Salt Stress of Jeursalim Artichoke Plants in Relation to The Presoaking Tubers with Hydrogen Peroxide

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ABSTRACT: Two main experiments were carried out during the growing seasons 2018 and 2019 on Jerusalem artichoke cv. Fuseau at the Soil Salinity Laboratory Research, Agricultural Research Center, Alexandria, Egypt. This investigation is being proposed to assess the advantages of H₂O₂-soaked tubers and salt stress exposure on growth, yield components, guality and tubers' elemental composition. Tubers were planted in cemented-butaminzed lysimeters (2*2*1m) in rows, 80 cm in wide and 2 m in length. The treatments were comprised of two variables in split-plot design with three replicates. Four levels of saline irrigation water were prepared by mixing the fresh tap and sea waters, to create salt concentrations of 500 (control), 2000, 3500 and 5000mg/L, then, were arranged in the main plots. Whereas, three levels of H_2O_2 – soaking tubers before planting (control, 100 and 200mM) were arranged in the sub plots. The H₂O₂- treated tubers were seeded in the cemented lysimeters, in rows of 80cm in width, 2m in length. At the flowering growth stage, characteristics of vegetative growth were registered and finally at the harvest stage, yield, yield components and quality as well as the elemental tubers composition, including N, P and K% were recorded. The results showed that increasing the salinity levels of irrigation water up to 5000 mg/L negatively affected most of the growth characteristics (plant height, plant fresh weight). It also led to a decrease in the yield of tubers (the number of tubers per plant - the average weight of a tuber (g) and the total yield / fad. (tons). The results also showed that the increase in the salinity of irrigation water led to exhibited marked reductions in the tuber concentration content of elements N, P and K% as well as carbohydrate and inulin% during the two growing seasons. The data also revealed positive effects of 100mM H_2O_2 – soaked tubers on vegetative growth criteria and yield performance. Similar trend was achieved on carbohydrate, inulin tuber content and N% of tuber only in the first season of this study. The interaction study revealed that there are no marked significant variation appeared for the vegetative growth criteria during the respective seasons. The results detected on plant height and yield components indicated that growth performance was clearly manifested for the combined treatment of 100 mM H₂O₂-treated tubers and salinity exposure, defined from 500 up to 2000 mg/L. Similar trend was also registered on carbohydrate, inulin and only N% contents of tubers. In conclusion, it seems possibly, in particular under saline conditions, soaking the tuber in 100 mM H_2O_2 before planting for 2h would be essential to avoid the harmful effect of salt exposure and to achieve better records on yield component and quality and elemental composition content in tubers.

Keywords: Jerusalem artichoke, hydrogen peroxide solution, irrigation water salinity, salt stress.

INTRODUCTION

Jerusalem artichoke (*Helianthus tuberosus L*.) is a non-traditional tuberous crop ,which is recently introduced to Egypt for high nutritional and medicinal values. The tuber flesh of this plant is a rich source for nutrients and polysaccharides, particularly, inulin; that contains considerable levels of fructose sweetener that could be utilized for human-being without any side effects on blood sugar level (Seljåsen and Slimestad, 2007). Its protein has high food value due to

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the presence of almost all essential amino acids (Rakhimov *et al.,* 2003). Tubers are considred a suitable livestock feed (Seiler and Campbell, 2004). In the last decades Jerusalem artichoke has been concidered as a biomass crop for ethanol because it produces high levels of carbohydrates (Denoroy, 1996).

Salinity is one of the most important environmental stress variables that affects the growth and productivity of different crops (Lopez *et al.*, 2002). The negative impact of salinity is increasing dramatically in the arid and semi-arid regions of the world where the majority of developing countries are located (Khan *et al.*, 1999). Salinity not only exerts differences between average productivity and potential productivity, but also induces a marked drop the yield from year to year. It directly affects plant growth through its interaction with metabolic rates and pathways in plants (Rahimi *et al.*, 2012).

Hydrogen peroxide (H_2O_2) , produced naturally in plant cells under stress conditions, is characterized by high oxidative reactivity (Ogawa and Iwabuchi, 2001). Hydrogen peroxide is considered necessary for cell signaling, due to its important role in regulating oxidative stress (Rhee, 2006). Recently H₂O₂ has been regarded as a stress signaling molecule in regulating plant development and adaptation to abiotic and biotic stresses (Hung et al., 2005). Exogenous application of H_2O_2 at low concentrations (≤ 2.5 mM) had stimulatory effect on growth traits of plants, while the concentration up to 5 mM played an opposite role (Deng et al., 2012). Hydrogen peroxide is found to be involved in the acclimation and tolerance of plants grown under salt stress (Li et al., 2011; Wang et al., 2013). Therefore H₂O₂, at low concentrations, is considered one of the exogenous materials that used to induce the defense mechanisms in plant cells (Chen et al., 1993) and have a central role in improving plant tolerance to environmental stresses such as salinity (Azevedo Neto et al., 2005; Ashraf et al., 2013). Furthermore, H₂O₂ seems to be a "master hormone" that controls a variety of stress responses and plays a key role in primary plant metabolism (Ślesak et al., 2007).

This study was carried out to investigate the potential role of H_2O_2 to improve the performance of growth and productivity of Jerusalem artichoke plants under different salt stressed conditions.

MATERIALS AND METHODS

The present investigation was carried out during the two successive summer seasons on mid of April 2018 and 2019 at Soil Salinity Laboratory Research, Alexandria, Agricultural Research Center. Jerusalem artichoke tubers of Fuseau cv. were planted in cemented-butaminzed lysimeters (2*2*1m). Tuber seeds were planted in rows, 80 cm in wide, 2 m in length.

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Four levels of saline irrigation water, namely, tap water as a control, 2000, 3500 and 5000mg/L, were used as a source of irrigation, keeping the soil moisture content near the field capacity (27.85%). The saline water was prepared by mixing tap water (0.68 dS/m) with sea water (46 dS/m) at certain ratios. The tuber seeds were partitioned in three parts, the first and second parts of tubers were soaked in liquid H_2O_2 solutions (100 and 200 mM) for 2h. before planting. The third part of tubers (soaked tubers in distilled water) was used as the control.

Before planting, the following fertilizers were added to the soil at rates of kg, 20 m³ organic manure /fed. plus 150 P_2O_5 /fed.in the form of mono calcium phosphate, 15.5 % P_2O_5 . Nitrogen fertilizer was added in the form of ammonium nitrate (33.5% N) at the rate of 300 kg/fed. at three equal doses ; after 4, 8 and 12 weeks from planting date. Potassium sulphate (48% K₂O) was applied at rate of 90 kg/fed. in two equal doses after 8 and 12 week from the planting date. All other recommended agro-managements such as disease pests and weed control were performed whenever they appeared to be necessary.

The physical and chemical analyses of the experimental soil are presented in Table (1) according to (Page *et al.*, 1982).

	Physical properties											
year	Sand%	Silt%	Clay%	Texture		рΗ	EC dS/m	CaCO3 %		O.M%)	
2018	38.5	21.0	40.5	Clay loam		7.87	1.69	2.32		2.15		
2019	38.2	21.1	40.7	Clay I	oam	7.86	1.72	2.35		2.17		
Soluble cations (meq/L) S						oluble a	anions (meq/L)	<i>A</i> nutr	Availab ients n	ole ng/kg	
year	Ca ⁺⁺	Mg⁺⁺	Na⁺	K⁺	CO₃ ⁼	HCC	0 ₃ ⁻ Cl⁻	SO₄ [⁼]	Ν	Р	Κ	
2018	5.48	4.66	9.88	0.23		8.46	6 3.67	7 8.12	80.0	17.9	38.2	
2019	5.51	4.68	9.65	0.25		8.4 ⁻	1 3.76	5 7.94	86.4	18.2	39.1	

Table (1). Physical properties and chemical analyses of the experimental soil

Vegetative characteristics were measured and recorded at the initial stage of flowering (150 days after planting). A random sample of three plants from each experimental plot was taken to estimate plant height (cm), number of main stems and plant fresh weight (kg).

At the harvest time (180 days old plants), each individual plant in the all treatment lysimeters was removed to measure the number of tubers, average tuber fresh weight (g) and total tubers' yield / feddan(ton) .

Random samples of ten fresh tubers per treatment were used to determine their dry matter percentage, at 70 °C. Inulin content was determined in tubers according to the method of Winton and Winton (1958). Total carbohydrate were

determined colorimetrically as in terms of gram of glucose/100g dry weight of tubers roots according the methods described by James (1995). In the digested dry matter of tubers nitrogen was determined according to the methods described by Pregl (1945) using micro-Kjeldahl apparatus. Meanwhile, phosphorus was determined colorimetrically following Murphy and Riley (1962). Potassium was determined against a standard using air propane flame photometer following Chapman and Pratt (1961). The concentration of N, P and K were expressed as percentage.

The experimental design used was a split-plot design with three replicates, where the four levels of saline irrigation water, namely, tap water as a control, 2000, 3500 and 5000 mg/L were arranged in the main plots, whereas, tuber seeds was soaked with hydrogen peroxide (0,100 and 200 mM) were arranged in the sub plots. Each sub plot contained 2 rows. Collected data from the experiments were statistically analyzed, using the analysis of variance method. Comparisons among the means of different treatments were assessed, using least significant differences (L.S.D) test procedure at $p \le 0.05$ level of probability, as illustrated by Snedecor and Cochran (1980) using Co-Stat software program.

RESULTS AND DISCUSSIONS

1. Mean performances of vegetative growth parameters of Jerusalem artichoke crop

a- The main effect of irrigation water salinity concentrations

The results given in Table 2 showed significant effects at $p \le 0.05$ on plant height and foliage fresh weight/plant characters during the two seasons. Evidently, increasing irrigation water salinity concentration from 500 mg/L up to 5000 mg/L negatively affected plant height and foliage fresh weight/plant characters during the two seasons of this study. Unlike, the number of stems per plant was not significantly affected under the salt-stressed conditions at $p \le 0.05$. Plant growth, as revealed from the data of plant height and foliage fresh were more superior in the control treatment. As the salinity of irrigation increased up to 5000 mg/L, adversable effects were clearly manifested. These results falls in line with the data reported by Mahmoud (2012) on potato plants. The reduction in plant growth under salinity stress conditions is consistent with the fact that salinity induces accumulation of certain ions and deficiency of the others and lowers the external water potential in the cell (Salem et al., 2017). Furthermore, the reduction in plant growing may be due to the interruption in metabolic activities affected by the decrease of water absorption and disturbance in water balance (Fahad et al., 2015).

b- The main effect of soaked tubers in H_2O_2 concentration

The results of Table (2) showed that most of the studied vegetative characters were significantly affected $p \le 0.05$ with soaking tubers in H₂O₂

concentration treatments, except for No. of stems / plant. However, increasing H_2O_2 concentration exerted a significant positive effect on both plant height and foliage fresh weight/plant (g) traits during the two seasons. In this respect the highest mean values for plant height (cm) and fresh foliage weight/plant (g) were scored when the tubers soaked in 100mM H_2O_2 treatment, followed by with the soaking treatment of 200mM H_2O_2 treatment. In contrast, the lowest values were clearly noted in the control treatment. The results of Attia *et al.* (2017) showed that H_2O_2 priming can induce plants by modulating physiological and metabolic processes such as photosynthesis, proline accumulation detoxification, and that this ultimately leads to better growth and development.

c- The effect of tested interaction

The 2-way interaction of saline irrigation and H_2O_2 - soaked tubers (table 2) imposed significant effects on growth traits at $p \le 0.05$. In general, soaking tubers with H_2O_2 concentration treatment gave the best results for the characteristics of plant height at any given level of salt stress treatment. As mentioned earlier, the characteristic of foliage fresh weight (g) and No. of stems /plant was not affected by the two independent variables, this trait was also not affected by the interaction between these two variables (Table, 2). A number of studies on plants have demonstrated that the pre-treatment with an appropriate level of H_2O_2 can enhance abiotic stress tolerance through the modulation of multiple physiological processes, such as photosynthesis, and by modulating multiple stress-responsive pathways (Hossain and Fujita 2013; Wang *et al.*, 2014).

2. Yield and yield components of Jerusalem artichoke crop

a- The main effect of irrigation water salinity concentrations

The documented results in Table 3 revealed clearly that total tuber yield and the corresponding its components e. i., number and weight of tuber /plant, average tuber weight as well as total yield/fed. were significantly affected ($p \le 0.05$) by irrigations water salinity concentrations during the two seasons of the growth. In this respect, highest records in tubers yield/fed. and their related characters that include i.e., average tuber weight, number of tubers/plant and tubers weight yield/plant were clearly manifested in non-salt-stressed plants (control, 500 mg/L). With increasing the salinity levels of irrigation water, remarkable decrements on vield criteria were gradually noted, being the least at 5000 mg/L. As far as the total yield and its components are closely correlated with the vigorous of the vegetative growth. Therefore the reduction of the total yield and its components can be attributed to the fact that the vegetative characters were negatively affected by the high salinity of irrigation water (Table, 2). These results are in agreement with those reported by Elkhatib et al. (2004), Mahmoud (2012), Al-Hamdany and Mohammed (2014), Arafa and El-Howeity (2017) on potato. Abu-Muriefah (2015) attributed these results to changes in osmotic capacity due to reduction of water content in addition to the specific toxic effects resulting from the accumulation of sodium and chloride ions as observed in many plants. It was observed that salinity gradually reduced the size and number of marketable tubers per plant. In this

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respect, Ghosh *et al.* (2001) attributed the yield decrement in salt-stressed plants to the reduction of the tuber number per plant. It should be emphasized that the drop in yield, associated with salt- stress could be interpred to the nutritional imbalance, which consequently caused the inactivation of enzymes such as nitrate reductase (NR).

Troatmonte		Vegetative growth characters / plant							
Ireau	nents		2018		2019				
water salinity levels (ppm)	H ₂ O ₂ Soaking levels (mM)	Plant height (cm)	No. of main stem/ plant	Plant fresh weight (kg)	Plant height (cm)	No. of main stem/ plant	Plant fresh weight (kg)		
	control	186.23a	9.26a	5.63a	188.31b	9.54a	5.26a		
control	100	188.91a	9.84a	6.12a	196.43a	9.96a	5.86a		
	200	165.32c	9.23a	6.84a	178.10c	9.45a	6.41a		
	Control	175.63b	8.56a	4.51a	189.36b	8.12a	5.45a		
2000	100	187.25a	8.32a	5.65a	197.21a	9.45a	5.28a		
	200	186.96a	9.54a	5.85a	179.36c	9.89a	5.63a		
	control	169.25b	8.56a	4.56a	174.23c	8.52a	4.51a		
3500	100	174.56b	8.23a	4.78a	163.25e	8.96a	4.56a		
	200	163.23c	8.21a	4.96a	173.21d	8.42a	5.12a		
	control	121.23f	7.23a	4.52a	145.63f	7.41a	4.02a		
5000	100	146.23d	7.98a	4.48a	149.23e	7.76a	4.22a		
	200	132.45e	7.12a	4.65a	146.12f	7.89a	4.45a		
		Main effe	ct of saline ir	rigation v	vater (A)				
Control		210.51a	9.23a	5.894a	243.52a	10.23a	6.175a		
2000 mg/L		200.43b	8.12a	5.190b	214.56b	10.25a	5.635b		
3500 mg/L		196.21c	8.25a	4.653c	195.34c	10.03a	5.056c		
5000 mg/L		185.63d	7.36a	4.285d	186.23d	9.87a	4.229d		
		Main effe	ct of soaked	tuber in I	H_2O_2 (B)				
control		223.15c	8.56a	4.82c	186.21c	7.56a	4.63b		
100 mM		246.45a	9.51a	5.86a	238.23a	8.23a	5.32a		
200 mM		238.23b	9.26a	5.19b	206.58b	8.36a	4.89b		

Table	(2).	Mean	values	of	vegetative	growth	indices	of	Jerusalem
		artich	oke plan	ts d	uring 2018 a	nd 2019 s	seasons		

Significant at 0.05 level of probability.

b. The main effect of soaked tubers in H_2O_2 concentration

The results of Table (3) appeared that tubers yield/plant were significantly affected ($p \le 0.05$) by soaking tubers in H₂O₂ solutions during the two growing seasons. Regarding to the number of tubers/plant and total yield per feddan, the data showed that both traits were significantly decreased by soaking tubers in H₂O₂ with 100mM, only in 2019. The result obtained on the fresh tubers did not appear any significant variations along the H₂O₂-soaking treatments during both seasons. Similar results were recorded on wheat (Hameed *et al.*, 2004), indicating that exogenous application of H₂O₂ provided more vigorous root system in wheat. H₂O₂

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applied in low doses can increase roots weight and length (Narimanov and Korystov, 1997). Recently, has been reported that intensive root growth acted well for higher nitrogen uptake in wheat (Liao *et al.*, 2004). More vigorous root grown will cause higher nitrogen uptake, creating better growth development of wheat plant. Niu and Liao (2016) showed that H_2O_2 mediates various developmental and physiological processes in plants. Also, the change of H_2O_2 level may impact metabolic and antioxidant enzyme activity related to plant growth and development (Barba-Espín *et al.*, 2014).

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Treatn	nents		2018			2019	
water salinity levels (ppm)	H ₂ O ₂ Soaking levels (mM)	No. of tubers/ plant	Tuber fresh weight (g)	Total yield (ton/fed.)	No. of tubers/ plant	Average tuber fresh weight (g)	Total Yield (ton/fed.)
	control	54.26b	39.26c	13.26b	52.63b	49.54a	14.54a
control	100	59.32a	49.84a	14.54a	58.12a	50.96a	14.96a
	200	48.25cd	39.23c	12.56c	51.56b	39.45c	12.45a
	Control	44.23e	48.56a	12.69c	50.12b	48.12a	13.56a
2000	100	50.23c	48.32a	14.25a	53.60b	49.45a	14.87a
	200	47.84d	39.54c	13.25b	45.23e	40.89a	12.41a
	control	52.36bc	38.56c	12.74c	35.21f	38.52c	12.54a
3500	100	54.63b	48.23a	13.56b	46.23d	46.96b	12.89a
	200	47.21d	38.21c	11.45d	33.25g	38.42a	11.23a
	control	41.25e	37.23c	11.35d	33.85g	37.41c	10.52a
5000	100	36.98f	42.98b	12.36c	42.03e	39.76c	10.96a
	200	32.58g	36.12d	10.25e	36.25f	37.89c	10.12a
		Main eff	ect of saline	irrigation w	/ater (A)		
Control		76.41a	48.21a	12.96a	70.12a	42.36a	12.36a
2000 mg/L		65.45b	41.52b	12.63a	63.45b	38.14b	12.12a
3500 mg/L		61.56c	36.84c	11.25a	54.32c	32.69c	11.52b
5000 mg/L		43.85d	30.58d	10.23b	46.23d	30.52d	10.56b
		Main ef	fect of soake	d tuber in H	H_2O_2 (B)		
control		55.02a	45.13a	13.24a	48.45a	47.56a	14.25a
100 mM		58.41a	49.51a	13.85a	50.69a	48.23a	14.85a
200 mM		48.65a	39.26a	11.87a	39.69b	38.36a	12.55b
*							

Table (3). The average values of Jerusalem artichoke tuber yield and its component characters as affected with water salinity concentrations, soaked tubers in H_2O_2 treatments and their interaction during the two study seasons

Significant at 0.05 level of probability.

c. The interaction effect of combined treatment

The interaction effect of two combined treatments exerted significant trend at $p \le 0.05$ on the number of tubers/plant and average tuber fresh weight traits during the two study seasons (Table, 3). In 2018, optimal results were realized when the tubers were soaked in 100mM H₂O₂ and irrigated with 500mg/L saline water, followed by water salinity concentrations of 2000 and 3500 mg/L, respectively. While the lowest positive results were at water salinity level of 5000 mg/L. As for the first season the significantly highest mean values for total yield / fed. were recorded with the saline irrigation , namely, 500 mg/L and 2000 mg/L , when the tubers were soaked in 100 mM H_2O_2 solution.

Table (4).	. The average values of Jerusalem artichoke tubers' quality traits as
	affected with water salinity concentrations, soaked tubers in H ₂ O ₂
	treatments and their interaction treatments during the two study
	seasons

Treatm	ents		2018			2019	
water salinity levels (mg/L)	H ₂ O ₂ - Soaking Levels (mM)	Total carbohydr ates (%)	Inulin (%)	Tubers dry matter (%)	Total carbohydrates (%)	Inulin (%)	Tubers dry matter (%)
<i> i</i>	control	55.62a	22.54a	23.45a	58.42a	23.45a	23.54a
control	100	57.91a	23.41a	24.15a	61.23a	23.65a	24.32a
	200	51.32b	21.45a	24.28a	53.21b	22.10a	23.15a
	control	50.63b	21.56a	22.84a	52.36b	22.85a	22.45a
2000	100	58.16a	22.74a	24.85a	59.81a	23.41a	24.78a
	200	50.28b	21.36a	22.36a	57.22ab	21.35a	21.41a
	control	51.08b	21.52a	22.51a	50.21b	21.74a	22.54a
3500	100	52.41b	20.98a	22.87a	53.84b	21.53a	23.12a
	200	49.85c	20.74a	21.46a	47.63cd	20.71a	21.54a
	control	46.23d	21.56a	21.65a	49.36c	20.36a	21.54a
5000	100	48.53c	21.05a	21.85a	48.53c	20.17a	21.23a
	200	47.75c	19.45a	20.14a	47.23cd	20.67a	20.09a
	Main e	effect of irrig	ation wat	er salinity co	oncentrations	(A)	
control		55.89a	21.36a	22.45a	57.96a	22.23a	22.41a
2000 mg/L		52.96b	20.54b	21.36b	51.85b	21.25b	21.64b
3500 mg/L		48.21c	20.04c	20.54c	49.68c	20.03c	20.45c
5000 mg/L		45.86d	19.36d	19.78d	44.83d	18.87d	19.02d
		Main effe	ct of soal	ked tuber in	H ₂ O ₂ (B)		
control		59.21b	23.56a	20.74a	58.21a	22.56a	20.63a
100 mM		63.45a	23.51a	21.56a	61.23a	22.23a	20.32a
200 mM		54.28c	21.26b	19.54a	53.58a	21.36a	19.89a

Significant at 0.05 level of probability.

Treatme	nts		2018			2019	
water salinity levels (mg/L)	H ₂ O ₂ - Soaking levels (mM)	N (%)	P (%)	K (%)	N (%)	P (%)	K (%)
control	control 100 200	1.521a 1.598a 1.536a	0.231a 0.245a 0.236a	4.325a 4.155a 4.361a	1.425a 1.486a 1.205cd	0.24a 0.243a 0.251a	4.263ab 4.562a 3.814bc
2000	Control 100 200	1.421ab 1.325b 1.254c	0.243a 0.250a 0.232a	4.184a 4.815a 4.264a	1.496a 1.523a 1.352b	0.232a 0.241a 0.223a	4.204b 4.851a 3.296d
3500	100 200	1.342b 1.305b 1.145d	0.214a 0.216a 0.205a	3.514a 3.874a 3.026a	1.265c 1.312b 1.156d	0.214a 0.232a 0.182a	3.025d 2.654e 2.346f
5000	100 200	0.954e 0.854ef 0.836e	0.184a 0.173a	2.815a 2.148a	0.890e 0.892e 0.853e	0.174a 0.179a 0.165a	2.410er 2.169f 2.135f
control	Main effec		on water s		1 6250	S (A)	4 4260
2000 mg/L 3500 mg/L 5000 mg/L		1.524b 1.356c 0.949d	0.207a 0.242a 0.154b 0.125b	4.233a 3.725b 3.286c 2.842d	1.478b 1.306c 1.021d	0.213a 0.205b 0.194c 0.173d	4.436a 3.687b 3.045c 2.874d
g	Ν	lain effect	of soaked	tuber in H	₂ O ₂ (B)		
control 100 mM 200 mM		1.478a 1.458a 1.280b	0.189a 0.221a 0.170a	3.654a 3.436a 3.412a	1.529a 1.635a 1.486a	0.232a 0.214a 0.192a	3.512a 3.457a 3.419a

Table (5). The averages of tubers' mineral element contents as affected with water salinity concentrations, soaked tubers in H_2O_2 treatments and their interaction treatments during the two study seasons

Significant at 0.05 level of probability.

3. The performances of tubers' quality and elemental composition a. The main effect of saline irrigation water

The properties of tuber quality traits and the elemental composition of Jerusalem artichoke tubers are given in Tables 4 and 5. As for the tested water salinity irrigation concentrations, the results revealed that the tested tubers' quality traits expressed as total carbohydrate , inulin and dry matter percentages and tuber elemental composition, including N, P and K content were significantly affected ($p \le 0.05$) with increasing saline irrigation water during the two seasons. The results clearly showed a gradual decline in the mean values of the quality traits and tubers' elemental contents of nitrogen, phosphorus and potassium elements with increasing the level up to 3500 mg/L and severely dropped at the highest salinity exposure. Total dry matter production significantly decreased with increasing water salinity level. In contrast, Ghosh *et al.* (2001) illustrated that tuber N content increased by salt stress presumably due to the decrease in the carbohydrate content in the tubers. The decreases in K⁺ could be attributed to the antagonism of Na⁺ and K⁺ at uptake positions in the roots, the effect of Na⁺ on K⁺

transport into the xylem or the inhibition of uptake processes (Hu and Schmldhlter, 2005). In a saline environment, plants take up an excessive amount of sodium at the expense of K^+ and Ghosh *et al.* (2001) illustrated that the decrease of dry matter production as a result of increasing salinity was relatively more pronounced in tubers than in the other parts of the plant. This result is consistent with El-Hedek (2013), who found a decrease in wheat plant phosphorus content with increased salt concentration in the soil.

b- The main effect of H₂O₂-soaked tubers

The results given in Table 5 indicated that except N% in 2018 & 2019 as well as K% in, 2019, no marked variations were detected at $p \le 0.05$. Regarding to the tubers quality, including; carbohydrate and inulin %, the results reported in Table 4 indicated that, only in 2018, significant differences were appeared between the 2 concentration levels of H₂O₂ 100 and 200 mM soaked-treated tubers. The results in Table 4 clarified that although the differences in the dry yield of tubers and P and K% between H₂O₂ –treated tubers along the all exposed salinity treatment did not impose any significant variations at $p \le 0.05$ different results were reported by Samah *et al.* (2012), indicating that the highest percentage of tubers dry matter was possessed when potato plants were sprayed with 40mM hydrogen peroxide.

c- The effect of combined Treatments

The 2-way interaction of the combined treatment effects exhibited significant variations on total carbohydrate and nitrogen tubers content affected during the two growing seasons (Tables 4 and 5). Tubers potassium content was significantly affected only during the second season. The remaining estimated element P, inulin and dry matter percentage in tubers were not significantly affected by this interaction during the two growing seasons. In general, soaked tubers with H_2O_2 (100mM) generally gave the best values for the estimated elements and tubers' quality traits, even if this increase was not significant in some cases.

Generally, it could be concluded that soaked Jerusalem artichoke tubers on concentration of 100mM hydrogen peroxide before planting was promising to achieve better results and was effective to alleviate the adverse effects of irrigation water salinity on the vegetative growth criteria ad inducing progressive in increases in total tubers yield per feddan, and tubers' quality characteristics.

REFERENCES

- Abu-Muriefah, S. S. (2015). Effect of sitosterol on growth, metabolism and protein pattern of pepper (*Capsicum annuum* L) plants grown under salt stress conditions. Int. J. Agric. Sci., 8(2): 94- 106.
- Al-Hamdany, S.A. and M.S. Mohammed (2014). Effect of salinity of irrigation water and spraying amino acids (proline, arginine) in the growth and holds potato (*Solanum tuberosum* L). Diyala Agric. Sci., 6(2): 154-163. (In Arabic)
- Arafa, M.M. and M.A. El-howeity (2017). Response of some potato (Solanum tuberosum L.) cultivars to different salinity levels of irrigation water under sandy soil. Middle East J. Agric. Res., 6(1): 57-66.
- Ashraf, M.A.; M. Rasool; Q. Ali; M.Z. Haider; A. Noman and M. Azeem (2013). Salt-induced perturbation in growth, physiological attributes, activities of antioxidant enzymes and organic solutes in mungbean (*Vigna radiata* L.) cultivars differing in salinity tolerance. Archives Agron. Soil Sci., 59(12): 1695-1712.
- Attia, G.M.; A.H. Mahmoud and A. A. El-Araby (2017). Response of wheat plants to seed pre- soaking in hydrogen peroxide solution under salt stress conditions. Alex. Sci. Exch. J., 38(2): 314-325.
- Azevedo Neto, A.D.; J.T. Prisco; J. Eneas-Filho; J.V.R. Medeiros and E. Gomes-Filho (2005). Hydrogen peroxide pretreatment induces stress acclimation in maize plants. J. Plant Physiol., 162: 1114–1122.
- Barba-Espín G., P. Diaz-Vivancos, D. Job, M. Belghazi, C. Job and J.A. Hernández (2014). Understanding the role of H2O2 during pea seed germination: a combined proteomic and hormone profiling approach. Plant Cell Environ., 34 :1907–1919.
- Chapman, H. D. and P.F. Pratt (1961). Methods of Analysis for Soil, Plant and Water. University of California, Division of Agricultural Sciences. Riverside California.
- Chen, Z.; H. Silva and D.F. Klessing (1993). Active oxygen species in the induction of plant systemic acquired resistance by salicylic acid. Sci., 262: 1883–1886.
- Elkhatib, H.A., E.A. Elkhatib, A.M. Khalf-Allah and A.M. El-Sharkawy, (2004). Salt tolerance of four potato cultivars. J. Pl. Nutr. 27(9):1575–1583.
- **EI-Hedek, K.S. (2013).** Effect of foliar applications of salicylic acid and potassium silicate on tolerance of wheat plants to soil salinity. J. Soil Sci. Agric. Eng., Mansoura Univ., 4(3): 335 357.
- Deng, X.P.; Y.J. Cheng; X.B. Wu; S.S. Kwak; W. Chen and A.E. Eneji (2012). Exogenous hydrogen peroxide positively influences root growth and exogenous hydrogen peroxide positively influences root growth and metabolism in leaves of sweet potato seedlings. Aust. J. Crop Sci., 6: 1572-1578.
- **Dnoroy, P. (1996).** The crop physiology of *Helianthus tuberoses L*.: a model oriented view. Biomass Bioenergy., 11(1):11-32.

.30

- Fahad, S., S. Hussain, A. Matloob, F. A. Khan, A. Khaliq, S. Saud and M. Faiq (2015). Phytohormones and plant responses to salinity stress: a review. Pl. Grow. Regul., 75: 391-404.
- **Ghosh, S.C., K. Asanuma, A. Kusutani and M. Toyota (2001).** Effect of salt stress on some chemical components and yield of potato. Soil Sci. Pl. Nutr., 47(3): 467-475.
- Hameed, A., S. Farooq, N. Iqbal and R. Arshad (2004). Influence of Exogenous Application of Hydrogen Peroxide on Root and Seedling Growth on Wheat (*Triticum aestivum* L.). Intr. J. Agric. And Biol., 6(2): 1-11.
- Hossain, M.A., and M. Fujita (2013). Hydrogen peroxide priming stimulates drought tolerance in mustard (*Brassica juncea* L.). Plant Gene Trait., 4:109–123.
- Hu,Y. and U. Schmidhalter (2005). Drought and salinity: A comparison of their effects on mineral nutrition of plants. J. Pl. Nutr. Soil Sci., 168: 541-549.
- Hung, S.H.; C.W. Yu and C.H. Lin (2005). Hydrogen peroxide functions as a stress signal in plants. Bot. Bull. Acad. Sin., 46: 1–10.
- James, C.S. (1995). Analytical Chemistry of Foods. Blokie Academic & Professional, London.
- Khan, M.J., H. Rashid, R. Ali (1999). Inter-varietal variability in wheat grown under saline conditions. Pak. J. Biol. Sci., 2: 693-696.
- Liao, M.; I. R.P. Fillery and J. A. Palta (2004). Early vigorous growth is a major factor influencing nitrogen uptake in wheat. Functional plant Biology, 31(2):121-129.
- Li, J.T., Z.B. Qui, X.W. Zhang and L.S. Wang (2011). Exogenous hydrogen peroxide can enhance tolerance of wheat seedlings to salt stress. Acta Physiol. Plant., 33:835-842.
- Lopez, C.M.L., H. Takahashi, S. Yamazaki (2002). Plant-water relations of kidney bean plants treated with NaCl and foliar applied glycinebetaine. J. Agron. Crop Sci., 188: 73-80.
- Mahmoud, M.M.S.(2012). Differential response of some potato varieties to irrigation with saline water. Ann. Agric. Sci. Moshtohor, 50(3): 327-340.
- Murphy, J. and J. P. Riley (1962). A modified single solution for the determination of phosphate in natuural waters. Anal. Chem. Acta., 27: 31- 36.
- Narimanov, A.A. and Y.N. Korystov (1997). Low doses of ionizing radiation and hydrogen peroxide stimulate plant growth. Biologia (Bratislava), 52: 121–124.
- Niu, L. and W. Liao (2016). Hydrogen Peroxide Signaling in Plant Development and Abiotic Responses: Crosstalk with Nitric Oxide and Calcium. Front Plant Sci., 7: 230.
- **Ogawa, K. and M. Iwabuchi (2001).** A mechanism for promoting the germination of Zinnia elegans seeds by hydrogen peroxide. Plant Cell Physiol., 42(3): 286–291

- Page, A. L., R. H. Miller and D. R. Keeney (1982). Methods of Soil Analysis, Part
 2: Chem. and Microbiological Properties. Am. Soc. Agron., Madison, Wisconsin, U.S.A.
- Pregl, E. (1945). Quantitative Organic Microanalysis. 4th Ed. J. Chundril, London.
- Rahimi, R., A. Mohammakhani, V. Roohi and N. Armand (2012). Effects of salt stress and silicon nutrition on chlorophyll content, yield and yield components in fennel (*Foeniculum vulgar* Mill.). Int. J. Agri. Crop Sci., 4(21):1591-1595.
- Rakhimov, D.A. ,A. O. Arifkhodzhaev, L. G. Mezhlumyan, O. M. Yuldashev, U.A. Rozikova, N. Aikhodzhaeva, and M. M. Vakil (2003). Carbohydrates and proteins from Helianthus tuberosus. Chem . Nat. Comp., 39(3): 312-313.
- **Rhee, S.G. (2006).** H₂O₂, a necessary evil for cell signaling. Science, 312:1882–1883.
- Salem, H.M., Y. Abo-Setta, M.A. Aiad, H.A. Hussein and R.A. El-Awady (2017). Effect of potassium humate and potassium silicate on growth and productivity of wheat plants grown under saline conditions. J. Soil Sci. Agric. Eng., Mansoura Univ., 8 (11): 577 – 582.
- Sameh, A.M. M., N.I. Abo-Elfadel and N.F.Agamy (2012). Role of hydrogen peroxide in improving potato tuber quality. Alex. Sci. Exch. J. 33(2):75-88.
- Seiler, G. J. and I. G.Campell (2004). Genetic variability for mineral element concentrations of wild Jerusalem artichoke forage. Crop Sci., 44:289-292.
- Ślesak, I.; M. Libik, B. Karpinska, S. Karpinski and Z. Miszalski (2007). The role of hydrogen peroxide in regulation of plant metabolism and cellular signalling in response to environmental stresses. Acta Biochemica Polonica, 54: 39– 50.
- Seljåsen,R. and R.Slimestad (2007). Fructooligosaccharides and phenolic in flesh and peel of spring harvested Helianthus tuberoses. ISHS Acta Horticulture 744:1 International Symposium on Human Health Effects of Fruits and Vegatables.
- Snedecor, G.W. and W.G. Cochran (1980). *Statistical Methods*. 7th Ed., Iowa State Univ., Press, Ames., Iowa, USA.
- **U.S. Salinity Laboratory Staff (1954).** Diagnosis and improvement of saline and alkali soils. U.S. Dep. Agri. Handbook 60.U.S. Gov. Printing Office, Washington, DC.
- Wang, X., C. Hou, J. Liu, W. He, W. Nan, H. Gong, and Y. Bi (2013). Hydrogen peroxide is involved in the regulation of rice (*Oryza sativa* L.) tolerance to salt stress. Acta Physiologiae Plantarum, 35: 891-900.
- Wang,Y., J. Zhang, J. L. Li and X. R. Ma (2014). Exogenous hydrogen peroxide enhanced the thermo tolerance of *Festuca arundinacea* and *Lolium perenne* by increasing the antioxidative capacity. Acta Physiol. Plant, 36: 2915– 2924.
- Winton, A.L. and K.B. Winton (1958). The analysis of foods. John Wiley and Sons. Inc. London.pp.857.

الملخص العربى

الاجهاد الملحي لنباتات الطرطوفة وعلاقته بنقع الدرنات قبل الزراعة في محلول فوق اكسيد الهيدروجين

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تم إجراء تجربتين رئيسيتين خلال موسمي النمو ٢٠١٨ و ٢٠١٩ على نبات الطرطوفة صنف فيوزا في معمل بحوث الاراضي الملحية والقلوية ، مركز البحوث الزراعية ، الإسكندرية ، مصر . تم اقتراح هذا البحث لتقييم مزايا نقع الدرنات في محلول H_2O_2 والتعرض للإجهاد الملحي على النمو ومكونات المحصول والجودة ومحتوى الدرنات من العناصر الغذائية. استخدم في تطبيق التجربة نظام القطع المنشقة في ثلاث مكرارات حيث وزعت ٤ مستويات من مياه الري المالحة ، تم تحضيرها بخلط ماءالصنبور ومياه البحر ، لتكوين تركيزات ملحية (٥٠٠ (كنترول)، ٢٠٠٠ ، ٢٠٠٠ و المالحة ، تم معر ، تم تحضيرها بخلط ماءالصنبور ومياه البحر ، لتكوين تركيزات ملحية (٥٠٠ (كنترول)، ٢٠٠٠ ، ٢٠٠٠ و المالحة ، تم تحضيرها بخلط ماءالصنبور ومياه البحر ، لتكوين تركيزات ملحية (٥٠٠ (كنترول)، ٢٠٠٠ ، ٢٠٠٠ و معرف معمر/لتر) عشوائيا على القطع الرئيسية بينما وزعت ٣ مستويات من محلول فوق اكسيد الهيدروجين (كنترول ، ١٠٠ و ١٠٠ و ٢٠٠ و ٢٠٠ معرفي فوق اكسيد الهيدروجين (كنترول ، ١٠٠ و ١٠٠ و ١٠٠ و ٢٠٠ محم/لتر) عشوائيا على القطع الرئيسية بينما وزعت ٣ مستويات من محلول فوق اكسيد الهيدروجين (كنترول ، ١٠٠ و معرفيا في القطع الرئيسية بينما وزعت ٣ مستويات من محلول فوق اكسيد الهيدروجين (كنترول ، ١٠٠ و ٢٠٠ و ٢٠٠ مليمول) لنقع الدرنات قبل الزراعة عشوائيا في القطع تحت المنشقة. تم زراعة الدرنات المنقوعة بمحلول فوق اكسيد الهيدروجين في معرول ٢ متر و معرفي فوق اكسيد الهيدروجين في احواض أسمنتية (٢ * ٢ * ١ م) ، في خطوط بعرض ٥٠ سم ، وطول ٢ متر وقياس بودة المحصول بالإضافة إلى قياس محتوى الدرنات ميانيات النمو الخاصة بالنبات.و في مرحلة الحصاد ، تم تسجيل بيانات المحصول وقياس جودة المحصول بالإضافة إلى قياس محتوى الدرنات من العناصر المغذية N = 0

أوضحت النتائج أن زيادة ملوحة مياه الري حتى ٥٠٠٠ مجم/لتر أثر سلبا على غالبية صفات النمو الخضرى (ارتفاع النبات ، وزن النبات الطازج) بالاضافة للتأثير السلبي على عدد الدرنات / نبات ، ووزن الدرنات الطازج والمحصول الكلي / فدان (الطن). كما أوضحت النتائج أن النباتات المعاملة بالملوحة أظهرت انخفاض ملحوظ في محتوى الدرنات من العناصر الغذائية Nو P و X وكذلك محتواها من الكربوهيدرات والإنيولين خلال موسمي النمو . وكشفت النتائج أي أيضًا عن العناصر الغذائية ماد و X وكذلك محتواها من الكربوهيدرات والإنيولين خلال موسمي النمو . وكشفت النتائج أي أيضًا عن العناصر الغذائية Nو P و X وكذلك محتواها من الكربوهيدرات والإنيولين خلال موسمي النمو . وكشفت النتائج أيضًا عن الاثر الإيجابي للنقع في محلول فوق اكسيد لهيدروجين (١٠٠ ملليمول) على مقابيس النمو الخضري و أيضًا عن الاثر الإيجابي للنقع في محلول فوق اكسيد لهيدروجين (١٠٠ ملليمول) على مقابيس النمو الخضري و أيضًا عن الاثر الإيجابي للنقع في محلول فوق اكسيد لهيدروجين (١٠٠ ملليمول) على مقابيس النمو الخضري و أيضًا عن الاثر الإيجابي للنقع في محلول فوق اكسيد لهيدروجين (١٠٠ ملليمول) على مقابيس النمو الخضري و معطول. كما اثرت المعاملة السابقة نفس التأثير الايجابي على محتوى الدرنات من الكربوهيدرات والانيولين و N، موسمي النمو الخضري و N، موسمي النمو الخضري و N، موسمي النمو معلول فوق اكسيد لهيدروجين (١٠٠ ملليمول) على مقابيس النمو الخضري و المحصول. كما اثرت المعاملة السابقة نفس التأثير الايجابي على محتوى الدرنات من الكربوهيدرات والانيولين و N، موسمي النمو الخصري خلال موسمي النمو . مولحة مياه الري ونقع الدرنات أنه لا يوجد اختلاف معنوي في مقلو في موسم النمو الخصري خلال موسمي النمو . ما موحم الترى معاملة الدرنات الموحم المولي و الايبيد وموسمي النمو . موسمي النمو . مولي المولي والتري ونقع الدرنات المولي المولي المولي فوق اكسيد موسمي النمو الخصري خلال موسموي المولي واليبي . مولي مولي مولي والذيبي . ومولي فوق اكسيد موسمول ومحتوي الده . مولي المولي والني والذيبي والنتيروجين.

بناء على النتائج السابقة وتحت ظروف هذه التجربة فانه يوصى بنقع درنات الطرطوفة فى محلول فوق اكسيد الهيدروجين تركيز ١٠٠ (ملليمول) قبل الزراعة لمدة ساعتين سيكون ضروريًا لتجنب التأثير الضار للاجهاد الملحي على محصول درنات الطرطوفة وخصائص جودة الدرنات.