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Association of Hepatitis C viral load with liver functions and risk factors among HCV patients, Minia governorate, Egypt

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Abstract

Hepatitis C virus (HCV) is one of the blood transmitted hepatitis viruses. HCV infections have been identified as major causes of chronic hepatic diseases, and hepatocellular carcinoma. The aims of the current study were to determine the HCV viral load between 35 hepatitis C patients in Minia governorate, Egypt, and to assess association of the viral load with abnormal liver functions including; Alanine aminotransferase (ALT), prothrombin activity and platelet count. In addition to assessing if there are any risk factors associated with the population group, sex, age and other factors. About 35 blood samples were collected from hepatitis C patients randomly selected from the outpatient clinic at the Viral Hepatitis Management Center, Minia governorate, Egypt; including males and females of different ages. Viral load was determined using Real-time polymerase chain reaction (RT-PCR). All relevant information was collected from each patient including personal and clinical data. Current results showed that 68.60 % of the samples were from males and 31.4 % were from females, and most of them aged between 51 and 70 years. Approximately 11 (31.4 %) of the HCV patients had viral loads of <106, 12 (34.3 %) recorded viral loads of <105, and about 12 cases (34.3 %) had a viral load of < 104. HCV infection has been associated with 4 risk factors representing high HCV transmission routes including; dental intervention (80.0 %), history of hospital admission (65.7 %), previous surgeries (57.1 %) and family history of HCV (48.6 %). However, history of Schistosomiasis and blood-transfusion showed low association with HCV infection; recording (31.4 %) and (22.9 %), respectively.

Keywords: Hepatitis, HCV, Liver functions, RT-PCR, Risk factors

1. Introduction

Hepatitis C virus (HCV) is a viral pandemic, which represents the most common blood-borne viral

infection that preferentially replicate in the liver (Karoney and Siika, 2013; Tabata et al., 2020).

Previous studies conducted by <u>Heim et al.</u>, (2016); <u>Na and Song</u>, (2019) reported that HCV is a major cause of chronic viral hepatitis, which can lead to severe liver fibrosis, cirrhosis and hepatocellular carcinoma (HCC).

According to Blach et al., (2017), chronic HCV infection is estimated globally to affect 71 million people, corresponding to 3% of the world's population. Messina et al., (2015); Okasha et al., (2015) revealed that Egypt has the highest HCV prevalence rate in the world, with a rate of ~14.7 % of the Egyptian population, 26 % in the Nile Delta and 28 % in Upper Egypt. Previous studies of Strickland, (2006); Waked et al., (2014) highlighted that the widespread use of tartar emetic injections, which were used to treat schistosomiasis in the 1950s to the early 1980s, is the most likely reason for the high prevalence of HCV currently detected. High prevalence of HCV is not the sole problem; however, high incidence of HCV is another important issue that reflects the newly occurred HCV infections, as about 160,000 to 500,000 new HCV infections were recorded annually, as reported by Miller and Abu-Raddad, (2010).

According to Mohamoud et al., (2013); El-Ghitany et al., (2015), HCV primarily spread through direct contact with infected human blood; blood and blood product transfusion, unsafe medical treatment, healthcare exposure including; injury by needle stick, intravenous use of drugs, vertical and sexual transmission. A recent study of Raad et al., (2018) revealed that new HCV infections caused by nosocomial transmission are a major problem in developing countries, owing to the re-use of infected or improperly sterilized syringes and/or devices used in various medical purposes. Elghitany, (2019) added that elderly people are a significant global risk factor. Male sex and low education; however, represent high risk of HCV infection in Egypt. Furthermore, residence in rural areas is another risk factor in Egypt.

Briefly, real-time PCR system detects and quantifies the fluorescent reporter signal that increases

in direct proportion to the amount of PCR product in a reaction. A previous study conducted by <u>Kleiber et al.</u>, (2000) reported that, as the template is present at the beginning of the reaction, it takes lesser cycles to reach the point; where the fluorescent signal is first reported above the background, which is statistically important. This point is known as the threshold cycle (Ct) that always occurs during the amplification exponential phase.

Standard curves are generated by plotting the different Ct values of the four quantitation external standards against their RNA concentrations. From this standard curve, the viral load of the HCV-RNA is extracted based on the fluorescent intensity of the sample. The viral load is measured in international unit per ml (IU/ ml), as reported by Pawlotsky et al., (2000). The objectives of the current study were to determine the HCV viral load between hepatitis C patients in Minia governorate, Egypt, and to assess association of the viral load with abnormal liver functions. In addition to assessing if there are any risk factors associated with the population group, sex, age and other factors.

2. Material and methods

2.1. Samples collection and processing

During January, 2019-August, 2019, about 27 blood samples were collected from selected treatmentnaïve HCV infected patients, and 8 patients who failed to achieve an early viral response on combined treatment with sofosbuvir (400 mg/d) and daclatasvir (60 mg/d) for 3 months, at the Viral Hepatitis Management Center, Minia governorate. All relevant information were collected from each patient including personal data such as; age, sex, and residence; in addition to medical data including; history of blood transfusion, history of Schistosomiasis and antischistosomiasis treatment, previous surgical interference and dental interventions.

Approximately, 3 ml of blood were collected from each patient, centrifuged at 1500 rpm; sera were separated then distributed in aliquots, and then stored

at -20°C for further investigations, in reference to Halfon *et al.*, (1996).

2.2. Extraction of HCV-RNA

Viral RNA was extracted from patients' sera using QIAamp DSP virus Kit (cat#60704, Qiagen, Germany) according to the manufacturer's instructions, and the eluted RNA was stored at -80°C until further use in HCV-RNA quantification.

According to the method conducted by Taha et al., (2017), there were 4 stages during the extraction procedure mainly; lysis, wrapping, cleaning, and elution. A proteinase K and a lysis buffer have been used, which ensure inactivation of the RNase enzyme and digestion of the viral coat proteins. The lysed samples were applied to the binding buffers and binding conditions were adjusted. To allow optimum adsorption of the viral nucleic acids to the silica-based membrane of the spin column, lysates were thoroughly mixed with magnetic particles (carrier of RNA). The viral nucleic acids remained attached to the magnetic particles during a series of washing steps using first wash buffer 1, wash buffer 2 and then ethanol, while the contaminants were effectively washed away. Highly purified viral nucleic acids were eluted in the adsorbed viral nucleic acid elution buffer (AVE).

2.3. Quantification of the HCV-RNA

The HCV-RNA was amplified on Stratagene Mx3000 PTM using Artus HCV QS-RGQ-PCR Kit reagents. Amplification followed by simultaneous detection was carried out using RT-PCR, with TaqMan assay for a particular region of the HCV genome.

The reaction mixture for RT-PCR was prepared in a single eppendorf tube containing all components needed for PCR assay including; buffers, reverse transcriptase, and Taq Polymerase enzyme. The amplification reaction was carried out as follows; 6 μ l of HCV RG Master A, 9 μ l f HCV RG Master B and 10 μ l of the extracted RNA of a serum sample were

added to complete the reaction final volume to 25 μ l, in reference to Elsawaf *et al.*, (2015).

The reaction occurred under the following temperature conditions; incubation for 30 min. at 50°C, to reverse transcription (RT) of HCV viral RNA, to cDNA, followed by AmpliTaq gold activation for 15 min. at 95°C, 45 cycles of three PCR amplification steps, denaturation for 30 sec at 95°C, annealing for 1 min. at 50°C, extension for 30 sec at 72°C, and finally fluorescence detection.

The HCV probe is a small linear oligonucleotide that is labeled with a 5'-end fluorophore and a 3'-end quencher. The target of the PCR primers and probe is the conserved 5' un-translated region of the HCV genome. The HCV probe hybridizes preferentially to the HCV target sequence, which allows fluorescence emission and detection (Halfon *et al.*, 2006).

2.4. Statistical analysis

Sets of patient's data were analyzed using Microsoft® Excel software to produce charts and tables shown in the section of results. Moreover, data statistical analysis was done using the SPSS version 16. Qualitative variables were described as numbers and percentages. T-test of independent samples was used for parametric quantitative data, Mann Whitney test for non-parametric quantitative data, while Fisher's exact test was used for comparison between groups as appropriate. Significant level at P value < 0.05.

3. Results

3.1. Extraction and quantification of HCV-RNA by real time RT-PCR

Real time RT-PCR amplification curves of HCV-RNA are shown in Fig. (1). Values for HCV-RNA are reported in IU/ml. Of our 35-HCV patients 12 (34.3%) have viral loads of < 104, 12 patients (34.3%) their viral loads are of <105, and 11 cases (31.4%) have a viral load of < 106 (Table 1, Fig. 2).

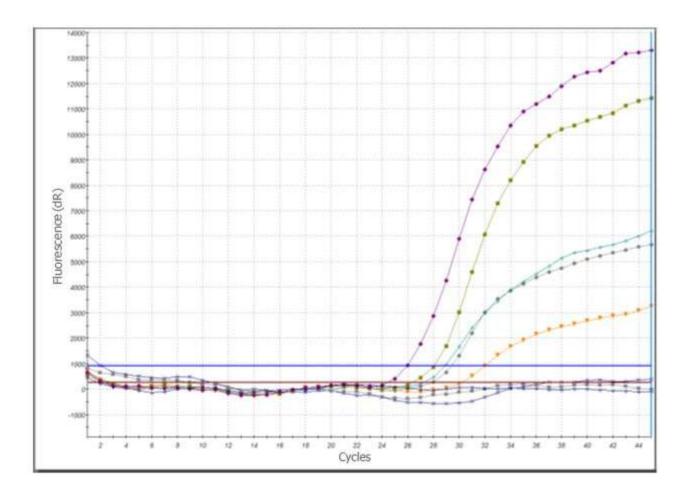


Fig. 1: Real time amplification curves of HCV-RNA by real time RT-PCR assay. The figure shows amplification plots of HCV cases showing positive cases with different threshold cycles (Cts)

Table 1: Viral load of the 35 HCV patients

Viral load (IU/ml)	No.	%
$> 10^3 - < 10^4$	12	34.3
$> 10^4 - < 10^5$	12	34.3
$> 10^5 - < 10^6$	11	31.4
Total	35	100

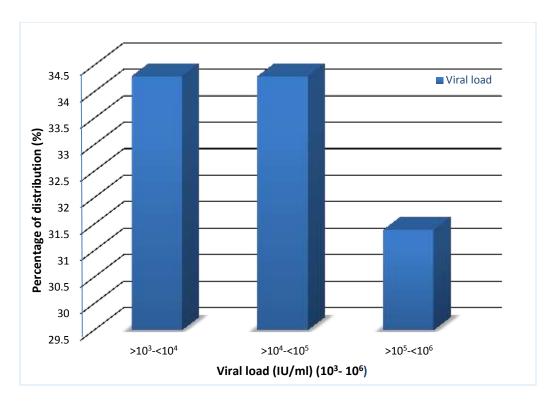


Fig. 2: Distribution of the 35 studied patient's samples viral load

3.2. Data analysis

Ages of patients involved in the study ranged from 23-75 years old. Out of these samples, 24 (68.60 %) were males, and 11 (31.40 %) were females. About 60.0 % of the patients aged 51-70 years, 4 (11.4%) patients were present in each age group of 20-30, 31-40 and 71-80, while those in the age group 41-50 years were 5.7% (Table 2).

The highest prevalence of HCV 9 (25.7 %) is in males of age groups of 51-60 years, followed by 61-70 years age-groups with prevalence of 6 (17.1 %); however, age groups from 31-40 and 71-80 years showed HCV prevalence of 3 (8.6 %) for each.

Age-groups from 20-30 and 41-50 years showed lowest prevalence of 5.7% and 2.9%, respectively. On the other hand, females predominant cases are in age groups of 51-60 years with 5 (14.2 %), followed by age groups of 20-30 and 41-50 years with 2 (5.7 %)

prevalence in each. The prevalence of age groups from 31-40 and 61-70 years is 1 (2.9 %) in each, whereas no female cases (0.0 %) are recorded in the 71-80 years age group, as demonstrated in Fig. (2, 3).

Out of all the HCV patients, about 25 (71.4%), 27 (77.1%) and 28 (80.0%) cases had normal ALT, platelet count and prothrombin levels, respectively (Fig. 4). The viral load is not significantly associated with the elevated ALT, platelet count and prothrombin abnormalities, recording; P = 0.152, P = 0.154 and P = 0.087, respectively (Table 3).

Many health care risk factors are associated with HCV infection showing different distribution among the current 35 HCV patients (Fig. 5). These risk factors include; history of dental intervention 28 (80.0 %), history of hospital admission 23 (65.7%), surgery 20 (57.1 %), family history of HCV infection 17 (48.6 %), history of schistosomiasis and blood transfusion 11 (31.4 %), 8 (22.9 %), respectively (Table 4).

Table 2: Distribution of the 35 HCV patients according to their ages and gender

Age -	Male		Female		Total	
	No	%	No	%	No	%
20-30	2	5.7	2	5.7	4	11.4
31-40	3	8.6	1	2.9	4	11.4
41-50	1	2.9	2	5.7	3	5.7
51-60	9	25.7	5	14.2	14	40
61-70	6	17.1	1	2.9	7	20
71-80	3	8.6	0	-	3	11.4
Total	24	68.6	11	31.4	35	100

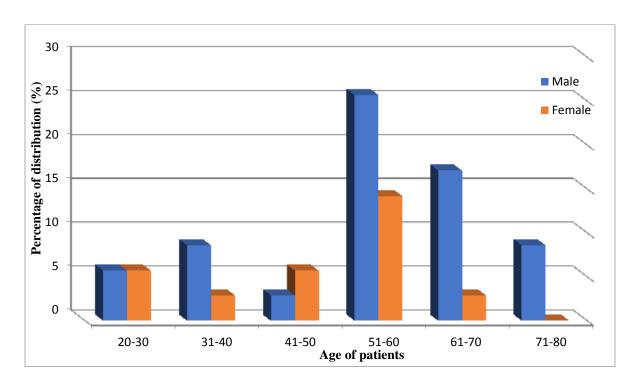


Fig. 3: Distribution of the 35 HCV patients according to their age and sex

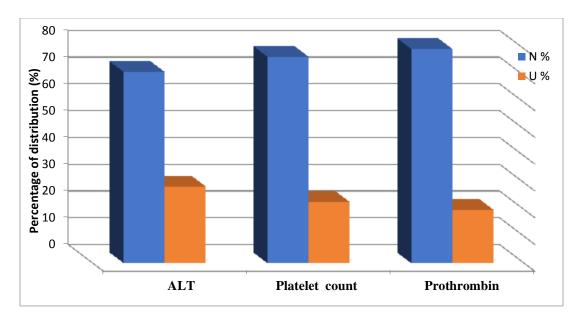


Fig. 4: The ALT, Platelet count and Prothrombin activity among the 35 HCV patient's cases. Where; N: Normal, U: Up-normal

Table 3: HCV viral loads correlations with liver functions abnormalities

	Viral load		
	r	P value	
ALT level abnormality (S)	0.247	0.152	
Platelets count abnormality (S)	0.246	0.154	
Prothrombin activity abnormality (S)	0.294	0.087	

Where; (S) Spearman's correlation; r: correlation coefficient, significant level at P value < 0.05

Table 4: Distribution of risk factors among the 35 HCV patients

Risk factors	No	%
Dentistry	28	80.0
Hospital admission	23	65.7
Surgery	20	57.1
Family History of HCV	17	48.6
Anti-schistosomal treatment	11	31.4
Blood Transfusion	8	22.9

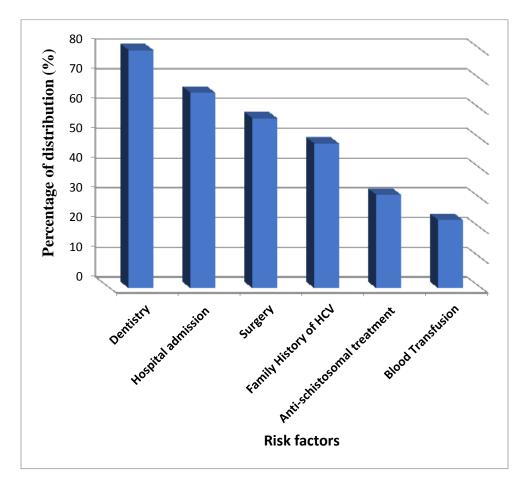


Fig. 5: Distribution of risk Factors among the 35 HCV patients

For community-based dental interventions, the strength of correlations is 19 out of 24 (5.72 %) male cases, compared to 9 out of 11 (3.45 %) female cases, with non-statistical significance (P = 1). The admission to hospital is related to 16 HCV male status (69.6 %) compared to 7 females (30.4 %) with non-statistical relevance (P=1). Undergo to previous surgery associate in 13 (65 %) males and 7 (35 %) females of current HCV patients, showing non-statistical significance (0.721). History of HCV infections among families and history of anti-

shistosomiasis each show more distribution between males than females, presenting non-statistical significance (P = 0.725), (P = 1), respectively. In those patients who have undergone blood transfusion (5 patients), HCV is less common than in those who have not received blood (30 patients), demonstrating non-statistical significance in male to female distribution (P = 0.64). Different reported risk factors associated with HCV infections show non-statistical significance correlation with the ages and viral loads, as shown in Table (5).

Table 5: Association of HCV infection with different risk factors among 35 HCV cases

Risk factors		Sex		Age	Viral load (×10 ³)
		Male	female		
Dentistry	Yes	19(67.9%)	9(32.1%)	53±14	201/(93-1075)
	No	5(71.4%)	2(28.6%)	53±17	93/(90-1030)
P value		1		0.922	0.523
Hospital	Yes	16(69.6%)	7(30.4%)	55±14	520/(107-1200)
Admission	No	8(66.7%)	4(33.3%)	50±16	96/(83-420)
P value		1		0.378	0.095
Surgery	Yes	13(65%)	7(35%)	51±15	257/(92-1075)
	No	11(73.3%)	4(26.7%)	56±13	112/(75-1030)
P value		0.7	21	0.284	0.629
Family history	Yes	11(64.7%)	6(35.3%)	52±15	182/(93-550)
of HCV	No	13(72.2%)	5(27.8%)	54±15	235/(79-1100)
P value		0.7	25	0.696	0.934
Anti-schistosomal	Yes	8(72.7%)	3(27.3%)	53±15	545/(98-1100)
Treatment	No	16(66.7%)	8(33.3%)	53±14	144/(81-807)
P value		1		0.948	0.268
Blood	Yes	3(60%)	2(40%)	60±9	550/(182-563)
Transfusion	No	21(70%)	9(30%)	52±15	111/(87-1100)
P value		0.6	54	0.257	0.409

Where; Independent Samples (T) test for parametric quantitative data (expressed by mean) between the two groups; Mann Whitney test for non-parametric quantitative data (expressed by median) between the two groups; Fisher's exact test for qualitative data between the two groups. -Significant level at P value < 0.05

4. Discussion

This study was performed in Minia governorate of Egypt, where the majority of population lives in rural areas. Seropositive cases in the current study remained higher in 51-60 years (38.9 %), followed by 61-70 years (22.2 %). Several other studies have also demonstrated high sero-prevalence of anti-HCV antibody among adult populations. In a recent study conducted by Vilas et al., (2018), the highest percentage of positive HCV was in the age group 31- 40 years. A study in India conducted by Verma et al., (2014) reported that adults of 21-60 years had a maximum (70 %) incidence of HCV. Moreover, Bharadwaj et al., (2014) revealed that the highest HCV prevalence was detected among people of 41-60 years age group. The prevalence of HCV among high age individuals suggests that they may substantially contribute to the ongoing HCV transmission in Egypt (Breban et al., 2014).

In this study, HCV sero-prevalence among males was higher (77.8 %) compared to females (22.2 %). Current results are in agreement with the study of El-Zanaty, (2016), who estimated that the prevalence of HCV among males are more than females, and to the findings of Bharadwaj et al., (2014) who also reported higher prevalence of HCV in males (0.7 %), compared to females (0.66 %). This male preponderance could be attributed to their high level of exposure to different risk factors causing transmission of HCV, due to male lifestyle.

Results of this study which demonstrated higher HCV prevalence in males than females are similar to the recent study of El-Adly et al., (2020), who also reported that among the tested samples, prevalence of 78.4 % were in males and 21.6 % were in females. Similarly, Niya et al., (2017) also recorded that out of 70 HCV patients; 54 were males and 16 were females, as statistical analysis did not record any meaningful link between age, sex and genotype variables.

On the contrary, current results are not compatible with other studies in which the prevalence of anti-HCV antibodies among females was higher than among males (Ramarokoto et al., 2008; Ayele and Gebre-Selassie, 2013; Abo-Amer et al., 2018). This study demonstrated that dental intervention is strongly associated with infection with HCV. Arafa et al., (2005) highlighted that in Egypt, periodontal therapy has also been reported as a possible source of HCV transmission. Also, detection of HCV-RNA on dental equipment used for treatment of HCV infected patients was reported by Piazza et al., (1995). Thus, strict guidelines for cleaning and sterilizing instruments between patients should be applied to decrease the risk of HCV transmission. Similarly, several other reports of Kalil et al., (2010); Barakat and El-Bashir, (2011) have shown the continuing occurrence of HCV in dental and medical facilities.

In our findings other routes of infection including previous hospital admission and surgeries have been recorded; 65.7 % and 57.1 %, respectively. Current results are higher than those reported earlier by Omran et al., (2009), who stated that previous surgeries as a possible route of infection by HCV recorded 108 (36 %). In this study, blood transfusion and history schistosomiasis showed a low HCV association recording; 22.9 % and 31.4 %, respectively. These results agreed the recent results of El-Adly et al., (2020), who detected low prevalence of HCV among patients with blood transfusion and schistosomal history. Similar results obtained by Omran et al., (2009) reported that 114 (38 %) of the 300 HCV studied patients had a history of schistosomiasis infection.

Several reports of El-Gohary *et al.*, (1999); Omran *et al.*, (2009) have documented the role of blood transfusion as a risk factor for transmission of HCV infection, in spite of screening for anti-HCV antibodies in the blood donors. Although several

earlier studies reported that, in developed countries such as the USA and Europe, the prevalence of HCV antibodies in blood donors was relatively low <1 %, while in developing countries such as Nigeria, Central African countries and Egypt; the prevalence of these antibodies ranged from 6 to 28% (Tanaka et al., 1992; Kowo et al., 1995; Halim and Ajayi, 2000). Nowadays, the high sensitivity and specificity of methods used to detect HCV antibodies in blood donors decreases the chance of considering blood transfusion as a risk factor. The 35 HCV patients included in this study have a viral load ranging between >103 - <106. Several studies conducted by McGuinness et al., (1996); De Moliner et al., (1998); Araujo et al., (2002) have argued that there is no correlation between serum levels of HCV-RNA and the serious liver disorders. Araujo et al., (2002) revealed that there was no correlation between hepatitis C viral (HCV) load and the histological abnormalities of the disease. In addition, Anand and Velez, (2004) highlighted that there is little correlation between HCV viral loads and the severity or progression of the liver disease. On the contrary, Adinolfi et al., (2001) reported that liver damage is correlated with the HCV-RNA levels. However, a previous study conducted by Kato et al., (1993) indicated that HCV-RNA titers were significantly higher in patients with chronic hepatitis and cirrhosis, compared to those with milder histological changes. In the present study, viral load is not correlated significantly with abnormal liver functions including; ALT, prothrombin activity and Platelet count. According to Agrawal et al., (2016), the severity of HCV infection can't be predicted solely by liver function tests, and normal results do not exclude the risk of a liver disease.

Conclusion

The current study provides information on hepatitis C viral load association with liver functions and risk factors among HCV patients in Minia governorate, Egypt. The HCV sero-prevalence found to be higher among males (68.6 %) as compared to females (31.4 %). Prevalence increased with age,

and there was a sharp increase among 51-70 years age group. Abnormal liver functions including ALT, prothrombin activity and Platelet count were not significantly associated with the viral load. HCV infections showed various associations with different risk factors representing high HCV transmission routes. The dental intervention showed strong association with HCV infection (80.0 %). However, history of Schistosomiasis and blood-transfusion presented low associations with HCV infection; recording (31.4 %) and (22.9 %), respectively. The severity of HCV infection can't be predicted solely by liver function tests, and normal results do not exclude the risk of a liver disease.

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Conflict of interest

No conflict of interests exists between the authors of this study.

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Ethical approval

This research was carried out in compliance with the ethical principles. The approval of Faculty of Pharmacy, Minia University Ethical Committee was provided before starting this study (reference number 65/2019). The patient's consents and statement of protection of the patient's privacy are provided.

5. References

Abo-Amer, Y.E.E.; Abd-Elsalam, S.; Eldosoky, H.; ELShenawy, A.K.; Awny, S.; Elagawy, W.

- and Negm, M.S. (2018). Declining prevalence of hepatitis C virus among university students in one of the main governorates in Egypt. Infection and Drug Resistance. 11: 2435-2441.
- Adinolfi, L.E.; Utili, R.; Andreana, A.; Tripodi, M.F.; Marracino, M.; Gambardella, M. and Ruggiero, G. (2001). Serum HCV RNA levels correlate with histological liver damage and concur with steatosis in progression of chronic hepatitis C. Digestive diseases and sciences. 46(8): 1677-1683.
- **Agrawal, S.; Dhiman, R.K. and Limdi, J.K.** (2016). Evaluation of abnormal liver function tests. Postgraduate Medical Journal. 92(1086): 223-234.
- **Anand, B.S. and Velez, M. (2004).** Assessment of correlation between serum titers of hepatitis C virus and severity of liver disease. World Journal of Gastroenterology: 10(16): 2409-2411.
- Arafa, N.; El Hoseiny, M.; Rekacewicz, C.; Bakr, I.; El-Kafrawy, S.; El Daly, M. and Fontanet, A. (2005). Changing pattern of hepatitis C virus spread in rural areas of Egypt. Journal of Hepatology. 43(3): 418-424.
- Araujo, E.S.A.D.; Cavalheiro, N.D.P.; Leitao, R.M.C.; Tosta, R.A.B. and Barone, A.A. (2002). Hepatitis C viral load does not predict disease outcome: going beyond numbers. Revista do Instituto de Medicina Tropical de São Paulo. 44(2): 71-78.
- Ayele, A. and Gebre-Selassie, S. (2013). Clinical Study Prevalence and Risk Factors of Hepatitis B and Hepatitis C Virus Infections among Patients with Chronic Liver Diseases in Public Hospitals in Addis Ababa, Ethiopia. International Scholarly Research Notices. 213: 1-7.
- **Barakat, S.H. and ElBashir, N. (2011).** Hepatitis C virus infection among healthy Egyptian children: prevalence and risk factors. Journal of Viral Hepatitis. 18(11): 779-784.

- Bharadwaj, B.V.G.; Vazhavandal, G.; Uma, A. and Rajalakshmi, P.C. (2014). Seroprevalence of hepatitis C virus infection among patients attending a rural teaching hospital in South India: a three year study. International Journal of Current Microbiology and Applied Sciences. 3(2): 123-127.
- Blach, S.; Zeuzem, S.; Manns, M.; Altraif, I.; Duberg, A.S.; Muljono, D.H. and Negro, F. (2017). Global prevalence and genotype distribution of hepatitis C virus infection in 2015: a modelling study. The Lancet Gastroenterology and Hepatology. 2(3): 161-176.
- Breban, R.; Arafa, N.; Leroy, S.; Mostafa, A.; Bakr, I.; Tondeur, L. and Mohamed, M.K. (2014). Effect of preventive and curative interventions on hepatitis C virus transmission in Egypt (ANRS 1211): a modelling study. The Lancet Global Health. 2(9): 541-549.
- De Moliner, L.; Pontisso, P.; De Salvo, G.L.; Cavalletto, L.; Chemello, L. and Alberti, A. (1998). Serum and liver HCV RNA levels in patients with chronic hepatitis C: correlation with clinical and histological features. Gut. 42(6): 856-860.
- **El-Adly, A.M.; Wardany, A. and Morsy, M.F.M.** (2020). Prevalence and risk factors for Hepatitis C virus infection among general population in luxor governorate, Egypt. Al-Azhar Medical Journal. 49(1): 33-44.
- **Elghitany, I.** (2019). Hepatitis C virus infection in Egypt: current situation and future perspective. Journal of High Institute of Public Health. 49(1): 1-9.
- El-Ghitany, E.M.; Abdel Wahab, M.M.; Abd El-Wahab, E.W.; Hassouna, S. and Farghaly, A.G. (2015). A comprehensive hepatitis C virus risk factors meta-analysis (1989–2013); Do they differ in Egypt?. Liver International. 35(2): 489-501.
- EI-Gohary, A.; Hosny, A.M.; Fawaz, N.; Heiba, A.; Abdalla, E.; Kawano, F. and Kiyokawa, T.

- (1999). HCV-RNA and anti-HCV IgM in Egyptian subjects bearing IgG anti-HCV antibodies. Japanese Journal of Infectious Diseases. 52(3): 113-116.
- Elsawaf, G.E.D.A.; Mahmoud, O.E.K.; Shawky, S.M.; Mohamed, H.M.M. and Alsumairy, H. H.A. (2015). Impact of occult hepatitis B virus infection on antiviral therapy in chronic hepatitis C patients. Alexandria Journal of Medicine. 51(3): 241-246.
- **El-Zanaty, F. (2016).** Egypt Health Issues Survey 2015, Cairo, Egypt and Rockville, Maryland, USA: Ministry of Health and Population and ICF International. 5(4): 74.
- Halfon, P.; Bourlière, M.; Pénaranda, G.; Khiri, H. and Ouzan, D. (2006). Real-time PCR assays for hepatitis C virus (HCV) RNA quantitation are adequate for clinical management of patients with chronic HCV infection. Journal of Clinical Microbiology. 44(7): 2507-2511.
- Halfon, P.; Khiri, H.; Gerolami, V.; Bourliere, M.; Feryn, J.M.; Reynier, P. and Cartouzou, G. (1996). Impact of various handling and storage conditions on quantitative detection of hepatitis C virus RNA. Journal of Hepatology. 25(3): 307-311.
- **Halim, N. and Ajayi, O. (2000).** Risk factors and seroprevalence of hepatitis C antibody in blood donors in Nigeria. East African Medical Journal. 77(8): 410-412.
- **Heim, M.H.; Bochud, P.Y. and George, J. (2016).** Host–hepatitis C viral interactions: The role of genetics. Journal of Hepatology. 65(1): S22-S32.
- Kalil, K.; Farghally, H.; Hassanein, K.; Abdel Sayed, A. and Hassanein, F. (2010). Hepatitis C virus infection among paediatric patients attending University of Assiut Hospital, Egypt. Eastern Mediterranean Health Journal. 16 (4): 356-361.
- **Karoney, M.J. and Siika, A.M. (2013).** Hepatitis C virus (HCV) infection in Africa: a review. Pan African Medical Journal. 14(1): 44.

- Kato, N.; Yokosuka, O.; Hosoda, K.; Ito, Y.; Ohto, M. and Omata, M. (1993). Quantification of hepatitis C virus by competitive reverse transcription—polymerase chain reaction: Increase of the virus in advanced liver disease. Hepatology. 18(1): 16-20.
- Kleiber, J.; Walter, T.; Haberhausen, G.; Tsang, S.; Babiel, R. and Rosenstraus, M. (2000). Performance characteristics of a quantitative, homogeneous TaqMan RT-PCR test for HCV RNA. The Journal of Molecular Diagnostics. 2(3): 158-166.
- Kowo, M.P.; Goubau, P.; Ndam, E.C.N.; Njoya, O.; Sasaki, S.; Seghers, V. and Kesteloot, H. (1995). Prevalence of hepatitis C virus and other blood-borne viruses in Pygmies and neighbouring Bantus in southern Cameroon. Transactions of the Royal Society of Tropical Medicine and Hygiene. 89(5): 484-486.
- McGuinness, P.H.; Bishop, G.A.; Painter, D.M.; Chan, R. and McCaughan, G.W. (1996). Intrahepatic hepatitis C RNA levels do not correlate with degree of liver injury in patients with chronic hepatitis C. Hepatology. 23(4): 676-687.
- Messina, J.P.; Humphreys, I.; Flaxman, A.; Brown, A.; Cooke, G.S.; Pybus, O.G. and Barnes, E. (2015). Global distribution and prevalence of hepatitis C virus genotypes. Hepatology. 61(1): 77-87.
- Miller, F.D. and Abu-Raddad, L.J. (2010). Evidence of intense ongoing endemic transmission of hepatitis C virus in Egypt. Proceedings of the National Academy of Sciences. 107(33): 14757-14762.
- Mohamoud, Y.A.; Mumtaz, G.R.; Riome, S.; Miller, D. and Abu-Raddad, L.J. (2013). The epidemiology of hepatitis C virus in Egypt: a systematic review and data synthesis. BMC Infectious Diseases. 13(1): 1-21.

- Na, S.K. and Song, B.C. (2019). Development and surveillance of hepatocellular carcinoma in patients with sustained virologic response after antiviral therapy for chronic hepatitis C. Clinical and Molecular Hepatology. 25(3): 234-244.
- Niya, M.H.K.; Salman-Tabar, S.; Bokharaei-Salim, F.; Behmanesh, M. and Keyvani, H. (2017). Prevalence of resistant associated variants (RAVs) in the naïve HCV patient candidate for direct acting antiviral (DAA) therapy. Microbial Pathogenesis. 105: 166-170.
- Okasha, O.; Munier, A.; Delarocque-Astagneau, E.; El Houssinie, M.; Rafik, M.; Bassim, H. and Fontanet, A. (2015). Hepatitis C virus infection and risk factors in health-care workers at Ain Shams University Hospitals, Cairo, Egypt. EMHJ-Eastern Mediterranean Health Journal. 21(3): 199-212.
- Omran, M.H.; Youssef, S.S.; El-Garf, W.T.; Tabll, A.A.; Bader-Eldin, N.G.; Atef, K. and El-Awady, M.K. (2009). Phylogenetic and genotyping of hepatitis C virus in Egypt. Australian Journal of Basic and Applied Sciences. 3(1): 1-8.
- Pawlotsky, J.M.; Bouvier-Alias, M.; Hezode, C.; Darthuy, F.; Remire, J. and Dhumeaux, D. (2000). Standardization of hepatitis C virus RNA quantification. Hepatology. 32(3): 654-659.
- Piazza, M.; Borgia, G.; Picciotto, L.; Nappa, S.; Cicciarello, S. and Orlando, R. (1995). Detection of hepatitis C virus-RNA by polymerase chain reaction in dental surgeries. Journal of Medical Virology. 45(1): 40-42.
- Raad, I.I.; Chaftari, A.M.; Torres, H.A.; Ayoub, E.M.; Narouz, L.I.; Bartek, J. and Hachem, R. (2018). Challenge of hepatitis C in Egypt and hepatitis B in Mauritania. World Journal of Hepatology. 10(9): 549-557.
- Ramarokoto, C.E.; Rakotomanana, F.; Ratsitorahina, M.; Raharimanga, V.; Razafindratsimandresy, R.; Randremanana, R.

- and Richard, V. (2008). Seroprevalence of hepatitis C and associated risk factors in urban areas of Antananarivo, Madagascar. BMC Infectious Diseases. 8(1): 1-7.
- **Strickland, G.T.** (2006). Liver disease in Egypt: hepatitis C superseded schistosomiasis as a result of iatrogenic and biological factors. Hepatology. 43(5): 915-922.
- **Tabata, K.; Neufeldt, C.J. and Bartenschlager, R.** (2020). Hepatitis C Virus Replication. Cold Spring Harbor Perspectives in Medicine. 10(3): a037093.
- **Taha, A.B.; Ali, K.S. and Sabeer, F.T. (2017).** Quantitation assay of hepatitis C virus RNA using real-time PCR technique. Cihan University-Erbil Scientific Journal. (Special-2): 242-252.
- Tanaka, E.; Kiyosawa, K.; Sodeyama, T.; Hayata, T.; Ohike, Y.; Nakano, Y. and Watanabe, J. (1992). Prevalence of antibody to hepatitis C virus in Japanese schoolchildren: comparison with adult blood donors. The American Journal of Tropical Medicine and Hygiene. 46(4): 460-464.
- Verma, R.; Behera, B.K.; Jain, R.; Arora, V.; Chayal, V. and Gill, P. (2014). Hepatitis C, a silent threat to the community of Haryana, India: a community-based study. The Australasian Medical Journal. 7(1): 11-16.
- Vilas, B.; Lyra, P. and Venkatesha, D. (2018). Prevalence of Hepatitis C Virus Infection among Chronic Liver Disease Patients in a Tertiary Care Hospital. International Journal of Current Microbiology and Applied Sciences. 7(2): 2489-2493.
- Waked, I.; Doss, W.; El-Sayed, M.H.; Estes, C.; Razavi, H.; Shiha, G. and Esmat, G. (2014). The current and future disease burden of chronic hepatitis C virus infection in Egypt. Arab Journal of Gastroenterology. 15(2): 45-52.