Original article



The role of N-Acetyl Cysteine in ameliorating Doxorubicin induced cardiotoxicity in rats

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Abstract

*Corresponding author Emad Altaher e-mail: emadeldeen@aswu.edu.eg Doxorubicin (DOX) is an anthracycline antibiotic and a quinonecontaining chemotherapeutic drug used for various types of solid and hematological cancers. However, it causes many toxic side effects,

including cardiac, renal, hematological, and testicular. The current study investigated the prophylactic and ameliorative effect of n-acetylcysteine (NAC) against DOX-induced cardiotoxicity in albino rats. Sixty rats were divided into six groups. Negative Control group: rats received food pellets and tap water ad libitum. NAC treated (positive control) group: rats received NAC (400 mg/kg/p.o.) Every other day for 28 days. DOX-treated group: rats received DOX (5 mg/kg, i.p.) for four weeks on days 1, 7, 14 and 21. NAC pretreated group: rats received NAC (400 mg/kg/p.o.) every other day one week earlier and then along with DOX (5 mg/kg/ i.p.) for the next four weeks on days 1, 7, 14 and 21). NAC1+DOX treated group : rats received NAC (400 mg/kg/ p.o.) every other day started at the first day of the experiment till the end of the experiment and DOX (5 mg/kg/ i.p.) for four weeks on day 1, 7, 14 and 21. NAC2+DOX treated group: rats received NAC (800 mg/kg, p.o.) every other day started at the first day of the experiment till the end of the experiment and DOX (5 mg/kg, i.p.) for four weeks on day 1, 7, 14 and 21. The present results showed a significant increase in the serum levels of LDH, CK-MB, CTn-I, plasma level of malondialdehyde (MDA). Whilst, plasma level of reduced glutathione (GSH) and Erythrocyte lysate level of glutathione peroxidase (GSH-Px) were significantly decreased. Moreover, there were histopathological abnormalities in the cardiac tissue of DOX-treated rats, was recognized by disrupted cardiac muscle fibers and foci of degeneration of the myocardium. The present study demonstrated that NAC has a cardioprotective effect on the cardiac damage induced by DOX, through inhibition of inflammation and oxidative stress. This cardio protective effect was more pronounced in both NAC pre-treated and concurrent (NAC1+DOX & NAC2+DOX), as it produced a significant increase in GSH and GSH-Px levels, a significant decrease in MDA, LDH, CK-MB, and CTn-I, and more improvement in the histopathological abnormalities, without a significant difference either between NAC pre-treated & concurrent groups or between groups of concurrent one (NAC1+DOX & NAC2+DOX). Keywords: Doxorubicin; N-Acetyl cysteine; cardiotoxicity; Oxidative stress.

I. INTRODUCTION:

II.

Doxorubicin (DOX) is one of the most effective 1) Drugs and extensively used chemotherapeutic agents for different types of solid and hematological tumors. Unfortunately, the usage of this drug leads to severe side effects of cardiotoxicity (Yu et al., 2018).

Cardiotoxicity is one of the hazardous chemotherapy side effects and was defined as a reduction in left ventricular ejection fraction (LVEF) of more than 10% to a value lower than 50% (Cardinale et al., 2015).

DOX acts by numerous mechanisms of action. Currently, it is unclear which of these 2) mechanisms are the most responsible for its a) associated cardiac toxicity. Many articles support that the generation of reactive oxygen species (ROS) due to DOX treatment and its consequential peroxidation. lipid calcillm dysregulation, and intervention in energy transfer could cause heart failure(Winningmalthet al., 2019).

While the molecular processes behind the anticancer effects of anthracyclines (ANTs) such b) as doxorubicin are well known and studied, the Imechanisms underlying their cardiotoxic effects are still poorly understood and controversial. It is action by directly targeting and hindering topoisomerase 2 (Top2) in cancer cells, more specifically the 2a isoform, halting DNA III-Glutathione peroxidase (GSH-Px) Tests were transcription, and replication (Murabito et al., 2020).

N-acetylcysteine (NAC), a precursor of reduced glutathione (GSH), was widely used as **3**) an antioxidant against reactive oxygen species (ROS) in several disorders related to oxidative stress (Teodorczyk and Schmidt, 2014).

N-Acetylcysteine (NAC) is an acetylated precursor of the amino acid L- cysteine. It used as an antidote for paracetamol intoxication and as a mucolytic agent (Elbini etal., 2016).

This study aimed to evaluate cardiac toxicity of doxorubicin at biochemical and histopathological levels in rats; and assessment of the protective role of N- Acetylcystiene in both pre-treatment and co-administration with doxorubicin against its induced cardiotoxicity.

Materials & Methods

1-N-acetyl-cysteine $(C_5H_9NO_3S)$ was purchased from (AK Scientific, Inc. company) with purity 98%, Stored at 2 to 8° C.

2-Doxorubicin (Adriblastina vials, Pharmacia Italia S.P.A., Italy) was purchased as an injectable commercial product. Each vial contains doxorubicin hydrochloride as a 50 mg dried powder stored at 2 -8°C. The contents of each vial were freshly dissolved in a sterile saline solution just before use in 25 ml so each one ml contains 2mg DOX and the rest is discarded after use.

Chemicals and reagents

Cardiac markers assessment Kits

Lactate dehydrogenase enzyme (LDH) Kits were purchased from the Egyptian Company of Biotechnology, EGYPT.

(CK-MB) kits VITRO KINETIC

SCIENT Germany

Troponin I ELISA Kits were purchased from IMMUNOSPEC Corporation, USA, CAT. No. E29-061.

- **Oxidative stress parameters**
- Lipid peroxide (Malondialdehyde) Kits was purchased from Bio-diagnostic Company, Egypt CAT. No., MD 25 29.
- well established that ANTs do their anticancer II- Glutathione Reduced (GSH) Kits were purchased from Bio-diagnostic company. Egypt CAT No. GR 25 11.
 - purchased from Bio-diagnostic Company. Egypt CAT. No. GP 25 24.

Animals:

The study was carried out on (60) male adult (two-month-old) albino rats, with an average weight (150-200 grams). They wereobtained from the animal house of the national center for research, Giza, Egypt. They were housed in an animal house, Faculty of Medicine, Assiut University, Egypt. The animals were kept under routine healthy laboratory conditions andwere fed normal food pellets ad libitum and tapwater, with room temperature being maintained

 $(25\pm2 \text{ °C})$. The ethics and husbandry conditions of animal research were considered according to the guidelines approved by the ethical committee of Aswan Faculty of Medicine, Aswan University.

III. Experimental design:

The rats were divided into six groups ten rats each. The experiment lasted for five weeks to investigate the cardiac toxicity induced by DOX and protective effects of NAC on heart organs.

Animal grouping

• Group A: (negative control): Rats have received food pellets and tap water ad libitum.

Group B: (NAC treated group): Rats were received NAC orally by gastric tube at adose of 400 mg/kg every other day for four weeks (*Dauletbaev et al., 2009*).

• Group C: Rats were received DOX intraperitoneal (i.p.) at a dose of 5mg/kg once a week for four weeks days (*Kulkarni and Swamy, 2015*).

• Group D: (NAC pre-treated) Rats were received NAC one week earlier orally by gastric tube at a dose of 400mg/kg every other day for 35 days and then with (DOX) 5mg/kg I.p. once for four weeks from the second week.

• Group E: Concurrent treatment (NAC1 + DOX) Rats were received NAC orally by gastric tube at a dose of 400mg/kg everyother day and (DOX) i.p. at a dose of 5mg/kg every week for 28 days.

• Group F: Concurrent treatment (NAC2 + DOX) Rats were received NAC orally by gastric tube at a dose of 800mg/kg everyother day and DOX i.p. at a dose of 5 mg/kg for 28 days.

At the end of the study, animals were anesthetized by urethane (1.2 g/kg) (*Cocchetto and Bjornsoon, 1983*), and sacrificed, then blood samples were collected and animals were dissected to expose the heart.

IV. Methods

A. Samples collection, preparation and preservation

Blood samples (**4ml**) from each rat were withdrawn from the retro-orbital venous plexus; each sample was divided into two tubes. (**2ml**) inside plan tube (no anticoagulant) for preparation of serum and (**2ml**) inside heparinized tube (**2ml**) for preparation of plasmaand erythrocyte lysate.

These samples prepared in Metabolic and Genetic disorders Unit. Faculty of Medicine. Assiut University as follow: -

1. **Serum preparation:** the samples in plan tubes allowed to clot for 30 min at 25°C, and then centrifuged at 2000 rpm for 20 min. Then pipette off the top yellow serum layer without disturbing the white buffy layer and preserved at - 20°C in Clinical Toxicology laboratory to the time of assay. Serum is used for estimation of cardiac enzymes (LDH, CK-MB, and CTnI).

2. **Plasma and Erythrocyte lysate preparation:** the samples in heparinized tube centrifuged at 3000 rpm for 20 min then pipette off the yellow plasma layer intubes and preserved at -20°C to the time of assay. Plasma samples were used for estimation of MDA and GSH levels.

After withdrawal of plasma, the cells were washed once with ten volumes of coldsaline. Then lyse the red cell pellets by adding four volumes of cold deionized water to the estimated pellet volume. Remove the red cell stroma by centrifuging at (4000) rpm for 10 min. The resulting clarified supernate (**Erythrocyte lysate**) was collected and preserved at - 20°C in Clinical Toxicology laboratory to the time of assay. **Erythrocyte lysate** is used for estimation of GSH-Px levels.

B. Biochemical analysis

1) Cardiac enzymes assessment

• LDH (lactate dehydrogenase enzyme) level in the serum of male albino rats was estimated by kinetic ultraviolet method using commercial available kits. according to manufacturer instructions.

• CK-MB (creatine kinase MB) level in the serum of male albino rats was estimated by kinetic ultraviolet method using commercial available kits according to manufacturer instructions (Vitro SCIENT) Germany.

• Troponin-I (CTn-I) level in serum of male albino rats was estimated by using commercial available Troponin I enzyme immunoassay test kits according to manufacturer instructions.

2) Oxidative stress markers assessment

• Lipid peroxide MDA (Malondialdehyde) level in plasma of male albino rats was estimated by using colorimetric method Lipid peroxide (Malondialdehyde) test kits. It was determined according to the method of (*Satoh K et al.*, *1978*).

• Glutathione reduced (GSH) level in plasma of male albino rats was estimated by using colorimetric method using test kits. It was determined according to the method of (*Beutler and Kelly, 1963*).

• Glutathione peroxidase (GSH-Px) level in erythrocyte lysate of male albino rats was estimated by using colorimetric method using test kits. It was determined according to the method of (*Paglia and Valentine, 1967*).

C. Histopathological studies

Autopsy sections were taken from the hearts of rats in different groups and fixed in 10% neutral buffered formalin for 24 h. washing was done with tap water, and then dehydration was carried out using serial dilutions of alcohol (methyl, ethyl, and absolute ethyl). Specimens were cleared in xylene and embedded in paraffin at 56°C in a hot air oven for 24 h. Paraffin wax tissue blocks were prepared for sectioning at 5µm thickness by a sledge microtome. The obtained tissue sections were collected on glass slides, deparaffinized, stained with hematoxylin, and eosin, and then examination was done through the light electric microscope (Banchroft et al., 1996).

D. Statistical analysis of data

Study data were analyzed using IBM_SPSS. Statistical package for social sciences Ver. 23. (Standard version copyright © SPSS Inc., 2011-2012. NY, USA. 2012). Kruskal-Wallis test was used as the data were abnormally distributed by using test of normality, then data were analyzed by Mann-Whitney U comparison test amongst different groups.

Results

1. Biochemical results.

a) Changes in serum LDH level

The serum level of LDH of the DOX- treated group showed a significant increase (*P*.

< 0.05) by 70.9% in comparison to the negative group (A). The serum level of LDH of (NAC pretreated, NAC1+DOX & NAC2+DOX) groups showed a significant decrease (P. < 0.05)by 20.3%, 40.1%, and 8.5% respectively when compared to the DOX-treated group. While the LDH level of groups (NAC1+DOX & NAC2+DOX) showed insignificant change (P.>0.05) when compared to NAC pre-treated group and that of the group (NAC2+DOX) showed insignificant change (P. >0.05) when compared to group (NAC1+DOX) (**as shown in Table 1&2) (Fig. 1**)

b) Changes in serum CK-MB level

The serum level of CK-MB of the DOX- treated group showed a significant increase (*P*.

< 0.05) when compared to the negative group (A). The levels of serum CK-MB of groups (NAC pretreated, NAC1+DOX & NAC2+DOX) showed a significant decrease (P.<0.05) when compared to the DOX-treated group, Also, the CK-MB level of groups (NAC1+DOX & NAC2+DOX) showed insignificant change (P.>0.05) when compared to NAC pre-treated group and that of a group (NAC2 + DOX) showed insignificant change (P.>0.05) when compared to (NAC1+DOX) group (**as shown in Table 1&2**) (**Fig. 1**)

c) Changes in serum cardiac troponin I level

The serum level of CTn-I of DOX- treated group showed a significant increase (P.< 0.05) when compared to the negative group (A). The levels of of groups (NAC pre-treated, serum CTn-I NAC1+DOX & NAC2+DOX) showed a significant decrease (P. < 0.05) when compared to the DOX-treated group while the serum levels of groups (NAC1+DOX & NAC2+DOX) showed insignificant change (P. > 0.05) when compared to the Pre-treated group. Also, the serum CTn-I level of the group (NAC2+DOX) showed insignificant change (P. > 0.05) when compared to (NAC1+DOX) group (as shown in Table 1&2) (Fig. 1)

d) Changes in plasma MDA levels

The plasma level of malondialdehyde (MDA) of DOX-treated showed a significant increase (P. < 0.05) by 238.4% when compared to the negative group (A). The levels of plasma MDA of NAC pre-treated showed a significant decrease (P. < 0.05) by 77.7 when compared to the DOX-treated group

while the levels of groups (NAC1+DOX & NAC2+DOX) showed insignificant change (P.> 0.05) when compared to the DOXtreated group. In addition, the MDA levels of the group (NAC1+DOX) showed a significant increase (P=0.001) when compared to NAC pre-treated group, and that of (NAC2+DOX) group showed insignificant change (P.>0.05)in comparison to (NAC1+DOX) as shown in Table 3&4) (Fig. 2)

e) Changes in plasma GSH levels The plasma level of GSH of the DOX- treated group showed a significant decrease (*P*. < 0.05) by 46.4% in comparison to the normal group (A). The plasma level of GSH of groups (NAC

pretreated, NAC1+DOX & NAC2+DOX) showed a significant increase (P. < 0.05) by 64%, 91.8%, and 93.32% when compared to group C (DOX treated). While the plasma levels of groups (NAC1+DOX & NAC2+DOX) showed a significant increase (P. < 0.05) by 17% and 17.8% respectively when compared to the (NAC pre-treated) group, but that of the group (NAC2+DOX) showed insignificant change (P.> 0.05) when compared to (NAC1+DOX) group **as shown in Table 3&4**) (Fig. 2).

f) Changes in erythrocyte lysate GSH-Pxlevels

The GSH-Px level in the erythrocyte lysate solution of the DOX-treated group showed a significant decrease (P. < 0.05) by 39.4% when compared to the negative group (A). The level of erythrocyte lysate GSH-Px of groups (NAC pretreated, NAC1+DOX) showed a significant increase (P. < 0.05)by 23.7% and 72.06% when compared to DOX-treated group while the the groups ervthrocvte lvsate levels of (NAC1+DOX & NAC2+DOX) showed insignificant change (P.> 0.05) when compared to the NAC pre-treated group. The plasma level of (NAC2+DOX) group showed insignificant change (P > 0.05) in comparison to (NAC1+DOX) group as

shown in Table 3&4) (Fig. 2)

2. Histopathological results (shown in figure 3)

Examination of cardiac sections of the negative control group showed normal architecture of cardiac muscles (Fig 3 A). Cardiac sections of NAC treated group showed normal findings (Fig. 3 B). Sections from cardiac muscle tissues in DOXtreated group showed histopathological changes was recognized by disrupted cardiac muscle fibers and foci of degeneration of the myocardium (Fig. 3 C). Sections of rat cardiac muscle tissue, which received (NAC pre-treated) & (NAC1+ DOX) group, showed small foci hemorrhage and mild disruption of cardiac muscles (Fig. 3 D&E). Sections from rat hearts of NAC2+DOX treated group (showed restoration of most of the normal cardiac tissue architecture and cardiac muscles showing no significant pathological changes (Fig. 3 F).

Table (1) The effects of i.p. DOX and oral administration of NAC on serum levels of cardiac markers LDH, CK-MB and CTn-I in the sixstudied groups.

Group	Α	B	С	D	Ε	F	p-valueby				
Variable		Median(IQR)									
LDH unit (U/L)	981(470- 2244.25)	1165(1089.25- 2040)	1677.5(1435- 1834)	1336.5(914- 1771.5)	1004(843-1811)	1535(1210.75- 1971.25)	0.004**				
CK-MB (U/L)	136(111.45- 229.5)	129(80.45-203)	261(123.88- 367.5)	163(28-178.5)	143 (105-234)	141(24.98-276)	0.042*				
CTn-I (ng/ml)	0.3(0.17-0.64)	0.37(0.21-0.46)	0.64(0.27-0.84)	0.53(0.44-0.79)	0.48(0.27-0.48)	0.52(0.37-0.99)	0.038*				

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Statistical analysis was carried out by Kruskal-Wallis test among all groups and shows statistically significant differences in cardiac enzymes betweengroups.

* Statistically significant difference (p. <0.05)

** Highly statistically significant difference (p. <0.01).

Table (2): The significance changes in cardiac markers assessment.

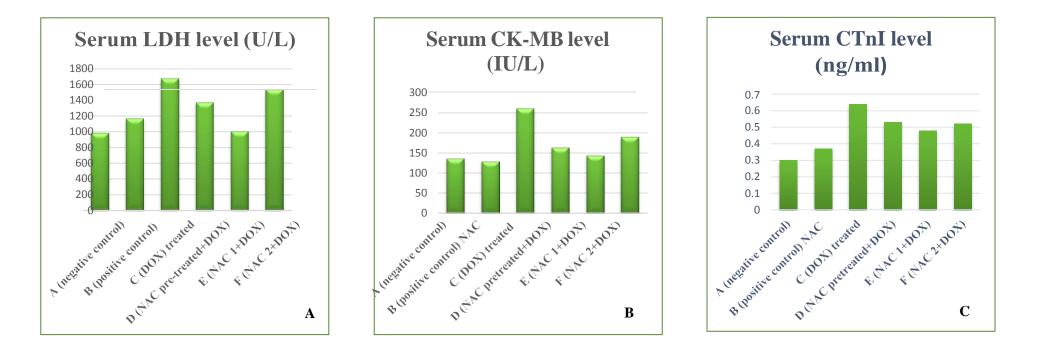
Group			P-va	alue							
	B VersusA	C VersusA	D VersusA	E VersusA	VersusA	D VersusC	E VersusC	F VersusC	E VersusD	F VersusD	F VersusE
LDH	0.387	0.040*	0.022*	0.005**	0.0.36*	0.035*	0.012*	0.004**	0.671	0.419	0.701
СК-МВ	0.283	0.045*	0.274	0.868	0.369	0.001**	0.011*	0.034*	0.353	0.804	0.494
CTn-I	0.400	0.007**	0.022*	0.552	0.018*	0.009**	0.003**	0.001**	0.452	0.499	0.654

Statistical analysis was done using Mann-Whitney U multiple comparison test.

* Statistically significant difference (*p*. <0.05)

** Highly statistically significant difference (*p*. <0.01).

DOX (Doxorubicin), **NAC** (N-Acetylcystiene), **i.p** (Intraperitoneal), **LDH** (Lactate dehydrogenase enzyme), **CK-MB** (creatine kinase isoenzyme in blood), **CTn-I** (cardiac troponin – I).



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Figure: (1) (A, B &C) Effect of oral administration of (NAC) on the serum level of (LDH (A), CK-MB (B), and CTn-I (C)) in doxorubicin-induced cardiac toxicity in rats. Each value represents the median ± IQR. Data were analyzed by Kruskal-Wallis test followed by MannWhitney U comparison test amongst different groups.

Group	A	B	C	D	E	F	p-valueby	
Variable	Median(IQR)							
MDA (nmol/ml)	80.88(56.75- 150)	87.5(51.44- 111.81)	273.75(111.25- 712.5)	61.13(25- 107.88)	92.5 (87.5-163)	180.25(32.5- 352.5)	0.012*	
GSH (mg/dl)	93.33(84.99- 103.32)	95.66(76.66- 100.99)	50(44-52.99)	82(79.99-88.66)	95.99(95.32- 97.32)	96.66(89.16- 101.82)	0.000**	
GSH-Px (Mu/L)	37.2(13.89- 26.87)	39.91(5.74- 29.16)	22.55(39.98- 54.44)	27.9(5.23-23.89)	38.8(12.84- 30.14)	41.7(6.76-65.9)	0.277	

Table (3) The effects of i.p. DOX and oral administration of NAC on serum levels of oxidative stress markers MDA (Malondialdehyde), GSH, and GSH-Px in six studied groups. Each value represents the median value and interquartile range.

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Statistical analysis was carried out by Kruskal-Wallis test among all groups and shows statistically significant differences regarding MDA&GSH betweengroups.

* Statistically significant difference (*p*. <0.05)

** Highly statistically significant difference (*p*. <0.01).

Table (4): The significance changes in oxidative stress parameters assessment.

Group	P-value										
Variable	B VersusA	C VersusA	D VersusA	E VersusA	F VersusA	D VersusC	E VersusC	F VersusC	E VersusD	F VersusD	F VersusE
MDA	0.401	0.004**	0.967	0.006**	0.600	0.001**	0.050	0.394	0.001**	0.571	0.094
GSH	0.569	0.000**	0.012*	0.416	0.866	0.000**	0.000**	0.000**	0.001**	0.007**	0.514
GSH-Px	0.312	0.001**	0.824	0.969	0.123	0.003**	0.001**	0.896	0.855	0.079	0.119

Statistical analysis was done using Mann-Whitney U multiple comparison test.

* Statistically significant difference (*p*. <0.05).

** Highly statistically significant difference (*p*. <0.01).

DOX (Doxorubicin), NAC (N-Acetylcystiene), i.p (Intraperitoneal), MDA (Malondialdehyde), GSH (Reduced glutathione), GSH-Px (Glutathione peroxidase enzyme).



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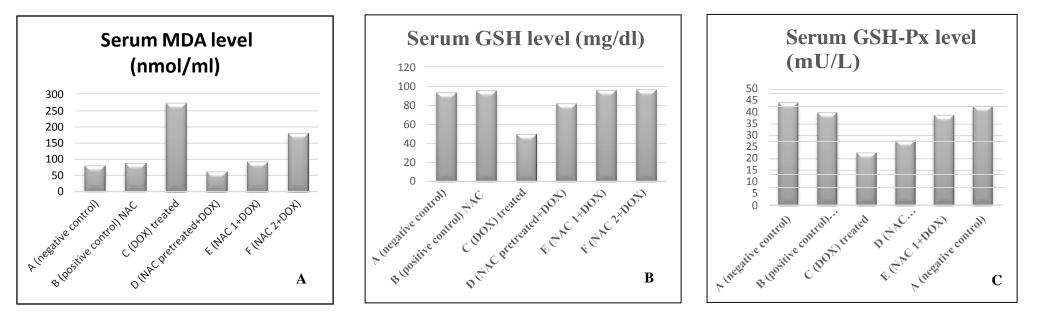


Figure: (2) (A, B &C) Effect of oral administration of (NAC) on the plasma level of (MDA (A), GSH (B) and RBCs lysate level of (GSH-PX) (C) in doxorubicin-induced oxidative stress in rats. Each value represents the median \pm IQR. Data were analyzed by Kruskal-Wallis testfollowed by Mann Whitney U comparison test amongst different groups.

А

С

D

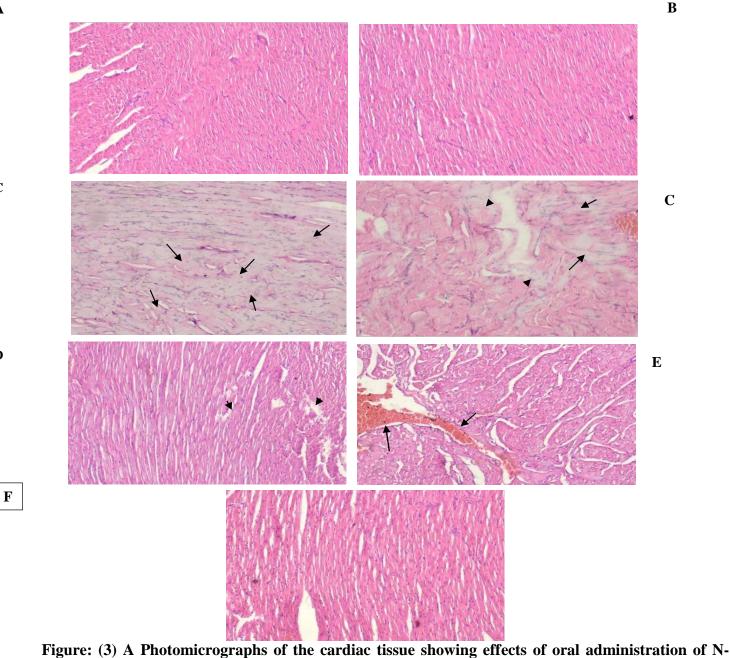


Figure: (3) A Photomicrographs of the cardiac tissue showing effects of oral administration of Nacetyl cysteine in doxorubicin- induced cardiotoxicity in rats. (A) cardiac tissue of the control negative group showing normal cardiac architecture (H&E x200). (B) cardiac tissue of NAC only treated group showing normal cardiac muscle architecture (H&E x200). (C) cardiac tissue of the DOX-treated group showing disrupted cardiac muscle fibers (left one-arrows) and disrupted cardiac muscle fibers & foci of degeneration of the myocardium (right one-arrows & arrow heads). Sections of rat heart from (D) which received (NAC pre-treated) &(E) group which received NAC1+ DOX, cardiac muscles showing mild disruption of cardiac muscles (arrow heads) and small foci hemorrhage (arrows). Sections from rat hearts of group (F) which received NAC2+DOX showed restoration of most of the normal cardiac muscle tissue architecture and cardiac muscles showing no significant pathological changes.

Discussion:

Doxorubicin (DOX) is an anthracycline antibiotic and one of the most powerful and widely used chemotherapeutic agents for different types of solid and hematological tumors. Unfortunately, clinical application of this drug leads to severe cardiotoxicity (*Yu et al., 2018*).

In the present study, the protective and ameliorative effects of N-acetylcysteine (NAC) against (DOX) induced cardiac toxicity was investigated in male albino rats. The rats were divided into six groups ten rats each.

The present study showed a significant increase in serum levels of LDH, CK-MB & CTn-I levels (p. < 0.05) by 70.9%, 91.1%, and113.3 respectively in DOX treated group when compared to negative control group.

The most severe side effect of DOX is dosedependent cardiotoxicity, which takes place through inducing oxidative stress apoptosis (*Abushouk et al., 2017*). Also, the cellular mechanisms underlying DOX-induced cardiotoxicity include free-radical damage to cardiac myocytes, leading to mitochondrial injury and subsequent death of myocytes (*Nonaka et al., 2017*).

Elevations in baseline high-sensitivity cardiac troponin T (hscTn-T) levels are suggestive of an oncology subgroup at high risk of developing cardiac complications from their chemotherapy. (*Blaes et al., 2015*).in agreement with the present results recently, *Olorundare et al. (2020)* results revealed that serum cardiac troponin I and LDH were significantly elevatedby the DOX treatment.

The results obtained in *Abd El-Gawad and El-Sawalhi*, (2003) study agree with that obtained in this current study when DOX was given in a dose of (2.5 mg/kg/BW) twice per week for two weeks, he found that cardiotoxicity was manifested by a

marked increase in serum LDH and CPK in addition to the sharp increase in MDA reaching eightfold the basal level.

In *Saad et al. (2004)* DOX was given ina dose of (5mg/kg) twice per week for two weeks, the induced cardiotoxicity was manifested by abnormal biochemical changes including marked increases in serum (CK-MB),(LDH), glutathione peroxidase (GSH-Px).

The current study results agree with that of *Alkreathy et al. (2010)* study, where a single dose of DOX (25mg/kg) caused increased both serum cardiac enzymes LDH and CPK activities and a significant increase in plasma level of MDA.

Similar findings were present in *Al- Harthi et al.*, (2014), the tested dose of DOX (20 mg/kg) caused a significant increase in the serum activities of the cardiac enzymes lactate dehydrogenase (LDH), creatine phosphokinase (CPK), and the level of malondialdehyde (MDA) in the heart tissue. In addition, there was a significant decrease in the glutathione level inthe heart tissue.

The current study results agree with those gained in *El- Sayed., et al (2016)* study where the tested dose of DOX was given (10 mg/kg) i.v. injection once in 16 days, it induced marked and significant increase in serum levels of LDH, CK-MB & CTn-I, these changes were attributed to DOX induced cardiac damage.

The present study results agree with that obtained from *Bai et al.*, (2019) Who investigated the cardio-toxic effects of DOX in a dose of (5 mg/kg) I.P injection once per week for four weeks and found significant increase in CTn-I level when compared to control.

DOX-induced cytotoxic effects on the heart could be clarified by free radicals created by DOX and interacts with the mitochondrial and endoplasmic reticulum membranes leading to their degeneration and consequent cell apoptosis (Zhan et al., 2015).

The current study showed that there was significant decrease in serum levels of LDH, CK-MB and CTn-I in NAC pre-treated group, (NAC1+DOX) & (NAC2+DOX) by (20.3%, 40.1, and 8.5%) (37.5%, 45.5, and 45.6) (17.2%, 25%, and 18.75%) respectively when compared to DOX-treated group and this indicates that

NAC therapy either pre-treatment or concurrently administrated with DOX can ameliorate and suppresses DOX-induced elevations of cardiac markers.

The serum levels of LDH, CK-MB and CTn-I in groups (NAC1+DOX & NAC2+DOX) showed insignificant change when compared to (NAC pretreated) group, we infer that no difference in effect between NAC concurrent treatment and pretreatment with NAC. The serum levels of LDH, CK-MB and CTn-I of (NAC2+DOX) group showed insignificant change when compared to group (NAC1+DOX), that means no difference noticeable between protective effect of NAC with different doses.

NAC prevented the negative inotropic effect produced by DOX on isolated rat atria. A good relationship exists between the cardio- protective effects of NAC and its ability to enhance the nonprotein sulfhydryl group content of the myocardium (*Villani et al., 1990*).

The results of the current study agree with *Pereira et al.*, (2019) who concluded that N-acetylcysteine play a protective role in cardiac surgery it was safely administered to patients in all included studies in this review. Cardiac markers LDH, CK-MB, and CTn-I levels were significantly improved in those patients with marked restoration of GSH cardiactissue stores

Reddy et al. (2011) stated that the use of

substances with antioxidant properties has been proven effective at protecting the cardiac muscle against damage caused by reactive oxygen species (ROS) generation. In this sense, several studies (*Arica et al., 2013; Al-Harthi et al., 2014; Abushouk et al., 2017*) have shown that NAC can reduce the effects caused by DOX They reported that NAC was able to restore the anti- and prooxidant balance of the cardiac muscle and prevent the harmful effects of doxorubicin treatment.

NAC largely reduced CK-MB release in diabetic rats. NAC alone reasonably decreased cardiac troponin I levels in diabetic rats (*Lin et al.*, *2016*).

The present study results revealed that sections from cardiac muscles of DOX-treated group showed evidence of toxic effects demonstrated as areas of disrupted cardiac muscle fibers with foci of degeneration. However, with NAC therapy, cardiac muscles of groups (NAC pretreated & NAC1+DOX) exhibit small foci hemorrhage and mild disruption of cardiac muscles. Whereas, Cardiac muscles of group (NAC2 + DOX) showed significant pathological changes. no The pathological changes occur after administration of DOX were in concordance with previous studies as in study by Swamy and his colleagues (2012). In their study DOX was given in dose of 2.5mg/kg/BW i.p three times per week for two weeks Results revealed that DOX-treated group showed loss of myofibrils and vacuolization of the cytoplasm were observed.

Shivakumar et al. (2012) results were in agreement with current study. Examination of sections from the heart in male rats treated with DOX (2 mg/kg/BW) once per week for four weeks showed inter-fibrillar hemorrhages, congestion, and focal areas of disrupted cardiacmuscle fibers. DOX produced necrosis, hemorrhages and leukocyte infiltration in the myocardium. The injury to the

myocardium can be explained by oxidative stress induced by the reactive intermediates doxorubicin semiquinone formed from doxorubicin. The anthracyclines are reported to form semiquinone radical intermediates, which react with molecular oxygen to form reactive oxygenspecies that interact with macromolecules of the cells to induce tissue damage.

The present study results agree with that obtained from (Farshid et al., 2014). Who investigated the protective effects of NAC against DOX -induced cardiotoxicity, NAC administered as 40mg/kg/BW I.P; doxorubicin was given 4mg/kg/BW for four weeks. Results showed reversal of serum elevated LDH, CPK and MDA induced by DOX. Also cardiac sections showed partial reduction in leucocyte infiltration and necrosis. Also, Arica et al. (2013) who studied the effect of NAC protection against DOX-induced cardiotoxicity. NAC was given in a dose of 200mg/kg/BW once every day for five days and DOX was given as 20 mg/kg/BW as a single dose once. Results showed that preservation of the general architecture of cardiac muscle tissue. However, edema, disorganization, and myofibril loss was minimal than DOX-treated group. This could be explained by the antioxidant activity of NAC and this is confirmed by modulation of cardiac enzyme back to normal in NAC treated groups in comparison to DOX-treated group. Moreover, NAC showed a protective role in the cardiac tissue of rats submitted to hemorrhagic shock, mainly in lessening oxidative stress and histologic injury (Oliveira Filho et al., 2015).

Lu et al. (2016) study that investigated Doxorubicin-induced myocardial damage (2.5 mg/kg/BW) i.p. injection once per week for six weeks Results revealed serious myocardial damage characterized by disorganization of myo-fibrillar arrays, interstitial fibrosis, massive cardiomyocyte loss, cytoplasmic vacuolization, eosinophilic degeneration and forceful infiltration of neutrophil granulocytes.

In agreement with these results, Mostafa and Raafat, (2019) found that in Wister adult male albino rats that received NAC 250mg/kg/BW by nasogastric tube every day. DOX (2.5mg/kg/BW) was given three times a week for two weeks. Cardiac stained sections revealed that the cardiac muscle fibers showed improvement in pathological findings after administration of NAC. There was improvementin vacuolations and fibrosis. NAC can attenuate the effects caused by DOX and able to restore the anti- and pro-oxidant balance of the cardiac muscle and prevent the injurious effects of DOX treatment. This may be explained by enriching glutathione (GSH), an important enzyme of the cellular antioxidant system that is able to stimulate and sustain its intracellular levels, which detoxify reactive oxygen species.

On the other hand, (*Shi et al., 2009*) who studied the ability of NACA to protect cardiomyocytes in tissue culture from DOXinduced toxicity. N-acetylcysteine amide (NACA), a structural analogue of NAC, was synthesized and evaluated in certain in vivo and in vitro models. They found that although NACA was able to provide oxidative relief; it only had minimal protective effect on DOX-induced cell death.

To summarize, our results indicate that NAC induces ameliorative and protective effects against DOX-induced cardiotoxicity as NAC restores the elevated cardiac enzymes in NAC treated groups (pre-treated and concurrent) in comparison to DOX-treated group. These results were confirmed by the histopathological findings as NAC was able to restore cardiac muscle architecture to near normal except some muscle disruption and few foci of hemorrhage in few areas in NAC pretreated and NAC1+DOX groups. Furthermore, NAC2+DOX group sections showed no pathological abnormalities. This can be explained by doubling the dose of NAC can restore histological changes to normal.

The present study showed a significant increase in serum level of MDA by 238.4% and a significant decrease in serum GSH & GSH-Px levels by 46.4% and 39.4% in DOX treated group when compared to control groups.

These results can be explained by DOX undergoes antioxidant redox cycling resulting in the production of ROS; DOX generates semiquinone radicals, which in turn react with molecular oxygen and provide other ROS at an early stage after administration (*Bulucu et al., 2010*).

The current study agrees with most studies that had recorded elevation of MDA in hearttissue after acute and chronic application of DOX (*Mokni et al.*, 2012; Ammar et al., 2013).

Frijhoff et al. (2015) stated that GSH is an intracellular non-enzymatic antioxidant and one of the most important scavengers of free radicals. In addition, it is a co-factor of many detoxifying enzymes against oxidative stress as GSH-Px and glutathione reductase (GR).

The current study agree with results obtained from *Kwatra et al. (2016)* study in which DOX was given in a dose of (15 mg/kg)

I.P injection at 10th day, results revealed that Doxorubicin-induced cardiotoxicity confirmed by significant increase in MDA level, significant decrease in GSH level.

The present study agrees with *Rashid et al.* (2013); *Kwatra et al.* (2016) and Song et al. (2019) studies that DOX administration significantly decreased the levels of antioxidant enzymes, namely, superoxide dismutase (SOD), GSH-Px,

GR, and catalase (CAT) in rats in response to free radical formation, supporting the serious role of oxidative stress in DOX hepatotoxicity.

Lipid peroxidation includes oxidation of lipids, yields an aldehyde (MDA). Administration of (DOX) was closely associated with lipid peroxidation which is attributed to increase in free radical formation, thereby, suggesting a state of oxidative injury as reported previously in the heart and liver of animals after Dox administration (*Rashid et al., 2013; Rehman et al., 2014; Kuzu et al., 2019*).

The current study results agree with *Kuzuet al.* 2019 who investigated the DOX induced effects on oxidative status by using DOX in a dose of (40mg/kg/BW) I.p. injection on the 8th day and the study continued for 10 days. The results showed a significant increase in MDA levels and a significant decrease in GSH and GSH-Px levels, this may be due to the high dose (40 mg/kg) indicating the highly toxic profile of DOX even if there is short duration (48 hours) interval between the dose and samples taken. Current results agreement with aforementioned studies proves DOX-induced toxicity against the normaloxidative status.

Lipid peroxidation is the hallmark of oxidative stress. It results in increase of fibrogenic

cytokines by stimulating the formation of collagen and liver stellate cells. DOX administration results in a substantial increase in the lipid peroxidation. Therefore, leading to marked increase of ROS as well as exhaustion of cellular antioxidant defense system markers including GSH, GSSG, CAT, SOD, GSH-Px, GR, and H2O2 levels (*Afsar et al., 2019*). The results of the presenting study agree with that obtained from *Walaa et al. (2019*) study, DOX was given in a dose of (4mg/kg) for four weeks, there was significant increase in MDA level and a massive decrease in the brain GSH & GSH-Px levels in the DOX-treated group compared to the control group. Current obtained results agree with *Wali et al.* (2020) in that there was a significant increase in MDA (a well-known biomarker of oxidative stress & lipid peroxidation) level, also a significant decrease in both GSH & GSH-Px activity when compared to control group.

Results of *Wali et al. (2020)* study who investigated the toxic effects of DOX on the liver. DOX was given in a dose of (20 mg/kg) in the 20th day of 21-day study), it showed that ROS were significantly higher in (Dox-treated group) when compared with the control group. Revealing that (DOX) increases oxidative stress by instigating ROS & RNS that play role in number of cellular mechanisms like redox cycling of quinine moiety of doxorubicin and disrupt iron homeostasis.

The present study showed insignificant change in plasma MDA levels in groups (NAC1+DOX &NAC2+DOX) when compared to (DOX treated) group. There is a significant decrease in plasma MDA level of (NAC pre- treated) group by 77.7% when compared to DOX treated, and this indicates that effect of NAC pre-treatment has a better result to decrease lipid peroxidation than concurrent treatment.

Plasma GSH levels in groups (NAC pretreated, NAC1+DOX & NAC2+DOX)) showed significant increase by 64%, 91.8%, and 93.32% respectively when compared to (DOX treated) group. Furthermore, a significant

increase in plasma GSH levels of groups (NAC1+DOX & NAC2+DOX) when compared to (NAC pretreated) group. This reflects effects of concurrent more than NAC pre-treatment, there is no statististically significant change in GSH level in group (NAC2+DOX) when compared to group (NAC1+DOX), so no value added by increasing the dose of NAC to ameliorate GSH level.

Erythrolysate level GSH-Px levels results showed significant increase in groups (NAC pretreated & NAC1+DOX) by 23.7% and 72.06% respectively when compared to DOX treated. While Erythrolysate GSH-Px levels showed insignificant change in groups (NAC1+DOX & NAC2+DOX) when compared to NAC pretreated, and insignificant change in group (NAC2+DOX) when compared to group (NAC1+DOX), so these results refers to the NAC effect to restore GSH-Px activity after DOX induced oxidative changes without significant restorative effects differences between NAC pre-treatment or concurrent treatment options.

The present study agrees with *Koçkar et al.* (2010) who investigated NAC modulation effect on DOX-induced oxidative stress in rat liver. DOX was given in a dose (2.5 mg/kg/BW)once and NAC (10 mg/kg/BW) every day for 10 days (experiment duration). Results showed serum GSH level significantly increased in the (DOX+NAC) group than in control and DOX group. GSH-Px levels showed insignificant changes in the three groups by DOX and NAC administrations. In conclusion, N-acetylcysteine induced modulator effects on the doxorubicin- induced hepatoxicity by inhibiting free radical production and supporting the antioxidant vitamin levels.

Sathish et al., (2011) showed that Posttreatment with NAC 50 mg/kg daily for 7 days effectively prevent the elevation in oxidative stress marker enzymes and LPO, and restoring the activities of both the enzymic (SOD, CAT and GPx) and non-enzymic (GSH, Vit.C and Vit.E) antioxidants and membrane bound ATPases towards normal.

Walaa et al. (2019) found that administration of NAC produced a statistically significant increase in GSH and GPx levels. The uniqueness of NAC is most probably due to its serving as a precursor of L-cysteine for GSH synthesis (Samuni et al., 2013).

The present study results agree with *Mansour* and *Hasan* (2015) study as NAC given in

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(200mg/kg) i.v for five days then a single dose of cyclophosphamide given after 1hour of last NAC dose. After one day samples were taken, the results showed that marked and significant decrease in MDA level, also significant increase in both GSH & GSH Px.

The normalization of MDA following NAC treatment is very likely due to its anti- peroxidative properties (*Zafarullah et al., 2003*), as the presence of acetyl and sulfhydryl groups makes NAC an effective inhibitor of lipid peroxidation (*Dhouib et al., 2015*).

Conclusion:

• DOX administration in cumulative dose 20mg/kg/BW induced cardiac toxicity at biochemical and histopathological levels.

• NAC treatment did not induce any observed toxic effects in any dose used in our study.

• NAC induces ameliorative and protective effects against DOX-induced cardiotoxicity as NAC restores the elevated cardiac enzymes in NAC treated groups (pre-treated and concurrent) in comparison to DOX- treated group. These results was confirmed by NAC induced effects in restoring cardiac muscle architecture to near normal except some muscled isruption and few foci of hemorrhage infew areas in NAC pre-treated and NAC1+DOX groups.

• Furthermore, NAC2+DOX group sections shows no pathological changes in cardiac tissues. This can be explained by adding more protective effect of doubling the dose of NAC.

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الملخص العربى

دور مادة إن أستيل سيستايين في الوقاية من سمية عقار دوكسرويسين على أنسجة القلب للفئران عماد الدين محمد الطاهر محمد محمود¹ رندة حسين عبد المهادى² صفاء ماهر جورج² عبير الرفاعى محمد الرفاعى³ رانيا أحمد رضوان⁴ أقسم الطب الشرعي و السموم الإكلينيكية – كلية الطب – جامعة أسوان – مصر ²قسم الطب الشرعي و السموم الإكلينيكية – كلية الطب – جامعة أسيوط – مصر ³ قسم الباثولوجيا – كلية الطب – جامعة أسيوط مصر ⁴ قسم الطب الشرعي و السموم الإكلينيكية – كلية الطب – جامعة مصر.

مقدمة البحث يعتبر دواء دوكسروبسين من أهم مضادات السرطان التي تستخدم في علاج العديد من الأورام ولكن الاستخدام طويل المدى له غالبا ما يكون محدودا نظرا لما يسببه من سمية القلب وكذلك العديد من أجهزة الجسم الأخرى. **الهدف من البحث** هو الاستدلال والتعرف على التأثير الوقائي المحتمل لعلاج إن أستيل سيستايين وذلك بجرعات مختلفة في الاعتلال والسمية المحدثة بعقار دوكسروبسين فى الفئران . **طريقة البحث** أجريت هذه الدراسة على عدد (60) فأرا من ذكور الفئران البيضاء البالغة مقسمة إلى (6) مجموعات (10) فئران في كل مجموعة. المجموعة الأولى: المجموعة الضابطة. المجموعة الثانية: تم إعطاؤها محلول إن أسيتيل سيستايين عن طريق أنبوب بالفم بجرعة (400) مجم لكل كجم يوم بعد يوم. المجموعة الثالثة: تم حقن الفئران بجرعة واحدة من دوكسروبيسين(5) مجم لكل كجم مرة واحدة أسبوعيا لمدة أربعة أسابيع.المجموعة الرابعة: تم البدء بإعطاء جرعات إن أستيل سيستايين بالفم بجرعة (400) مجم لكل كجم يوم بعد يوم لمدة أسبوع ثم من الأسبوع الثاني تم حقنها بعقار دوكسروبيسين داخل الغشاء البروتوني بجرعة (5) مجم لكل كجم من وزن الجسم أسبوعيا لمدة أربعة أسابيع بالتزامن مع إعطاء مادة إن أستيل سيستايين لنهاية مدة الدراسة. المجموعة الخامسة: تم حقنها بعقار دوكسروبيسين بجرعة (5) مجم لكل كجم من وزن الجسم بالتزامن مع جرعات إن أسينيل سيستايين بالفم بجرعة (400) مجم لكل كجم من وزن الجسم يوم بعد يوم لمدة أربعة أسابيع. المجموعة السادسة: تم حقنها بعقار دوكسروبيسين بجرعة(5) مجم لكل كجم من وزن الجسم أيام بالتزامن مع جرعات إن أسيتيل سيستايين بالفم بجرعة (800) مجم لكل كجم يوم بعد يوم لمدة أربعة أسابيع. النتائج تسبب عقار ان اسيتيل سيستايين في وجود فرق ذو دلالة إحصائية عند فحص أنسجة القلب ما بين عينات المجموعة الثالثة وباقي المجموعات في الوقاية من التغيرات المحدثة بواسطة عقار دوكسروبيسين المتمثل في نقص إنزيمات القلب ونقص في عوامل الضغوط الاكسيدية وهي خفض الدهون فائقة التأكسد مع زيادة ملحوظة في الجلوتاثيون وانزيم بيروكسيدات الجلوتاثيون وكذلك وجود فرق ليس ذو دلالة إحصائية بين المجموعة الرابعة والتي تمثل إعطاء العقار قبل عقار دوكسروبيسين بأسبوع والخامسة والسادسة التي تم إعطاء العقاران بالتزامن مع بعضهما في نفس الوقت. ووجود فرق ذو دلالة إحصائية لبعض العوامل في المجموعة السادسة عن الرابعة والخامسة. و قد أظهر التحليل الباثولوجي إستعادة معظم الشكل و التركيب الطبيعي لعضلة القلب مع عدم وجود أي دلائل باثولوجية بألياف عضلة القلب للمجموعة السادسة. **الخلاصة**: لم يتسبب عقار ان أسيتيل سيستابين بحدوث أثار سمية معينة في أنسجة القلب و لم يتبين أفضلية أي من الجرعتين عن الأخرى (400و 800مجم) التي تم إعطاءهم بالتزامن مع عقار دوكسروبيسين. **التوصيات** يمكن استخدام عقار ان اسيتيل سيستايين للوقاية من الأثار السمية الناجمة عن عقار دوكسروبيسين أثناء إستخدامه لعلاج بعض الأورام وتكون هذه الوقاية ناتجة عن خفض تكوين الدهون فائقة التأكسد واستعادة حالة منع التأكسد الانزيمية وغير الإنزيمية. يجب إجراء دراسات أخرى على استخدام عقار إن أسيتيل سيستايين في الوقاية من سمية عقار دوكسروبيسين وذلك بإستخدام جرعات مختلفة ولمدة زمنية مختلفة وقياس كفاءة هذه الوقاية و كذلك في الوقايلة من سمية أدوية علاج السرطانات الأخرى.