





ISSN: 2357-0210

VOLUME 4, ISSUES 1, 2018

Antitubercular Activity of Some Substituted Phenothiazine Derivatives

Mohammad Asif*

Department of Pharmacy, GRD (P.G) Institute of Management & Technology, Dehradun, (U.K), 248009, India.

Received: 12 Nov. 2017, Revised: 23 Dec. 2017, Accepted: 27 Dec. 2017. Published online: 1 May 2018.

Abstract: In search of newer and potent antitubercular agents, a series of phenothiazine derivatives were evaluated for antitubercular activity. This study may prove to be helpful in development and optimization of existing antitubercular activity of this class of compounds.Quaternized chlorpromazine, triflupromazine, and promethazine derivatives were tested as antitubercular agents against both actively growing and non-replicating Mycobacterium tuberculosis H37Rv. A slight variation in the substitution pattern on the phenothiazine nucleus often causes a marked difference in activities and therefore phenothiazines with various substituents are being synthesized and tested for activities in search of better medicinal agents.

Keywords: Phenothiazines, antitubercular, M. tuberculosis, Promethazine; Chlorpromazine.

1 Introduction

The chemistry of nitrogen-sulfur heteroatom containing aromatic compounds is becoming more popular as an area of research. From medicinal chemistry perspective, phenothiazines are an important group of condensed threering heterocycles (Gupta and Kumar.1988). Phenothiazine derivatives and their analogues containing 1,4-thiazine structural fragment show diverse biological activities including as tranquilizers (El-Said.1981), anti-inflammatory (Sharma et al., 2005; Tilak, et al., 1998), antimalarial (Dominguez, et al., 1997), antipsychotropic (Lin, et al., 1991), antimicrobial (Raval, and Desai 2005;Kaatz et al., 2003), antitubercular (Viveros and Amaral. 2001; Amaral and Kristiansen. 2000) antitumour (Motohashi et al., 2000; Motohasho, et al., 1999; Kurihara, et al., 1996; Kurihara, et al., 1999) and stimulation of the penetration of anticancer agents via the blood-brain barrier (BBB). However, solid cancers of the brain and stomach are generally resistant to chemotherapeutic agents (Ghosh and Chattopadhyay. 1993). Phenothiazines are inexpensive and widely available, and therefore have been examined as antitubercular drugs. It has been reported (Floyd, et al., 1993) that some phenothiazines inhibit intracellular replication of viruses including human immunodeficiency viruses (HIV). Furthermore, some of these derivatives have been reported to exhibit significant anti-TB activities (Viveros and Amaral. 2001; Amaral and Kristiansen. 2000) and great interest has arisen in the design and synthesis of

new phenothiazines to explore their antitubercular activities. Phenothiazine derivatives that contain aminoalkyl substituents at the thiazine nitrogen atom are used as antipsychotropic and antihistamine drugs (Isaacson. 1998). Extensive search has been conducted regarding new methods of synthesizing potentially useful phenothiazine analogs having pharmacological activities (Ziĺba et al., 2010).

Tuberculosis (TB), the disease caused by Mycobacterium tuberculosis (Mtb), infects about two billion people. The WHO estimates that about two million people die each year from TB due to the lack of inability to afford proper health care (Cande, et al., 2009; Maher, and Raviglionem. 1999).Overcrowding and ill-nourishment of poor people living in large cities leads to a high incidence of the disease due to the ease at which the infection can be transferred (Lowell. 1999). This situation contributes to the accelerated speed at which TB spreads in underdeveloped countries. TB has become a serious worldwide problem, infecting in synergy with human immunodeficiency virus (HIV) infection (Upadhayaya, et al., 2009). There is also an alarming increase in cases of TB caused by multidrugresistant tuberculosis (MDR-TB), due in part to inadequate drug therapy as a result of incorrectly selected medications or suboptimal drug dosing (Savini, et al., 2002; Bearing, et al., 1999).WHO declared TB as a global health emergency and aimed at saving 14 million lives between 2006 and 2015 (Eswaran, et al., 2010). TB is difficult to treat due to residence of bacteria within the macrophages and its unusual cell wall barrier. Moreover, MDR-TB and extensively drug resistant (XDR) TB have emerged recently (Bairwa, et al., 2010). One of the most important issues in current medical practice is antibiotic-resistant bacterial infections (Chambers. 1997; Livermore. 2000; CDC. 2002). Their pervasiveness justifies the search for innovative antimicrobial agents featuring novel chemical structures and mechanisms of action, helpful in combating infections (Fung et al., 2001; Wright and Sutherland. 2007; Girdhar et al., 2010; Ziĺba et al., 2010). In this article, describe some phenothiazine derivatives as antitubercular agent (Zieba et al., 2012).

Thus, there is a need for new drugs targeting enzymes essential to mycobacterial survival. One such target is type II NADH-menaquinone dehydrogenase (ndh-2). By inhibiting ndh-2, the electron transport chain in Mtbbecomes blocked and shuts down (Weinstein, et al., 2005). Ndh-2 is also found in a number of other bacteria such as Staphylococcus aureus and Enterococcus faecalis but is not expressed in humans (Melo, et al., 2004). Humans rely only on type I NADH dehydrogenase (ndh-1) and thus minimal toxicity in humans is predicted with ndh-2 inhibitors. The Mtbis an obligate aerobe that is capable of long-term persistence under conditions of low oxygen tension. Analysis of the Mtbgenome predicts the existence of a branched aerobic respiratory chain terminating in a cytochrome bdsystem and a cytochrome aa3 system. Both chains can be initiated with type II NADH: Menaquinoneoxidoreductase. Α biochemical characterization of the aerobic respiratory chains from Mtb and show that phenothiazine analogs specifically inhibit NADH: Menaquinoneoxidoreductase activity. Type-II NADH-Menaquinoneoxidoreductase(NDH-2) is an essential respiratory enzyme of the pathogenic bacterium Mtbthat plays a vital role in its growth (Miesel, et al., 1998; Yano, et al., 2006; Melo, et al., 2004; Amaral, et al., 2007; Agarwal et al., 2013).

In the 1950s and 1960s psychiatrists noticed a more pronounced inhibition of Mtb occurring in TB-infected, schizophrenic patients taking higher doses of an antipsychotic drug, chlorpromazine, as opposed to patients who were taking lower doses of this drug (Kardos, et al., 1964). The N-(benzyl)-chlorpromazinium inhibited Mtb in vitro at an even lower concentration than chlorpromazine itself. This was the first quaternizedpromazine derivative (QPD) discovered to be an antibacterial agent against Mtb. Chlorpromazine and its QPD were both found to selectively inhibit ndh-2; there was no inhibition of ndh-1 (Yano, et al., 2006). Other phenothiazines were also described to have anti-TB activity (Ratnakar, et al., 1995; Gadre, and Talwar. 1999).

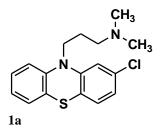
Biological activities of Phenothiazine derivatives: Phenothiazine is also a bioactive heterocyclic compound of pharmaceutical importance and possesses different biological activities viz. antibacterial (Rawat and Srivastava 1998; Trivediet al., 2008), antifungal (Weng and Tan. 2003), antitubercular (Rajasekaran and Tripathi. 2003), and anti-inflammatory (Kasmi-Mir, et al., 2006) etc.

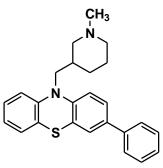
Antituber activities of Phenothiazine derivatives: Phenothiazines have been reported for their antitubercular(anti-TB) activity for many years, and the phenothiazine drug chlorpromazine (CPZ) (1a) is reported to have been successfully used to treat a TB patient. In this concern, aseries of psychotropicphenothiazines were tested as anti-TB agents againstM. tuberculosis (Mtb)H37Rv. Among all, three compounds (1b-d) exhibited promising activity with a mean MIC of 2.13µg/mL (Madridet al, 2007). Whereas quaternized CPZ,triflupromazine (2a) and promethazine (2b) derivatives inhibited non-replicating Mtbat concentrations equal to or double their MICs against the actively growingstrain. All the active compounds (2c-f) were non-toxic toward Vero cells (IC50>128µM). The or substituted benzyl benzyl groups, anelectron withdrawing substituent on the phenothiazine ring improved the potency. The optimum anti-TB structures possessed N-(4- or 3-chlorobenzyl)substitution on triflupromazine(Bate et al, 2007).

Some 2-heterocycle-substituted phenothiazines having a pyrazolo[3,4-d]pyrimidines were tested for their anti-TB activity againstMtbH37 Rv (Trivedi et al., 2008).Several pyrimidine derivatives containing a phenothiazine ring, 3methyl-1-(10H-phenothiazin-2-yl)-4-phenyl-6-hydroxv-4,5-dihydro-1H-pyrazolo[3,4d]pyrimidines(3a-h)3-Methyl-1-(10H-phenothiazin-2-yl)-4-phenyl-6hydroxy-4.5dihydro-1H-pyrazolo[3,4-d] pyrimidine (3a), 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(2-hydroxyphenyl)-6-hydroxy-4,5-dihyd-ro-1Hpyrazolo[3,4-d]pyrimidine (3b), 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(4-hydroxyphenyl)-6hydroxy-4,5-dihydro-1H-pyrazolo[3,4-d]pyrimidine (3c). 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(2-chlorophenyl)-6hydroxy-4,5-dihydro-1H pyrazolo[3,4-d]pyrimidine (3d), 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(4-chlorophenyl)-6hvdroxy-4.5-dihvdro-1H-pvrazolo[3.4-d]pvrimidine (3e), 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(2-nitrophenyl)-6hydroxy-4,5-dihydro-1H-pyrazolo[3,4-d]pyrimidine (3f), 3-Methyl-1-(10H-phenothiazin-2-yl)-4-(3-nitrophenyl)-6hydroxy-4,5-dihydro 1H-pyrazolo[3,4-d]pyrimidine (3g) and3-Methyl-1-(10H-phenothiazin-2-yl)-4-(4-methoxyphenyl)-6-hydroxy-4,5-dihydro-1H-pyrazolo[3,4-d] pyrimidine (3h) were testedat >6.25 µg/ml against MtbH37Rv in BECTEC 12B medium. The anti-TB activities in Table 1.

The anti-TBactivity, all the compounds showed mild to moderate activity. Compounds 3c, 3d and 3e were found to be predominantly active againstMtbH37Rv strain (Trivedi eta l., 2008).

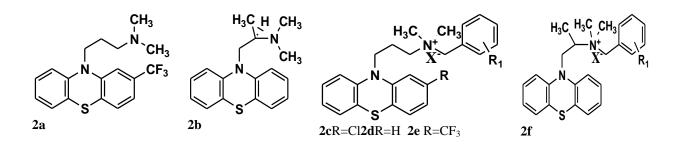
Several phenothiazine analogs inhibited non-replicating Mtb at concentrations equal to or double their MICs against the actively growingstrain. All active compounds were non-toxic toward Vero cells (IC50>128IM).







1b



2	SNo.	R	1	2	3	4	5
CH ₂ 3 D	3a	Η	Н	Н	Н	Н	Н
$\Gamma_{3} = \Gamma_{1} - R$	3b	2-OH	OH	Н	Н	Н	Н
	3c	4-OH	Н	Н	OH	Н	Η
	3d	2-C1	Cl	Н	Н	Н	Η
	3e	4-C1	Н	Н	Cl	Н	Н
	3f	$2-NO_2$	NO_2	Н	Н	Н	Η
	3g	$3-NO_2$	Н	NO_2	Н	Н	Н
OH	3h	4-OCH ₃	Η	Н	OCH ₃	Н	Η

Compounds 3a-h

Table 1	.Antitubercular	activity	of 3a-h

SNo.	R	MIC	% Inh	Activity	SNo.	R	MIC	% Inh	Activity
3 a	Н	>6.25	79	-	3e	4-CI	<6.25	94	+
3b	2-OH	>6.25	75	-	3f	2-NO ₂	>6.25	74	-
3c	4-OH	< 6.25	94	+	3g	3-NO ₂	>6.25	77	-
3d	2-C1	< 6.25	92	+	3h	4-OCH ₃	>6.25	63	-

N-Allylchlorpromazinium bromide was only weaklyanti-TB, but replacing allyl with benzyl or substituted benzyl improved potency. An electron-withdrawing substituent onthe phenothiazine ring was also essential. Branching at the carbon chain decreased anti-TB activity. The optimum anti-TBstructures possessed N-(4- or 3-chlorobenzyl) substitution on triflupromazine (Bate et al., 2007).

The quaternized derivatives of promazine, chlorpromazine, and triflupromazine, and to measure their MICs againstboth actively growing and non-replicating Mtb.

The minimum inhibitory concentration (MIC) of QPDs against actively growing Mtb H37Rv was determined using the microplate alamarblue assay (MABA) (Collins and Franzblau. 1997; Falzari, et al., 2005). The activities of three compounds (4c, 6c, and 7c)were confirmed for both and LORA using acolony-forming MABA unit determination by subculturingfrom the microplate onto drug-free 7H11 agar. MICsfor actively growing Mtb were 1.9-, 2.1-, and 2.9-fold higher than the MABA MICs, and 1.7-,1.9-, and 2.1-fold higher than the correspondingLORA MICs.The in vitro cytotoxicity for VERO cells was determined for all compounds with a MABA MIC of lessthan 10 lM using a dye reduction assay following 3days exposure to test compounds as previously described. The IC50 of all compounds was >128 lM(except for 3b which was >64 lM, the highest concentrationtested).

From the MICs in Table 2, N-benzyl substitution inQPDs is a requirement for significant anti-TBactivity (1a vs. 2). Alkyl chain branching (8a, 8f–h)decreases potency, three of the QPDs havingMICs both<4 IM against actively growing Mtb and <8 IM againstnon-replicating Mtb possess N-(4- or 3-chlorobenzyl)groups and electron-withdrawing substituents on thephenothiazine ring (4c, 7b–c). Based on this MIC dataand the lack of in vitro mammalian cell toxicity, we willattempt to improve these leads by studying the anti-TBpotency of other electron-withdrawing substituentson the phenothiazine ring and various halogen substitutions on the benzyl group. If these improvedleads also lack mammalian cell toxicity, animal studieswill be warranted (Bate et al., 2007).

Aseries of 10-(3-chloropropyl)-10Hphenothiazine 9, N-[3-(10H-phenothiazin-10-yl)propyl]urea10, N-[3-(10H-phenothiazin-10-yl)propyl]-N'-[(substituted

phenyl)methylidene]urea, 11a–s, N-[3--(10H-phenothiazin-10-yl)propyl]-2-(substituted phenyl)-4-oxo-3thiazolidinecarboxamide13a–sand N-[3-(10H-phenothiazin-10-yl)propyl]-2-(substituted phenyl)-4-oxo-5-(substituted benzylidene)-3-thiazolidinecarboxamide, 13a–s were tested for their anti-TB activity against the Mtb(Sharma et al., 2012).All the compounds were exhibited anti-TB activity against Mtb.

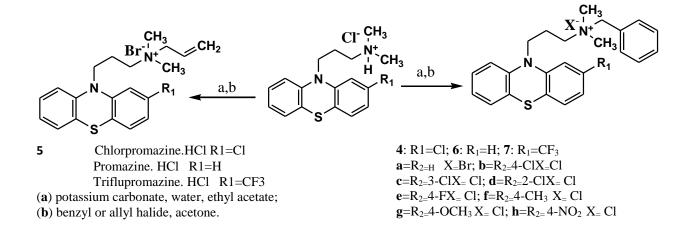
The anti-TB activities of compounds 9, 10, 11a–s, 12a–s and 13a–s were assayed in vitro against MtbH37Rv strain(Sharma, et al., 2011)at 25 and 50 μ g mL-1 and lower concentrations. For the anti-TB activity, isoniazid and

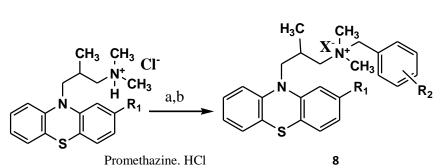
rifampicin were taken as standards.The compounds 13c, 13d, 13e, 13f, 13h, 13i and 13j displayed high activity, compounds 12h, 12j, 13b, 13g and 13q showed moderate

activity and the other compounds showed less activity against all the strains compared with standard drugs. The activity of compounds varies with substitution. The nitro group-containing compounds 13h, 13i and 13j showed higher activity than the chloro group- (5c and 13d) or the bromo group-containing compounds (13e and 13f). The chloro- and bromo-derivatives also had a higher activity than the other tested compounds.

It could be concluded that the activity of compounds depended on the electron withdrawing nature of the substituent groups (Sharma et al., 2012). The sequence of the activity is following: $NO_2 > Cl > Br > OCH_3 < OH > CH_3$.Standards drugs, isoniazid and rifampicin, showed 100 % activity at both tested concentrations

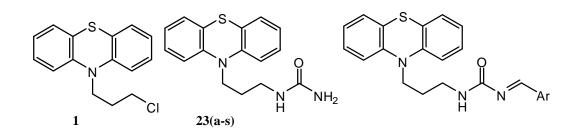
The compounds 9, 10, 11a–s, 12a–s and 13a–ssome compounds displayed good biological activities while the others showed lower anti-TB activities.





1 Tomethazh	10.1101
Tabla 2	OPD antit

Table 2. QPD antitubercular activity								
Compound (D.)	MIC (SD) in l tuberculos		Compound (D.)	MIC (SD) in IM versus M. tuberculosis that is				
Compound (R ₂)	Actively growing	Non- replicating	Compound (R ₂)	Actively growing	Non- replicating			
4a (H)	7.33(0.4)	11.1(2.4)	6e (4-F)	14.6	32.9			
4b (4-Cl)	6.06(1.9)	6.7(0.9)	6f (4-CH ₃)	9.9(2.0)	15.0(0.2)			
4c (3-Cl)	4.5(1.3)	6.7(0.4)	7b (4-Cl)	3.81(0.1)	6.1(0.4)			
4d (2-Cl)	5.6(1.8)	7.6(0.3)	7c (3-Cl)	3.8(0.1)	5.8(0.9)			
4e (4-F)	7.6(0.2)	13.7(0.9)	7d (2-Cl)	7.3(0.3)	7.5(0.2)			
4f (4-CH ₃)	4.7(1.0)	7.5(0.3)	7e (4-F)	6.4(2.2)	10.0(1.5)			
4g (4-OCH ₃)	8.5(1.3)	11.9(1.2)	7f (4-CH ₃)	6.8(0.2)	6.9(0.6)			
4h (4-NO ₂)	12.3(4.0)	27.5(1.6)	8a (H)	31.6	99.7			
5	30.6	105.9	8f (4-CH ₃)	12.1	34.7			
6b (4-Cl)	9.31(2.2)	15.4(3.2)	8g (4-OCH ₃)	14.4	105.2			
6c (3-Cl)	7.5(0.3)	13.0(0.6)	8h (4-NO ₂)	20.7	>128.0			
6d (2-Cl)	14.3	24.6						



	S N N	O Ar	S N N	N H S	— Ar ₁
		4(a-s)	5(a-s)		
Compound	$Ar = Ar_1$	Compound	Ar = Ar1	Compound	Ar = Ar1
11a, 12a, 13a	C_6H_5	11h, 12h, 13h	$4-NO_2C_6H_4$	110, 120, 130	$3-CH_3OC_6H_4$
11b, 12b, 13b	$4-ClC_6H_4$	11i, 12i, 13i	$3-NO_2C_6H_4$	11p, 12p, 13p	$2-CH_3OC_6H_4$
11c, 12c, 13c	$3-ClC_6H_4$	11j, 12j, 13j	$2-NO_2C_6H_4$	11q, 12q, 13q	$4-HOC_6H_4$
11d, 12d, 13d 11e, 12e, 13e 11f, 12f, 13f 11g, 12g, 13g	2-ClC ₆ H ₄ 4-BrC ₆ H ₄ 3-BrC ₆ H ₄ 2-BrC ₆ H ₄	11k, 12k, 13k 11l, 12l, 13l 11m, 12m, 13m 11n, 12n, 13n	4-CH ₃ OC ₆ H ₄ 3-CH ₃ OC ₆ H ₄ 2-CH ₃ OC ₆ H ₄ 4-CH ₃ C ₆ H ₄	11r, 12r, 13r 11s, 12s, 13s	3-HOC ₆ H ₄ 2-HOC ₆ H ₄

Table3. Antitubercular activity of compounds 1, 2, 3a–s, 4a–s and 5a–s

Compd	Compd Anti-TB activity, %		Comp		activity, %	Compd	Anti-TB activity, %		
1	MtbH3	7Rv strain	d	MtbH37Rv strain		1	MtbH37Rv strain		
	<i>c</i> /µg	g mL-1	<i>c</i> /μg mL		mL-1		<i>c</i> /µg 1	nL-1	
9	13	20	10	10	18	13a	22	45	
11a	18	22	12a	20	35	13b	32	74	
11b	25	32	12b	25	55	13c	36	80	
11c	27	34	12c	30	60	13d	32	80	
11d	30	35	12d	30	60	13e	30	78	
11e	28	40	12e	30	68	13f	30	79	
11f	27	50	12f	32	70	13g	29	76	
11g	25	52	12g	30	75	13h	32	82	
11h	32	65	12h	30	70	13i	27	83	
11i	35	68	12i	32	68	13j	28	81	
11j	38	66	12j	35	70	13k	28	60	
11k	25	40	12k	30	50	13l	30	63	
111	28	42	12l	32	53	13m	31	65	
11m	23	43	12m	30	50	13n	22	45	
11n	20	38	12n	29	41	130	18	49	
110	24	35	120	28	42	13p	20	47	
11p	25	38	12p	30	45	13q	24	76	
11q	28	50	12q	33	70	13r	27	70	
11r	30	52	12r	34	70	13s	25	65	
11s	32	55	12s	33	65				

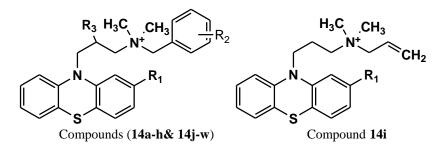


Table 4.Structure and Antitubercular activity data of compounds (Agarwal et al., 2013).

S No.	R ₁	\mathbf{R}_2	R ₃	MIC	S No.	R ₁	\mathbf{R}_2	R ₃	MIC
14a	Cl	Н	Н	7.33	14m	Н	p-F	Н	14.6
14b	Cl	p-Cl	Н	6.06	14n	Н	p-CH ₃	Н	9.9
14c	Cl	m-Cl	Н	4.5	140	CF ₃	p-Cl	Н	3.81
14d	Cl	o-Cl	Н	5.6	14p	CF ₃	m-Cl	Н	3.8
14e	Cl	p-F	Н	7.6	14q	Н	p-Cl	Н	7.3
14f	Cl	p-CH ₃	Н	4.7	14r	CF ₃	p-F	Н	6.4
14g	Cl	p-OCH ₃	Н	8.5	14s	CF ₃	p-CH ₃	Н	6.8
14h	Cl	p-NO ₂	Н	12.3	14t	Н	Н	CH ₃	31.6
14i				30.6	14u	Н	p-CH ₃	CH ₃	12.1
14j	Н	p-Cl	Н	9.3	14v	Н	p-OCH ₃	CH ₃	14.4
14k	Н	m-Cl	Н	7.5	14w	Н	p-NO ₂	CH ₃	20.7
14l	Н	o-Cl	Н	14.3					

4 Conclusions

This study may prove to be helpful in development and optimization of existing anti-TB activity of phenothiazine compounds.Development of new chemotherapeutic drugs is the need of the hour to improve TB control. In the last fourty years no new compound has been brought to the market for the treatment of TB. However, in recent years there is an enhanced activity in the research and development of new drugs for TB. Some compounds are presently in clinical development, while others are being investigated pre-clinically in an attempt to explore new molecules for the target based treatment of TB. Simultaneously some new targets are being identified and validated for their practical usefulness. The review provides an overview of the drugs that are being used and the compounds that are in clinical or preclinical studies and also attempted to highlight the efforts that are being made the development of new molecules.Various in phenothiazine analogs were developed. Other compounds of this group are presently under investigation. The phenothiazines nucleus, which has a useful structure for further molecular exploration for the development of new derivatives with different biological activities, has received much attention in recent years.

References

- Amaral L, Kristiansen. Int. J. Antimicrob. Ag. 2000, 14, 173.
- [2] Amaral L, Martins M, Viveiros M.J Antimicrob Chemother,2007, 59, 1237-1246.
- [3] Bairwa R, Kakwani M, Tawari NR, Lalchandani J, Ray MK, Rajan MGR, Degani MS.Bioorg Med Chem Lett,2010, 20, 1623-1625.
- [4] Bate BA, Kalin JH, Fooksman EM, Aramose EL, Price CM, Williams HM, Rodig MJ, Mitchell MO, Cho, SH, Wang Y, Franzblau SG.Bioorg Med Chem Lett,2007, 17, 1346-1348.
- [5] Bearing SE, Peloquin CA, Patel KB. In Tuberculosis and Nontuberculous Mycobacterial Infections; Schlossberg, D, Ed, Fourth ed, Saunders: Philadelphia, 1999; p 83.
- [6] Cande ALP, FerreiraMDL, Pais KC, Cardoso LNDF, Kaiser CR, Henriques MDGMO, Lourenc MCS, Bezerra FAFM, Souza MVND, Bioorg Med Chem Lett,2009, 19, 6272-6274.
- [7] CDC. Staphylococcus aureus resistant to vancomycin-United States. MMWR, 2000, **51**, 565.
- [8] Chambers HF. Clin. Microbiol. Rev.1997, **10**, 781.

- [9] Collins L, Franzblau SG. Antimicrob. Agents Chemother. 1997, **41**, 1004.
- [10] Dominguez JN, Lopez S, Charris J, Iarruso L, Lobo G, SemenowA, Olson JE, Rosenthal PJ. J. Med. Chem. 1997, 40, 2726.
- [11] El-Said MK. Pharmazie 1981, 36, 678.
- [12] Eswaran S, Adhikari AV, Kumar RA, Eur J Med Chem 2010, 45, 957–966.
- [13] Falzari K, Zhu Z, PanD, LiuH, Hongmanee P, Franzblau SG. Antimicrob. Agents Chemother. 2005, 49, 1447.
- [14] Floyd RA, Scheider JE, Zhu YQ, North TW, Schinazi F. Proc. Am. Assoc. Cancer. Res. 1993, 34, 359.
- [15] Fung HB, Kirschenbaum HL, Ojofeitimi BO. Clin. Ther. 2001, 23, 356.
- [16] Gadre DV, Talwar VJ. Chemother. 1999, 11, 203.
- [17] Ghosh N, Chattopadhyay U. In Vivo 1993, 7, 435.
- [18] Girdhar A, Jain S, Jain N, Girdhar S. Acta Pol. Pharm. Drug Res. 2010, 67, 211.
- [19] Gupta RR, Kumar M. Synthesis, properties and reactions of phenothiazines, in, Phenothiazines and 1,4-Benzothiazines: Chemical and Biomedical Aspects, Gupta R.R. Ed, pp. 1-146, Elsevier, Amsterdam 1988.
- [20] Isaacson EI. Central nervous system depressants, in Wilson and Gisvoldís Textbook of Organic Medicinal and Pharmaceutical Chemistry, 10 edn, Delgado J.N, Remers W.A. Eds, pp. 435-461, Lippincott-Raven Publishers, Philadelphia 1998.
- [21] Kaatz GW, Moudgal VV, Seo SM, Kristiansen JE. Antimicrob. Agents Chemother. 2003, **47**, 719.
- [22] Kardos G, Boszormenyi Z, Vamos G. Intern. Congr. Chemother. Proc, 3rd, Stuttgart 1964, 194.
- [23] Kasmi-Mir S, Djafri A, Paquin L, Hamelin J, Rahmouni M.Molecules 2006, **11**, 597.
- [24] KuriharaT, Motohasho N, Pang GL, Higano M, Kiguchi K, Molnar J. Anticancer Res. 1996, **16**, 2757.
- [25] Kurihara T, Nojima K, Sakagami H, MotohashiN, Molnar J. Anticancer Res 1999, **19**, 3895.
- [26] Lin G, Midha KK, Hawes EM. J. Heterocycl.Chem. 1991, 28, 215.
- [27] Livermore DM. Int. J. Antimcrob. Agents. 2000,16, 3.
- [28] Lowell AM. In Tuberculosis and Nontuberculous Mycobacterialc Infections; Schlossberg, D, Ed, fourth ed, Saunders: Philidelphia, 1999; p 3.
- [29] Madrid PB, Polgar WE, Toll L, Tangaa MJ. Bioorg. Med. Chem. Lett. 2007, 17(11), 3014-3017.

- [30] Maher D, RaviglionemMC. In Tuberculosis and Nontuberculous, Mycobacterial Infections; Schlossberg, D, Ed, fourth ed, Saunders: Philadelphia, 1999; p 104.
- [31] Melo AMP, Bandeiras TM, Teixeira M, Microbiol Mol Biol Rev 2004, **68**, 603-616.
- [32] Miesel L, Weisbrod TR, Marcinkeviciene JA, Bittman R, Jacobs WR, J Bacteriol 1998, 180, 2459-2467.
- [33] Motohasho N, Kawase M, Saito S, Sakagami H. Curr. Drug Targets 2000, 1, 237.
- [34] Motohasho N, Kurihara T, Satoh K, Sakagami HH, Mucsi I, Pusztai R, Szabo M, Molnar J. Anticancer Res. 1999, 19, 1837.
- [35] Rajasekaran A, Tripathi PP.Acta Pharm. Turc. 2003, 45, 235.
- [36] Ratnakar P, RaoSP, Sriramarao P, Murthy PS. Int. Clin. Psychopharmacol. 1995, **10**, 39.
- [37] Raval JP, Desai KR, Arkivoc 2005, 21.
- [38] Rawat TR, Srivastava SD.Indian J. Chem, 1998, B 37, 91.
- [39] Savini L, Chiasserini L, Gaeta A, Pellerano C.Bioorg Med Chem 2002, 10, 2193-2198.
- [40] Sharma R, Samadhiya P, Srivastava SD, Srivastava SK.Acta Chim. Slov. 2011, 58, 110.
- [41] Sharma R, Samadhiya P, Srivastava SD, Srivastava SK. J. Serb. Chem. Soc. 2012, 77 (1), 17–26.
- [42] Sharma S, Srivastava VK, Kumar A. Pharmazie2005, 60, 18.
- [43] Tilak SR, Tyagi R, Goel B, Saxena KK. Indian drugs 1998, 35, 221.
- [44] Trivedi AR, Siddiqui AB, Shah VH. Arkivoc,2008, (ii) 210-217.
- [45] Upadhayaya RS,Vandavasi JK,Vasireddy, NR,SharmaV, Dixit SS, Chattopadhyaya J.Bioorg Med Chem 2009, 17, 2830-2841.
- [46] Viveros M, Amaral L. Int. J. Antimicrob. Ag. 2001, 17, 225.
- [47] Weinstein EA, Yano T, Li LS, Avarbock D, Avarbock A, HelmD, McColm AA, Duncan K, Lonsdale JT, Rubin H.Proc Natl Acad Sci USA 2005, 102, 4548-4553.
- [48] Weng F, Tan J.Acta Pharmacol. Sin. 2003, 24, 1001.
- Wright GD, Sutherland AD. Trends Mol. Med. 2007, 13, 260.
- [49] Yano T, Li SL, Weinstein E, Teh J. S, Rubin H.J Biol Chem 2006, 281, 11456-11463.

- [50] Zieba A, Czuba ZP, Krol W. Acta Pol Pharm-Drug Res, 2012, 69(6), 1149-1152.
- [51] ZiÍba A, Sochanik A, Szurko A, Rams M, Mrozek A, Cmoch P. Eur. J. Med. Chem.2010, 45, 4733.
- [52] Ziĺba A, Wojtyczka R.D, Kĺpa M, Idzik D. Folia Microbiol. 55, 3 (2010).