



Structure and Biochemical Analysis of Phytoplankton in the Wadi El-Rayan Lakes, El-Fayoum, Egypt.

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ABSTRACT

Wadi El-Rayan lakes are an important source of irrigation water and fisheries in El-Fayoum Governorate, Egypt. Phytoplankton is an important bioindicator of the physicochemical and biological changes in natural ecosystems including the Wadi El-Rayan lakes. Seasonal and regional variations lead to concomitant changes in the total protein, carbohydrate, and lipid contents of phytoplankton; such changes affect the nutritional quality of food available in the aquatic system. In the present study, these changes were determined in the phytoplankton of Wadi El-Rayan lakes. The maximum protein (27.95 & 11.36 mgL⁻¹) contents were detected during summer and winter respectively. The maximum carbohydrate content was detected in autumn and winter (1.76 and 1.12 mgL⁻¹ at site 8 and 4 respectively). The maximum lipid content was observed in winter (0.73 mgL⁻¹ at site 6). Phytoplankton density was much higher in the upper lake than in the lower lake. In total, 92 species were identified; the phytoplankton community in the two lakes included typical groups such as Chlorophyceae, Cyanophyceae, Bacillariophyceae, Cryptophyceae, Dinophyceae, Chrysophyceae, and Euglenophyceae. The green alga was the dominant group, constituting 44.4%–45.0% of the phytoplankton standing crop across seasons. Cyanophyceae (29%–39%), Bacillariophyceae (9.7%–16.1%), and Dinophyceae (4.0%–9.2%) were the next most abundant classes; Cryptophyceae, Chrysophyceae, and Euglenophyceae were rarely found. The major peak of phytoplankton density (7684×10⁴ Cells L⁻¹) was observed during winter, whereas the minor peak was detected during summer (4879×10⁴ Cells L⁻¹). *Cosmarium nitidulum* (Brebisson) Ralfs, *Dictyosphaerium pulchellum* Wood, *Oocystis solitaria* Wittrock, *Gomposphaeria compacta* (Lammermann) Ström, *Lyngbya*

limnetica Lemmermann, *Microcystis aeruginosa* Kützing, *Cyclotella meneghiniana* Kutz., and *Syndra ulna* (Nitzsch) Ehr. were the most abundant species identified. More detailed studies on the biochemical structure of phytoplankton are needed to better understand Wadi El-Rayan lakes' response to the many environmental changes.

Keywords: Wadi El-Rayan Lakes; phytoplankton; biochemical analysis.

INTRODUCTION

The Wadi El-Rayan lakes are a natural protected area and important source of irrigation water and fisheries in El-Fayoum Governorate, Egypt. Wadi El-Rayan depression (Southwest of Cairo in the Western desert of Egypt) holds two man-made lakes that receive the wastewater drainage of El-Fayoum depression. Phytoplankton plays an important role in the equilibrium of many aquatic ecosystems. The biochemical content of phytoplankton is an important indicator of their nutritional quality (Müller-Navarra *et al.*, 2000). Seasonal and regional variations lead to changes in the total protein, carbohydrate, and lipid contents of phytoplankton; these changes affect the nutritional quality of food available in the aquatic system (Boëchat and Giani, 2008). The water quality of a water body depends on many physical, chemical, and biological indicators (Poonam *et al.*, 2013). The health of all living organisms in an aquatic ecosystem is affected by the quality of water (Doherty *et al.*, 2010). Specifically, Saeed and Ibrahim (2008) reported that inorganic and organic chemical pollution of the aquatic environment poses a serious threat to the survival of aquatic organisms. In addition, several industrial and agricultural processes cause environmental pollution and contribute to the contamination of water ecosystems (Mohamed *et al.*, 2017). Wadi El-Rayan lakes receive agricultural wastewater drainage from the El-Wadi Drain and vary in their physical and chemical characteristics (Ali *et al.*, 2007). According to the evaporation of water from the Wadi El-Rayan lakes, the gradual increase in concentrations of salts, heavy metals, pesticides, and other pollutants is changing their water quality and likely affecting aquatic life (Mansour and Sidky, 2003; Paleczny *et al.*, 2007). Thus, the physicochemical and biological properties of the Wadi El-Rayan lakes have been extensively studied (Abd El-Karim, 2004; Abdel Hameed *et al.*, 2007; Ali *et al.*, 2007; Abdel-Satar and Goher, 2015). In terms of phytoplankton, the upper lake has a more condensed population than the lower one, while maximum and minimum phytoplankton production has been recorded in winter and spring, respectively (Taha and Farghaly, 1994). To build on these previous studies, the current phytoplankton

community structure of the Wadi El-Rayan lakes and its biochemical content across seasons were analyzed.

MATERIALS AND METHODS

Study area:

Wadi El-Rayan holds two main lakes at different elevations that are connected by a swampy channel. They lie between 30°20′–30°25′E and 29°05′–29°20′N (**Fig. 1**).

Samples locations:

Ten stations were chosen to cover the area under investigation. Stations 1–5 and stations 6–10 represent the upper and lower lakes, respectively; and the El-Wadi Drain station (**Fig. 1**). This study was seasonally conducted from autumn 2019 to summer 2020.

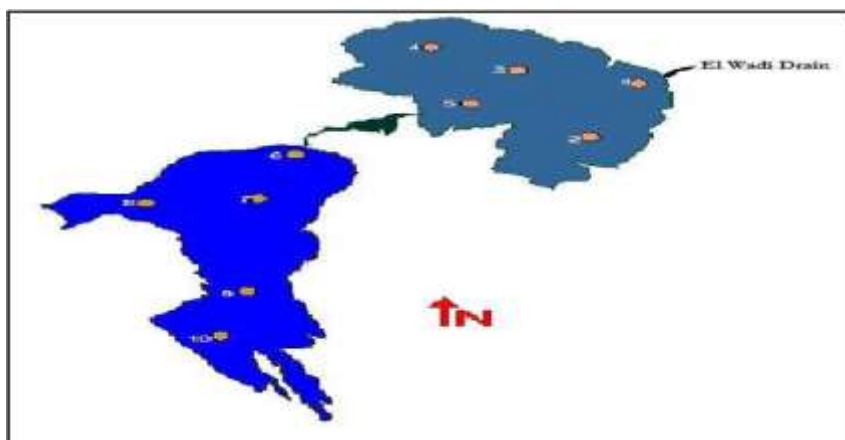


Fig. 1: Map of Wadi El-Rayan lakes showing the samples' locations.

Qualitative estimation of phytoplankton:

Water samples were collected from the sites described previously; phytoplankton in the sample was concentrated by settling 500 ml water sample in a volumetric cylinder for about 5 days after being preserved with Lugol's solution (10 g of pure iodine, 20 g of potassium iodide KI, 200 mL of distilled water, and 20 ml of glacial acetic acid). This solution facilitates the sedimentation and staining of phytoplankton cells. During time phytoplankton organisms were allowed to settle by gravitation (APHA, 1992). The supernatant was then carefully siphoned off with a small plastic tube including a fine net (20µm mesh diameter) until the samples were concentrated to about 50 mL. The remaining volume was adjusted to exactly 50 mL, and the samples were kept at 4° C in a dark plastic vial until microscopic examination. Phytoplankton counts were conducted using a wild inverted microscope (Zeiss

Axiovert 25C); 5 μL of the reduced-volume sample was placed in a counting chamber and examined with a 10 \times eyepiece and 40 \times objective. The drop method (APHA, 1992) was applied for both the counting and identification of different algal species from different samples. Algal taxa were identified according to standard references including Kofoid (1911), Kofoid and Swez (1921), Lebour and Marie (1930), Hannford and Britton (1952), Henedy (1964), Bourrely (1968), Prescott (1978), Mizuno (1990), and Krammer and Lang Bertalot (1991).

Biochemical analysis:

The total protein content was determined using the biuret method (David and Hazel, 1993). The hydrolysis of carbohydrates was conducted using the method of Myklestad and Haug (1972), and the carbohydrate content was determined according to the work of Dubois *et al.*, (1956). The total lipid content was determined according to the method of Chabrol and Castellano (1961).

Statistical analysis:

Relation between phytoplankton and the three biochemical parameters (proteins, lipids, and carbohydrates) was calculated by principal component analysis (PCA), using XLSTAT (2021) program.

RESULTS AND DISCUSSION:

There was a large variation in the biochemical content of phytoplankton among species and locations across the seasons. Generally, total protein was the major biochemical constituent of the phytoplankton community; the protein content of algae is also an important criterion for their use as food (Fawzy *et al.*, 2017). In contrast, the total lipid content was the minor constituent of the biochemical composition of the communities. As shown in Fig. 2, the maximum protein (27.95 & 11.36 mgL^{-1}) contents were detected in summer and winter respectively. Proteins are important components of all membranes, involved in the transport of other molecules and ions across the membranes. Most enzymes are proteins that organize, construct and receive signals, while the structural proteins maintain the shape of the cell (Jónasdóttir 2019). The maximum carbohydrate content was detected in autumn and winter (1.76 and 1.12 mgL^{-1} at sites 8 and 4 respectively). The maximum lipid content was observed in winter (0.73 mgL^{-1} at site 6). The biochemical content of phytoplankton cannot explain the variation observed in the consumer's growth when fed different phytoplankton types. So, it is necessary to look further into the more detailed structure of the classes.

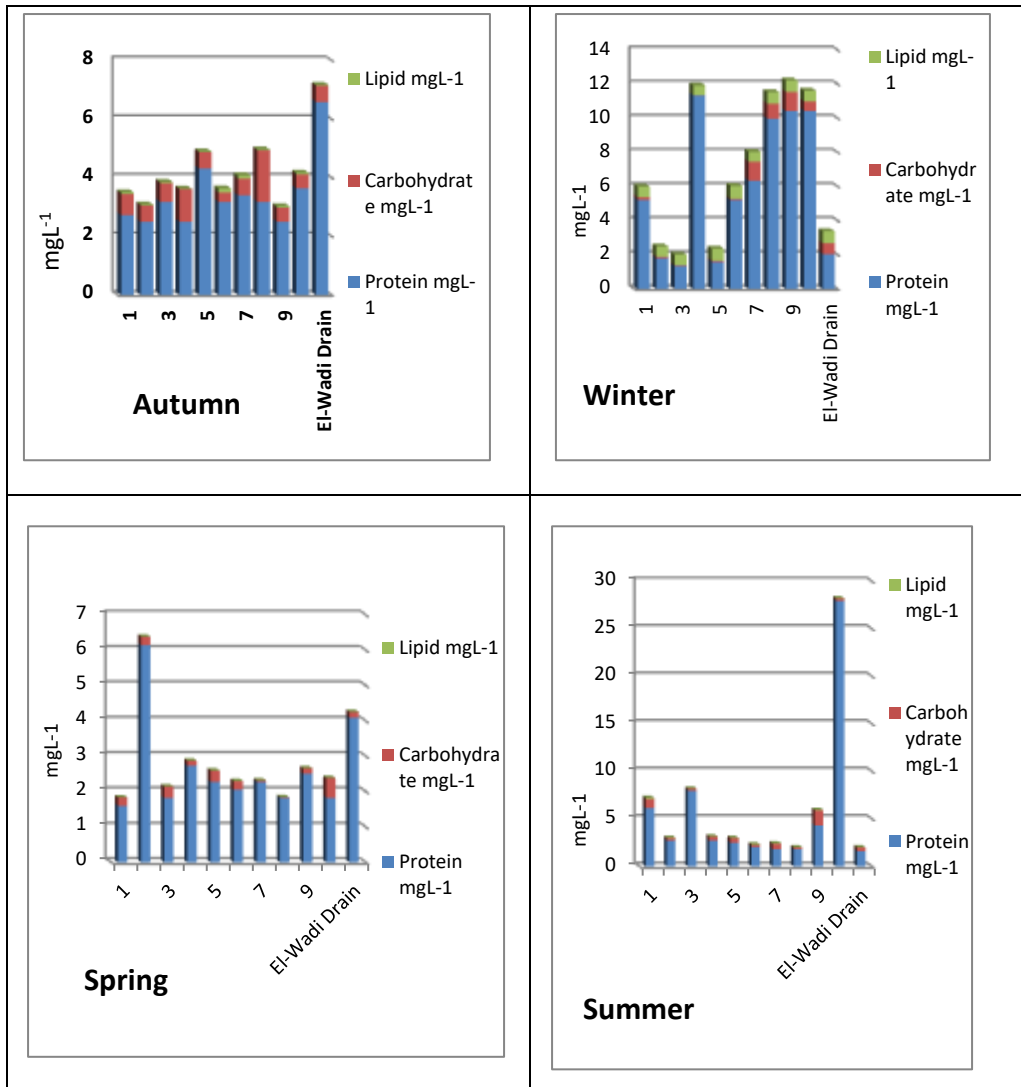


Fig. 2: The biochemical content of phytoplankton in Wadi El-Rayan lakes across the seasons (from autumn 2019 to summer 2020).

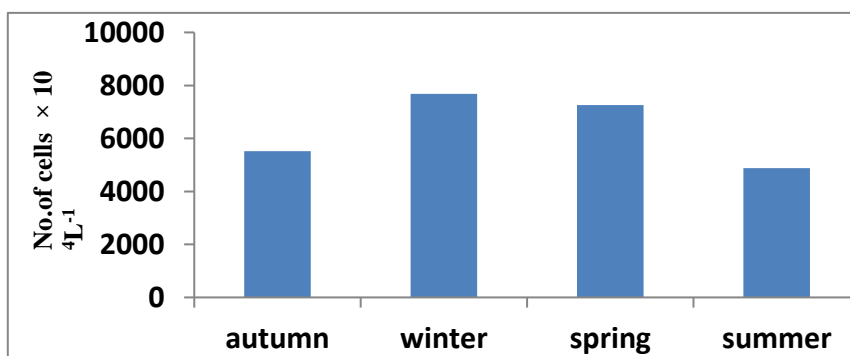


Fig. 3: Seasonal variation in total phytoplankton density ($\times 10^4$ Cells L⁻¹) in Wadi El-Rayan lakes (from autumn 2019 to summer 2020).

The seasonal distribution of phytoplankton showed two major peaks in density: 7684 and 7260 $\times 10^4$ Cells L⁻¹ in winter and spring, respectively. The lowest peak in density was detected in summer (4879 $\times 10^4$ Cells L⁻¹). In the present study, phytoplankton density was much higher in the upper lake than in the lower lake (**Table 1**); this was likely due to the high nutrient concentrations received by the upper lake from drainage water via the El-Wadi Drain, which is again similar to that reported by **Konsowa (2007)**.

In total, 92 species were identified in the present study (**Table 2**). The phytoplankton community in the lake was composed of typical groups (**Fig. 4 and Table 1**), e.g., Chlorophyceae, Cyanophyceae, Bacillariophyceae, Cryptophyceae, Dinophyceae, Chrysophyceae, and Euglenophyceae. The green alga was the most dominant class (44.4%–45.0% of the phytoplankton standing crop across seasons), followed by Cyanophyceae (29%–39%), Bacillariophyceae (9.7%–16.1%), and Dinophyceae (4.0%–9.2%). In contrast, Cryptophyceae, Chrysophyceae, and Euglenophyceae were rarely found.

Chlorophyceae (green algae) was the dominant phytoplankton group, with 34 Chlorophyceae taxa identified. The most common green algal taxa were *Cosmarium nitidulum* (Brebisson) Ralfs, *Oocystis borgei* Snow, *Dictyosphaerium pulchellum* Wood, *Planktonema lauterbornii* Schmidle, and *Oocystis solitaria* Wittrock (**Fig.5**). The presence of *Tetraedron minimum* (A. Braun) Hansgirg and *Scenedesmus* spp. in the lake water indicated that it was a eutrophic environment (**El-Sheekh et al., 2018**).

Cyanophyceae (Blue-green algae) including 17 taxa, was the second major constituent group in the study area; the dominant species were *Gomphosphaeria compacta* (Lammermann) Ström, *Lyngbya limnetica* Lemmermann, and *Microcystis aeruginosa* Kützing. These species can tolerate a wide range of salinity and fluctuations in Wadi El-Rayan water chemistry. These findings are in agreement with those of **Konsowa and Taha (2002)**. In

addition, **Konsowa and Abd Ellah (2002)** observed *M. aeruginosa* in the first lake during winter and *L. limnetica* in the second lake during summer.

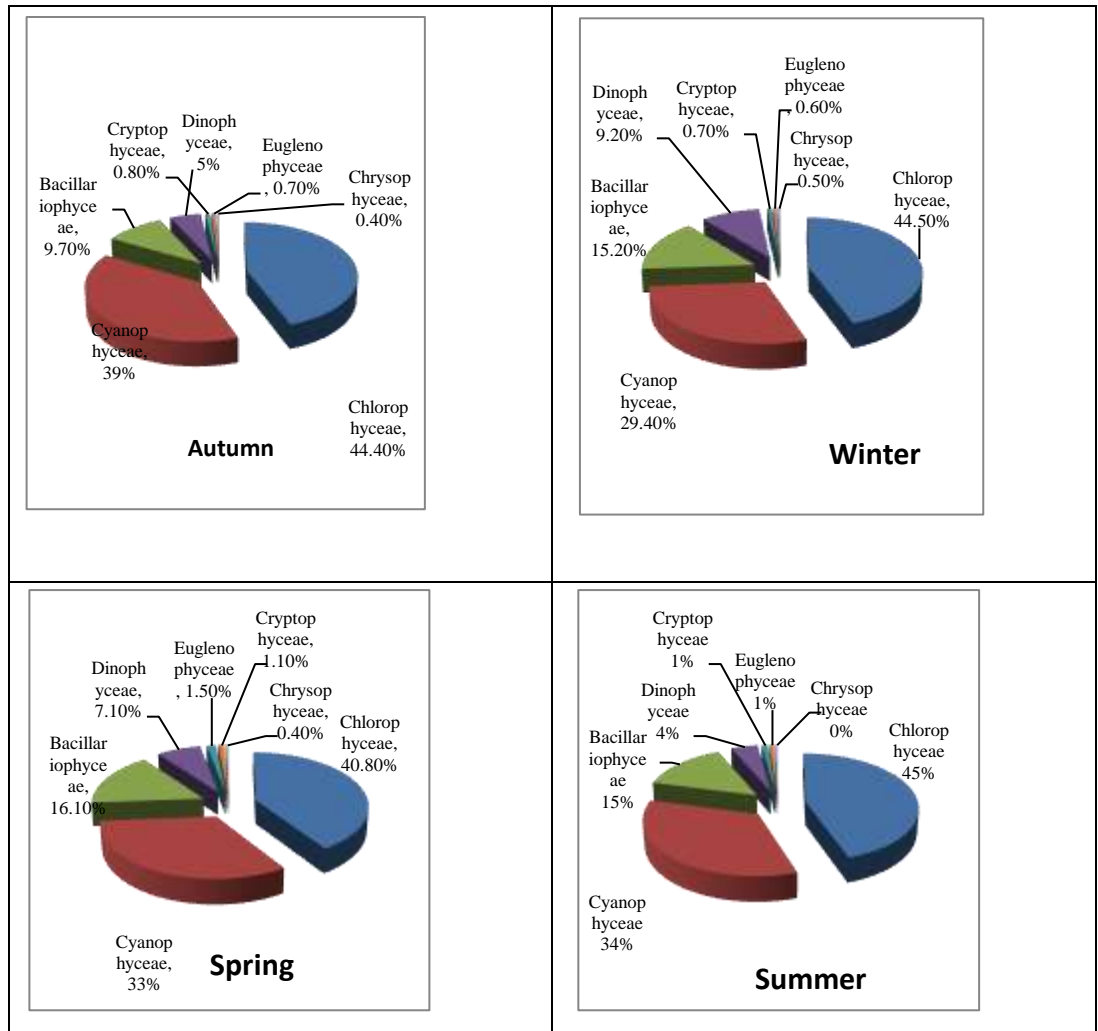


Fig. 4: Phytoplankton community composition across the seasons (from autumn 2019 to summer 2020) in the Wadi El-Rayan lakes.

Bacillariophyceae (Diatoms) was the third most predominant algae after green and blue-green (**Fig. 4**). In total, 28 taxa related to Bacillariophyta were identified (**Table 2**). The most abundant species of diatoms were *Cyclotella meneghiniana* Kutz., *Nitzschia paleacea* Grum, and *Syndra ulna* (Nitzsch) Ehr. Four phytoplankton classes, Chrysophyceae, Cryptophyceae, Dinophyceae, and Euglenophyceae, were rarely found; thus, their standing

crops were low (**Table 2**). These classes are related to the increase in water salinity in the lower lake (**Konsowa, 2005**). **Abd Ellah (2016)** stated that the lower lake has faced extreme water loss over the last 20 years due to the extension of agricultural land and fish farms in the depression.

Table 1: Spatial and seasonal variations in the class composition of phytoplankton Count ($\times 10^4$ Cells L⁻¹) in Wadi El-Rayan lakes and their drain.

Site	1	2	3	4	5	6	7	8	9	10	Drain	Total	Average
Autumn													
Chlorophyceae	409	540	390	484	229	40	89	91	74	65	44	2455	223.2
Bacillariophyceae	98	10	110	80	76	2	34	38	20	20	50	538	48.9
Cyanophyceae	492	398	417	281	113	87	151	82	29	8.7	90	2148	195.3
Dinophyceae	20	130	21	24	10	35	4	13	11	10	0	278	25.3
Cryptophyceae	8	10	4	4	5	4	2	4	2	0	0	43	3.9
Euglenophyceae	7	0	4	2	8	10	2	2	4	0	0	39	3.5
Chrysophyceae	2	0	6	0	10	0	1	0	4	0	0	23	2.1
Total phytoplankton	1036	1088	952	875	451	178	283	230	144	104	184	5524	502.2
Winter													
Chlorophyceae	526	502	546	453	220	80	257	141	49	54	590	3418	310.7
Bacillariophyceae	66	85	98	76	106	96	43	50	20	18	508	1166	106.0
Cyanophyceae	196	454	468	221	120	77	256	152	66	70	182	2262	205.6
Dinophyceae	6	40	47	16	80	110	17	14	20	14	340	704	64.0
Cryptophyceae	2	0	2	8	20	10	2	4	2	0	0	50	4.5
Euglenophyceae	3	0	2	3	10	0	4	1	2	2	22	49	4.5
Chrysophyceae	0	0	6	0	10	10	2	0	5	2	0	35	3.2
Total phytoplankton	799	1081	1169	777	566	383	581	362	164	160	1642	7684	698.5

Con. Table 1:

Site	1	2	3	4	5	6	7	8	9	10	Drain	Total	Average
Spring													
Chlorophyceae	746	482	415	346	201	161	212	113	123	89	76	2964	269.5
Bacillariophyceae	294	188	118	132	58	68	60	64	56	72	60	1170	106.4
Cyanophyceae	620	260	390	297	164	184	188	85.7	96.9	35.4	75.4	2395	217.8
Dinophyceae	126	62	65	64	43	38	24	36	30	16	10	514	46.7
Euglenophyceae	26	12	14	16	6	4	8	16	10	0	0	112	10.2
Cryptophyceae	21	14	12	2	6	6	4	4	8	0	0	77	7.0
Chrysophyceae	10	10	4	0	4	0	0	0	0	0	0	28	2.5
Total phytoplankton	1843	1028	1018	857	482	461	496	319	324	212	221	7260	660.0
Summer													
Chlorophyceae	590	396	337	319	160	32	88	82	66	90	32	2192	199.3
Bacillariophyceae	158	28	121	74	58	20	48	50	40	44	66	707	64.3
Cyanophyceae	377	275	306	197	94.9	87	141	53.5	30	21.1	93.1	1676	152.3
Dinophyceae	56	30	22	22	24	17	12	9	7	5	5	209	19.0
Cryptophyceae	13	6	2	6	4	0	4	2	2	0	0	39	3.5
Euglenophyceae	8	4	8	2	0	2	0	10	4	0	0	38	3.5
Chrysophyceae	4	2	6	0	0	0	6	0	0	0	0	18	1.6
Total phytoplankton	1206	741	802	620	341	158	299	207	149	160	196	4879	443.5

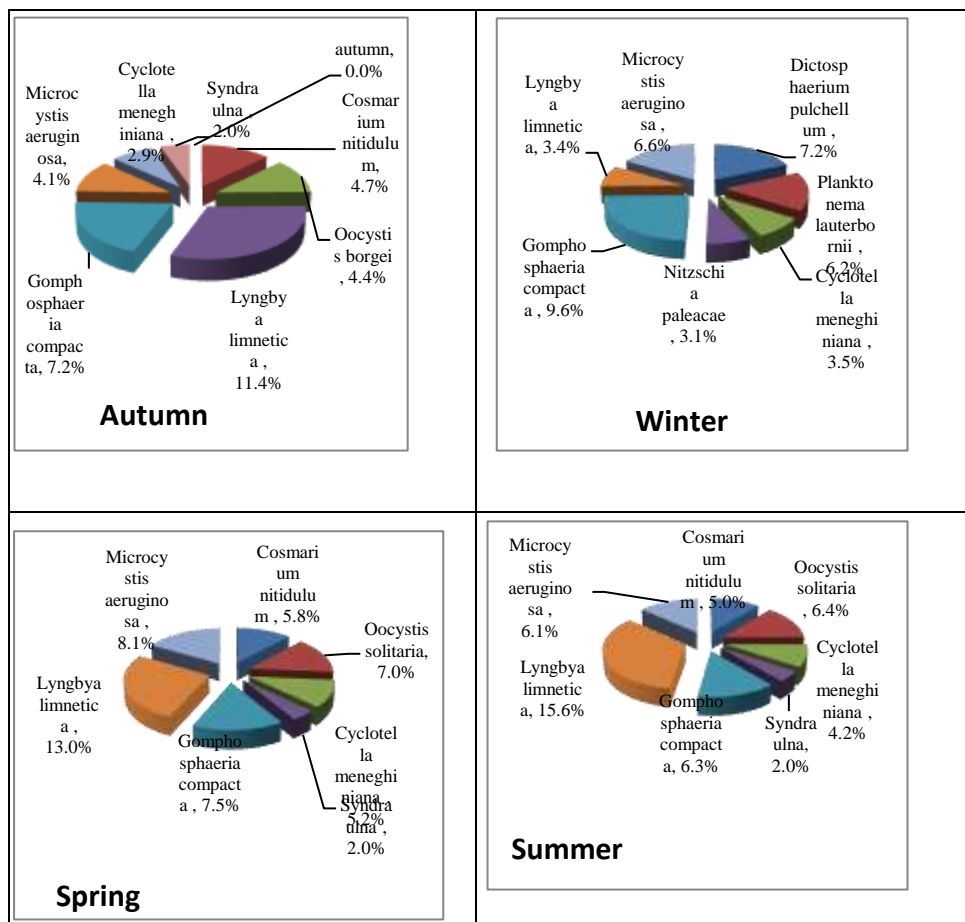


Fig. 5: Percentage abundance of the most common species of phytoplankton in Wadi El-Rayan lakes.

As shown in **Fig. 6**, the principal component analysis indicated that total phytoplankton was directly related to Chlorophyceae, Cyanophyceae, and *L. limnetica*, while Bacillariophyceae had high lipid levels relative to protein and carbohydrate levels. Overall, these results suggest that the biochemical content of phytoplankton can be a good indicator of their nutritional value.

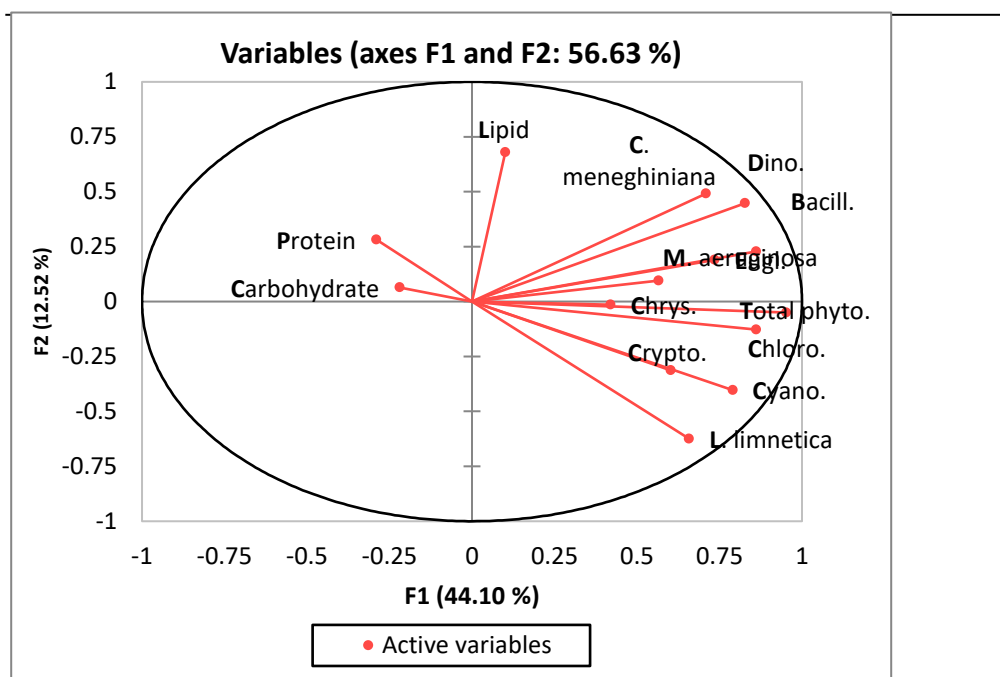


Fig. 6: Principal component analysis of biochemical and biological variables measured at the various stations of Wadi El-Rayan lakes and drain. Chloro, Chlorophyceae; Bacill, Bacillariophyceae; Cyano, Cyanophyceae;Cryp, Cryptophyceae; Chrys, Chrysophyceae; Eugl, Euglenophyceae; Dino, Dinophyceae; Total Phyto, Total phytoplankton; *M. aeruginosa*, *Microcystis aeruginosa*; *L. limnetica*, *Lyngbya limnetica*; *C. meneghiniana*, *Cyclotella meneghiniana*.

Table 2: List of phytoplankton species in the Wadi El-Rayan lakes and drain.

Chlorophyceae

<i>Actinastrum hantzschii</i> var. <i>fluviatile</i> J.B.L.Schröder	<i>Nephrocystium subsolitaria</i>
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs	<i>Oocystis borgei</i> Snow
<i>Ankistrodesmus falcatus</i> v. <i>acicularis</i> (Braun.) West	<i>Oocystis elliptica</i> W. West
<i>Ankistrodesmus fusiformis</i> Corda	<i>Oocystis gigas</i> var. <i>minor</i> West & G.S.West.
<i>Chlamydomonas globosa</i> Snow	<i>Oocystis parva</i> W.&G.S. West
<i>Cosmarium amgulosum</i> Brebisson	<i>Oocystis solitaria</i> Wittrock
<i>Cosmarium depressum</i>	<i>Planktonema lauterbornii</i> Schmidle
<i>Cosmarium nitidulum</i> (Brebisson) Ralfs.	<i>Scenedesmus bicudatus</i> Dedusenko
<i>Dictosphaerium pulchellum</i> Wood	<i>Scenedesmus dimorphus</i> (Turpin) Küzing
<i>Elakatothrix gelatinosa</i> Wille	<i>Scenedesmus ecornis</i> (Ehrenberg) Chodat
<i>Gonium sociale</i> (Dujardin) Warming	<i>Scenedesmus opoliensis</i> P.G.Richter
<i>Keratococcus suecicus</i> Hindák	<i>Scenedesmus quadricauda</i> (Turpin) Brébisson
<i>Kirchneriella contorta</i> (Schmidle) Bohlin	<i>Scenedesmus quadricauda</i> v. <i>quadrispina</i> (Chod.) Smith
<i>Kirchneriella contorta</i> var. <i>elegans</i> (PLAYFAIR) KOMÁREK	<i>Staurostrum chaetoceras</i>
<i>Kirchneriella irregularis</i> (Petyk.) Printz	<i>Staurostrum paradoxum</i> Meyen ex Ralfs
<i>Legerheimia ciliata</i> . (Lag.) Chodat. L.	<i>Tetraëdron minimum</i> (A.Braun) Hansgirg

Monoraphidium contortum (Thuret) Komárková-
Legnerová

Treubaria triappendiculata C. Bernard

Con. Table 2:

Bacillariophyceae

<i>Amphora coffeaeformis</i> Kutz.	<i>Fragillaria capucina</i> Desm
<i>Bacillaria paradoxa</i> Gemelin	<i>Gomphonema parvulum</i> var. <i>longenula</i> Ktz.
<i>Chaetoceros curvsetus</i> Cleve	<i>Navicula confervacea</i> (Kützing) Grunow
<i>Cocconies placentula</i> Ehr.	<i>Navicula cryptocephala</i> Kützing
<i>Cosinodiscus divisus</i> Grun.	<i>Navicula festiva</i> Krasske
<i>Cyclotella bodanica</i> Eulenstein ex Grunow	<i>Nitzschia closterium</i> (Ehrenberg) W. Smith
<i>Cyclotella glomerata</i> Bachmann	<i>Nitzschia filiformis</i> (W. Smith) Van Heurck
<i>C. kützingiana</i> Thwait.	<i>Nitzschia frustulum</i> (Kützing) Grunow
<i>Cyclotella ocellata</i> Pant	<i>N. gracilis</i> Hantz
<i>Cyclotella operculata</i> (Ag.) kutz.	<i>N. palea</i> (Kutz.) W. Smith
<i>Cyclotella meneghiniana</i> kutz.	<i>N. tryblionellae</i> Hant
<i>C. stelligera</i> Cleve & Grun.	<i>Stephanodiscus astraeta</i> (Ewhren.) Grun
<i>Cymbella microcephala</i> Grunow in Van Heurck	<i>Synedra tabulata</i> var. <i>acuminata</i> (Grun) Hust.
<i>Epithemia proboscidea</i> Kützing	<i>Syndra ulna</i> (Nitzsch) Ehr.

Cyanophyceae

<i>Chroococcus minutus</i> (Kützing) Nägeli	<i>Microcystis flosaquae</i> (Wittrock) Kirchner
<i>Cylindrospermopsis raciborskii</i> Woloszynska	<i>Microcystis grevillei</i> (Hassall) Elenkin
<i>Gomphosphaeria compacta</i> (Lammermann) Ström	<i>Oscillatoria brevis</i> (Kutz.) Gom.
<i>Gomphosphaeria fusca</i> Skuja	<i>Phormidium interruptum</i> Kutz
<i>Limnococcus limneticus</i> (Lammermann) Komárková	<i>phormidium frigidum</i> Fritsch
<i>Lyngbya limnetica</i> Lemmermann	<i>Phormidium laminosum</i> Gomont ex Gomont
<i>Lyngbya profundalis</i> Lindstedt	<i>Pseudanabaena papillaterminata</i> (Kisselev) Kukk
<i>Merismopedia elegans</i> A. Braun ex Kützing	<i>Rhabdoderma lineare</i> Schmidle & Lauterborn
<i>Microcystis aeruginosa</i> Kützing	

Cryptophyceae

Cryptomonas obovata
Ehrenberg

Cryptomonas phaseolus Skuja
Chroomonas salina (Wills.)
Butch

Dinophyceae

Gymnodinium biconica Skuja
Peridinium bipes Stein
Peridinium penardii (Lemm.) Lemm.
Prorocentrum micans Ehr

Chrysophyceae

Mallomonas heterospina

Euglenophyceae

Euglena pisciformis Klebs
Euglena proxima P.A. Dangeard
Phacus pleuronectes (O. F. Muller)
Dujardin

Protoperidinium brevipes (Paulsen) Balech

Prorocentrum dentatum Stein

CONCLUSION

The phytoplankton communities, and the biochemical content of these communities, can be considered ecosystem biomarkers at different sites in Wadi El-Rayan lakes. The biochemical composition of phytoplankton could be a valid integrator of surrounding environments in which phytoplankton grow and can be a good indicator of their nutritional value.

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التركيب والتحليل الحيوي الكيميائي للعوالق النباتية في بحيرات وادي الريان ، الفيوم، مصر.

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الملخص العربي

تعتبر بحيرات وادي الريان مصدرا هاما لمياه الري والثروة السمكية في محافظة الفيوم ، مصر. وتعتبر العوالق النباتية مؤشراً حيوياً مهماً للتغيرات الكيميائية والبيولوجية في النظم البيئية الطبيعية بما في ذلك بحيرات وادي الريان. تؤدي الاختلافات الموسمية والإقليمية إلى تغييرات في محتوى البروتين والكربوهيدرات والدهون في العوالق النباتية. تؤثر هذه التغييرات على الجودة الغذائية في النظام المائي. في هذه الدراسة تم تحديد هذه التغيرات في العوالق النباتية لبحيرات وادي الريان. تم الكشف عن أقصى محتوى من البروتين (27.95 و 11.36 ملغم لتر⁻¹) في الصيف والشتاء على التوالي. وكان الحد الأقصى لمحتوى الكربوهيدرات في الخريف والشتاء (1.76 و 1.12 ملغم لتر⁻¹ في المواقع 8 و 4 على التوالي). ولوحظ الحد الأقصى لمحتوى الدهون في الشتاء (0.73 ملغم لتر⁻¹ في الموقع 6). كانت كثافة العوالق النباتية أعلى بكثير في البحيرة العليا منها في البحيرة السفلى. تم تحديد 92 نوعاً من العوالق النباتية؛ تضمنت العوالق النباتية في البحيرة 7 مجموعات وهي Chlorophyceae و Cyanophyceae و Bacillariophyceae و Cryptophyceae و Dinophyceae و Chrysophyceae و Euglenophyceae. كانت الطحالب الخضراء (Chlorophyceae) هي الصنف السائد وشكلت 44.4-45.0% من المحصول الكلي للعوالق النباتية عبر الفصول. ومثلت الطحالب الخضراء المزرقة (Cyanophyceae) النسبة 29-39%، وكانت نسبة الدياتومات (Bacillariophyceae) 9.7-16.1% والسوطيات (Dinophyceae) 4.0-9.2% من المجموع الكلي للعوالق النباتية. نادراً ما تم العثور على Cryptophyceae و Chrysophyceae و Euglenophyceae. لوحظت اعلي كثافة للعوالق النباتية (7684 × 10⁴ خلايا لتر⁻¹) في الشتاء. كانت الأنواع الأكثر وفرة التي تم تحديدها هي *Cosmarium nitidulum* (Brebisson) Ralfs و *Dictyosphaerium pulchellum* Wood و *Oocystis solitaria* Wittrock و *Lyngbya limnetica* و *Gomphosphaeria compacta* (Lammermann) Ström و *Lemmermann* و *Microcystis aeruginosa* Kützing و *Cyclotella meneghiniana* و *Syndra ulna* (Nitzsch) Ehr. و Kutz. يتضح مما سبق ان هناك حاجة إلى مزيد من الدراسات التفصيلية حول التركيب الكيميائي الحيوي للعوالق النباتية لفهم استجابة بحيرات وادي الريان بشكل أفضل للتغيرات البيئية العديدة.

الخلاصة:

يمكن اعتبار مجتمعات العوالق النباتية ، والمحتوى الكيميائي الحيوي لها، من المؤشرات الحيوية للنظام البيئي في مواقع مختلفة في بحيرات وادي الريان. ويمكن أن يكون التركيب الكيميائي الحيوي للعوالق النباتية مؤشراً جيداً على قيمتها الغذائية.