.
- Padma, I. and Prabhasankar, A. (2013). Fructooligosaccharide- Retention during baking and its influence on biscuit quality. *Food Bioscience*, 4: 68-80
- Pearson, D. (1976). *The Chemical Analysis of Food*, 1 (Ed). pp. 446- 447. J.A. Churchill, 104 Gloucesterplace, London.
- Peleg M. and Bagley, E.B. (1983). *Physical Properties of Foods*. Westport: CN: AVI Publishers Co.,pp.103-115
- Pellegrini, N. and Agostoni, C., (2015). Nutritional aspects of gluten-free products. *J. Sci. Food Agric.* 95 (12): 2380–2385.
- Pomeranz, Y. (1985). *Functional properties of food components*. New York: Academic Press, Inc.
- SAS Program (1996). *SAS/STAT User's Guide Release 6.12^{ed}*. Cary, NC, USA: SAS Inst. Inc.
- Singh, P. and Srivastava, S. (2006). Nutritional composition of sixteen new varieties of finger millet. *J. Community Mobilization Sustainable Dev.*,1(2): 81-84.
- Singh, S. and Rai, T. (1998). Storage studies of whey powder. *Beverage and Food World*, 25(20): 52–55.
- Tang, Y.; Li, X.; Chen, P.X.; Zhang, B., and *et al.*, (2015). Characterisation of fatty acid, carotenoid, tocopherol/tocotrienol compositions and antioxidant activities in seeds of three *Chenopodium quinoa* Willd genotypes. *Food Chem.* 174, 502–508.
- Tilman, J. S.; Colm, M.O.; Denise, M.; Anja, D.; Elke, K. and Arendt, P. (2003). Influence of gluten-free flour mixes and fat powders on the quality of gluten-free biscuits. *Eur. Food Res. Technol.*, 216:369–376.
- Ubbor, S.C. and Akobundu, E.N.T. (2009). Quality characteristics of cookies

Chemical and functional properties of free-gluten biscuit making from corn, quinoa and millet flours

- made from composite flours of watermelon seed, cassava and wheat. *Pak. J. Nutrition*, 8: 1097-1102.
- USDA (2015). National Nutrient Database for Standard Reference Release 27. At: www.ndb.nal.usda.gov/ndb.
- Valencia-Chamorro, S.A. (2003). Quinoa. In: Caballero B.: *Encyclopedia of Food Science and Nutrition*, Vol. 8. Academic Press, Amsterdam: 4895–4902.
- Vega-Gálvez, A.; Miranda, M.; Vergara, J.; Uribe, E.; Puente, L., and Martínez, E.A. (2010). Nutrition facts and functional potential of quinoa (*Chenopodium quinoa* willd.), an ancient Andean grain: a review. *J. Sci. Food Agric.*, 90(15):2541-2547.
- Vijayakumari, J.; Mushtari, B.J.; Shamsad, B. and Sumangala, G. (2003). Sensory attributes of ethnic foods from finger millet. Paper presented at CCSHAU, Hisar. *Recent Trends in Millet Processing and Utilization*: 7–12.
- Vilche, C.; Gely, M. and Santalla, E. (2003). Physical properties of quinoa seeds. *Biosystems Engineering*, 86: 59–65.
- Villa, D.Y.G.; Luigi, R.; Khawla, K.; Maddalena, L. and Luca, R. (2014). Chemical and nutritional characterization of *Chenopodium pallidicaule* (cañihua) and *Chenopodium quinoa* (quinoa) seeds. *Emir. J. Food Agric.*, 26(7):609-615.
- Wang, S. and Zhu, F. (2016). Formulation and quality attributes of quinoa food products. *Food Bioproc. Technol.*, 91: 49–68
- Wu, G. (2015). Nutritional properties of quinoa. *Quinoa: Improvement and Sustainable Production*. pp. 193–210. <http://dx.doi.org/10.1002/9781118628041.ch11>.
- Yang, X.; Wan, Z.; Perry, L.; Lu, H.; Wang, Q.; Hao, C.; Li, J.; Xie, F.; Yu, J.; Cui, T.; Wang, T.; Li, M., and Ge, Q. H., (2012). Early millet use in northern China. *Proc. Nat. Acad. Sci. USA*, pp. 1–5.
- Zhu, F. and Li, G. (2018). Quinoa starch: Structure, properties, and applications. *Carbohydrate polymers*, 181: 851-861.

الخصائص الكيميائية والوظيفية لبسكويت خالي الجلوتين مصنع من دقيق الذرة والكينوا والدخن

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المستخلص

تهدف هذه الدراسة إلى تقييم الخصائص الكيميائية والوظيفية لخلطات مختلفة للبسكويت خالي الجلوتين وعلاقتها بجودة المنتج النهائي المصنع من نسب مختلفة من دقيق الذرة والكينوا والدخن. أظهرت النتائج أن بذور الكينوا تحتوي على نسبة سابونين (0.035%) ووصلت بعد الغسل والنقع الي (0.22%) لذلك فهي في المعدل المقبول به. كما أظهرت النتائج أن دقيق الذرة سجل أعلى نسبة رطوبة (6.86%) بينما كانت نسبة الرطوبة في الكينوا (5.41%) وفي الدخن (4.87%). كما سجل الذرة أعلى نسبة في الدهون (9.72%) يليها الدخن (7.9%) والكينوا (6.55%). والجدير بالذكر أن كل من الكينوا والدخن يعتبران أهم مصادر للبروتين في الحبوب فقد سجلا (15.10% و 12.50% على الوزن الجاف) على التوالي بينما كانت نسبة البروتين في الذرة (9.20%). كما أظهرت النتائج أن كل من الدخن والكينوا لديهم محتوى عالي من الالياف (4.28%) ، (3.94%) على التوالي مقارنة بالذرة (2.76%). ويعتبر النشا هو أهم جزء في الكربوهيدرات وسجلت النتائج نسبته في الذرة (41.29%) بينما في الكينوا (46.97%) والدخن (43.85%).

يحتوي الدخن عموماً على كميات محدودة من الاحماض الامينية الاساسية وخاصة المجموعات الكبرى ومنها الميثونين والسيسنتين بنسب (2.87% و 3.60% على التوالي) مقارنة بالكينوا والذرة. كما تعتبر الكينوا مصدراً جيداً للمعادن وخاصة الحديد (4.47) والكالسيوم (82.78) والماغنسيوم (169.55) والبوتاسيوم (1508.63 ملجم/100جم). كما أظهرت النتائج ان الذرة تحتوي على اعلى نسبة من (فيتامين أ وفيتامين هـ) يليها الكينوا ثم الدخن. وأوضحت النتائج أن القدرة على الأرتباط بالماء WHC قد ازدادت في (الكينوا+الذرة) ويتبعها الدخن. وزيادة نسبة التبلل wettability في خلطات الذرة والدخن والكينوا بينما سجل خليط الكينوا والدخن اقل النسب.

ومن خلال النتائج يتضح الآتي: بزيادة نسبة دقيق الذرة في الخلطات يزداد القبول العام للبسكويت كما هو واضح في الخلطة (25% كينوا + 75% ذرة) تليها (75% كينوا + 25% ذرة) ((9.6±74.3) و (8.6±71.4) على التوالي. بينما سجلت عينة البسكويت الكنترول (100% ذرة) أعلى قيم في جميع المقاييس الحسية والقبول العام (87.2±9.8) مقارنة بعينات البسكويت الاخرى المختبرة. والخلاصة هي أن اضافة دقيق الذرة بكل نسبه سواء للكينوا أو للدخن سجل قيما جيدة وقبول مرضي في العينات المخلوطة من الحبوب الثلاث معا.

الكلمات المفتاحية: الذرة- الكينوا- الدخن- البسكويت- خالي الجلوتين- الخصائص الوظيفية