

Plant Production Science

Available online at http://zjar.journals.ekb.eg http:/www.journals.zu.edu.eg/journalDisplay.aspx?Journalld=1&queryType=Master



EFFECT OF SPRAYING NANO-CHITOSAN AND NANO-SILICON ON PHYSICOCHEMICAL FRUIT QUALITY AND LEAF MINERAL CONTENT OF FLORIDA PRINCE PEACH TREES

Nabil G. Soliman, Safaa A.A. Nomier, M.M. Ibrahim^{*} and M.M. Gad

Hort. Dept. Fac. Agric., Zagazig Univ., Egypt

Received: 29/08/2021 ; Accepted: 17/10/2021

ABSTRACT: This work was carried out in the two successive seasons of 2016 and 2017 on fiveyear-old peach Florida prince cv. trees (*Prunus persica* L.) grown in sandy silt soil at 3×5 m apart under drip irrigation system of a private peach orchard located at Belbies district, Sharkia Governorate, Egypt. The tested trees sprayed with nano-chitosan at 10, 20, 30 and 40 ppm, nanosilicon at 200, 400 and 600 ppm and potassium silicate at 1000, 2000 and 3000 ppm as well as water (control treatment). The results showed that spraying of nano-chitosan at all rates exhibited the highest TSS/acid ratio in the fruit juice without significant differences between them in the two seasons. Untreated trees had a highest vit. C content in fruits compared to other treatments. Also, 3000 ppm potassium silicate at 3000 ppm gained highest leaf mineral contents of N, P, K, Fe, Zn and Mn, while, lowest leaf mineral contents of Fe, Zn and Mn were recorded with control and nanochitosan at 10 ppm.

Key words: Peach, Nano, silicon, chitosan, fruit quality, leaf mineral content.

INTRODUCTION

Peach fruits are delicious taste and unique flavor with high nutritional value have popularized it across the world. It's the most popular stone fruits in the world because of its high nutrient level and pleasant flavor. Peach fruits are enriched with ascorbic acid, carotenoids (provitamin A), phenolic compounds and are considered prime sources for antioxidants (Tomas-Barberan et al., 2001; Byrne, 2002). Florida Prince is an early ripening cultivar under the Egyptian environmental conditions it starts to ripe in April, two months earlier than the European peaches cvs (Stino et al., 2010). The production and commercialization of stone fruits like peaches have increased briskly throughout the world.

The use of Nano applications on fruit trees contributes very effectively to improving the quality of fruits and increasing the productivity of trees by improving nutrient management in modern agriculture as well as increasing the storing potential of fruits, as it was noted that the use of Nano fertilizer in the agricultural field preserves the soil. It reduces their pollution by reducing the amount of fertilizer used, which is positively reflected in the increased economic return of the farmer (Malerba and Cerana, 2016; Al-Hchami and Alrawi, 2020).

Chitosan has been used in agriculture as a coating material for vegetables, fruits and seeds (**Photchanachai** *et al.*, **2006**). Chitosan, a polycationic polymer of (3-1,4, linked D-glucosamine chemically derived from crustaceans and soluble in organic acids is one of a range of natural compounds that have been successfully used to maintain the quality of harvested fruits and vegetables (**Li and Yu, 2001 and Dong** *et al.*, **2004**). Plants treated with chitosan may be less prone to stress evoked by unfavorable conditions, such as drought, salinity, low or high

^{*} Corresponding author: Tel. :+201005456118 E-mail address: mohamednasr79@yahoo.com

temperature (Liu *et al.*, 2011 and Shao *et al.*, 2015). Application of chitosan increased key enzymes activities of nitrogen metabolism (nitrate reductase, glutamine synthetase and protease) and improved the transportation of nitrogen (N) in the functional leaves which enhanced plant growth and development and increase the yield (Mondal *et al.*, 2013).

Silicon considered an essential element for higher plants because silicon deprived plants tend to grow abnormally, whereas silicon supplemented plants grow normally (Artyszak, 2018). The application of silicon in fertilization of plants has a positive influence on the growth, development and yield of plants (Hogendorp, 2008; Górecki and Danielski-Busch, 2009 and Mohaghegh *et al.*, 2010).

The aim of this work is improving growth, fruit weight and its quality of Florida prince peach trees by using some natural growth promoters as alternatives to synthetic growth regulators.

MATERIALS AND METHODS

This study carried out during two seasons of 2016 and 2017 on five-year-old peach Florida prince cv. trees (Prunus persica L.). The trees are grown in a private peach orchard at Belbies district, Sharkia Governorate, Egypt. The trees were planted at 3 x 5 m apart, in silt sandy soil under drip irrigation system. The usual agriculture practices for peach trees in the orchard will be adapted to all trees. The tested treatments could be summarized as follows: Spray trees with tap water (control), spray trees with nano-chitosan at 10, 20, 30 and 40 ppm, spray trees with nano-silicon at 200, 400 and 600 ppm and spray trees with potassium silicate at 1000, 2000 and 3000 ppm. The selected trees were sprayed three times at 25% of full bloom (on 15 Dec.), 50% of full bloom (on 30 Dec.) and 75% of full bloom (on 10 Jan.), in addition, fourth spray after fruit thinning (on 15 Feb.). Each of the previous 11 spraying treatments has been supplied to 3 Florida prince peach trees.

Nano Chitosan and Silicon Preparation

The stock solution of chitosan (2% W/V) was prepared by dissolving chitosan powder in

2% acetic acid as described by **Park** *et al.* (2002). Chitosan nanoparticles was prepared by addition of 1ml aqueous tripolyphosphate solution (0.25%, w/v) to 3mL of chitosan solution under magnetic stirring. The nano chitosan particle size was characterized and described by **Qi** *et al.* (2004).

Potassium silicate of nano crystallite powder synthesized by high-energy ball milling by prof. Dr. Osama M. Hemeda at Central lab., department of physics, faculty of science, Tanta University, Egypt.

The responses of the tested trees to the applied treatments were evaluated through the following characteristics:

Fruit characteristics

At time of harvesting (end of April in both seasons), 10 fruits were randomly collected from each replicate to determine the following fruit characteristics:

- 1. Fruit length (cm) and diameter (cm) were measured by using Vernier caliper.
- 2. Fruit volume (cm³): was determining by immersing fruits in water in a graduated cylinder.
- 3. Fruit firmness (g/cm²) was determined by using a push pull Dynamometer.
- 4. Total soluble solids: acidity ratio (TSS: acid ratio).
- 5. Vitamin C content as mg ascorbic acid / 100 ml juice was determined by titration against 2, 6-dichlorophenol endo phenol dye as index (AOAC, 2006).
- 6. Total sugars (%): were determined in juice according to the method of Lane and Eynon as described in the **AOAC** (2006).
- 7. Carbohydrate percentage: were determined colorimetrically according to the method described by **Smith and Dubois** (1956).

Leaf mineral content

The sample of leaves were taken from the third of shoot top. The middle part of the blade free from the midrib was cut. Samples of 200 g of fresh leaves were cleaned and washed several times with tap water, the leaf samples be air dried then put in an electrical furnace at 60 - 70°C for 48 h. till constant weight and finally ground. An adequate processed sample was providing to determine the following minerals:

- Total nitrogen was determined by modified microkjeldahl method as outlined by **Black** *et al.* (1965).
- Phosphorus content was determined calorimetrically according to Chapman and Pratt (1975).
- Potassium content was determined by using flame photometer (Browen and Lilleland, 1964).
- Iron, zinc and manganese were determined according to the standard method described by **Jackson** (1958). The concentration was expressed as a percentage of dry weight bases.

Statistical Analysis

The obtained data were statistically analyzed according to the randomized complete block design with 3 replicates and one tree for each replicate and subjected to analysis of variances (ANOVA) according to **Snedecor and Cochran** (1990) using CoStat program. Furthermore, means were compared using mean comparison at 0.05 level (**Duncan**, 1958).

RESULTS AND DISCUSSION

Effect of Foliar Spray on Fruit Quality

Fruit length and diameter (cm)

It is quite evident from Table 1, that fruit dimensions; i.e.; length and diameter were significantly affected by the tested treatments in both seasons. However, trees sprayed with nanochitosan, nano-silicon and potassium silicate markedly increased fruit dimensions in comparison with those of water sprayed trees. Treatments of nano-chitosan, nano-silicon and potassium silicate recorded insignificantly differences between them in the first season only. While, in the second season the longest fruits were from nano-chitosan at 20 (5.72 cm) and 30 (5.76 cm) and trees sprayed with potassium silicate at 2000 (5.82 cm) and 3000 ppm (5.84 cm) without significant differences between them. The trees sprayed with water produced the shortest length (4.97 and 4.33 cm)

and diameter (5.22 and 4.75 cm) values in the first and second seasons, respectively. As a general, trees spayed with 3000 ppm potassium silicate exhibited higher dimensions than other spraying treatments, while that water spayed produced fruits with smallest length and diameter.

These results are in parallel with those reported by Lalithya *et al.* (2014a&b) on sapota; El-Gioushy (2016) and Kotb and Abdel-Adl (2017) on orange; Mohamed (2016) on olives; Patil and Jagadeesh (2016) on banana; El Kholy *et al.* (2018) on loquat. They indicated that, silicon or potassium silicate enhanced fruit physical properties as fruit dimensions (length and diameter).

Alwea (2018) and Elsheery *et al.* (2020) reported that, nano-silicon foliar spray improved physiochemical characteristics of mango fruits.

Grapevines were sprayed with nano chitosan exhibited significantly higher berry length (**Ibrahim** *et al.*, **2019**). Also, mango foliar spray with nano chitosan improved fruit physical properties (Alwea, 2018).

Fruit volume (cm³)

Data presented in Table 1 emphasized that, fruit volume was significantly affected by the studied treatments in both seasons. Fruit volume of Florida prince peach fruits ranged between 85.00 - 120.00 and 54.67 - 115.67 cm³ in the first and second seasons, respectively. The largest fruit volume was recorded by potassium silicate at 3000 ppm (120.00 and 115.67 cm³) in the two seasons, respectively, without significant differences between it and those treated with 600 ppm nano-silicon (113.00 cm³) in the first season and treatments of potassium silicate at 2000 ppm (109.33 cm³) and nano-chitosan at 20 & 40 ppm (110.00 & 114.00 cm³) in the second one, respectively. Unsprayed trees (control) produced the lowest fruit volume (85.00 and 54.67 cm^3) in the first and second seasons, respectively. The other treatments gained intermediate fruit volumes.

Similar results were stated by application of potassium silicate or nano silicon enhanced fruit quality and increased fruit size (Lalithya *et al.*, 2014 a &b on sapota; Verma *et al.*, 2017 on mandarin; Youssef, 2017 on date palms; El Kholy *et al.*, 2018 on loquat; Elsheery *et al.*, 2020 on mango).

Soliman, et al.

| Spraying treatments | | First season (2015) | | | | Second season (2016) | | | |
|------------------------|----------|-------------------------|---------------------------|---|---------------------------------------|-------------------------|---------------------------|---|---------------------------------------|
| | | Fruit length (cm) | Fruit diameter (cm) | Fruit firmness (g/cm ²) | Fruit volume (cm ³) | Fruit length (cm) | Fruit diameter (cm) | Fruit firmness (g/cm ²) | Fruit volume (cm ³) |
| Control | | 4.97 c | 5.22 c | 1774.0 a | 85.00 f | 4.33 d | 4.75 d | 1780.6 cd | 54.67 g |
| Nano-chitosan | 10 ppm | 5.55 ab | 5.76 ab | 1729.0 ab | 102.80 de | 5.13 b | 5.66 abc | 1772.2 d | 94.67 def |
| | 20 ppm | 5.58 ab | 5.83 ab | 1658.5 b | 104.00 cde | 5.72 a | 5.93 ab | 1797.2 bc | 110.00 abc |
| | 30 ppm | 5.23 bc | 5.55 bc | 1707.0 ab | 101.00 e | 5.76 a | 5.96 a | 1798.3 bc | 114.00 ab |
| | 40 ppm | 5.43 ab | 5.66 ab | 1706.0 ab | 111.10 bc | 4.73 c | 4.90 c | 1788.9 bcd | 82.00 f |
| Nano-silicon | 200 ppm | 5.49 ab | 5.61 ab | 1774.0 a | 108.47 b-e | 4.72 c | 5.29 c | 1829.2 a | 87.33 ef |
| | 400 ppm | 5.60 a | 5.82 ab | 1450.0 c | 105.00 cde | 5.31 b | 5.46 bc | 1805.6 b | 102.00 bcd |
| Na | 600 ppm | 5.61 a | 5.75 ab | 1768.0 a | 113.00 ab | 4.63 cd | 5.33 c | 1801.4 b | 102.00 bcd |
| otassium silicate | 1000 ppm | 5.33 abc | 5.48 bc | 1743.7 a | 110.00 bcd | 5.27 b | 5.62 abc | 1797.2 bc | 97.33 cde |
| | 2000 ppm | 5.36 ab | 5.48 bc | 1757.0 a | 103.00 de | 5.82 a | 6.01 a | 1787.5 bcd | 109.33 abc |
| | 3000 ppm | 5.67 a | 5.94 a | 1787.3 a | 120.00 a | 5.84 a | 6.05 a | 1798.6 bc | 115.67 a |

 Table 1. Effect of spraying treatments on some physical characteristics of Florida prince peach fruits in 2015 and 2016 seasons

Means having the same letter(s) within the same column are not significantly different according to Duncan's multiple range test at 5% level of probability.

Foliar spray of chitosan or nano-chitosan improved fruit physical quality (Zagzog *et al.*, 2017, Zahedi *et al.*, 2020 on mango; Mohamed and Ahmed, 2019 on orange and Ibrahim *et al.*, 2019 on grapevine).

Fruit firmness (g/cm²)

It is clear from Table 1 that, peach fruit firmness was significantly affected by the spraying treatments in the two seasons. Generally, the hardness of Florida prince peach fruits was significantly affected by spraying treatments, where the values ranged between 1450.0 - 1787.3 and 1772.2 - 1829.2 g/cm² in the first and second seasons, respectively. Anyhow in the first season, the highest fruit firmness was gained by trees sprayed with potassium silicate at 3000 ppm (1787.3 g/cm²) followed by other spraying treatments without significant differences between them, except the trees treated at 400 ppm nano-silicon which recorded the lowest value (1450.0 g/ cm²). But in the second season, the trees sprayed with 200 ppm nano-silicon gave the highest fruit firmness (1829.2 g/cm²) compared by those sprayed by water and nano-chitosan at 10 ppm which gave the lowest values (1780.6 and 1772.2 g/cm²), respectively without significant differences between them, and the other treatments produced significantly differences in-between fruit firmness.

These results agreed with those reported by El-Badawy (2012) on peach, Giacalone and Chiabrando (2015) on nectarines, Ahmed *et al.* (2016) on orange, Gad *et al.* (2016) on peach, Zagzog *et al.* (2017) and Zahedi *et al.* (2020) on mango and Mohamed and Ahmed (2019)

on orange. They stated that chitosan or nanochitosan applications mostly had positive effects on improving fruit quality and maintained fruit pulp firmness.

The hardness or firmness and anti-stress abilities of fruits were improved and increased by using silicon (Jia, 2011; Jia *et al.*, 2011 on apple and grape; Kotb and Abdel-Adl, 2017 on orange; Youssef, 2017 on date palms; El Kholy *et al.*, 2018 on Loquat; Elsheery *et al.*, 2020 on mango). While, Su *et al.* (2011) said that, the apple fruit hardness did not affect.

TSS/ acid ratio

It is clear from Table 2 that, the tested spraying treatments significantly affected TSS/acid ratio in fruit juice in the two seasons. The other treatments gave intermediate insignificantly different ratios. The TSS/acid ratio in the juice of Florida prince peach fruits ranged between 5.93 - 14.04 and 5.77 - 10.70 in the first and second seasons, respectively. Anyhow, spraying of nano-chitosan at all rates (10, 20, 30 and 40 ppm) (12.77 & 10.70, 14.04 & 8.59, 12.45 & 9.36 and 12.90 & 7.68) and nano-silicon at 200 ppm (11.28 & 8.59) and 400 ppm (12.48 & 7.91) exhibited the highest TSS/acid ratio in the fruit juice without significant differences between them in the two seasons, respectively, and those treated with potassium silicate at 2000 ppm (7.86) and at 3000 ppm (9.10) in the second season only. Control trees gained the lowest TSS/acid ratios (5.93 and 5.77) in the two seasons, respectively. TSS/acid ratio in the fruit juice was markedly increased due mainly to reducing juice total acidity percentage in each season.

These results came in line with those of Mondal et al. (2013) on mungbean, Zagzog et al. (2017), Alwea, (2018) and Zahedi et al. (2020) on mango, Mohamed and Ahmed (2019) on orange, Ibrahim et al. (2019) on grapevines, they reported that application of chitosan or nano-chitosan increased TSS/acid ratio in fruit juice. On the other hand, Gad et al. (2016) found that decreased TSS/acid ratio in peach fruit juice.

Similar results were stated by application of potassium silicate in normal or nano form enhanced fruit quality and increased TSS/acid ratio in fruit juice (Su *et al.*, 2011 on apple; El-Gioushy, 2016 on orange; Youssef, 2017 on date palms; Abd-Elall and Hussein, 2018 on orange; El Kholy *et al.*, 2018 on loquat; Alwea, 2018 and Elsheery *et al.*, 2020 on mango).

Juice Vitamin C content (mg ascorbic acid/ 100 ml juice)

Data illustrated in Table 2, indicated that ascorbic acid (vit. C) content in the fruit juice was significantly affected by the tested treatments in both tested seasons. However, vit. C content in Florida prince peach fruits ranged between 23.00 - 31.67 and 26.40 - 46.80 mg/ 100 ml juice in the first and second seasons, respectively. Fruits on unsprayed trees (control) produced the highest vit. C content (31.67 and 46.80 mg/100 ml juice) of juice in the two seasons, respectively, followed by those trees treated by potassium silicate at 1000, 2000 and 3000 ppm (28.67, 28.00 and 28.00 mg/100 ml juice) in the first season, respectively without significant differences between them and control and 10 nano-chitosan treatment (34.00 mg/100 ml juice) in the second one only. The lowest vit. C content (23.00 and 26.40 mg/100 ml juice) was found in fruit juice on trees treated by 20 ppm nano-chitosan in the both seasons, respectively, and trees sprayed with 200 ppm nanosilicon without significant differences between them in the first season only. The other treatments gave intermediate contents and recorded insignificant differences lower vit. C contents.

The obtained data were in line with those stated by **Jitareerat** *et al.* (2007) on mangoes and **Xing** *et al.* (2015) on jujube fruits. They showed that chitosan reduced ascorbic acid in fruit juice. On the contrast, chitosan or nano-chitosan increased ascorbic acid in fruit juice according to Zagzog *et al.* (2017), Alwea (2018) and Zahedi *et al.* (2020) on mango and Mohamed and Ahmed (2019) on orange.

Jia (2011) and Jia *et al.* (2011) on nectarine, Su *et al.* (2011) on apple fruit, Youssef (2017) on date palms, El Kholy *et al.* (2018) on loquat, Alwea (2018) and Elsheery *et al.* (2020) on mango, they found that application of silicon or nano-silicon increased ascorbic acid in fruit juice. Soliman, et al.

| Spraying treatments | | First season (2015) | | | | Second season (2016) | | | |
|------------------------|----------|------------------------|------------------------------|------------------------|---------------------|----------------------|------------------------------|------------------------|---------------------|
| | | TSS / acid ratio | Vitamin C (mg/ 100 ml) | Total sugars (%) | Carbohydrate (%) | | Vitamin C (mg/ 100 ml) | Total sugars (%) | Carbohydrate (%) |
| Control | | 5.93 c | 31.67 a | 12.11 k | 7.03 k | 5.77 c | 46.80 a | 12.69 k | 7.29 k |
| Nano-chitosan | 10 ppm | 12.77 a | 26.00 bcd | 14.05 j | 7.15 ј | 10.70 a | 34.00 a | 14.67 j | 7.65 j |
| | 20 ppm | 14.04 a | 23.00 d | 14.17 i | 7.33 i | 8.59 abc | 26.40 c | 14.93 i | 7.92 i |
| | 30 ppm | 12.45 a | 26.67 bcd | 14.61 h | 7.51 h | 9.36 ab | 35.60 b | 15.06 h | 8.02 h |
| | 40 ppm | 12.90 a | 25.67 bcd | 14.79 g | 7.89 g | 7.68 bc | 32.00 bc | 15.32 g | 8.18 g |
| Nano-silicon | 200 ppm | 11.28 ab | 23.33 d | 15.28 f | 8.01 f | 8.59 abc | 30.40 bc | 15.64 f | 8.55 f |
| | 400 ppm | 12.48 a | 24.67 cd | 15.44 e | 8.19 e | 7.91 abc | 29.60 bc | 15.93 e | 8.92 e |
| | 600 ppm | 8.50 bc | 26.33 bcd | 16.02 d | 8.42 d | 7.46 bc | 36.80 b | 16.36 d | 9.16 d |
| Potassium silicate | 1000 ppm | 8.28 bc | 28.67 ab | 16.29 c | 8.75 c | 6.75 bc | 30.00 bc | 16.93 c | 9.37 c |
| | 2000 ppm | 8.79 bc | 28.00 abc | 17.05 b | 9.20 b | 7.86 abc | 32.00 bc | 17.37 b | 9.72 b |
| | 3000 ppm | 8.24 bc | 28.00 abc | 18.78 a | 9.42 a | 9.10 ab | 31.60 bc | 19.08 a | 10.18 a |

Table 2. Effect of spraying treatments on vitamin C, total sugars % and carbohydrate % ofFlorida prince peach fruit juice in 2015 and 2016 seasons

Means having the same letter(s) within the same column are not significantly different according to Duncan's multiple range test at 5% level of probability.

Total Sugars and Carbohydrate Percentages

Data presented in Table 2 indicated that the total sugars and carbohydrate percentages in fruit juice of Florida prince peach trees was significantly affected with spraying nanochitosan, nano-silicon and potassium silicate in two seasons. The highest total sugars and carbohydrate percentages (18.78, 9.42, 19.08 and 10.18%) were found in fruit juice of trees sprayed with 3000 ppm potassium silicate in the first and second seasons, respectively, followed by those sprayed with 2000 and 1000 ppm potassium silicate with significant differences between them in both seasons. The lowest total sugars and carbohydrate percentages (12.11, 7.03, 12.69 and 7.29%) were recorded for trees spraved with water (cont.) in the two seasons,

respectively. The other treatments gained intermediate significantly different total sugars and carbohydrate percentages in each season.

Generally, Florida prince peach trees sprayed with potassium silicate gave total sugars and carbohydrate percentages in fruit juice (20.54, 11.49, 18.68 and 11.26% for total sugars (%), and 22.09, 11.08 and 22.92, 9.91% for carbohydrate %) higher than those treated with nano-chitosan and nano-silicon.

These results were in accordance with those found by **Ahmed** *et al.* (2016) on orange, **Mohamed and Ahmed** (2019) on sugars orange and **Hidangmayum** *et al.* (2019) on many plants. They cleared that, foliar spray of chitosan or nano-chitosan had a significant improvement of chemical fruit properties and induces production sugars in fruits.

1220

In addition, Si applications were very effective on improving fruit quality increased sugars (Shi *et al.* (2010) on grapevine; Jia, 2011 and Jia *et al.* (2011) on apple and grapes; Rong (2011) on cherry; Hanumanthaiah *et al.* (2015) on banana; Badran (2016) on date palms; El-Gioushy (2016) on orange; Patil and Jagadeesh (2016) on banana; Youssef (2017) on date palms; El Kholy *et al.* (2018) on loquat; Elsheery *et al.* (2020) on mango).

Leaf Mineral Content

Nitrogen (N), phosphorus (P) and potassium (K) percentages

As shown in Table 3, there are significant varietal differences in leaf mineral content in both seasons. Since, N percentage ranged between 1.87 - 2.44% & 1.97 - 2.91% and P percentage ranged between 0.220 - 0.387% & 0.226 – 0.396 % as well as K percentage ranged between 1.05–1.86% & 1.08 – 1.88% in the first and second seasons, respectively. The treatment of potassium silicate at 3000 ppm gained highest leaf mineral contents of N (2.44 & 2.91%), P (0.387 & 0.396%), K (1.86 & 1.88%) and significantly increase of values with other treatments in two seasons, respectively, except N% in the first season only, whereas the trees sprayed with potassium silicate at 2000 ppm gave high value insignificant differences with potassium silicate at 3000 ppm in the first season. Generally, the lowest mineral contents of N (1.87 & 1.97%), P (0.220 & 0.226%), K (1.05 & 1.08%) were recorded with nano-chitosan treatment at 10 ppm in the two seasons, respectively, also, nano-chitosan treatment at 20 ppm gained low N % (1.93%) in the first season and K% (1.13%) in the second one without significant differences with nano-chitosan at 10 ppm. The leaves of other treatments gave intermediate values of N, P and K% in both seasons.

These results were in accordance with those found by Ahmed *et al.* (2012) on K mango; Al-Wasfy (2013) on date palms; Al-Wasfy (2014) on grapevines; Lalithya *et al.* (2014 a &b) on Sapota; Abd El-Rahman (2015) on mango; Mohamed *et al.* (2015) on mango; Rizwan *et al.* (2015) on many plants; Mohamed (2015) on pomegranate; Nagy-Dina (2015) on grapevines; El-Gioushy (2016) on orange; Kotb and Abdel-Adl (2017) on orange; Mohamed (2017) on grapevines; Verma *et al.* (2017) on mandarin; Elsheery *et al.* (2020) on mango. They concluded that foliar sprays potassium silicate in normal or nano form increased leaf mineral content. The application of silicic acid enhanced uptake of essential nutrient and improving nutrient content (**Bhavya** (2010) on grapevines; Neeru *et al.* (2016) on rice plants and Javaid and Misgar (2017) on apple).

Foliar application of chitosan or nano chitosan increased leaf mineral content (Ahmed *et al.* (2016) on orange; Khafagy (2018) on grapevines; Abdel-Aziz *et al.* (2016) on orange; Alwea (2018) on mango).

Iron (Fe), zinc (Zn) and manganese (Mn) contents (ppm)

Data in Table 4 demonstrated that, leaf Fe. Zn and Mn contents (ppm) were significantly affected by the tested treatments in the two seasons. Fe content ranged between 193.25 -284.2 & 201.60 - 292.54 ppm and Zn content ranged between 26.17 - 51.22 & 29.23 - 54.16 ppm as well as Mn content ranged between 0.228 - 0.377 & 0.237 - 0.442 ppm in the first and second seasons, respectively. The leaves on the trees were sprayed with potassium silicate at 3000 ppm recorded highest leaf Fe (284.21 & 292.54 ppm), Zn (51.22 & 54.16 ppm) and Mn (0.377 & 0.442 ppm) contents in the first and second season, respectively. Also, in the second season only leaf Mn content at all levels of nano-chitosan, nano-silicon and potassium silicate were insignificant differences between them. The least mineral contents of Fe (193.25 & 201.60 ppm) and Zn (26.17 & 29.23 ppm) from leaves trees were sprayed with nanochitosan at 10 ppm in the two seasons, respectively, and also leaves of control gave lowest Mn content (0.228 ppm & 0.237 ppm) in the first and second season, respectively. The leaves of other treatments gave intermediate values of Fe, Zn and Mn ppm in both seasons.

The obtained findings are in agreement with those reported by **Al-Wasfy (2014)** on grapevines, **El-Gioushy (2016)** on orange, **Kotb and Abdel-Adl (2017)** on orange and **Alwea (2018)** on mango. They indicated that leaf Fe, Zn, Mn and Cu contents were increased by all investigated silicate spray treatments in normal or nano form.

Ahmed *et al.* (2016) on orange and Alwea (2018) on mango, they mentioned that foliar spray of chitosan or nano-chitosan increased values of Zn in leaves.

Soliman, et al.

| Spraying treatments | | Fir | rst season (20 |)15) | Second season (2016) | | | |
|------------------------|---------------|------------------|----------------|--------|----------------------|------------------|------------------|--|
| | | N content (%) | | | N content (%) | P content (%) | K content (%) | |
| Control | | 2.19 d | 0.320 d | 1.23 h | 2.24 fg | 0.327 d | 1.30 e | |
| Nano-chitosan | 10 ppm | 1.87 g | 0.220 k | 1.05 k | 1.97 i | 0.226 j | 1.08 g | |
| | 20 ppm | 1.93 g | 0.232 j | 1.12 j | 2.15 h | 0.247 i | 1.13 g | |
| | 30 ppm | 2.08 ef | 0.254 i | 1.18 i | 2.29 e | 0.263 h | 1.20 f | |
| | 40 ppm | 2.18 d | 0.276 h | 1.29 f | 2.40 d | 0.288 g | 1.31 e | |
| Nano-silicon | 200 ppm | 2.05 f | 0.287 g | 1.24 g | 2.19 gh | 0.299 f | 1.25 ef | |
| | 400 ppm | 2.14 de | 0.293 f | 1.38 e | 2.27 ef | 0.3030 f | 1.38 d | |
| | 600 ppm | 2.27 c | 0.304 e | 1.42 d | 2.35 d | 0.314 e | 1.43 cd | |
| Potassium silicate | 1000 ppm | 2.33 b | 0.349 c | 1.46 c | 2.52 c | 0.352 c | 1.46 c | |
| | 2000 ppm | 2.44 a | 0.361 b | 1.62 b | 2.73 b | 0.374 b | 1.64 b | |
| | 3000 ppm | 2.44 a | 0.387 a | 1.86 a | 2.91 a | 0.396 a | 1.88 a | |

Table 3. Effect of spraying treatments on N, P and K percentages in Florida prince peach leaves in 2015 and 2016 seasons

 Table 4. Effect of spraying treatments on Fe, Zn and Mn contents (ppm) in Florida prince peach leaves in 2015 and 2016 seasons

| Spraying treatments | | Fi | rst season (2 | 015) | Second season (2016) | | | |
|------------------------|----------|---------------------|---------------------|---------------------|----------------------|---------------------|---------------------|--|
| | | Fe content (ppm) | Zn content (ppm) | Mn content (ppm) | Fe content (ppm) | Zn content (ppm) | Mn content (ppm) | |
| Control | | 221.13 e | 42.22 d | 0.228 i | 229.46 e | 44.98 d | 0.237 b | |
| e | 10 ppm | 193.25 k | 26.17 k | 0.254 h | 201.60 k | 29.23 k | 0.371 ab | |
| hitosa | 20 ppm | 200.36 j | 27.37 ј | 0.272 g | 208.55 j | 31.03 ј | 0.384 ab | |
| Nano-chitosan | 30 ppm | 209.86 i | 30.76 i | 0.299 e | 217.65 i | 33.39 i | 0.307 ab | |
| Z | 40 ppm | 218.78 g | 34.12 g | 0.311 c | 226.70 g | 37.03 g | 0.323 ab | |
| con | 200 ppm | 215.13 h | 33.03 h | 0.291 f | 223.45 h | 36.09 h | 0.300 ab | |
| Nano-silicon | 400 ppm | 220.43 f | 36.52 f | 0.305 d | 228.67 f | 39.28 f | 0.314 ab | |
| Nar | 600 ppm | 237.27 d | 40.712 e | 0.312 c | 245.40 d | 43.37 e | 0.320 ab | |
| E . | 1000 ppm | 244.12 c | 44.09 c | 0.304 d | 252.67 c | 47.01 c | 0.312 ab | |
| Potassium silicate | 2000 ppm | 266.28 b | 46.37 b | 0.319 b | 274.77 b | 49.20 b | 0.327 ab | |
| | 3000 ppm | 284.21 a | 51.22 a | 0.377 a | 292.54 a | 54.16 a | 0.442 a | |

REFERENCES

- AOAC (2006). Official method of analysis of the association of agricultural chemists, Twelfth edition, Washington DC, USA.
- Abd El-Rahman, M.M.A. (2015). Relation of spraying silicon with fruiting of Keitte Mango trees growing under upper Egypt conditions. Stem Cell, 6: 1–5.
- Abd-Elall, E.E.H. and M.A. Hussein (2018). Foliar Application of Micro Silica, Potassium Chloride and Calcium Chloride Enhances Yield and Fruit Quality of Balady Orange Tree. Alex. Sci., 39 (3): 387-392.
- Abdel-Aziz, H.M.M., M.N.A. Hasaneen and A.M. Omer (2016). Nano chitosan-NPK fertilizer enhances the growth and productivity of wheat plants grown in sandy soil. Spanish J. Agric. Res., 14 (1): 1 – 9.
- Ahmed, A.H.H., M.R. Aboul-Ella Nesiem, H.A. Allam and A.F. El-Wakil (2016). Effect of pre-harvest chitosan foliar application on growth, yield and chemical composition of Washington navel orange trees grown in two different regions. Afr. J. Biochem. Res., 10 (7): 59-69.
- Ahmed, F.F., A.E.M. Mansour, A.Y. Mohamed, E.A.M. Mostafa and N.E. Ashour (2012). Using silicon and salicylic acid for promoting production of Hindy Bisinnara mango trees grown under sandy soil. Middle East J. Agric. Res., 2: 51–55.
- Al-Hchami, S. H. J. and T. K. Alrawi (2020). Nano fertilizer, benefits and effects on fruit trees: A review. Plant Archives, 20 (1):1085-1088.
- Al-Wasfy, M.M. (2013). Response of Sakkoti date palms to foliar application of royal jelly, silicon and vitamins B. J. Ame. Sci., 9 : 315–321.
- Al-Wasfy, M.M.M. (2014). The synergistic effects of using silicon with some vitamins on growth and fruiting of flame seedless grapevines. Stem Cell, 5: 8–13.
- Alwea, T.G.A. (2018). Using some nano particle materials to enhance growth, fruiting and resistance malformation of mango. M.Sc.

Thesis, Plant Prod. Dept. Fac.Techno. and Develop. Zagazig Univ., 121.

- Artyszak, A. (2018). Effect of silicon fertilization on crop yield quantity and quality-A literature review in Europe. Plants, 7(3): 54.
- Badran, M.A. (2016). Effect of spraying seaweed extracts and silicon on yield and fruit quality of Zaghloul date palms grown under sandy soil conditions. Assiut J. Agric. Sci., 47 (5): 165-174.
- Bhavya, H.K. (2010). Effect of foliar silicic acid and boron in Bangalore blue grapes. Univ. Agric. Sci. GKVK, Bangalore, Thesis M.Sc., 111.
- Black, C.A., D.D. Evans, L.E. Evsminger, J.L. White and F.E. Clark (1965). Methods of soil analysis. Ame. Soc. Agron., Inc., Publ. Madison, Wisconsin, USAP, 1162 – 1168.
- Browen, J.D. and O. Lilleland (1946). Rapid determination of potassium and sodium in plant material and soil extracts by flame photometer. Proc. Amer. Soc. Hort. Sci., 48: 341 346.
- Byrne, D.H. (2002). Peach breeding trends. Acta Hort., 592: 49–59.
- Chapman, H.D. and P.F. Pratt (1975). Methods of analysis for soils, plants and water. Univ. of California. Divison. Agric. Sci., 172–173.
- Dong, H., L. Cheng, J. Tan, K. Zheng and Y. Jiang (2004). Effect of chitosan coating on quality and shelf life of peeled litchi fruit. J. Food Eng., 64: 335-358.
- Duncan, D.B. (1958). Multiple range and Multiple F test. Biomet., 11:1-42.
- El Kholy, M.F., A.A. Mahmoud and S.M.A. Mehaisen (2018). Impact of potassium silicate sprays on fruiting, fruit quality and fruit storability of Loquat trees. Mid. East J. Agric. Res., 7(1): 139-153.
- El-Badawy, H.E.M. (2012). Effect of Chitosan and Calcium Chloride Spraying on Fruits Quality of Florida Prince Peach Under Cold Storage. Res. J. Agric. and Biol. Sci., 8(2): 272-281.
- El-Gioushy, S.F. (2016). Productivity, fruit quality and nutritional status of Washington

Navel orange trees as influenced by foliar application with salicylic acid and potassium silicate combinations. J. Hort. Sci. and Ornamen. Plants, 8 (2): 98-107.

- Elsheery, N.I., M.N. Helaly, H.M. El-Hoseiny and S.M. Alam-Eldein (2020). Zinc oxide and silicone nanoparticles to improve the resistance mechanism and annual productivity of salt-stressed mango trees. Agron., 10 (558): 1-20.
- Gad, M.M., O.A. Zagzog and O.M. Hemeda (2016). Development of nano-chitosan edible coating for peach fruits cv. Desert Red. Int. J. Environ., 5(4): 43-55.
- Giacalone, G. and V.Chiabrando (2015). Effect of preharvest and postharvest application of chitosan coating on storage quality of nectarines. Acta Hortic., 1084: 675-680.
- Górecki, R.S. and W. Danieski-Busch (2009). Effect silicate fertilizers on yielding of greenhouse cucumber (*Cucumissativus* L.) in container cultivation. J. Elementol., 14 (1): 71-78.
- Hanumanthaiah, M.R., K.R.C. Vijendrakumar, D.M. Renuka, K.K. Kumar and K.V. Santhosha (2015). Effect of soil and foliar application of silicon on fruit quality parameters of banana cv. Neypoovan under hill zone. Plant Archives, 15 (1): 221-224.
- Hidangmayum, A., P. Dwivedi, D. Katiyar and A. Hemantaranjan (2019). Application of chitosan on plant responses with special reference to abiotic stress. Physiol. Mol. Biol. Plants, 25(2): 313–326.
- Hogendorp, B.K. (2008). Effect on silicon-based fertilizer application on the development and reproduction of insect pests associated with greenhouse-grown crops. Ph.D. Diss., Univ. Illinois, Urbana–Champaign, IL, USA.
- Ibrahim, M.M., A.A. Ali and N.K.H. Serry (2019). Effect of nano trace elements and nano chitosan foliar application on productivity and fruits quality of grapevine cv. 'Superior Seedless'. Hort. Sci. and Ornamen. Plants, 11(1): 07-13.
- Jackson, M.L. (1958). Soil chemical analysis. Constable and Co. Ltd. London, 498.

- Javaid, K. and F.A. Misgar (2017). Effect of foliar application of orthosilicic acid on leaf and fruit nutrient content of apple cv. "Red Delicious. Advance Res. J. Multidisciplinary Discoveries. 20.0, C-7 (1): 30-32.
- Jia, J.X. (2011). Improvement of crop quality by silicon fertilizers: Effects and its possible mechanisms. Int. Conf. Silicon in Agric. September 13-18, Pp 76 Beijing, China.
- Jia, J.X., D.L. Cai and Z.M. Liu (2011). New progress in silicon-improvement of quality of crops. Int. Conf. Silicon in Agric. Sep.13-18, 77 Beijing, China.
- Jitareerat, P., S. Paumchai, S. Kanlayanarat and S. Sangchote (2007). Effect of chitosan on ripening, enzymatic activity, and disease development in mango (*Mangifera indica*) fruit. N. Z. J. Crop Hort. Sci., 35: 211–218.
- Khafagy, A.M.M. (2018). The beneficial effects of using chitosan and glutathione on the fruiting of Red Roomy grapevines M. Sc. Thesis. Fac. Agric. Minia Univ., Egypt.
- Kotb, F.A. and M.D. Abdel-Adl (2017). Effect of silica compounds on vegetative growth, yield, fruit quality and nutritional status of Olinda Valencia orange. Mid. East J. Agric. Res., 6 (1): 45-56.
- Lalithya, K.A., H.P. Bhagya and R. Choudhary (2014a). Response of silicon and micro nutrients on fruit character and nutrient content in leaf of sapota. Biolife, 2 (2): 593-598.
- Lalithya, K.A., H.P. Bhagya, A. Taj, K. Bharati and K. Hipparagi (2014b). Response of soil and foliar application of silicon and micro nutrients on leaf nutrient status of Sapota. The Bioscan., 9(1): 159-162.
- Li, H. and T. Yu (2001). Effect of chitosan coating on incidence of brown rot, quality and physiological attributes of postharvest peach fruit. J. Sci. Food and Agric., 81: 269-274.
- Liu, J.M., C. Han, X.B. Sheng, S.K. Liu and X. Qi (2011). Potassium-containing silicate fertilizer: Its manufacturing technology and agronomic effects. Oral presentation at 5th

Int. Conf. Silicon in Agric.; September 13– 18, Beijing.

- Malerba, M. and R. Cerana (2016). Chitosan effects on plant systems. Int. J. Mol. Sci., 17 (7): 996.
- Mohaghegh, P., M. Shirvani and S. Ghasemi (2010). Silicon application effect on yield and growth of tomato fruit grown on straw or rockwool substrate. Acta Sci. Pol. Hort. Cult., 11(3): 79 89.
- Mohamed, H.M.A. (2017). Promoting the productivity of early sweet grapevines grown under sandy soil conditions by using glutamic acid and potassium silicate. J. Hort. Sci. and Ornamen. Plants, 9 (3): 138-143.
- Mohamed, M.A.A. (2016). Physiological studies on the effect of some silicon, boron and amino acid treatments on some olive cvs. Ph. D. Thesis Fac. Agric. Al-Azhar Univ. Assiut, Egypt.
- Mohamed, R.H.M. (2015). Studies on the effect of spraying potassium silicate and vitamin B on fruiting of Manfalouty pomegranate trees. M.Sc. Thesis, Fac.Agric. Minia Univ. Egypt.
- Mohamed, S.S.A. and H.S. Ahmed (2019). Study effect of chitosan and gibberellic acid on growth, flowering, fruit set, yield and fruit quality of Washington Navel orange trees. Mid. East J. Agric. Res., 8 (1): 255-267.
- Mondal, M.M.A., M.A. Malek, A.B. Puteh and M.R. Ismail (2013). Foliar application of chitosan on growth and yield attributes of mungbean (*Vigna radiata* (L.) Wilczek). Bangladesh J. Bot., 42(1): 179-183.
- More, S.S., N.B. Gokhale, M.C. Kasture, S.E. Shinde and N. Jain (2017). Comparison of different sources of silica on the yield and quality of "Alphonso" mango in Kokan Region of Maharashtra. In Proc. 7th Int. Conf. Silicon in Agric., Bengaluru, India, 140.
- Nagy-Dina, A.M. (2015). Response of Flame seedless grapevines to spraying silicon. M. Sc. Thesis Fac. Agric. Minia Univ. Egypt.
- Neeru, J., C. Shaliesh, T. Vaishali, S. Purav and R. Manoherlal (2016). Role of orthosilicic acid (OSA) Based formulation in improving plant growth and development. Silicon, 1–5.

- Park, S., K. Marsh and J. Rhim (2002). Characteristics of different molecular weight chitosan films affected by the type of organic solvents. J. Food Sci., 67(1): 194-7.
- Patil, R.M. and S.L. Jagadeesh (2016). Effect of silicon bunch spraying and bunch bagging on yield, quality and shelf life of banana var. Grand Naine. Hort. Flora Res. Spectrum, 5 (3): 195-200.
- Photchanachai, S., J. Singkaew and J. Thamthong (2006). Effects of chitosan seed treatment on Colletotrichum sp. and seedling growth of chili cv. Jinda. Acta Hort., 712: 585-590.
- Qi, L., Z. Xu, X. Jiang, C. Hu and X. Zou (2004). Preparation and antibacterial activity of chitosan nanoparticles. Carbohydrate Res., 339 (16): 2693-700.
- Rong, J.F. (2011). Experimental study of the impact of silicon fertilizers on the growth, quality and output of big cherries. Int. Conf. on Silicon in Agric., Beijing, China, 182.
- Shao, X., B. Cao, F. Xu, S. Xie, D. Yu and H. Wang (2015). Effect of postharvest application of chitosan combined with clove oil against citrus green mold. Postharvest Biol. and Technol., 99: 37-43.
- Shi, Y.Z., J.F. Rong, L. Su, C. Li and Y. Feng (2010). Study on effects of silicon on grapevine physiology and fruit quality. Jilin Agric., 11: 98–100.
- Smith, F. and M. Dubois (1956). Colorimetric method for determination of sugars and related substances. Anal. Chem., 29:350.
- Snedecor, G.W. and G.W. Cochran (1990). Statistical Methods.7th Ed the Iowa state Univ., Press, Ames, Iowa, USA, 593.
- Stino, R., T. Fayed, M. Ali and S. Alaa (2010). Enhancing fruit quality of Florida Prince peaches by some foliar treatments. J. Hort. Sci. and Ornamental Plants, 2: 38-45.
- Su, X.W., S.C. Wei, Y.M. Jiang and Y.Y. Huang (2011). Effects of silicon on quality of apple fruit and Mn content in plants on acid soils. Shandong Agric Sci., 6: 59–61.
- Tomas-Barberan, F.A., M.I. Gil, P. Cremin, A.L. Waterhouse, B. Hess-Pierce and A.A.

Kader (2001). HPLC–DAD–ESIMS analysis of phenolic compounds in nectarines, peaches, and plums. J. Agric. Food Chem., 49: 4748–4760.

- Verma, V., V. Goyal, P. Bubber and N. Jain (2017). Effect of foliar spray of stabilized orthosilicic acid (OSA) on the fruit quality and quantity of Kinnow mandarin. In Proceedings of the 7th Int. Conf. Silicon in Argric., Bengaluru, India, 147.
- Xing, Y., H. Lin, D. Cao, Q. Xu, W.Han, R.Wang, Z. Che and X. Li (2015). Effect of chitosan coating with cinnamon oil on the quality and physiological attributes of China

jujube fruits. Bio. Med. Res. Int., 2015, Article ID 835151, 10.

- Youssef, M.S.M. (2017). Effect of spraying silicon on fruiting of Sakkoti date palms. M.Sc. Thesis Fac. Agric. Minia Univ. Egypt.
- Zagzog, O.A., M.M. Gad and N.K. Hafez (2017). Effect of nano-chitosan on vegetative growth, fruiting and resistance of malformation of mango. Trends in Hort. Res., 7 (1): 11-18.
- Zahedi, S.M., M. Karimi and J.A. Teixeira da Silva. (2020). The use of nanotechnology to increase quality and yield of fruit crops. J. Sci. Food Agric., 100 (1):25-31.

تأثير رش النانوشيتوزان والنانوسيليكون على صفات الجودة الطبيعية والكيماوية للثمار والمحتوى المعدني لأوراق أشجار الخوخ فلوريدا برنس

نبيل جودة سليمان – صفاء عبدالغني أحمد نمير – محمد محمود إبراهيم – محمد ممتاز جاد قسم البساتين - كلية الزراعة - جامعة الزقازيق - مصر

أجريت هذه الدراسة خلال موسمين متتاليين 2016 و 2017 على أشجار خوخ صنف فلوريدا برنس عمر 5 سنوات ومنزرعة في تربة رملية سلتية على مسافة 3 × 5 متر تحت نظام الري بالتنقيط لبستان خوخ خاص بمركز بلبيس بمحافظة الشرقية ، مصر. تم رش الأشجار المختبرة بالنانوشيتوزان بمعدلات 10، 20، 30 و 40 جزء في المليون ، النانوسيليكون بمعدلات 20، 20، 30 و 200 جزء في المليون ، النانوسيليكون بمعدلات 20، 20، 100 و 2000 جزء في المليون ، النانوسيليكون بمعدلات 20، 20، 20، 20، 20، 20، 200 و 300 جزء في المليون ، النانوسيليكون بمعدلات 200، 200 و 2000 جزء في المليون و سليكات البوتاسيوم بمعدلات 20، 200، 2000 و 3000 جزء في المليون و سليكات البوتاسيوم بمعدلات 2000، 2000 و 3000 جزء في المليون ، النانوسيليكون بمعدلات 200، 2000 و 3000 جزء في المليون و سليكات البوتاسيوم بمعدلات 2000، 2000 و 3000 جزء في المليون و سليكات البوتاسيوم بمعدلات 2000، 2000 و 3000 جزء في المليون ما بلأضافة للرش بالماء (معاملة الكنترول). أظهرت النتائج أن رش النانوشيتوزان بجميع معدلاته أعطي أعلى نسبة مواد صلبة ذائبة/ الحموضة في عصير الثمار دون وجود فروق معنوية بينهم في الموسمين. احتوت ثمار الأشجار غير المعاملة على أعلى معدل 2000 جزء في المليون أعلى محتوى من فيتامين ج مقارنة بالمعاملات الأخرى. كما أعطى رش سليكات البوتاسيوم بمعدل 3000 جزء في المليون أعلى نسبة من السكريات الكلية والكربو هيدرات. وأعطت معاملة سليكات البوتاسيوم بمعدل 3000 جزء في المليون أعلى محتوى من فيتامين ج مقارنة بالمعاملات الأخرى. كما أعطى رش سليكات البوتاسيوم بمعدل 3000 جزء في المليون أعلى محتوى من فيتامين ج مقارنة بالمعاملات الأخرى. كما أعطى رش سليكات البوتاسيوم بمعدل 3000 جزء في المليون أعلى محتوى لما لمريون أعلى محتوى لعناصر النيتروجين، الفوسفور، البوتاسيوم، الحديد، الزنك والمنجنيز بالأوراق، بينما أعطت معاملة معاملة المليون أعلى محتوى لعناصر العدين المون أعلى محتوى لعناصر النيتروجين، الفوسفور، البوتاسيوم، الحديد، الزنك والمنجنيز بالأوراق، بينما أعطت معاملة المليون أعلى محتوى لعناصر الحديون والموليون أعلى محتوى لمان الحديون والول الخوى مول والنانوشيون أعلى محتوى لعن مروسي معاملة معاملة الحديون والزي والنوون والموليون أمل محتوى من عناصر الحديو والزي واليوليون.

أستاذ الفاكهة - كلية العلوم الزراعية البيئية - جامعة العريش.

المحكمون :

أ.د. هاني عبدالله العلاقمي
 أ.د. فريد محمد سامي

م أستاذ الفاكهة – كلية الزراعة – جامعة الزقازيق