BIOLOGICAL CONTROL OF *BLACK MOLD* OF ONION CAUSED BY *Aspergillus niger* BY USING *Saccharomyces cervisiae*

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ABSTRACT

Aspergillus niger (Tieghem) attacked onion plants causing black mold. The tested bioagent *Saccharomyces cervisiae* at three concentrations 0.25%, 0.5% and 1%) were evaluated as bioagent on the mycelial growth, incidence and disease severity of black mold. The bioagent *Saccharomyces cervisiae* was more effective on inhibiting mycelial growth, disease severity and disease incidence at high concentration 1% than 0.25% and 0.5%.

Keywords: Aspergillus niger - Saccharomyces cerevisiae - Biological control - Onion - Black mold

INTRODUCTION

Onion (*Allium cepa. L.*) is one of the most economical field crops in Egypt. Black mold caused by (Aspergillus niger) is one of the most important onion storage diseases. Mousa *et al.*, (1973), Yoo, *et al.*, (1989), Chalutz and Wilson (1990), Tanaka, (1991); Hayden *et al.*, (1994); Köycü and Özer (1997); Chand Goyal and Spotts (1997); Droby *et al.*, (1998); Özer *et al.*, (1999), and Eman, Farrag (2005).

Several studies have documented onion storage losses (Tucker and Drew, 1982 and Suzuki and Cutcliffe 1989).

Mousa *et al.*, (1973) reported up to 80% of bulb stored in sudan were infected by black mold. In Japan, black mold caused more than 60% storage losses during summer (Tanaka 1991).

Therefore disease managements are important for reducing storage losses in Egypt. The modern trends are directed to study the effect of the yeast *(Saccharomyas cerevisiae)* against some pathogens causes plant diseases Chalutz and Wilson 1990; Teixido (1998) Johnes Prusky (2002); Hammouda *et al.*, (2005) Gianluca *et al.*, (2006) Fatten Mansour and Soltan (2006).

The aim of the present work was to study the effect of yeast in controlling black mold disease of onion

MATERIALS AND METHODS

1- Isolation and identification of the causal pathogen:

Samples of onion black mold disease were collected from Assiut Governorate. Isolation procedures was carried out using infected onion bulbs Giza 6 cultivar. Soaking the infected plant parts in 1% sodium hypochlorite solution for 10 minutes followed by complete washing in sterile distilled water for surface disinfestation of infected sample before plating on PDA medium at 25°C for 5-7 days. Pure culture of the developing fungi was obtained by single spore isolation. Primary identification was carried out according Moubasher (1993) Hoog *et al.*, 2000 then was confirmed by Assiut University Mycological Center (AUMC).

2- Pathogenicity test:

Apparantly healthy onion bulbs, free from obvious infection by Aspergillus niger Giza 6 cultivar, were washed thoroughly with tap water, surface sterilized by dipping in 1% sodiumn hypochlorite solution for 2-3 minutes. Then rewashed with sterile distilled water and left for arial at room temperature. Inoculum of Aspergills niger was prepared by growing the two tested isolates on 100ml Potato Dextrose liqued in 250ml. concical flasks, then incubated at 25°C. After 6 days, incubated culture filtrates were decanted and the growing mycelium of each tested isolate was washed using sterile distilled water, suspended in 100ml of distilled water and blended in warning blender for two minutes. Healthy onion bulbs were sprayed with the mycelial suspension 4×10^5 propagulus/ml. Fungal suspension was replaced by distilled water and the bulbs were treated in the same manner as a control treatment. Twenty five of bulbs were put in 30×40 cm. Jute sacks for each treatment replicated four times. Sacks were kept in rows at room temperature about 32°C for three months. A piece of wet cotton with sterilized distilled water was put in each bag to maintain suitable relative humidity around the bulbs. The percentage of infection were recorded after 3 months and disease index of bulbs were recorded after the end of experiment by using scale of o to 4 was used by AICRIP (1968).

Source of Saccharomyces cervisiae

Extract of commericall yeasts (Saccharomyces cervisiae).

In vitro effect of Saccharomyces cerevisiae on linear growth of Aspergillus niger:

Quarter or half or one gram of *Saccharomyces cerevisiae* was dissolved in 100ml of PDA broth medium to give final concentration of 0.25, 0.5 and 1% respectively. Flask (250ml) contained 90ml of sterilized PDA medium were melted, cooled to about 52°C and 10ml of each yeast concentration were separately added to them, mixed and poured in sterilized Petri dishes (20ml/plate). Inoculation was done with fungal discs, (5mm) in diameter obtained from *Aspergillus niger* 7 days old culture. Three replicates were used for each tested concentration. Another group of PDA plates free from *Saccharomyces cerevisiae* inoculated with the fungus as control. All plates were incubated at (25°C) for 7days. Linear growth was recorded until the full growth of comtrol plates. The obtained data were statistically analyzed according to Snedeor and Cochracn (1967).

Effect of *Saccharomyces cerevisiae* on the disease incidence of black mold during storage:

This experiment was carried out in 2005 and 2006 growing seasons. Two isolates of *Aspergillus niger* were used. Healthy onion bulbs were sprayed with the mycelial suspension (4×10^5) propagulus/ml and soaked in different concentrations of *Saccharomyces cerevisiae* 0.25% or 0.5% and 0.1% respectively for 5 minutes. The previously treated bulbs were kept in 30 × 40cm. Jute sacks each containing 25 bulbs and a piece of wetted cotton to

maintain suitable humidity around bulbs. Then kept at room temp. (about 32°). Four replicates were used for each treatment. Healthy onion bulbs were sprayed with the fungal suspension only used as control. After 90 days from inoculation, percent of infection and disease index of bulbs were recorded after the end of experiment by using scale of 0 to 4 which was used by A.I.C.R.I.P. (1968).

Statistical analysis:

The collected data were statistically analyzed using two factors Completely Randomized Block Design. Treatments were compared at 0.05 and 0.01 level of probability L.S.D. (Fisher 1984).

RESULTS AND DISCUSSION

Data presented in Table (1) indicated that all isolates of *Aspergillus niger* causes black mould disease of onion during storge. Data also show that isolate (No.1) was the highly pathogenic than isolate (No. 2).

These results were in agreement with the finding of Mousa *et al.*, 1973; Yoo *et al.*, (1989); Tanaka (1991) Haydan *et al.*, (1994); Köycü and özer (1997); Özer *et al.*, (1999) and Eman Frrag 2005.

Table (1): Pathogencity test of *Aspergillus niger* causal pathogen of black mold of onion:

Isolates	Disease incidence %	Disease severity % 15.5	
I 1	59		
2	57	14.5	
Control	0	0	
L.S.D. 1%	5.243	2.141	
5%	3.461	1.413	

Data in Table (2) indicate that all concentrations of *Saccharomyces cerevisiae* inhibited linear growth of *Aspergillus niger* in *vitro* when compared with the control. The most reduction was obtained when using the concentration 1% followed by 0.5% and 0.25%.

Theses result are in agreement with finally Richard, Jones and Prusky 2002 and Gianluca et al., (2006).

Table (2): Effect of Saccharomyces cerevisiae on the linear growth of Aspergillus niger in vitro:

Saccharomyces	cervisiae	cervisiae Linear growth (m) Mean	
concentration		Isolate ₁	Isolate ₂	Wiedli	
0.25%		55	52	53.5	
0.5%		46.3	45	45.65	
0.1%		31	30	30.5	
Control		89	89	89	
L.S.D.		1%	5%		
I		0.484	0.354		
С		0.689	0.500		
IXC		0.975	0.707		

I Isolates C. concentration of Saccharomyces cerevisiae

IXC isolates interaction X concentration of saccharomyces cerevisiae

Data presented in Table (3) indicated that the tested Saceharomyces cerevisiae reduced the effect of the two testing isolates developing of onion black mold disease after 90 days of storage when compared with the control. The concentration 1% was the most effective in controlling disease incidence and severity through the two tested growing seasons followed by concentration 0.5% and 0.25% respectively this results agree with Jones and Prusky 2002 they found that Saccharomyces cerevisiae inhibited the growth of germinated Colletotrichum coccodes spores and inhibited the decay development caused by Colletotrichum coccodes in tomato fruits. Fatten, Mansour and Soltan 2006 they mentioned that Saccharomyces cerevisiae, Candida teunis, Bacillus subtilis and the bioagent Biozeid at high concentration were the most effective treatment for controling fruit rots caused by Botrytis cinerea and Alternaria alternatia naturally infected persimmon fruit with efficacy exceeded 80% and Gianluca et al., (2006) found that Antagonistic yeasts showing a killer activity against Aspergillus carbonarius and Aspergillus niger on grape.

The use of microorganisms particularly yeasts occurring naturally on the surface of fruits or vegetables, usually has been preferred for the control of post harvest disease Jeffries and Jeger 1990 Chalutz and Wilson 1990; Droby *et al.*, 1991. Janisiewicz 1991; Smilanick *et al.*, 1993 and Janisiewicz *et al.*, 1994.

Isolate		2005 season		2006 season	
	Concentrations	Disease	Disease	Disease	Disease
		incidence %	severity %	incidence %	severity %
1	0.25%	26	6.5	20	5
2		24	4	19	4.75
Mean		25	5.25	19.5	4.87
1	5.0%	19	4.75	18	4.5
2		17	4.25	16	4
Mean		18	4.5	17	4.25
1	1%	17	5.25	15	2.75
2		13	3.25	11	3.66
Mean		15	4.25	13	3.205
1	Control	60	15	56	14
2		56	14	52	13
Mean		58	14.5	54	13.5
S.D.		I 3.230	l 1.044	l 1.850	II 0.654
1%		C 4.567	C 1.477	C 2.616	C 0.654
		IXC 6.459	IXC 2.088	IXC 3.700	IXC 0.926
5%		I 2.383	I 0.771	I 1.365	I 0.342
		C 3.370	C 1.090	C 1.931	C 0.483
		IXC 4.766	IXC 1.541	IXC 2.730	IXC 0.463

Table (3): Effect of *Saccharomyces cerevisiae* on the disease incidence and disease severity of black mold during storage 2005 and 2006 growing seasons:

IXC isolates interaction X concentration of Saccharomyces cervisial.

Yeasts are suitable as bio-control agents of postharvest diseases because they rapidly (i) colonize and survive on fruit surfaces for long periods of time under different conditiones (ii) use available nutrients to proliferate

rapidly, limiting nutrient availability to the pathogen (iii) are generally unaffected by fungicides used commercially: In the end we must use bioagent to save our environment from pollution caused by fungicides.

REFERENCES

- A.I.C.R.I.P. (1968): Progress Report of the all India coordinated Rice Improvement Project Trials. Agronomy. [C.F. Singh, R.P., and S.C. Mdgal (1978). Influence of different levels of nitrogen on the incidence of bacterial leaf blight in Indica rices. India J. Agron., 174-175].
- Chalutz, E., and C.L. Wilson, (1990): Postharvest biocontrol of green and blue mold and sour rot of citrus fruit by *Debaryomyces hansenii*. Plant Dis. 74: 134-137.
- Chand-Goyal, T. and R.A. Spotts (1997): Biological control of postharvest diseases of apple and pear under semi-commercial and commercial conditions using three saprophytic yeast. Biol. Control 10: 282-291.
- conditions using three saprophytic yeast. Biol. Control 10: 282-291. Droby, S., E., Chalutz, and C.L.Wilson (1991): Antagonistic microorganisms as biological control agents of postharvest diseases of fruits and vegetables. Postharvest News Inf. 2: 169-173.
- Droby, S., L., Cohen, A., Daus, B., Weiss, B., Horev, E. Chalutz, H., Katz, M., Keren-Tzur, and A. Shachnai (1998): Commerical testing of Aspire: A yeast preparation for the biological control of postharvest decay of citrus. Biol. Control 12: 97-101.
- Eman Farrage, S.H. (2005): Relation of diseases incidence and storability in some Egyptian onion cultivars. Assiut Journal of Agricultural Science, Vol. 36: 29-36.
- Fatten Mansour S. and H.H.M.Soltan (2006): Controlling persimmon fruit rots in cold storage by pre & post harvest applications of certain Bioagents Minufiya. J. Agric. Res. Vol. 31 No. 4: 775-786.
- Fisher, R.A. (1984): Statistical methods for research workers. Oliver and Boy, London.
- Gianluca, B.; G; Francesco, C.; Giuseppe, L Antonio, and V. Angelo, (2006): Isolation of epiphytic yeasts with potential for biocontrol of *Aspergillus carbonarius* and *A niger* on grape. International Journal of food microbiology, 108n°2: 204-209.
 Hammouda, A. M., Mamdouha, M. Hussien, O. A. Poulot and Nabila, A.
- Hammouda, A. M., Mamdouha, M. Hussien, O. A. Poulot and Nabila, A. Mostafa (2005): Biochemical changes associated with systemica resistant to wheat steam rust, *Puccinia graminis. F.sp. tritici* induced by naturally compounds. Plant pathology research institute Agriculture research center part 1 page 15-34.
- Hayden, N. J.; R.B. Maude and F.J. Proctor (1994): Strategies for the control of black mold (*Aspergillus niger*) on stored tropical onion. Acta Hort., 358: 271-274.
- Hoog, G.S. de, J., Guarro J. Gene and M.J. Figueras (2000): Atlas of Clinical fungi. 2nd edn. Centraalbureau voor Schimmelcultures / Universitat I Virgili pp. 1126.
- Janisiewičz, W. J. (1991): Biological control of postharvest diseases. Pages 301-326 in: Hanbook of Applied Mycology: Soils and Plants. Vol. 1. D. K. Arora, B. Rai, K.G. Mukerju, and K.L. Knudsen, eds. Dekker, New York

Janisiewicz, W.J., D.L. Peterson, and R. Bors (1994): Control of storage decay of apples with *Sporobolomyces roseus*. Plant Dis. 78: 466-470.

Jeffries, P., and M. J. Jeger (1990): The biological control of postharvest diseases of fruit. Postharvest News Inf. 1: 365-368.

- Jones R. W. and D. Prusky (2002): Experssion of an antifungal peptide in Saccharomyces: A new approach for Biological control of the postharvest disease caused Colletotrichum by coccodes. Phytopathology: 92: 33-37. Köycü N.D., N., Özer (1997); Determination of seed borne fungi in onion and
- their transmission to onion sets. Phytoparasitica 25: 25-31.
- Moubasher, A. H. (1993): Soil fungi in Qatar and other Arab countries. The Scientific and Applies Research centre, University of Qatar PP.566.
- Mousa, S.K.; H.A. Habish; A.A. Abdalla and B.B. Adlan (1973): Problems of onion storage in the Sudan. Trop. Sci., 15: 319-327.
 Özer, N. G.chilosi and P.Magro (1999): Polygalacturonase production by
- Aspergillus niger: Expression in onion seeds and possible involvement in virulence, Journal of plant pathology 81 (1) 17-24. Richard W., Jones and D. Prusky (2002): Experssion of an antifungal peptide
- in Saccharomyces: A new approach for Biological control of the Postharvest Disease caused by Colletotrichum coccodes.
- Phytopathology: 92: 33-37. Sendecor, G. W. and W.G. Cochracn (1967): Statistical Methods. Oxford and J.B.H. Publishing Co. 6th edition.
- Smilanick, J.L., R., Denis-Arrue, J.R., Bosch, A. R., Gonzales, D.J., Henson, and W.J. Janisiewicz (1993): Biocontrol of postharvest brown rot of nectarines and peaches by Pseudomonas species. Crop Prot. 2: 513-520.

Suzuki, M. and J.A. Cutclifle (1989): Fructans in onion bulbs in relation to storage life. Can J.plant Sci., 69: 1327-1333.

- Tanaka, K. (1991): Studies on the black mold disease of onion bulbs caused by Aspergillus niger Van Tieghem. Bul. Fac. Agr. Saga Univ., 70: 1-54. Teixido, N; J.; Usall, O. Gutierrez, and I. Vinãs (1998): Effect of the
- antagonist Candida sake on apple surface microflora during cold and ambient (S life) storage. European Journal of Plant Pathology, 104: 387-398
- Tucker, W.G. and R.L.K. Drew (1982): Postharvest study on autumn bulb onions. The effect of harvest date, conditioning treatments and field drying on skin quality and on storage performance. J. Hort. Sci., 57: 339-348.
- Yoo, K. S.; C.R. Andersen; L.M. Pike and K. Sun Yoo (1989): Determination of postharvest losses and storage life of "Texas Grano 1015Y" onion. J. Rio Grande Valley Hort. Soc., 42: 45-50.

المقاومة البيولوجية للعفن الأسود في البصل المتسبب عن اسبرجلس نيجر

باستخدام السكاروميسس سيرفسيا

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يصاب البصل بمرض العفن الأسود ويهدف هذا البحث إلى اختبار الخميرة (سكاروميسس سيرفسيا) بثلاث تركيزات (٢٠,٠% و ٥,٠% و ١%) في مقاومة الفُطر المسبب للمرض في الدراسة المعملية لوحظ أن التركيز العالى من الخميرة ١% كان أكثر فاعلية في مقاومة العفن الأسود في البصل بالمقارنة بالتركيز ٥, • % و٢٥%. ونفس النتائج المتحصل عليها في المعمل أعطت نتائج مماثلة لها في المخزن تحت ظروف العدوى الصناعية.