

## Protective and detoxifying effects of selenium nanoparticles and thyme extract on the aflatoxin B1 in rabbits

Atef A. Hassan<sup>1</sup>, Mogda K. Mansour<sup>2</sup>, Essam M. Ibrahim<sup>3</sup>, Mahmoud M. Arafa<sup>2</sup>, and Ola F.A. Talkhan<sup>4</sup> and Noha H. Oraby<sup>1\*</sup>



### Address:

<sup>1</sup>Department of Mycology and Mycotoxins, Animal Health Research Institute, Agriculture Research Centre, Egypt

<sup>2</sup>Department of Biochemistry, Toxicology and Feed Deficiency, Animal Health Research Institute, Agriculture Research Centre, Egypt

<sup>3</sup>Department of Pathology, Animal Health Research Institute, Agriculture Research Centre, Egypt

<sup>4</sup>Department of Biochemistry, Regional Laboratory at El- Menofia Governorate, Animal Health Research Institute, Egypt

\*Corresponding author: **Noha H. Oraby**, e-mail: [nohaoraby25@gmail.com](mailto:nohaoraby25@gmail.com)

Received: 08-03-2022; Accepted: 16-04-2022; Published: 29-04-2022

doi: [10.21608/ejar.2022.111578.1214](https://doi.org/10.21608/ejar.2022.111578.1214)

### ABSTRACT

Nanotechnology's application in biomedicine has piqued the curiosity of all authorities throughout the world. Different fungus species and aflatoxins were recovered from rabbit feed at significant quantities up to 300 ppb in this investigation. Selenium nanoparticles (Se-NPs) were generated using a chemical process, described briefly, and utilized to reduce the harmful effects of aflatoxicosis in rabbits. Biochemical parameters in tissues and sera of all aflatoxicated rabbits revealed that it caused the development of reactive oxygen species (ROS), an increase in nitric oxide levels, and malonaldehyde levels. While Total Antioxidant Capacity (TAC) activities are reduced, additional antioxidants in rabbit homogenate liver tissues will be found. In aflatoxicosis, decreased serum retinol and a-tocopherol were also found. For aflatoxicated rabbits, oral treatment of SeNPs and thyme restored these alterations to normal levels. Furthermore, SeNPs and thyme triggered the breakdown of AFB1 residues in liver tissues, suggesting that SeNPs and thyme can be introduced to animal feeds as antioxidants with good effects on animal health and mycotoxins removal.

**Keywords:** Nanotechnology, Se-NPs, Aflatoxicosis, Thyme, Antioxidant, fungi.

### INTRODUCTION

Recently, the biomedical application of nanomaterials gained large attention of all authorities related to human and animal health (Khalaf *et al.*, 2019). Several diseases cause serious public health problems on a human resulting in carcinogenic potential and liver cells necrosis (Tiew *et al.*, 2020). Whereas, the majority of used traditional drugs in the control of these diseases are not effective due to elaboration of microbial resistance during their frequent use. This problem gained more attention in finding novel effective agents from metallic nanomaterial for treatments of these infectious diseases (Moghimi *et al.*, 2005). The aflatoxin B1 (AFB1) is detected in cereals and feed and produced by mycotoxigenic *Aspergillus flavus* and *A. parasiticus* that resulted in several significant health hazard effects (El Nahass *et al.*, 2019). In developing countries such as Egypt, liver cancers are associated with aflatoxicosis in humans and animals and it can be reacted with DNA and produced reactive oxygen species (ROS), resulting in damage and destruction of lipids, DNA, and proteins (Benkerroum, 2020). These adverse effects have potentiated the occurrence of genetic toxicity and apoptosis. Moreover, the high worldwide morbidity and mortality of mycotoxicosis in humans and animal resulted in serious economic losses in their health activities and production (El-Nahass *et al.*, 2019). Hence, several studies have been evaluated for the degradation and elimination of the toxic effects of aflatoxicosis. The majority of used traditional drugs in the control of mycotoxicosis are not effective during their frequent use. This problem gained more attention in finding novel effective agents from metallic nanomaterial for treatments of toxicities (El-Nahass *et al.*, 2019). In addition, the valuable uses of metals nanoparticles are due to their low doses toxicity and help in drugs delivery to target organs (Youssef *et al.*, 2019) and explore novel effective vaccines. Hence, the synthesis of nanomaterial by simple and safe methods and easy application has been taken all attention. They have significant properties such as large surface area, high bioavailability, and decrease toxicity than bulk materials such as SeNPs (Zhang *et al.*, 2012). The selenium nanoparticles can be produced by the green method and the concentration of (60 µg/ml) caused destruction of cells (Menon and Shanmugam, 2019). Other benefits of SeNPs include activation of the thyroid gland function, antioxidant activity, and preventing harmful products of metabolism (Ulaiwi, 2018). Moreover, the recent strategies directed to the elimination and treatments of aflatoxicosis via the use of natural products in the environment that are available and ecofriendly such as the Thyme plant (a native herb) (Swayeh *et al.*, 2014). It has several potentials as antioxidants and antimicrobial and can be used instead of chemical drugs (Rasooli *et al.*, 2006). In addition, thyme is a potent natural antioxidant agent and can be reduced oxidative stress of toxins (Grigore *et al.*, 2010 and Placha *et al.*, 2014) and have an effective reduction of tumor necrotic factor (TNF- α) (Abdel-Aziem *et al.*, 2014). Hence, the present work was undertaken for evaluation the prevalence of fungi and aflatoxins in rabbits' feeds and drinking water in farms at El Minofya governorate. The induction of aflatoxicosis and their degradation by SeNPs and thyme extracts for improving the biochemical and pathological adverse changes due to aflatoxicosis in rabbits was also investigated.

## MATERIAL AND METHODS

### Ethical approval:

The current work was done and accepted as recommended local laws and regulations.

### Feed samples:

A total 100 of rabbits' rations and consumed water samples (50 of each) were collected from rabbits farms El- Menofia governorate, Egypt, where, animals sustained signs of poisoning, diarrhea, and deaths in some cases. The mycological and aflatoxins residues in samples were detected.

**Thyme and chemicals:** They were purchased from ALDRIK Sigma chemical company.

### Prevalence of mold and yeasts in samples

Samples were processed and examined for recovering and characterization of molds as recommended by (ISO 2008; Pitt and Hocking, 2009)

**Synthesis and detection of aflatoxins:** The recovered *A. flavus* in the present study were used for the synthesis of aflatoxins on yellow corn as recommended by (Smith, 1997). In addition, the toxin content of rabbits' feeds and water samples were determined and quantified as the method as (AOAC, 2000; Refai and Hassan, 2013).

**Chemical production and identification of SeNPs** (Verma and Maheshwari, 2018). The SeNPs were produced by the chemical reduction method. Briefly, in sterile flask contained 9ml of double-distilled water, 3 ml (25 mM) sodium selenite, 3 ml (100 Mm) glutathione (reduced form), and 0.15gm bovine serum albumin as stabilizer were added. Continuous stirring the flask for 1 hour using a magnetic stirrer and Sodium hydroxide (1M) poured till pH reach to 9. The color of the solution changed to a red color indicating the formation of Se-NPs. The prepared Se-NPs were identified and their shape, particles size, and morphology were measured by UV-visible spectra, XRD patterns (Atul et al., 2010).

**Experimental Design** (Cam, 2008; Zhou and Wang, 2011; Abdel-Azeim et al., 2013; Hassan et al., 2016)

The Se-NPs and Thyme were evaluated for improving the adverse effects of aflatoxicosis in rabbits. A 35 healthy New Zealand male rabbits (aged 6-8 weeks) were divided into 5 groups as observed in Table 1:

Groups	Feeding	Treatment
G1 (Negative control)	Healthy Feed and water ad libitum	No treatment
G2 (Positive control)	Orally given AFB1 at a dose of 50 ug / animal/ day for 4 weeks	No treatment
G3	Orally given AFB1 at a dose of 50 ug / animal/ day for 4 weeks	Orally administration of Aquas extract of thyme (500mg/Kg body weight) once daily
G4	Orally given AFB1 at a dose of 50 ug / animal/ day for 4 weeks	Orally administration of Se-NPs (0.3mg/kg body weight/ 0.2 ml of buffer/ day (low dose)
G5	Orally given AFB1 at a dose of 50 ug / animal/ day for 4 weeks	Orally administration of Se-NPs (0.5mg/kg body weight/ 0.2 ml of buffer/ day (high dose)

### Biochemical study:

The blood samples we collected from all bird after experimental work and sera were separated for biochemical analysis. After scarification of animals, liver tissues were obtained for detection the antioxidant assays (Chitra et al., 1999). Also, other antioxidant were determined as catalase; lipid peroxidation as malonaldehyde (MDA) and reduced glutathione (GSH) in liver tissues homogenate ( Aebi, 1974; Ohkawa et al., 1979; Ellman, 1959), respectively. The measurements of total antioxidant capacity (TAC) (Koracevic et al., 2001), Superoxide Dismutase (SOD) (Nishikimi et al., 1972), and GSH-px (Paglia and Valentine, 1967) were undertaken, respectively. The protein content of tissues samples were measured by the method of (Bradford, 1976). The levels of vitamins were determined as method as (Henry et al., 1974), and nitric oxide (NO) levels (Burgner et al., 1999).

### Pathological study:

After the end of the experimental study, liver and kidney tissues of all animals were collected and subjected for histopathological study as recommended by (Suvarna et al., 2012).

### Statistical Analysis

The statistical interpretation of the results was performed with One-Way ANOVA test. The results were given as mean  $\pm$  standard error using SPSS 14 (2006). The value of  $p < 0.05$  was considered significant.

## RESULTS

**Table 2.** Prevalence of fungi in rabbits feeds and water samples.

	No.	%	No.	%
Total fungi	40	80	30	60
<i>A. flavus</i>	35	70	26	52
<i>A. niger</i>	9	18	6	12
<i>A. Ochraceus</i>	5	10	3	6
<i>A.fumigatus</i>	1	2	2	4
<i>Penicillium species</i>	22	44	12	24
<i>Fusariumspecies</i>	12	24	-	-
<i>Cladosporium species</i>	3	6	5	10
<i>Scopulariopsis species</i>	2	4	-	-
<i>Mucor Species</i>	-	-	-	-
<i>Candida species</i>	3	6	10	20

The tabulated data in Table (2), illustrated that out of 100 rabbits feed and water samples (50 of each), from private rabbits farms at El- Menofia governorate, Egypt. Forty samples of rabbit feed and thirty samples of consumed water were contaminated with fungi (80% and 60%), respectively. The highest incidence of *A.flavus* was recovered from 35 feed samples (70%) and 26 samples of water (52%), respectively. While, other *Aspergillus species* were isolated at relatively lower rates as *A. niger* recovered from (18% and 12%) and *A. ochraceus* from (10% and 6%), respectively.

**Table 3.** Levels of aflatoxins in rabbits feed and water (ppb).

Samples	Total Aflatoxins in samples (ug/kg)(ppb)				
	+ve	%	Max. levels	Min. levels	Mean levels
Feed(50)	30	60	42.5±2.63	12±0.1	35.0±1.5
Water (50)	0	0	ND	ND	ND

ND: not detected during measurement of toxins by TLC method

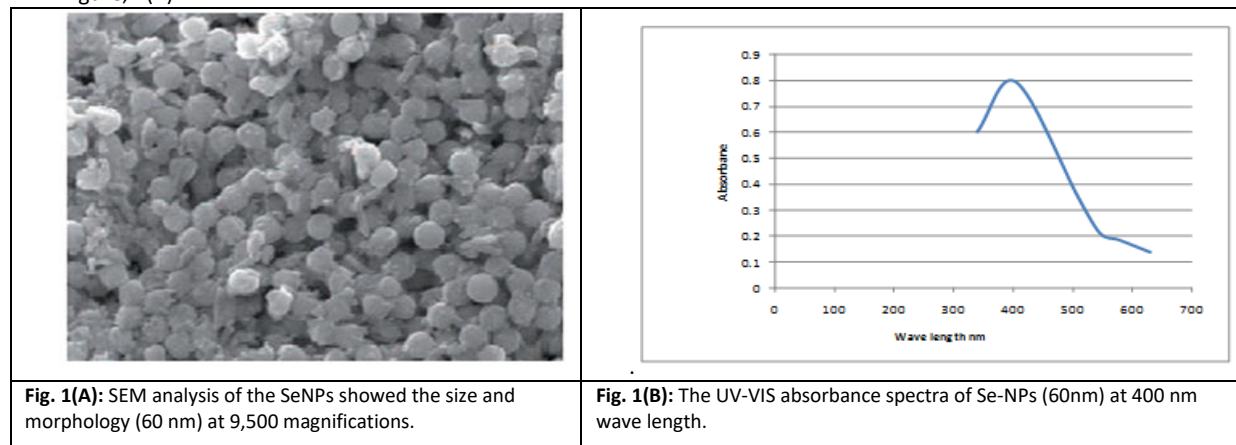
In the presented data in Table (3), we detected that the incidence of AFB1 residues in rabbits' feeds were reached 60% of feeds samples. However, the mean values of AFB1 residues in feeds was (35.0±1.5µg/kg) whereas, Afs residues did not detect in all water samples. Currently, most of the recovered *A. flavus* from the present rabbits' feed was aflatoxigenic.

**Table 4.** Levels of aflatoxins B<sub>1</sub> produced by *A. flavus* isolated from Rabbits rations.

Source of tested <i>A. flavus</i>	No. of tested isolates	Aflatoxigenic isolates		Levels of AFB <sub>1</sub> ppb		
		No.	%	Max	Min	Mean ± SE
Rabbits feed	20	15	75	300.0	50.0	100.5 ± 7.1

The recorded data in Table (4) illustrated that 15 of 20 tested isolates (75%) were produced significant levels of Afs on yellow. The mean levels of produced AFB1 were (100.5 ± 7.1 ppb) and the maximum value was (300 ppb) and the minimum was (50 ppb).

In recent years, nanotechnology has been used employed in several biomedical fields (Bai et al., 2018; Hassan et al., 2020, 2021). Herein, production and identification of Se-NPs proceeded and the obtained nanoparticles were spherical in forms and of (60 nm) in size Fig., 1(A). While, the UV-VIS detected that the maximum absorption occurred at a wavelength of 400 nm Figure, 1(B).



**Fig. 1(A):** SEM analysis of the SeNPs showed the size and morphology (60 nm) at 9,500 magnifications.

**Fig. 1(B):** The UV-VIS absorbance spectra of Se-NPs (60nm) at 400 nm wave length.

**Table 5.** Amelioration of AFB1-induced hepatocarcinogenesis by nano selenium and thyme (n = 5 for each group).

Groups	MDA nmol/g tissue	TAC Mmol/l	Nitric Oxid	GSH µmol/mg protein	CAT IU/mg protein	Gpx u/mg protein	Sod u/mg protein
Control	7.40 <sup>d</sup> ±0.36	6.40 <sup>a</sup> ±0.13	0.52 <sup>d</sup> ±0.01	411.60 <sup>a</sup> ±2.04	9.62 <sup>a</sup> ±0.07	17.12 <sup>a</sup> ±0.18	247.64 <sup>b</sup> ±8.70
AFB1	13.16 <sup>a</sup> ±0.48	3.63 <sup>d</sup> 0.08	0.76 <sup>a</sup> ±0.01	329.00 <sup>c</sup> ±9.66	5.10 <sup>d</sup> ±0.15	10.34 <sup>d</sup> ±0.41	224.38 <sup>c</sup> ±3.84
AFB1+low dose of SeNPs	9.16 <sup>c</sup> ±0.24	6.16 <sup>a</sup> ±0.06	0.63 <sup>c</sup> ±0.01	396.40 <sup>ab</sup> ±4.37	7.36 <sup>b</sup> ±0.10	14.21 <sup>b</sup> ±0.11	265.09 <sup>a</sup> ±4.87
AFB1+high dose of SeNPs	10.46 <sup>b</sup> ±0.28	5.58 <sup>b</sup> ±0.12	0.66 <sup>b</sup> ±0.01	355.2 <sup>b</sup> ±5.17	609 <sup>c</sup> ±0.17	13.69 <sup>b</sup> ±0.19	241.94 <sup>b</sup> ±6.06
AFB1+ Thyme	10.42 <sup>b</sup> ±0.07	4.74 <sup>c</sup> ±0.07	0.65 <sup>bc</sup> ±0.01	355.60 <sup>b</sup> ±8.06	5.83 <sup>c</sup> ±0.18	12.91 <sup>c</sup> ±0.21	257.29 <sup>ab</sup> ±0.59

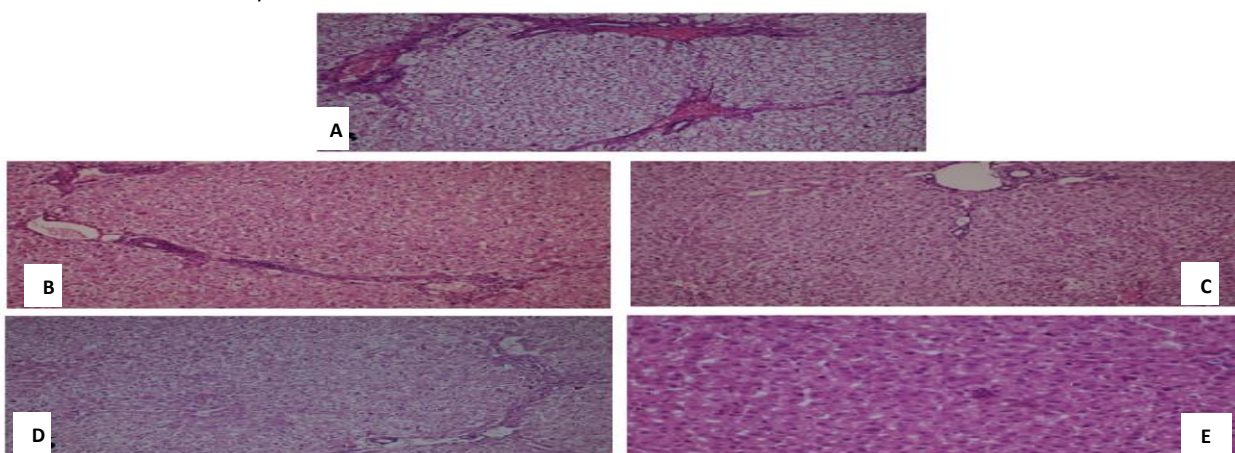
Significantly different (p<0.05) according to Duncan's test.

Our biochemical findings in the Table (5) illustrated that AFB1 increased the concentration of NO and MDA but reduced values of GSH and TAC. Also, the values of SOD, CAT, and GSH-PX in the homogenate of liver tissues of rabbits have been decreased.

**Table 6.** Amelioration of AFB1-induced hepatocarcinogenesis by nano selenium and thyme extract (n= 5 for each group).

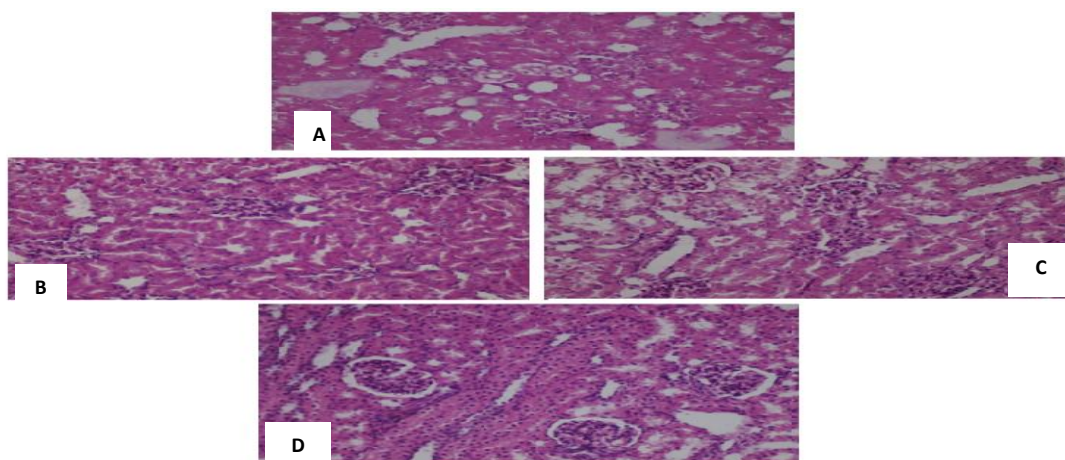
	Vit c ( $\mu\text{g}/\text{dl}$ )	Vit e ( $\mu\text{g}/\text{dl}$ )	Vit a ( $\mu\text{g}/\text{dl}$ )	$\beta$ carotene ( $\mu\text{g}/\text{dl}$ )
Control	0.70 <sup>a</sup> $\pm 0.01$	508.58 <sup>a</sup> $\pm 2.30$	43.54 <sup>a</sup> $\pm 0.92$	24.61 <sup>a</sup> $\pm 1.20$
AFB1	0.39 <sup>c</sup> $\pm 0.01$	364.52 <sup>c</sup> $\pm 3.32$	32.26 <sup>d</sup> $\pm 0.51$	13.46 <sup>c</sup> $\pm 0.21$
AFB1+ nanoselenium	0.57 <sup>b</sup> $\pm 0.02$	446.19 <sup>b</sup> $\pm 10.91$	40.04 <sup>b</sup> $\pm 0.28$	19.29 <sup>b</sup> $\pm 0.32$
AFB1+ nanoselenium	0.58 <sup>b</sup> $\pm 0.01$	438.55 <sup>b</sup> $\pm 6.01$	37.01 <sup>c</sup> $\pm 0.52$	18.35 <sup>b</sup> $\pm 0.76$
AFB1+ Thyme	0.57 <sup>b</sup> $\pm 0.02$	443.15 <sup>b</sup> $\pm 7.40$	37.60 <sup>c</sup> $\pm 0.45$	18.59 <sup>b</sup> $\pm 0.60$

Herein, results in Table (6) showed the low serum retinol, and  $\alpha$ -tocopherol in aflatoxicated rabbits compared to control animals. Currently, data observed. In Table (5 and 6) regarded that feed supplementation with SeNPs and Thyme extract in aflatoxicated rabbits resulted in ameliorating their toxic effects on biochemical functions of vital organs and increased the liver antioxidant activity.



**Fig.(2):** Aflatoxicated control group revealed thickening of portal tracts' areas, portal-portal bridge formation associated with severe hydropic degeneration of hepatocytes and congestion of hepatic blood vessel. H&E X 200 (A). High dose selenium-treated group showed improvement of hepatocytes status, reduction in the incidence of portal-portal bridge formation and less thickening of portal tracts' areas. Multiple foci of neutrophil infiltration were demonstrated (B). Low dose selenium-treated group had healthy hepatocyte, absence portal-portal bridge formation and normal portal tracts' areas H&E X 200 (C). Thyme-treated group showed mild portal bridge formation H & E x200 (D). Thyme treated group exhibited minute foci of neutrophils. H & E X 200 (E).

The histopathological status of renal tissues revealed that the aflatoxicated group showed severely swollen distal tubular lining epithelial and reduction of Bowman's space. Fig. 3 (A). Whereas the supplementation of thyme, Se-NPs in feeds of aflatoxicated rabbits resulted in a mild degree of tubular degeneration, and Bowman's spaces appeared normal. Fig. 3 (B, C, D).



**Fig 3.** Aflatoxicated group revealed highly eosinophilic tubular lining, severely swollen distal tubular lining epithelia. The renal corpuscles showed reduction of Bowman's space H&E X 200 (A). High dose selenium-treated group revealed mild degree of tubular epithelia hydropic degeneration associated with reduced Bowman's spaces (B). Low dose selenium-treated group had healthy tubules, absence portal-portal bridge formation and normal portal tracts' areas H&E X 200 (C). Thyme-treated group showed mild tubular degeneration. H&E X 20 (D)



**Table 7.** Levels of AFB1 residues in liver of rabbits in the different experimental groups (ppb).

Examined tissues	Trials of experimental design				
	T1	T2	T3	T4	T5
Liver (ug/kg) ppb	0.0	23.30±0.186	2.5±0.102	1.3±0.124	0.010±0.001

T1: Negative control - T2: aflatoxicated( Positive control) - T3: afla + Thym Extract

T4: afla + Low dose of SeNPs (0.3 mg/kg) - T5: afla + High dose of SeNPs (0.5 mg/kg obb.w.)

In the present work, the foregoing results in [Table \(7\)](#) significant beneficial effects of SeNPs and thyme supplementation against aflatoxicosis are also accompanied by decreasing AFB1 residues in liver tissues. The positive control rabbits of T2 had a high amount of AFB1 in the liver (23.30±0.186 ppb). Whereas, the aflatoxicated animals of T3 that supplemented with thyme extracts are also showed a reduction in the amount of AFB1 up to (2.5±0.102). The AFB1 residues Se-NPs treated aflatoxicated rabbits in T5 showed marked degradation toxin from liver higher ((0.010±0.001 ppb) than T4 (1.3±0.124 ppb).

## DISCUSSION

The fungal pollution of animal feed is associated with the adverse environmental factors of climates (Monda *et al.*, 2020). (Hassan *et al.*, 2016) isolated *Aspergillus* sp. as predominant molds in animal feeds(68%), while, the other fungi were also recovered as *Mucor* sp. (35%),*Penicillium* sp.(31%), *Fusarium* sp.(23%), *Rhizopus* sp.(21%),*Cladosporium* sp.(6%) and *Scopulariopsis* sp.(4%).(Similarly, Nooh *et al.*, 2014; Hassan *et al.*, 2017,2018), recovered the *Aspergillus* sp. as the most common isolated molds (100%) from feed in comparison to other molds.

Mycotoxins are of significant health hazard, especially in tropical districts and over 80% of the cereals foods have high levels of mycotoxins (Monda *et al.*, 2020). In addition, several adverse effects were detected due to consumption of contaminated food and feeds with fungi and aflatoxins as the occurrence of liver cancers (Nayak and Sashidhar, 2010, El-Nahass *et al.*, 2019). Moreover, various studies recovered AFs in animal and poultry feeds (El-Hamaky *et al.*, 2016) at levels ranging from (170-750 ppb) and (Hassan *et al.*, 2017) in poultry feed (97.5± 1.63 ppb) and yellow corn (31±1.36 ppb). (However, Hassan *et al.*, 2017) illustrated that he recovered *A. flavus* strains from ration samples produced a significant level of aflatoxin (100%) with the mean levels of (266±4.16).

In recent years, nanotechnology has been used employed in several biomedical fields (Bai *et al.*, 2018; Hassan *et al.*, 2020, 2021). The obtained results were similar to the detected findings of (Atul *et al.*, 2010; Jay and Shafkat 2018; Verma and Maheshwari, 2018). All updated studies detected the useful ability of nanomaterials as antioxidants, antitoxin, and other uses in the improvement of animal health (El-Sayed, 2020; Salem and Fouda, 2021; Hassan *et al.*, 2021).

In the present work, Se NPs were employed to improve the adverse carcinogenic effects Of aflatoxins in rabbits. Several studies were detected these changes in biochemical parameters in aflatoxicated rabbits (Hassan *et al.*, 2016; El-Nahas *et al.*, 2019). Aflatoxins B1 can also activate the oxidative reactions (ROS) and consequent lipid peroxidation in cells via inducing phospholipase that disrupts the integrity of cell membrane, DNA damage, and cancers (Lawrence, 2009; Aborehab and Waly, 2019). Moreover, the ability of AFB1 to induce DNA damage plays an important role in AFB1 carcinogenicity (Al-Anati and Petzinger, 2006; Yunus *et al.*, 2011).

Our biochemical findings illustrated that AFB1 increased the concentration of NO and MDA but reduced values of GSH and TAC. Also, the values of SOD, CAT, and GSH-PX in the homogenate of liver tissues of rabbits have been decreased. It is suggested that the elevated MDA as an oxidative stress system and lowered CAT and GPx activities as a protective system caused the development of cell damage (Benkerroum, 2020). Other studies detected that AFs caused liver cirrhosis due to decrease in the GSH stores (Mogda *et al.*, 2014, Neeff *et al.*, 2018). The decline in enzyme activities and increased MDA levels produced by AFB1 may be resulted from decreased ability of cells and tissues and also the nitric oxide formation in liver cells (Wang, 2009).

Herein, the low serum retinol, and a-tocopherol in aflatoxicated rabbits compared to control animals was also detected by (Mubarak *et al.*, 2009; Neeff *et al.*,2018; Dana *et al.*, 2018). They illustrated that aflatoxins cause a reduction in the antioxidant defense systems of cells (vitamins and glutathione) and disrupt the cell membrane integrity (Lee and Jacobs 2005; Kanchana *et al.*, 2012). Currently, data observed in our study regarded that feed supplementation with SeNPs and Thyme extract in aflatoxicated rabbits resulted in ameliorating their toxic effects on biochemical functions of vital organs and increased the liver antioxidant activity. Similarly, several studies reported that SeNPs have a significant functional role in GSH-PX activities which has huge importance during normal metabolic activity and stress during aflatoxicosis in rabbits (Jampilek *et al.*, 2020; Hamed and Selim, 2021). It has anti-apoptotic activities on oxidation-reduction, DNA synthesis and initiating benefit functions of carotenoids and vitamin A (Tayeb and Qader, 2012; Hamed and Selim, 2021). Currently, Se-NPs improved the adverse actions which resulted from aflatoxicosis as protein and DNA synthesis in rabbits and reduced destruction of lymphocyte cells and DNA damages (Bhattacharjee, 2016). Moreover, SeNPs have significant effects in the initiation of the formation of glutathione and cause less oxidative stress, and showed high biological activity and good (Cheng *et al.*, 2003; Peng *et al.*, 2007). Whereas, the thyme extracts are known antioxidant materials which have similar activity as a-tocopherol and other biological antioxidants (Lee and Shibamoto, 2002). Moreover, the administration of thyme compounds improves the adverse effects that resulted from aflatoxicosis, where, they prevent oxidative damage and oxidative stress as (lipid peroxidation which is associated with liver cells cancer (Esmail, 2018; Nazarizadeh *et al.*, 2019).These effects due to their contents of thymol and Phenolic compounds which have antioxidant activity (El-Nekeety *et al.*, 2011; El-Sherbeny *et al.*, 2021).

Herein, the observation of treatment effects on the tissues of vital organs was investigated via histopathological studies. The results indicated the large prevalence of liver cells cirrhosis and necrosis in aflatoxicated groups. It is suggested that these adverse effects of Afs are due to the liberation of large amounts of ROS that destroyed cells membrane, DNA, and death of liver cells (El-Nekeetya *et al.*, 2014; Hassan *et al.*, 2016, 2020, 2021; Hamed and Selim, 2021). Currently, the supplementation of aflatoxicated rabbits with high and low doses of SeNPs resulted in significant histological improvement of hepatic cells injury and neutrophil infiltration. These beneficial effects may be due to the SeNPs preventing cell membrane disruption and enhancement of antioxidant capacities (Shi *et al.*, 2015; Sun *et al.*, 2015) and its sequestration by the mitochondrial membrane and activation and hence no occurrence of cell apoptosis (Wang *et al.*, 2013, Hassan *et al.*, 2020, 2021; Hamed and Selim, 2021).

Similar results in our study were obtained in the thyme extracts treated group, where, the histological hepatocyte necrosis of aflatoxicated rabbits was significantly repaired in form of mild necrosis. These benefits of thyme were detected in several studies as (Lin *et al.*, 2009; Esmail, 2018; Nazarizadeh *et al.*, 2019) who recorded their potentials against inflammations of aflatoxicosis and prevention the liver cells necrosis and cause tissue restoration. They added that these effects were dose-dependent, where; the low concentrations prevent cell cirrhosis, while higher levels of thyme extracts resulted in apoptosis and fibrosis.

Herein, the histopathological status of renal tissues revealed that the aflatoxicated group showed severely swollen distal tubular lining epithelial and reduction of Bowman,s space. Whereas the supplementation of thyme, Se-NPs in feeds of aflatoxicated rabbits resulted in a mild degree of tubular degeneration, and Bowman,s spaces appeared normal. Similar findings were detected by (Hassan *et al.*, 2016, 2020; El-Nahas *et al.*, 2019; Hamed and Selim, 2021). In the present work, the foregoing results significant beneficial effects of SeNPs and thyme supplementation against aflatoxicosis are also accompanied by decreasing AFB1 residues in liver tissues Therefore, the feed supplementation with Se-NPs and thyme extracts has significant potential in activating the hepatic antioxidant actions and removing the toxic adverse effects of aflatoxins (Hamed and Selim, 2021).

## CONCLUSION

Animal mycosis and mycotoxicosis caused significant economic losses in animal health and output across the world. As a result, all authorities are concerned about effective management of unfavourable environmental variables that encourage the production of fungus and their toxin. The negative toxic effects of AFB1 on rabbit biological metrics were discovered in this investigation. These included potentially harmful oxidative effects on liver cells, which resulted in a reduction in antioxidant activity, leading to liver cirrhosis and death in toxicities rabbit cells. The toxic alterations of aflatoxicosis were greatly improved by supplementing rabbit diets with Se-NPs or thyme extracts. Metal nanoparticles, such as SeNPs, and natural plants, such as thyme, exhibit hepatoprotective properties due to their antioxidant activity. Furthermore, essential oils like thyme have a minimal toxicity for cattle and are a non-toxic environmental substance. Because the protective benefits of SeNPs are restricted in order to prevent the toxicity risk of nanomaterials by using low safe dosages, more laboratory research is needed to ensure their safe biomedical application in field animals to enhance human and livestock health and production.

## ACKNOWLEDGEMENT:

Great appreciation and heart thank to Prof. Dr. Hazem M. Mansour, for his advice in identification of prepared nanomaterials.

## AUTHOR'S CONTRIBUTION:

Atef Hassan conceived and designed the experiments; Mahmoud M. Arafa Mogda K. Mansour, Essam M. Ibrahim, Noha H. Oraby, and AlaFoad did the methodology, all authors did the manuscript preparation, analyzed the data and wrote this manuscript. Corresponding author: Noha H. Oraby. All authors read and approved the final manuscript.

## REFERENCES



- Abdel-Aziem, S. H., Hassan, A. M., El-Denshary, E. S., Hamzawy, M. A., Mannaa, F. A., and Abdel-Wahhab, M. A. (2014). Ameliorative effects of thyme and calendula extracts alone or in combination against aflatoxins-induced oxidative stress and genotoxicity in rat liver. *Cytotechnology*, 66(3), 457-470.
- Aborehab, N. M., and Waly, N. E. (2019). IL-6 and NFE2L2: A putative role for the hepatoprotective effect of *N. Sativa*, *P. Ginseng* and *C. Sempervirens* in AFB-1 induced hepatocellular carcinoma in rats. *Toxicology reports*, 6, 457-464.
- Al-Anati, L., & Petzinger, E. (2006). Immunotoxic activity of ochratoxin A. *Journal of veterinary Pharmacology and Therapeutics*, 29(2), 79-90. <https://doi.org/10.1111/j.1365-2885.2006.00718.x>
- AOAC (2000) Official Methods of Analysis. 17th Edition, The Association of Official Analytical Chemists, Gaithersburg, MD, USA. Methods 925.10, 65.17, 974.24, 992.16.
- Bai, D. P., Lin, X. Y., Huang, Y. F., and Zhang, X. F. (2018). Theranostics aspects of various nanoparticles in veterinary medicine. *International journal of molecular sciences*, 19(11), 3299.
- Benkerroum, N. (2020). Chronic and acute toxicities of aflatoxins: Mechanisms of action. *International journal of environmental research and public health*, 17(2), 423.
- Bhattacharjee, S. (2016). DLS and zeta potential –What they are and what they are not. *Journal of Controlled Release*. 235, 337-351. <https://doi.org/10.1016/j.jconrel.2016.06.017>

- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical biochemistry*, 72(1-2), 248-254.
- Burgner, D., Rockett, K., and Kwiatkowski, D. (1999). Nitric oxide and infectious diseases. *Archives of disease in childhood*, 81(2), 185-188.
- Çam, Y., Atasever, A., Eraslan, G., Kibar, M., Atalay, Ö., Beyaz, L., Inci, A. & Liman, B. C. (2008). Eimeria stiedae: experimental infection in rabbits and the effect of treatment with toltrazuril and ivermectin. *Experimental parasitology*, 119(1), 164-172. <https://doi.org/10.1016/j.exppara.2008.01.005>.
- Cheng, Y. N., Zhang, M., Xia, L., and Zhang, J. Xing (2012) The toxic effects and mechanism of CuO and ZnO nanoparticles. *Materials* 5(12): 2850–2871.
- Cheng, Y. J., Guo, W. W., Yi, H. L., Pang, X. M., and Deng, X. (2003). An efficient protocol for genomic DNA extraction from Citrus species. *Plant Molecular Biology Reporter*, 21(2), 177-178.
- Chitra, K. C., Latchoumycandane, C., and Mathur, P. P. (1999). Chronic effect of endosulfan on the testicular functions of rat. *DNA*, 1(11.131), 35.
- Dana, M. A., Kordbacheh, P., Ghazvini, R. D., Moazeni, M., Nazemi, L., & Rezaie, S. (2018). Inhibitory effect of vitamin C on Aspergillus parasiticus growth and aflatoxin gene expression. *Current Medical Mycology*, 4(3), 10. <http://doi.org/10.18502/cmm.4.3.170>.
- El-Hamaky, A. M., Hassan, A. A., El Yazeed, H. A., and Refai, M. K. (2016). Prevalence and Detection of Toxigenic *A. flavus*, *A. niger* and *A. ochraceus* by traditional and molecular biology methods in feeds. *International Journal of Current Research*, 8(1), 25621-25633.
- Ellman, G. L. (1959). Tissue sulfhydryl groups. *Archives of biochemistry and biophysics*, 82(1), 70-77.
- El-Nahass, E. S., Moselhy, W. A., Hassan, N. E. H. Y., and Hassan, A. A. (2019). Evaluation of the protective effects of adsorbent materials and ethanolic herbal extracts against aflatoxins hepatotoxicity in albino rats: histological, morphometric and immunohistochemical study. *Adv. Anim. Vet. Sci*, 7(12), 1140-1147.
- El-Nekeety, A. A., Abdel-Azeim, S. H., Hassan, A. M., Hassan, N. S., Aly, S. E., and Abdel-Wahhab, M. A. (2014). Quercetin inhibits the cytotoxicity and oxidative stress in liver of rats fed aflatoxin-contaminated diet. *Toxicology Reports*, 1, 319-329.
- El-Nekeety, A. A., Abdel-Azeim, S. H., Hassan, A. M., Hassan, N. S., Aly, S. E., and Abdel-Wahhab, M. A. (2014). Quercetin inhibits the cytotoxicity and oxidative stress in liver of rats fed aflatoxin-contaminated diet. *Toxicology Reports*, 1, 319-329.
- El-Sayed, A., and Kamel, M. (2020). Advanced applications of nanotechnology in veterinary medicine. *Environmental Science and Pollution Research*, 27(16), 19073-19086.
- El-Sherbeny E. A, Housseiny M.M., Aboelmagd H.I. & Gaber E. (2021). Effect of essential oil of wild thyme plant on the genetic behavior of aflatoxin. *Fresenius Environmental Bulletin* · February 2021, 47, 12A(9382-9391)
- Esmail, N. S. (2018). Effect of thyme and rosemary on liver functions in toxicated rats by aflatoxins. *Journal of Food and Dairy Sciences*, 2018, 59-65.
- Grigore, A., Paraschiv, I. N. A., Colceru-Mihul, S., Bubueanu, C., Draghici, E., and Ichim, M. (2010). Chemical composition and antioxidant activity of Thymus vulgaris L. volatile oil obtained by two different methods. *Romanian Biotechnological Letters*, 15(4), 5436-5443.
- Hamed, G. M. S., and Selim, M. A. (2021). Protective effect of nano-selenium against experimentally induced toxicity by aflatoxin B1 (AFB1) on the gingiva and periodontal ligament of albino rats (histological and immunohistochemical study). *Codex*, 46, 21-01.
- Hassan, A. A., El-Ahl, R. M. S., Oraby, N. H., El-Hamaky, A. M., and Mansour, M. K. (2021). Zinc nanomaterials: Toxicological effects and veterinary applications. In *Zinc-Based Nanostructures for Environmental and Agricultural Applications* (pp. 509-541). Elsevier.
- Hassan, A. A., El-Shafei, H. M., Sayed El-Ahl, R. M. H., and El-Hamaky, A. M. (2017). Molecular detection the influence of aflatoxin biosynthetic genes by Aspergillus flavus before and after bacillus subtilis and candida albicans biocontrol. *9th Scient. Congr. Egypt. J. Anim. Manag*, 1-19.
- Atef, H. A., El-Shafei, H. M., Mansour, M. K., Al Kalamawy, N. M., Hassan, N. E. H. Y., and Lashin, A. I. (2018). Prevalence of ochratoxigenic fungi and ochratoxin A residues in animal feeds and modulation of their toxic effects by glutathione. *International Journal of Current Microbiology and Applied Science* 7(4), 2559-2582.
- Hassan, A. A., Mansour, M. K., Sayed-ElAhl, R. M. H., El-Din, H. T., Awad, M. E. A., & Younis, E. M. (2020). Influence of selenium nanoparticles on the effects of poisoning with aflatoxins. *Adv. Anim. Vet. Sci*, 8(s2), 64-73. <http://dx.doi.org/10.17582/journal.aavs/2020/8.s2.64.7>
- Henry, H. L., Midgett, R. J., and Norman, A. W. (1974). Regulation of 25-hydroxyvitamin D3-1-hydroxylase in vivo. *Journal of Biological Chemistry*, 249(23), 7584-7592.
- Huang, W., Yan, M., Duan, H., Bi, Y., Cheng, X., and Yu, H. (2020). Synergistic antifungal activity of green synthesized silver nanoparticles and epoxiconazole against Setosphaeria turcica. *Journal of Nanomaterials*, 2020.

- ISO. (2008). *Microbiology of Food and Animal Feeding Stuffs: Horizontal Method for the Enumeration of Yeasts and Moulds*. ISO.
- Kanchana, K., Hedhayathullahkhan, H. B., Vedagiri, A., Palanivelu, S., and Panchanatham, S. (2013). Pharmacological effect of Kalpaamruthaa on renal and cardiac damage induced by ingestion of aflatoxin B1 studied in wistar rats. *Biomedicine & Preventive Nutrition*, 3(3), 233-240.
- Koracevic, D., Koracevic, G., Djordjevic, V., Andrejevic, S., and Cosic, V. (2001). Colorimetric method for determination of total antioxidant capacity. *J Clin Pathol*, 54(5), 356-361.
- Lawrence, T. (2009). The nuclear factor NF- $\kappa$ B pathway in inflammation. *Cold Spring Harbor perspectives in biology*, 1(6), a001651.
- Lee, K. G., and Shibamoto, T. (2002). Determination of antioxidant potential of volatile extracts isolated from various herbs and spices. *Journal of agricultural and food chemistry*, 50(17), 4947-4952.
- Lee, D. H., & Jacobs Jr, D. R. (2005). Association between serum gamma-glutamyltransferase and C-reactive protein. *Atherosclerosis*, 178(2), 327-330. <https://doi.org/10.1016/j.atherosclerosis>.
- Lin, Y. L., Lin, C. Y., Chi, C. W., & Huang, Y. T. (2009). Study on antifibrotic effects of curcumin in rat hepatic stellate cells. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 23(7), 927-932. <https://doi.org/10.1002/ptr.2764>
- Menon, S., and Shanmugam, V. (2020). Cytotoxicity analysis of biosynthesized selenium nanoparticles towards A549 lung cancer cell line. *Journal of Inorganic and Organometallic Polymers and Materials*, 30(5), 1852-1864.
- Mogda, K. M., and Hanan, A. Tag El-Din, Safaa, M. Abou El Soud Abo Norag, M.A. (2014). A trial to ameliorate the toxic effect of Aflatoxin in rabbits. *Journal of Veterinary Science* 15-25.
- Moghimi, S. J.M., Hunter, A. C. & Murry, J. C. (2005). Nanomedicine: current status and future prospects. *The FASEB journal*. 19(3), 311-330. <https://doi.org/10.1096/fj.04-2747rev>
- Monda, E., Masanga, J., and Alakonya, A. (2020). Variation in occurrence and aflatoxigenicity of *Aspergillus flavus* from two climatically varied regions in Kenya. *Toxins*, 12(1), 34.
- Mubarak, A., Rashid, A., Khan, I. A., and Hussain, A. (2009). Effect of Vitamin E and selenium as immunomodulators on induced aflatoxicosis in broiler birds. *Pakistan Journal of life Science* 7(1), 31-34.
- Nazarizadeh, H., Mohammad Hosseini, S., and Pourreza, J. (2019). Effect of plant extracts derived from thyme and chamomile on the growth performance, gut morphology and immune system of broilers fed aflatoxin B1 and ochratoxin A contaminated diets. *Italian Journal of Animal Science*, 18(1), 1073-1081.
- Nishikimi, M., Rao, N. A., and Yagi, K. (1972). The occurrence of superoxide anion in the reaction of reduced phenazine methosulfate and molecular oxygen. *Biochemical and biophysical research communications*, 46(2), 849-854.
- Nooh, A., Amra, H., Youssef, M. M., and El-Banna, A. A. (2014). Mycotoxins and toxigenic fungi occurrence in Egyptian maize. *International Journal of Advanced Research* 2(2), 521-532.
- Ohkawa, H., Ohishi, N., and Yagi, K. (1979). Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Analytical Biochemistry*, 95(2), 351-358.
- Paglia, D. E., and Valentine, W. N. (1967). Studies on the quantitative and qualitative characterization of erythrocyte glutathione peroxidase. *The Journal of laboratory and clinical medicine*, 70(1), 158-169.
- Peng, D., Zhang, J., Liu, Q., and Taylor, E. W. (2007). Size effect of elemental selenium nanoparticles (Nano-Se) at supranutritional levels on selenium accumulation and glutathione S-transferase activity. *Journal of inorganic biochemistry*, 101(10), 1457-1463.
- Pitt, J. I., and Hocking, A. D. (2009). *Fungi and food spoilage* (Vol. 519, p. 388). New York: Springer.
- Placha, I., Takacova, J., Ryzner, M., Cobanova, K., Laukova, A., Stropfova, V., ... and Faix, S. (2014). Effect of thyme essential oil and selenium on intestine integrity and antioxidant status of broilers. *British Poultry Science*, 55(1), 105-114.
- Rasooli, I., Rezaei, M. B., and Allameh, A. (2006). Ultrastructural studies on antimicrobial efficacy of thyme essential oils on *Listeria monocytogenes*. *International journal of infectious diseases*, 10(3), 236-241.
- Salem, S. S., & Fouda, A. (2021). Green synthesis of metallic nanoparticles and their prospective biotechnological applications: an overview. *Biological Trace Element Research*, 199(1), 344-370. <https://doi.org/10.1007/s12011-020-02138-3>.
- Shi, D., Liao, S., Guo, S., Li, H., Yang, M., and Tang, Z. (2015). Protective effects of selenium on aflatoxin B1-induced mitochondrial permeability transition, DNA damage, and histological alterations in duckling liver. *Biological trace element research*, 163(1), 162-168.
- Smith, J. E. (1997). Aflatoxins. *Handbook of Plant and Fungal Toxicants*. Edited by JP Felix D'mello, Crc Pres, 269-285.
- SPSS 14 (2006). "Statistical Package for Social Science, SPSS for windows Release14.0.0, and 12 June, 2006. Standard Version, Copyright © SPSS Inc. USA.



- Sun, L. H., Zhang, N. Y., Zhu, M. K., Zhao, L., Zhou, J. C., and Qi, D. S. (2015). Prevention of aflatoxin B1 hepatotoxicity by dietary selenium is associated with inhibition of cytochrome P450 isozymes and up-regulation of 6 selenoprotein genes in chick liver. *The journal of nutrition*, 146(4), 655-661. <https://doi.org/10.3945/jn.115.224626>
- Suvarna, K.S., Layton, C., & Bancroft, J.D. (2012). Bancroft's theory and practice of histological techniques. 7th ed. China: Churchill Livingstone.
- Swayeh, N. H., Abu-Raghif, A. R., Qasim, B. J., and Sahib, H. B. (2014). The protective effects of Thymus Vulgaris aqueous extract against Methotrexate-induced hepatic toxicity in rabbits. *Int J Pharm Sci Rev Res*, 29, 187-93.
- TAYEB, İ., and QADER, G. (2012). Effect of feed supplementation of selenium and vitamin E on production performance and some hematological parameters of broiler. *KSÜ Doğa Bilimleri Dergisi*, 15(3), 46-56.
- Tiew, P. Y., Mac Aogain, M., Ali, N., Thng, K. X., Goh, K., Lau, K. J., & Chotirmall, S. H. (2020). The mycobiome in health and disease: emerging concepts, methodologies and challenges. *Mycopathologia*, 185(2), 207-231. <https://doi.org/10.1007/s11046-019-00413-z>
- Ulaiwi, A. H. (2018). Effect of levamisole, Vitamin E, and selenium against aflatoxicosis in broilers chicken. *Veterinary World*, 11(2), 248.
- Verma, P., and Maheshwari, S. K. (2018). Preparation of silver and selenium nanoparticles and its characterization by dynamic light scattering and scanning electron microscopy. *Journal of microscopy and ultrastructure*, 6(4), 182.
- Wang, F., Shu, G., Peng, X., Fang, J., Chen, K., Cui, H., ... and Lai, W. (2013). Protective effects of sodium selenite against aflatoxin B1-induced oxidative stress and apoptosis in broiler spleen. *International journal of environmental research and public health*, 10(7), 2834-2844.
- Wang, Y. (2009). Differential effects of sodium selenite and nano-Se on growth performance, tissue Se distribution, and glutathione peroxidase activity of avian broiler. *Biological Trace Element Research*, 128(2), 184-190.
- Youssef, K., de Oliveira, A. G., Tischer, C. A., Hussain, I., and Roberto, S. R. (2019). Synergistic effect of a novel chitosan/silica nanocomposites-based formulation against gray mold of table grapes and its possible mode of action. *International Journal of Biological Macromolecules*, 141, 247-258.
- Yunus, A. W., Razzazi-Fazeli, E., & Bohm, J. (2011). Aflatoxin B1 in affecting broiler's performance, immunity, and gastrointestinal tract: A review of history and contemporary issues. *Toxins*, 3(6), 566-590. <https://doi.org/10.3390/toxins3060566>
- Zhang, L., Wang, L., Jiang, Z., and Xie, Z. (2012). Synthesis of size-controlled monodisperse Pd nanoparticles via a non-aqueous seed-mediated growth. *Nanoscale research letters*, 7(1), 1-6.
- Zhao, G., Wu, X., Chen, P., Zhang, L., Yang, C. S., and Zhang, J. (2018). Selenium nanoparticles are more efficient than sodium selenite in producing reactive oxygen species and hyper-accumulation of selenium nanoparticles in cancer cells generates potent therapeutic effects. *Free Radical Biology and Medicine*, 126, 55-66.
- Zhou, X., and Wang, Y. (2011). Influence of dietary nano elemental selenium on growth performance, tissue selenium distribution, meat quality, and glutathione peroxidase activity in Guangxi Yellow chicken. *Poultry Science*, 90(3), 680-686.

	<p><b>Copyright:</b> © 2022 by the authors. Licensee EJAR, EKB, Egypt. EJAR offers immediate open access to its material on the grounds that making research accessible freely to the public facilitates a more global knowledge exchange. Users can read, download, copy, distribute, print or share a link to the complete text of the application under <a href="https://creativecommons.org/licenses/by-nc-sa/4.0/">Creative Commons BY-NC-SA 4.0 International License</a>.</p>	
---	--	---

## التأثير الوقائي للجسيمات النانومترية للسيلينيوم ومستخلص الزعتر في إزالة الآثار السامة للأفلاتوكسين ب1 في الارانب

عاطف عبد العزيز حسن<sup>1</sup>، مجدة كامل منصور<sup>2</sup>، عصام محمد ابراهيم<sup>3</sup>، محمود محمد عرفة<sup>2</sup>، علا فؤاد طلخان<sup>4</sup> نهى حسن عرابي\*

<sup>1</sup> قسم الفطريات والسموم الفطرية، معهد بحوث الصحة الحيوانية، مركز البحوث الزراعية

<sup>2</sup> قسم الكيمياء الحيوية والسموم ونقص الغذاء، معهد بحوث الصحة الحيوانية، مركز البحوث الزراعية

<sup>3</sup> قسم الباثولوجيا والباثولوجيا الكلينيكية، معهد بحوث الصحة الحيوانية، مركز البحوث الزراعية

<sup>4</sup> قسم الكيمياء الحيوية، معمل فرعي المنوفية، معهد بحوث الصحة الحيوانية

\* بريد المؤلف المراسل: [nohaoraby25@gmail.com](mailto:nohaoraby25@gmail.com)

### الملخص

في هذه الدراسة تم عزل العديد من العترات التي لها القدرة علي افراز السموم في علائق الارانب حيث وصل مستوى الافلاتوكسين في العلائق الى 300 جزء من البليون. وقد تم استخدام جزيئات السيلينيوم المحضرة كيميائيا لتخفيف الآثار الضارة للسموم الفطرية في الارانب . كشفت القياسات البيوكيميائية في الانسجة والامصال لجميع الارانب التي تعاني من التسمم الفطري انها تسببت في تكوين الشوارد الحرة و ارتفاع مستويات أكسيد النيتريك والمالون الدهايد وانخفاض في مستوي المستوي الكلي لمضادات الاكسدة. بالإضافة الى انخفاض نسبة الريتنول في الدم والبيتا توكوفيرول . هذه التغيرات تبدلت الى القيم الطبيعية عن طريق الجرعات المختلفة عن طريق الفم لجزيئات السيلينيوم النانومترية (0.3- 0.5 مللجم/كجم) و الزعتر للارانب التي تعاني من التسمم بالافلاتوكسين . كما لوحظ دورهما في تراجع نسبة بقايا سموم الافلاتوكسين في الكبد. وبالتالي، يمكن إضافة جزيئات السيلينيوم النانومترية والزعتر الى الاعلاف الحيوانية كعوامل مضادة للاكسدة لها الدور في تنشيط الحالة الصحية للحيوان والقضاء على السموم الفطرية.

**الكلمات المفتاحية :** تقنية النانو ، جزيئات السيلينيوم النانومترية ، أفلاتوكسين ، الزعتر ، مضادات الأكسدة ، الفطريات