## ANTIBACTERIAL ACTIVITY OF FUROQUINOLONE ALKALOIDS AGAINST POTATO SOFT ROT BACTERIUM, Erwinia carotovora FROM Ruta chalepensis LEAVES

## Ahmed M. Emam<sup>1</sup> and Mohamed E. Mahmoud<sup>2</sup>

Biochemistry Department, Faculty of Agriculture, Fayoum Branch, Cairo Univ., Egypt.
Botany Department (Microbiology), Faculty of Science, Cairo Univ., Giza, Egypt.

#### ABSTRACT

Soft rot bacterium; *Erwinia carotovora* is one of the most severe post-harvest diseases of potatoes worldwide. This bacterium affects tubers during storage, transit and marketing. The aqueous ethanolic extract (80%) of *Ruta chalepensis* leaves (Rutaceae) showed antibacterial activity against *Erwinia carotovora* (MIC = 625 µg/ml). Bioassay-guided isolation of this aqueous extract by using a combination of different chromatographic methods (TLC and column chromatography) yielded two furoquinolone alkaloids for the first time from this plant. Their structures were characterized as 2, (hydroxy isopropyl) – 6, hydroxy –9, methyl-dihydrofuro [2,3] –quinol-4-one (I) and 3, (hydroxy methyl)-6, methoxy-2, 2, 9-trimethyl-dihydrofuro [2,3] quinol-4-one (II) by <sup>1</sup>H, <sup>13</sup>C NMR and MS spectral data. The isolated alkaloids I and II showed antibacterial activity against *Erwinia carotovora* with MIC at 20 and 35 µg/ml respectively.

# Key Words: Furoquinolone alkaloids, Soft rot bacterium, *Ruta chalepensis*, Rutaceae and *Erwinia carotovora*.

#### INTRODUCTION

Soft rot bacterium; *Erwinia carotovora* is one of the most severe postharvest diseases of potatoes worldwide. This bacterium affects tubers during storage, transit and marketing, causing a major problem to the potato industry worldwide.

In Egypt, potato is one of the most important vegetable crops. Its important is not due only to local consumption but also for exportation to European community, which represents about 42.7% of Egypt's agricultural exports. Synthetic bactericides like Tecto 5% D (1.25 kg/Ton tuber) have been used for control the soft rot disease. However, their use is restricted due to their harmful effects on human beings and environment, in addition to the chemical residues of potato diminished the potato exportation quantity to European community (Food and Veterinary Office, 2000).

Therefore, the replacement of synthetic by natural pesticides for pest control application has increased interest in the potential use of natural products in general.

Ruta chalepensis L. (Rutaceae) is a small shrub originating in southern Europe, but now spread over North America and some other places [Fischer et al. 1988] The leaves and roots are used in folk medicine against intestinal colic, spasmodic atonic amenorrhoea, rhematic diseases, headaches and wounds [Mahran, 1967]. This plant exhibited many activities such as antiinflammatory activity [Al-Said et al. 1990], antifertility activity [Ulubelen et

#### Ahmed M. Emam and Mohamed E. Mahmoud

al. 1994], anticonvulsant activity [Aguilar-Santamaria and Totoriello 1996], molluscicidal activity [Hmamouchi et al. 2000], antimicrobial activity [El-Sayed et al. 2000] larvicidal activity [Mookey et al. 2002] and repellent activity [Hadis et al. 2003]. Previous phytochemical research on this plant has resulted in the isolation of several acridone, quinoline and quinlone alkaloids and coumarines from the aerial parts [Mohr et al. 1982; Ulubelen et al. 1986; Ulubelen and Guner 1988 and Zobel et al. 1990] from the cellcultures [Fischer et al. 1988; Ulubelen et al. 1992] and from the roots [Ulubelen and Terem 1988; Ulubelen et al. 1988; Ulubelen and Tan 1990 and Elsayed et al. 2000].

In this paper we report for the first time the isolation and structural elucidation of two furoquinolone alkaloids responsible for the bactericidal activity of *Ruta chalepensis* leaves against the Potato soft rot; *Erwinia carotovora*.

## MATERIALS AND METHODS

#### Plant material

The leaves of *Ruta chalepensis* L. (Rutaceae) were collected from the experimental farm of the Faculty of Agriculture, Cairo University, Giza, Egypt and identified by the Botany Department, Faculty of Science, Cairo University. A voucher specimen deposited in the Biochemistry Department, Faculty of Agriculture, Cairo University, Giza.

#### Extraction

Ground air dried leaves (350 g) was extracted three times with 80% ethanol (each 700 ml) at room temperature ( $25 \pm 2^{\circ}$ C). After filtration, the combined extract was evaporated under reduced pressure to afford 55.2 g of dry extract.

#### Antibacterial test

Tester strain of *Erwinia caratovora* was obtained from Department of Plant Pathology, Faculty of Agriculture, Ain Shams University.

The in vitro antibacterial activity of the aqueous ethanolic extract (80%) and the isolated compounds were determined by bacterial broth dilution methods described by **Ellen** *et al* **1994** against the soft rot bacterium; *Erwinia carotovera*. Minimum Inhibitory Concentrations (MICs) were determined as the lowest concentrations preventing visible growth.

## Analytical Thin Layer Chromatography (TLC)

TLC analysis was carried out on precoated silica gel plates ( $F_{245}$  0.25 mm and  $F_{245}$  2.0 mm Merck) using the following solvent systems:

1) Chloroform – Methanol – Water (70:30:5).

- 2) Chloroform Methanol (80:20).
- 3) Chloroform Methanol (90:10).
- 4) Ethylacetate- Acetic acid- Formic acid- Water (100:11:11:27)
- 5) n-Butanol- Acetic acid Water (4:1:5) upper layer

Zones were detected under UV light (255 and 365 nm) and by spraying with: concentrated  $H_2SO_4$  followed by heating at 105°C for 5 min or with modified Dragendorff reagent (**Farnsworth 1966**).

A portion of the aqueous ethanolic extract (40 g) was suspended in water (150 ml) and extracted with  $CHCl_3$  (3 x 50 ml) to give  $CHCl_3$  soluble components (Fraction A, 6.5 g). The aqueous layer was freeze dried (33.5 g) and were then extracted with  $CHCl_3$ : MeOH:  $H_2O$  (70 : 30: 5; 150 ml). After centrifugation both the supernatant and the precipitate were dried under reduced pressure to afford 4.6 g (Fraction B) and 28.8 g (Fraction C) respectively. The three fractions A, B and C were tested for their antibacterial activity against the soft rot bacterium; *Erwinia carotovora*. The bioactive fraction B (4.5 g) was subjected to the isolation of the bioactive component (s) as follows:-

Fraction B (4.5 g) was chromatographed over silica gel column (100 g, 230-200 mesh, Merck) and eluted with the solvent mixtures of  $CHCl_3$ : MeOH : H<sub>2</sub>O (80:20:0 and 70:30:5, 200 ml for each eluent). Twenty fractions of each eluent were collected. The eluates were combined on the basis of similarity of TLC profiles to afford 7 fractions and were then tested for antibacterial activity.

The bioactive fractions No. 1 and 2 were further purified several times over Sephadex LH-20, silica gel column and PTLC as shown in Fig. (1) yielded two active compounds I and II. The purity of these two compounds were established by the resolution of each one as a single spot in four different TLC systems.

## Structure identification of the isolated compounds:

The isolated compounds were characterized by detection tests and spectroscopic methods.

## **Detection tests:**

The preliminary screening of the isolated compounds for saponins, flavonoids alkaloids and phenolic compounds were performed according to the methods described by **Farnsworth (1966)**.

## **Spectroscopic methods**

# Nuclear Magnetic Resonance (NMR) spectroscopy

<sup>1</sup>H and <sup>13</sup>C-NMR spectra were recorded in  $CD_3OD$  on a varion Mercury VXP 300 (300 MHz for <sup>1</sup>H and 75 MHz for <sup>13</sup>C). Chemical shifts (ppm) were related to that of the solvent.

#### Mass spectrometry (MS)

Mass spectra were recorded on a GCMS. QP 1000 Ex. Shimadzu mass spectrometer at 70 e.v.

#### **Ultraviolet spectrometry (UV)**

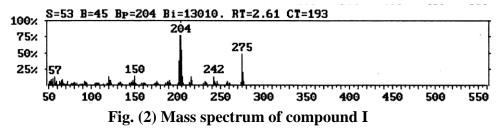
The UV-spectra were registered with a spectrophotometer CeCil 3000 series.

The aqueous ethanolic extract (80%) of the air dried leaves of *Ruta* chalepensis exhibited antibacterial activity against the soft rot bacterium, *Erwinia carotovora* (MIC: 625  $\mu$ g/ml). Bioassy- guided isolation of this aqueous extract by using chromatographic methods (see Malerials and Methods) yielded two pure compounds I (157 mg) and II (89 mg). The isolated compounds I and II exerted bactericidal activity against *Erwinia* carotovora with MIC values of 20 and 35  $\mu$ g/ml respectively. Thus, These compounds were in part responsible for the antibacterial activity of *Ruta* chalepensis leaves. The structure of these compounds (I and II) were characterized as follows:-

## **Compound I**

It was obtained as colorless neddles, which gave positive reaction with modified dragendorff reagent on TLC suggesting it is an alkaloid compound. Its structure was characterized as dihydrofuroquinolone alkaloid in accordance with the following considerations:-

The mass spectrum (Fig. 2) showed molecular ion peak ( $M^+$ ) at m/z 275 corresponding to molecular formula  $C_{15}H_{17}NO_4$ .



The <sup>13</sup>C–NMR spectrum (Fig. 3 and Table 1) showed 15 carbon atom signals out of which six carbon signals accounted for the aromatic group (between  $\delta$  97.88 to  $\delta$  154. 91 ppm). The remaining nine carbon atom signals were identified as three methyl groups ( $\delta$  22.03,  $\delta$  25.53 and  $\delta$  31.20 ppm; assigned to C-11,12 and 13 respectively), one methylene group ( $\delta$  26. 87 ppm; assigned to C-3), Oxymethine group ( $\delta$  84. 01 ppm; assigned to C-2) and four quaternary carbon atom signals including carbonyl group ( $\delta$  177.65 ppm; assigned to C-4), Two olefinic carbons ( $\delta$  109. 51 and  $\delta$  156. 15 ppm; assigned to C-3a and 9a) and Carbinol group ( $\delta$  69.11 ppm; assigned to C-10).

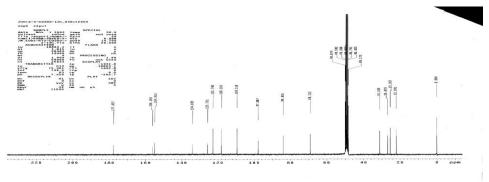


Fig.(3) <sup>13</sup>C-NMR spectrum of compound I in CD<sub>3</sub>OD Fayoum J. Agric. Res. & Dev., Vol.19, No.2, July, 2005

Table (1) <sup>13</sup> C and <sup>1</sup> H-NMR spectral data of compound I.				
C- atom No.	δС	<sup>13</sup> C	$^{1}$ H	
2	СН	84.01	3.87 t	
3	CH <sub>2</sub>	26.87	2.69, 2.97 dd	
3 a	С	109.51	-	
4	С	177.65	-	
4 a	С	118.11	-	
5	СН	97.88	7.63 d	
6	С	134.00	-	
7	СН	125.73	7.22 dd	
8	СН	122.74	7.56 d	
8 a	С	154.91	-	
9 a	С	156.15	-	
10	С	69.11	-	
11	CH <sub>3</sub>	25.35	1.45 d	
12	CH <sub>3</sub>	22.09	1.45 d	
13	N.CH <sub>3</sub>	31.20	3.75 s	

The <sup>1</sup>H-NMR spectrum of this compound (Fig. 4 and Table 1) supported the presence of substituted aromatic ring due to the three aromatic proton signals type ABX at  $\delta$  7.63 (1H, d, J = 3.0Hz, H- 5),  $\delta$  7.22 (1H, dd, J = 9.0, 3.0 Hz, H- 7) and  $\delta$  7.56 ppm (1H, d, J = 9.3 Hz, H- 8). Also the spectrum exhibited methyl proton signal at  $\delta$  3.75 ppm (3H, s) assigned to N- methyl group (Noshita *et al* 2001) and two geminal dimethyl signal at  $\delta$  1.45 ppm (6H, d, J = 3.3 Hz) assignable to the protons of C-11 and 12.

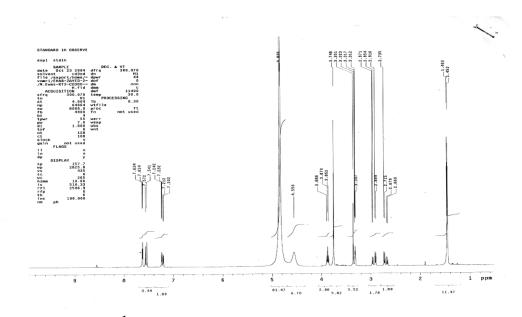


Fig. (4) <sup>1</sup>H-NMR spectrum of compound I in CD<sub>3</sub>OD

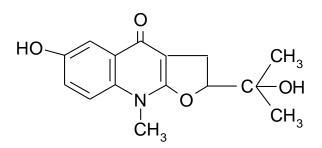
The presence of dihydrofuran ring was established by the appearance of three proton signals in the <sup>1</sup>H-NMR spectrum at  $\delta$  2.69 (1H, dd, J = 5.7, 16.8 Hz),  $\delta$  2.97 (1H, dd, J = 5.1, 16.5 Hz) and  $\delta$  3.87 ppm (1H, t) ascribed to the two protons of C-3 and the proton of C-2 (Boyd *et al.* 2000) as well as the carbon atom signals at  $\delta$  84.01,  $\delta$  26.87,  $\delta$  109. 51 and  $\delta$  156.15 ppm corresponding to C-2, 3, 3a and 9a.

The presence of hydroxy isopropyl group in the position of C-2 was established by the <sup>13</sup>C and <sup>1</sup>H-NMR spectral data due to the signals at  $\delta$  25.35,  $\delta$  22.09 and  $\delta$  69.11 ppm in <sup>13</sup>C-NMR spectrum and  $\delta$  1.45 ppm (6H, d, J = 3.3 Hz) in <sup>1</sup>H-NMR spectrum, as well as by comparing these signals with previously reported for this group (Boyd *et al.* 2000 and Noshita *et al.* 2001).

The diagnostic fragment ions of mass spectrum Fig. (2) at m/z 275 ( $M^+$ ; 47.7%), 242 ( $M^-$  CH<sub>5</sub>O; 13.2%), 204 ( $M^-$  C<sub>4</sub> H<sub>7</sub> O; 100%), 150 ( $M^-$  C<sub>7</sub> H<sub>9</sub> O<sub>2</sub>; 13.8%), 120 ( $M^-$  C<sub>8</sub>H<sub>13</sub>O<sub>2</sub>N; 12.1%) and 57 ( $M^-$  C<sub>12</sub>H<sub>12</sub>O<sub>3</sub>N) were further supported the above assigned structure. The <sup>1</sup>H and <sup>13</sup>C-NMR spectral data of this compound were similar to

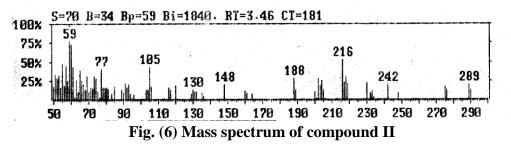
The <sup>1</sup>H and <sup>13</sup>C-NMR spectral data of this compound were similar to those of the dihydrofuroquinoline alkaloid (Ribalinium) isolated from the *Ruta graveolens* (Reisch *et al.* 1969 and Szendrei *et al.* 1969 and 1971) except the replacement of methoxyl group at C4 with carbonyl group.

From these data the structure of this compound was deduced to be 2-(hydroxy isopropyl) – 6-hydroxy- 9-methyl- dihydrofuro [2,3] – quinol – 4-one (Fig. 5). This compound was isolated for the first time from this plant.



# Fig (5) Structural formula of compound I ( $C_{15}H_{17}NO_4$ ). Compound II

It was obtained as a slightly yellow amorphous powder and gave positive colour with modified dragendorff's reagent on TLC suggesting it is an alkaloid compound. The mass spectrun (Fig. 6) of this compound showed a molecular ion peak ( $M^+$ ) at m/z 289 in accord with the molecular formula  $C_{16}H_{19}NO_4$ .



Fayoum J. Agric. Res. & Dev., Vol.19, No.2, July, 2005

#### Ahmed M. Emam and Mohamed E. Mahmoud

The <sup>1</sup>H- NMR spectrum (Fig. 7 and Table 2) showed signals for three aromatic protons formed an ABX system at  $\delta$  7.53 (1H, d, j = 2.4 Hz),  $\delta$  7.85 (1H, d, j = 9.0 Hz) and  $\delta$  7.45 (1H, d, j = 9.0 Hz) assigned to H 5, 7 and 8. The <sup>13</sup>C- NMR spectrum (Fig. 8 and Table 2) confirmed the presence of aromatic ring due to the six carbon signals between  $\delta$  95.16 to 163.43 ppm. The remaining ten carbon signals were identified as one methine group ( $\delta$  30.07 ppm; C-3), hydroxy methylene group ( $\delta$  64.29 ppm; C-12) four methyl groups (two geminal methyl at  $\delta$  24.94 and 26.02 ppm, methoxyl group at  $\delta$  60.28 ppm and N-methyl at  $\delta$  34.44 ppm) and four quaternary carbon atoms including carbonyl group ( $\delta$  171.99 ppm; C-4), two olefinic carbons ( $\delta$  108.62 and 166.12 ppm; C- 3a and 9a) and oxycarbon ( $\delta$  71.94 ppm; C-2). The presence of the four methyl groups were established by the <sup>1</sup>H-NMR spectrum due to the signals at  $\delta$  1.29 (3H, d, J= 5.7, Hz),  $\delta$  1.46 (3H, d, J= 5.7, Hz),  $\delta$ 3.60 (3H, s) and  $\delta$  4.05 ppm (3H, s) assigned to protons of C-10, 11, 13 and 14 respectively.

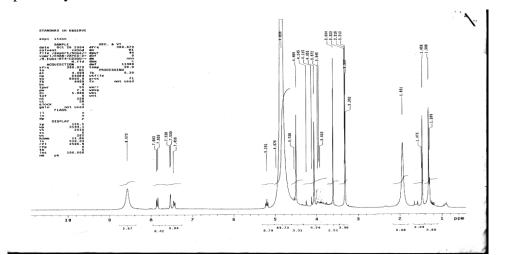
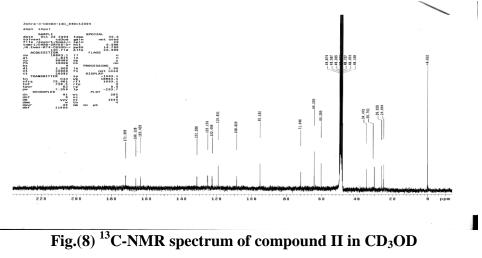


Fig. (7) <sup>1</sup>H-NMR spectrum of compound II in CD<sub>3</sub>OD



Fayoum J. Agric. Res. & Dev., Vol.19, No.2, July, 2005

C-atom No	δC	<sup>13</sup> C	$^{1}$ H
2	С	71.94	-
3	СН	30.07	5.2 t
3 a	С	108.62	-
4	С	171.99	-
4 a	С	119.03	-
5	СН	95.16	7.53 d
6	С	131.28	-
7	СН	125.15	7.85 d
8	СН	122.49	7.45
8 a	С	163.43	-
9 a	С	166.12	-
10	CH <sub>3</sub>	26.02	1.29 d
11	CH <sub>3</sub>	24.94	1.46 d
12	CH <sub>2</sub>	64.29	4.51 d
13	N-CH <sub>3</sub>	34.44	3.6 s
14	O-CH <sub>3</sub>	60.28	4.05 s

Table (2) <sup>13</sup>C and <sup>1</sup>H-NMR spectral data of compound II.

The presence of proton signal at  $\delta$  5.2 ppm (1H, t) due to the proton of methine group (C-3) and the signal at  $\delta$  4.51 (2H, d, J= 14.7, Hz) assigned to the protons of hydroxy methylene group (C-12) in the <sup>1</sup>H–NMR spectrum indicated that the two groups attached to each others.

The position of methoxyl group at C-6 was confirmed by the <sup>13</sup>C-NMR spectrum through the difference in the chemical shifts of the C-5 and C-6 in comparing with compound I and spectral data reported in the literature (**Noshita** *et al.* **2001 and Biavatti** *et al.* **2002**). Also the lack of a bathochromic shift on the addition of NaOAc in the UV spectrum indicated the absence of free hydroxyl group at the C-6 position. The presence of dihydrofuran ring was established by comparing carbon signals of this ring with previously reported (**Reisch** *et al.* **1969, Mohr** *et al.* **1982, Boyd** *et al.* **2000 and Biavatti** *et al.* **2002**). The position of the two geminal methyl groups at the C-2 were established by comparing the <sup>13</sup>C- signals with previously reported (**Boyd** *et al.* **2000**).

The diagnostic fragment ions of mass spectrum (Fig. 6) at m/z 289 (M<sup>+</sup>; 19.0%), 242 (M-  $C_2H_7O$ ; 19.0%), 216 (M-  $C_4H_9O$ ; 52.2%), 188 (M-  $C_5H_9O_2$ ; 27.7%), 148 (M-  $C_7H_{11}NO_2$ ; 19.6%), 130 (M-  $C_7H_{13}NO_3$ ; 12%), 105 (M-  $C_9H_{14}NO_3$ ; 42.4%), 77 (M-  $C_{11}H_{18}NO_3$ ; 39.7%), and 57 (M-  $C_{13}H_{14}NO_3$ ; 100%), were further supported the above assigned structure.

From these data, the structure of this compound was deduced to be 3-(hydroxy methyl)- 6-methoxy- 2, 2, 9-trimethyl-dihydrofuro [2,3] quinol -4one (Fig. 9). This compound was also isolated for the first time from this plant.

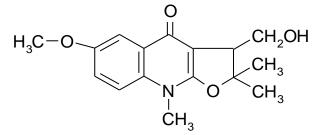


Fig. (9) Structural formula of compound II (C<sub>16</sub>H<sub>19</sub>NO<sub>4</sub>)

#### REFERENCES

- Aguilar-Santamaria L. and Tortoriello J. (1996). Anticonvulsant and sedative effects of crude extracts of *Ternstroemaic pringlei* and *Ruta chalepensis*. Phytotherapy Research 10: 531 533
- Al-Said M. S.; Tariq M.; Al-Yahya M.A.; Rafatullah S.; Ginnawi O.T. and Ageel A. M. (1990). Studies on *Ruta chalepensis* an ancient medicinal herb still used in traditional medicine. J. of Ethnopharmacology 28 : 305 – 312
- Baumert A.; Groger D.; Kuzovkina I.N. and Reisch J. (1992). Secondary metabolites produced by callus cultures of various *Ruta* species. Plant Cell Tissue and Organ Culture 28:159-162
- Biavatti M.W.; Vieira P.C.; Da Silva F.G.; Fernandes J.B.; Victor S.R.; Pagnocca F.C.; Albuquerque S.; Caracell I. and Schpector J.Z. (2002). Biological activity of quinoline alkaloids from *Echinata* and Xray structure of flindevsiamine. J. Braz. Chem. Soc. 13 (1) : 66-70
- Boyd D. R.; Sharma N. D.; Barr S. A.; Caroll J. C.; Mackerracher D. and Malon J.F. (2000). Synthesis and absolute stereochemistry assignment of enantiopure dihydrofuro and dihydropyrano-quinoline alkaloids. J. Chem. Soc. Perkin Trans I: 3397 – 3405
- Ellen J.; Peterson L.R. and Finegold S.M. (1994). Diagontive Microbiology 9<sup>th</sup> (ed.) Part 2 Chap. 14:168-188 Mosby Saint Louis USA
- El-Sayed K.; Al-Said M.S.; El-Feraly F.S. and Ross S.A. (2000). New Quinoline Alkaloids from *Ruta chalepensis*. J. Nat. Prod. 63:995–997.
- **Farnsworth N.R (1966).** Biological and phytochemical screening of plants. J. Farma. Sci. 55:225-276.
- **Fischer H.; Romer A.; Ulbrich B. and Arens H. (1988)**. A new biscoumarin glucoside ester from *Ruta chalepensis* cell cultures. Planta Med. 54 : 398 400
- **Food and Veterinary Office (2000)**. Report on a Mission carried out in Egypt From 1 to 6 February 2000 to assess the surveillance system for brown rot in potatoes for export to the EU, European commission, Health and Consumer Protection Directorate general . DG (SANCO) 1056/2000 – Mr Final 1-23

- Hadis M.; Lulu M.; Mekonnen Y. and Asfaw T. (2003). Field trials on the repellent activity of four plant products against mainly *Mansonia* population in western Ethiopia. Phytotherapy Research 17 : 202 – 205
- **Hmamouchi M.; Lahlou M. and Agoumi A. (2000).** Molluscicidal activity of some Moroccan medicinal plants. Fitoterapia 71 : 308 314
- Mahran G.E.H. (1967). Medicinal Plants P. 558, Anglo-Egyptian Book Shop, Cairo
- Mohr N.; Budzikiewicz H.; El-Tawil B. A. H. and El-Beih F. K. A. (1982). Further furoquinolone alkaloids from *Ruta chalepensis*. Phytochemistry 21:1838–1839
- Mookey K.; Young Su J.; YoungJoon A.; Dongkyu L.; HoiSeon L.; Kim M.K.; Jang Y.S.; Ahn Y.J.; Lee D.K. and Lee H.S. (2002). Larvicidal activity of Australian and Mexican plant extracts against *Aedes aegypti* and *Culex pipiens*. J. Asia Pacific Entomology 5 : 227 -231
- Noshita T.; Tando M.; Suzuki K.; Murata K. and Funayama S. (2001). New quinoline alkaloids from the leaves and stems of *Orixa japonica*, *o*rijanone, isopteleflorine and 3-o-methylorixine. Biosci. Biotechnol. Biochem. 56 (3): 710 – 713
- Reisch J.; Szendrei K.; Minker E. and Novak I. (1969). Quinoline alkaloids from *Ruta graveolens*. Pharmazie 24(11):699-700
- Szendrei K.; Reisch J.; Minker E. and Novak I. (1969). Structure of new compound separated from *Ruta graveolens*. Herba Hung. 8(1-2) :133-137
- Szendrei K.; Reisch J.; Novak I.; Simon L.; Rozsa Z.; Minker E. and Koltai M. (1971). Rutalinium, rutalinidin and other alkaloids from *Ruta graveolens*. Herba Hung 10 (2-3): 131-139
- Ulubelen A.; Ertugrul L.; Birman H.; Yigit R.; Erseven G. and Olgac V. (1994). Antifertility effects of some coumarins isolated from *Ruta chalepensis* var.latifolia in rodents. Phytotherapy Research 8:233–236.
- **Ulubelen A. and Guner H. (1988).** Isolation of dehydromoskachan C from *Ruta chalpensis* var. latifolia. J. Nat. Prod. 51:1012-1013
- **Ulubelen A.; Guner H. and Cetindag M. (1988)**. Alkaloids and coumarins from the roots of *Ruta chalepensis* var. latifolia. Planta Med. 54: 551-552.
- Ulubelen A. and Tan N. (1990). A moskachan from roots of *Ruta* chalepensis. Phytochemistry 29:3991-3992
- Ulubelen A. and Terem B. (1988). Alkaloids and coumarins from roots of *Ruta chalepensis*. Phytochemistry 27:650-651
- Ulubelen A.; Terem B.; Tuzlaci E.; Cheng K. F. and Kong Y. C. (1986). Alkaloids and coumarins from *Ruta chalepensis*. Phytochemistry 25 : 2692 – 2693.
- Zobel A.M.; Brown S.A. and Glowniak K. (1990). Localization of furanocoumarins in leaves, fruits and seeds of plants. Planta Med. 56:571-572

### Ahmed M. Emam and Mohamed E. Mahmoud

النشاط المضاد لمركبان من قلويدات الفيوروكوينولون المستخلصة من اوراق نبات الروتا كاليبنسس ضد بكتريا العفن الطرى للبطاطس ايرونيا كاروتوفورا

> **احمد معوض امام' ، محمد عويس محمود'** ١- قسم الكيمياء الحيوية – كلية الزراعة – جامعة القاهرة – فرع الفيوم. ٢- قسم النبات "الميكروبيولوجي" كلية العلوم – جامعة القاهرة – الجيزة.

يعتبر العفن الطرى الذى تسببه بكتريا "ايرونيا كاروتوفورا" من اكثر الامراض خطورة على درنات البطاطس بعد حصادها على النطاق العالمى وذلك اثناء عمليات التخزين والنقل والتسويق. ونظرا لان استخدام مبيدات البكتريا المخلقة صناعيا في مكافحة هذا المرض تؤدى الى تأثيرات ضارة على الانسان و البيئة علاوة على خفض كمية البطاطس المصدرة للاسواق الاوربية لذا فقد زاد الاهتمام بالمنتجات الطبيعية ذات الفعالية لاستخدامها ضد هذه البكتريا المسببة للمرض. وفى هذه الدراسة اظهر مستخلص الايثانول المائى (٥٠%) لاوراق نبات الروتا كاليينسس فعالية ضد هذه الدراسة اظهر مستخلص الايثانول المائى (٥٠%) لاوراق نبات الروتا كاليينسس فعالية ضد عملية الفصل المقرونة باختبار الفعالية للمكونات المفصولة باستخدام طرق التحليل الكروماتوجرافي (الطبقة الرقيقة و الاعمدة) عن فصل مركبان فعالان من قلويدات الفيوروكونيولون لأول مرة من هذا النبات وتم التعرف على التركيب الكيميائى لكلا منهما باستخدام طرق التحليل الطيفى (الرنين رالطبقة الرقيقة و الاعمدة) عن فصل مركبان فعالان من قلويدات الفيوروكونيولون لأول مرة من هذا المغناطيسى وتقدير الكتلة) حيث وجد أنهما ٢- (هيدروكسى ايزوبروبيل) ٦- هيدروكسى – ٩-ميثيل تلاثى ميثيل حايهيدروفيورو (٢، ٣) حكوينول ٤-اون (II). وكانت فعالية هذان المرين تلاثى ميثيل حايهيدروفيورو (٢، ٣) حكوينول ٤-اون (II). وكانت فعالية هذان المركبان ضد هذه البكتريا 2010 تساوى ٢، ٥٠ ميكروجرام/مل على الترتيب.