



The effect of clindamycin oral administration on some immunological and biochemical parameters of Newcastle vaccinated broilers

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ABSTRACT

Present work aimed to study some immunological and biochemical effects of Clindamycin on broiler vaccinated against Newcastle virus. Three groups of one day old Hubbard chicks each of 50 birds were used in this study. Group (1) was control non treated group (CG). Group C20 and C40 were given Clindamycin (20mg/kg and 40mg/kg body. wt) respectively in drinking water at 14th, 15th and 16th days of age. All groups were routinely vaccinated against Newcastle virus at 7 and 18 day of age. On the 25th day of age, blood samples were collected from ten birds of each group for immunological and biochemical tests. Five birds from each group were slaughtered at the same age then the lymphoid organs (thymus, bursa and spleen) were carefully separated & weighed and each organ relative weight was determined. Number of dead birds was recorded throughout the experiment period (45 day age). It was observed that administration of Clindamycin decreased the leukocytic count haemagglutination inhibition antibody titer (HI), total protein, globulin and the relative weight of bursa in C20 and C40. It did not induce any significant changes in the differential leukocytic count, liver and kidney function tests, spleen relative weight in C20 and C40. While, relative weight of thymus and body weight gain significantly decreased in C40 and non-significantly changed in C20. While, feed conversion rate significantly increased in C40 only. In addition, C20 and C40 had higher mortality rates than CG. Conclusion: Clindamycin administration suppresses the chicken's immune response to Newcastle disease virus vaccine.

Keywords: Newcastle vaccine, immunological parameters, biochemical parameters, broilers.

1. Introduction

Newcastle disease is an obstacle hinders the poultry industry development all over the world. Avian paramyxovirus attacks the respiratory system of the bird causing several symptoms as gasping, coughing, sneezing and rales (Brown and Bevins 2017; Desouky et al., 2020). Sometimes it causes nervous symptoms like tremors and complete paralysis. In-appetence and watery greenish diarrhea are general symptoms also related to the disease. The disease usually caused high morbidities and mortalities (Brown and Bevins 2017; Desouky et al., 2020). Moreover, ND virus can live under the bird skin and resist the freezing for several months and cause public health hazard for poultry consumers. Vaccination is the only effective tool can control the virus (Dakouo et al., 2020). Unfortunately, several factors can lead the vaccination failure as pollution, chemicals, infectious agents, hormones and nutrition. Chemotherapeutic agents also play a critical role in stimulation or suppression of the immune response against the vaccine (Mund, 2017; Kim and Lillehoj, 2019).

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Under field conditions, it is unusual to run a commercial poultry cycle without some medication as antibiotics & anticoccidials (Mund, 2017; Kim and Lillehoj, 2019). Clindamycin is a semi-synthetic lincosamide antibiotic, it is effective against anaerobic bacteria and gram-positive aerobic bacteria (including most Staphylococcus and Streptococci spp), and some protozoal infections (including Toxoplasma) (Wan et al., 2016; Dotel et al., 2019). It inhibits bacterial protein synthesis by binding to 23S RNA of the 50S subunit of the bacterial ribosome. It easily distributes in all body fluids and reaches bone. According to the dosage, it may be bacteriostatic or bactericidal (Okudo and Anusim, 2016). In comparison to other antibiotic groups it has a shorter half life time and easily eliminated from the bird body through the liver and kidneys (Sadek and Shaheen, 2014). This makes Clindamycin a good choice during broilers breeding. The present work aimed to shed the light on the effect of clindamycin oral administration on some immunological and biochemical parameters of Newcastle vaccinated broilers.

2. Materials and Methods

2.1. Drugs

Clindamycin (Ato Clinda 20%) are product of Atcopharma.

Experimental design

A total of 150 one day old Hubbard chicks were used in the present study. The chicks were divided into three equal groups, each of 50 chicks. First group was a control non treated group (CG). At 14th, 15th and 16th days of age, the second group (C20) and the third group (C40) were given Clindamycin (20 and 40mg/kg body. wt) respectively in drinking water as recommended by the producers. All groups were routinely vaccinated against Newcastle by Hitchner and Lasota at 7th and 18th day of age respectively. On 25th day of age, blood samples were collected from ten birds of each group for immunological and biochemical parameters evaluation. Five birds of each group were slaughtered at the same age then the lymphoid organs (thymus, bursa and spleen) were carefully separated & weighed and each organ relative weight was determined. Number of dead birds were recorded throughout the experiment period (45th day).

Laboratory examinations:

2.2. Immunological tests

Phagocytic activity was estimated according to Barry et al. (1989). Total leukocytic counts (TLC) were evaluated according to Natt and Herrick (1952). Blood film was prepared and stained with Giemsa stain for differential leukocytic count (DLC) according to Schalm et al., (1975). haemagglutination inhibition titer (HI) was determined according to Takatsy (1956).

2.3. Biochemical tests

Serum biochemical tests were measured spectrophotometrically in a private laboratory using commercial kits of Biodiagnostic company® commercial kits, Cairo, Egypt. Serum globulin concentrations were calculated by subtracting the value of albumin from the corresponding value of total protein and A/G ratio by division of albumin on the corresponding globulin value.

2.4. Statistical analysis

SPSS program version 24 was used to compare between means of different statistical parameters (one-way ANOVA test) and estimate the post-hoc differences between means (a multiple comparison Tukey's HSD test). A difference was considerable significant at $P < 0.05$.

3. Results

Table (1) displayed a significant ($P < 0.05$) decline in TLC, HI antibody titer, total protein and globulin levels in C20, C40 when compared to CG. While, A/G significantly ($P < 0.05$) decreased in C40 and non-significantly ($P \geq 0.05$) changed in C20 in relation to CG. On the other hand, no significant changes were observed between C20, C40 and CG in DLC (monocytes, lymphocytes, heterophils, eosinophils, basophils), phagocytic activity, albumin, liver enzymes (ALT, AST) and kidney function tests (uric acid, creatinine (Cr)).

Table (2) cleared a significant ($P < 0.05$) decrease in bursa relative weight of C20, C40 when compared to CG. While, thymus relative weight, body weight gain significantly ($P < 0.05$) depressed in C40 only and non-significantly ($P \geq 0.05$) changed in C20 in relation to CG. On contrast feed conversion rate significantly ($P < 0.05$) increased in C40 and non-significantly ($P \geq 0.05$) changed in C20 compared to CG. In comparison to CG number of dead birds and mortality rate increased in C20 and C40. The comparison between C20 and C40 revealed a significant ($P < 0.05$) inhibition of HI antibody titer and weight gain in C40 in relation to C20 (table 1 and 2).

4. Discussion

Vaccination is a necessary process in all animal husbandry section, including poultry. It is the most reliable mean for pandemic-highly infectious diseases control (Aldabagh et al., 2011). No doubt, the bird response towards the vaccine widely depends on his immune system soundness. Like mammals, avian immune system produces two types of immunity: innate immunity and adaptive immunity. Innate immunity (circulating leukocytes and macrophage) is non-specific, originated from bone marrow. Adaptive immunity has two parts: humeral specific immunity (immunoglobulin M, G, A developed in bursa of Fabricius from B-cells) and cell-mediated immunity (T-cell derived from thymus) (Birhan, 2019). Antibiotics administration usually restrains the immune system proper functions (Bystryzcka, et al., 2016; Jamal et al., 2017).

In this study, clindamycin affected both innate and adaptive immunity of the treated birds. Whereas, a pronounced depression was detected in the innate immunity of C20 and C40 chicks. This was indicated by the noticed leukopenia in both groups members. This leukopenia may be attributed to the myelosuppressive effect of clindamycin due to its direct hematopoietic tissue injury (Morales et al., 2014; Birhan, 2019). In the same way, clindamycin inhibits humeral immunity of C20 and C40 boilers, through reduction of the bursa relative weight in both groups and subsequently inhibits B cells maturation and antibody formation (Sureshkumar et al., 2013; Birhan, 2019). This was translated by the observed decrease in HI antibody titer and subordinate hypoglobulinemia, hypoproteinemia in C20 and C40 individuals and decreased A/G ration in C40 chicks. Similar results were reported before in Newcastle vaccinated boilers treated with antibiotics (Sureshkumar et al., 2013). While, the cell-mediated immunity was only suppressed in C40 broilers because of the remarkable decline of thymus relative weight in this group chicks (Birhan, 2019).

In addition to the clindamycin immune-inhibitory effect in treated animals, they suffered from lower body weight gain, higher feed conversion rates, higher number of dead birds and mortality rates (Jamal et al., 2017). Rationally, the lowered HI antibody titer and body weight gain in C40 in relation to C20 clarified that the clindamycin immunomodulatory effect correlated with its dosage.

On contrast, clindamycin has no effect on hepatic or renal function tests of C20 and C40 here. Although it is mainly metabolized and eliminated from the body through liver and kidney, it has a short-lived effect and rarely causes hepatic or renal injury (Moole et al., 2015; Subedi et al., 2017). These findings agree with Sadek and Shaheen (2014) results, who obtained physiological hepatic and renal functions in *E. Coli* experimentally infected broilers, within 2 weeks from clindamycin treatment.

5. Conclusion

It could be concluded that, Clindamycin oral administration interferes both innate and humeral broilers immune response against Newcastle disease virus vaccine and reduce body weight gain. So it is not recommended to use them few days before or after vaccination.

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Conflict of interest

There is no conflict of interest.

6. References

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Table 1: comparison between some immunological and biochemical parameters of C20, C40 in relation to CG. Values are means \pm SD.

Parameters	CG	C20	C40
TLC ($\times 10^3/\mu\text{l}$)	29.10 \pm 2.40 ^a	26.50 \pm 1.80 ^b	21.40 \pm 2.01 ^b
Monocytes (%)	9.70 \pm 0.65 ^a	10.10 \pm 0.54 ^a	10.20 \pm 0.53 ^a
Lymphocytes (%)	62.60 \pm 3.43 ^a	64.50 \pm 5.34 ^a	61.50 \pm 4.24 ^a
Heterophilis (%)	23.70 \pm 1.28 ^a	21.60 \pm 2.38 ^a	24.20 \pm 1.76 ^a
Eosinophils (%)	2.50 \pm 0.26 ^a	2.50 \pm 0.25 ^a	2.60 \pm 0.33 ^a
Basophils (%)	1.50 \pm 0.18 ^a	1.30 \pm 0.17 ^a	1.50 \pm 0.22 ^a
Phagocytic activity (%)	21.43 \pm 0.32 ^a	22.67 \pm 0.25 ^a	21.75 \pm 0.31 ^a
HI antibody titer (log ₂)	5.80 \pm 0.42 ^a	4.40 \pm 0.39 ^b	3.80 \pm 0.29 ^c
Total protein (g/dl)	4.77 \pm 0.46 ^a	4.30 \pm 0.35 ^b	4.14 \pm 0.42 ^b
Albumin (g/dl)	2.30 \pm 0.19 ^a	2.11 \pm 0.24 ^a	2.24 \pm 0.26 ^a
Globulin (g/dl)	2.49 \pm 0.07 ^a	2.03 \pm 0.07 ^b	1.90 \pm 0.06 ^b
A/G	0.92 \pm 0.05 ^a	1.04 \pm 0.08 ^{ab}	1.18 \pm 0.10 ^b
AST (U/L)	21.63 \pm 1.64 ^a	24.14 \pm 0.85 ^a	23. 67 \pm 1.38 ^a
ALT (U/L)	99.36 \pm 7.42 ^a	105.69 \pm 5.39 ^a	107.55 \pm 8.21 ^a
Uric acid (mg/dl)	11.72 \pm 0.57 ^a	11.47 \pm 0.65 ^a	12.21 \pm 0.68 ^a
Cr (mg/dl)	1.13 \pm 0.17 ^a	1.10 \pm 0.14 ^a	1.07 \pm 0.19 ^a

Means in the same row bearing different letters differ significantly (P<0.05).

CG: Control group, C20: Clindamycin 20mg/kg bodyweight, C40: Clindamycin 40mg/kg bodyweight.

TLC: total leukocytic count, A/G: Albumin/Globulin ratio, AST: Aspartate aminotransferase, ALT: Alanine aminotransferase, Cr: Creatinine.

Table 2: comparison between the relative weights of bursa, spleen and thymus body weight, feed conversion rates, no of dead birds and mortality rates of C20, C40 in relation to CG. Values are means \pm SD.

Parameters	CG	C20	C40
Bursa relative weight (%)	2.13 \pm 0.16 ^a	1.75 \pm 0 .15 ^b	1.65 \pm 0 .18 ^b
Spleen relative weight (%)	1.34 \pm 0 .14 ^a	1.36 \pm 0 .13 ^a	1.33 \pm 0 .16 ^a
Thymus relative weight (%)	2.30 \pm 0 .15 ^a	2.15 \pm 0 .21 ^{ab}	1.93 \pm 0 .16 ^b
Weight gain (gm)	1997 \pm 65.8 ^a	1879 \pm 61.2 ^a	1851 \pm 52.3 ^b
Feed conversion (%)	1.97 \pm 0.029 ^a	2.19 \pm 0.041 ^{ab}	2.41 \pm 0.078 ^b
Number of dead birds	5	6	8
Mortality rate (%)	7	9	15

Means in the same row bearing different letters differ significantly (P<0.05).

CG: Control group, C20: Clindamycin 20mg/kg bodyweight, C40: Clindamycin 40mg/kg bodyweight.