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GC/MS Volatile Constituents Analysis and Anticancer Activity of Moltkiopsis ciliata (Forssk.)

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# ABSTRACT

Methanolic extract of the aerial part of *Moltkiopsis ciliata* (Forssk.) (Boraginaceae) was obtained and successively fractionated into two main fractions (hexane fraction and methylene chloride fraction), which were phytochemically investigated using GC/MS technique. The whole of thirty-five compounds were identified and quantified. They are belonging to different classes; twenty-one acetogenins (Fat derivatives), six shikimates, five terpenes and one sterol. Beside the characterization of the volatile compounds profile,  $\beta$ -amyrin was isolated and identified using chromatographic and spectral techniques. The methanolic extract and two fractions were screened for their cytotoxic activity against breast cancer cell line MCF7 and hepatocellular carcinoma cell line HepG2. The methylene chloride fraction was proved to be the most potent cytotoxic agent. The major phyto-components of the two fractions were known by their various biological activity as antimicrobial, antiprofilative, antioxidant, Hypocholesterolemic, Antiandrogenic flavor, Nematicide, pesticides, anti-inflammatory, Hemolytic and have significant pharmaceutical importance especially as anticancer agents. The bioactive volatile constituents recommended phytopharmaceutical importance of the plant.

# Key Words:

Moltkiopsis ciliata, Boraginaceae, GC/MS technique, volatile constituents, cytotoxic activity

# 1. INTRODUCTION

*Moltkiopsis ciliata* (Forssk.) (Boraginaceae), is a perennial flowering shrub with a woody base, up to 25cm high, and has white beginning branches, the plant shielded with stiff white hairs. Flower varies in color significantly. Blue-violet, red, bluish limb with a red tube, white and pure yellow [1].

*M. ciliata* exposed carbohydrates, flavonoids, terpenes, sterols, nitrogenous bases, condensed tannins, and saponins [2]. Phenolics have attracted great consideration and get high importance due to their antioxidant activity. M. ciliata contains protocatechuic acid, caffeic acid, vanillic acid, p-coumaric acid, iso-ferulic acid, ellagic acid, salycilic acid, o-coumaric acid, e-vanillic acid, and p-OH-benzoic acid, apigenin 6-C-arabinoside-8-C-glactoside, apigenin 6-C-rhamnoside-8-C-glucoside, luteolin 6-C-glucoside, luteolin 6-C-arabinoside-8-C-glucoside, luteolin 6-C-arabinoside, luteolin 6-C-glucoside, luteolin 6-C-glucoside, kaempferol, acid, apigenin 7-O- $\beta$ -glucoside, kaempferol,

kaempferol 3,7-di-O-rhamnoside, quercetin, rutin, hesperidin, and naringenin were detected by using HPLC technique and compared with standards [3, 4].

In Veterinary medicine, *M. ciliata* used to improve lactation of camels [5]. Also, aqueous ethanolic extract of *M. ciliata* demonstrated moderate antibacterial activity against the Gram-negative *Moraxella catarrhalis* [2].

The Gas Chromatography-Mass Spectrum technique is applied for the identification and quantification of volatile constituents since this method provides high resolution, good sensitivity, as well as structural information about the analysis [6].

The main aim of the study was to extract and identify the bioactive volatile natural products of *Moltkiopsis ciliata* by using diverse solvents and then investigate their anticancer activity.

#### 2. EXPERIMENTAL

**General:** GC/MS analysis of the volatile moieties was carried out on a "Varian GC" connected to the selective mass detector (SMD); "Finnegan SSQ 7000 " with "ICIS V2.0" data module in order to identify the Mass spectra of the GC composites. The used column was DB-5 (J&W Scientific, Folosm, CA) interfaced with a fused silica capillary one (30 m long, 0.25 mm internal diameter) glazed the poly dimethyl-siloxane (0.5µm film thickness). The oven temperature was isothermally set at 50°C for 3 min, and after that was raised by 7°C /mi up to 250°C, then isothermally at the last temperature for 10 min. The temperature of the Injector was 200°C, whereas the injected volume was 0.5 µl. Ionization energy was adjusted at 70 eV. (Agriculture Research Center (NRC), Dokki, Cairo).

#### **3.1.**Solvents and chemicals

#### 3. MATERIALS AND METHODS

Hexane, methylene chloride, methanol petroleum ether (60–80°C), ethyl acetate, and anhydrous sodium sulphate were purchased from ADWIC Company. PTLC was performed on silica gel (Kieselgel 60, F 254) of 0.25mm thickness.

## **3.2.Plant material**

Moltkiopsis ciliata was collected from the north coastal road before Baltym by half Km. Egypt in April 2017 and was identified by prof. Dr. Ibrahim Mashaly, Department of Botany, Faculty of Science, Mansoura University.

#### **3.3.Extraction and Isolation**

The dried aerial part plant powder of *M. ciliata* (1.25 Kg) was extracted with methanol (10L x 5), then filtrated and evaporated using a rotary evaporator to its 1/3 volume. Exhaustive liquid–liquid extraction using Hexane (60-80°C) and methylene chloride was performed to yield hexane (16.8g), methylene chloride (2.81g) fractions. The hexane fraction was defatted using cold methanol, and then both fractions were subjected to GC/MS. The methylene chloride fraction was fractionated with the aid of column chromatography (silica gel, hexane: ethyl acetate, 70:30) to yield ten fractions. Fraction 3 (80 mg), was further purified on TLC (silica gel, petroleum ether: ethyl acetate, 8.5:1.5) to give compound 37 (25mg).

#### 3.4.Anti-proliferative activity against cancer cell lines

Cancer cell lines namely; breast cancer cell line MCF7 and hepatocellular carcinoma cell line HepG2 were obtained from" VACSERA", Egypt. The capability of cells in culture was calculated by the MTT assay [4]. As with all cells, the cell counts were adjusted to  $3 \times 103$  cells/well and plated in 100 µL of medium/well in 96-well plates. After incubation all overnight, various concentrations from different extracts were incubated with the cell line; 3 wells were involved in each concentration. After treatment for two days, 20 µL of 5 mg/ml MTT (pH 4.7) was added per well and cultivated for another 4 h, removing the supernatant fluid, and then 100 µL DMSO was added per well and shake well for 15 min. The absorbance measurements was reported at 570 nm with a microplate reader "Bio-Rad, Richmond, CA" using wells without cells as blanks. All experiments were performed in triplicate.

## 4. RESULTS AND DISCUSSION

The lipophilic volatile compounds of *Moltkiopsis ciliata* were extracted and obtained by the aid of two less polar organic solvents; hexane and methylene chloride.

GC/MS is the most befinicial technique designed for the identification and quantification of bioactive composites in plant extracts. GC/MS has been applied for the analysis of different medicinal herbs and was confirmed to be a valuable method for identifying different volatile oils, hydrocarbons, acids, alkaloids, esters, fatty acids, phytosterols and phenolic compounds [7, 8].

GC/MS analyses of hexane and methylene chloride fractions (Table 1 and 2) revealed the existence of important bioactive products, including acetogenins (fat derivatives), terpenes, sterols and shikimates derivatives (phenolic constituents) having many medicinal, pharmaceutical and antimicrobial properties.

The GC/MS characterization of hexane fraction has identified twenty components based upon a comparison of their EI-MS spectra with those of their analogous reported by the NIST database spectral library. Among the most prevailing matched constituents were oleic acid (22.49 %), (E)-9-octadecenoic acid methyl ester (14.76%), hexadecanoic acid methyl ester (11.47 %), (E,E)-9,12-octadecadienoic acid methyl ester (9.46 %), n-hexadecanoic acid (4.46%), Ar-tumerone (2.61 %), methyl stearate (2.15 %) which are used pharmaceutically as antimicrobial, antibacterial, anti-inflammatory, diuretic, antiasthma, antioxidant, inhibition of certain cancers, heart protection, antimalaria, dermatologic agent against acne, hypocholesterolemia, avoidance and treatment of diabetic retinopathy [7, 9, 10].

The methylene chloride fraction afforded fifteen phytochemicals and the more predominant constituents were eicosane (53.83%), n-hexadecanoic acid (12.54%) with antioxidant, hypocholesterolemic, antiandrogenic and 5-Alpha reductase inhibitor activities [7], squalene (2.56%) which has the property of antioxidant and anticancer activities [11], hexadecanoic acid methyl ester (2.27%), 6-hydroxy-4,4,7a-trimethyl-5,6,7,7a-tetrahydrobenzofuran-2(4H)-one (1.07%) beside, the phenolic constituents (3.58%) which are considered to be natural antioxidants due to their capacity to react with free radicals (scavenging) [12].

No	No. Compound name	RT	Area	Mol.	Mol.	m/z (%)		
110.	Compound name	(min)	%	formula	Wt.	III/Z (76)		
	Acetogenins (Fat Derivatives)							
1	E-15-Heptadecenal	34.84	0.14	C <sub>17</sub> H <sub>32</sub> O	252	$\begin{array}{c} 252\ (100\%)\ [M]^+,\ 83\ (100\%)\ [C_5H_7O]^+,\\ 55\ (83\%)\ [C_3H_3O]^+,\ 97\ (79\%)\ [C_6H_9O]^+,\\ 41\ (77\%)\ [C_3H_5]^+,\ 43\ (75\%)[C_3H_7]^+ \end{array}$		
2	Tetradecanoic acid methyl ester	37.82	0.43	C <sub>15</sub> H <sub>30</sub> O 2	242	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
3	Tetradecanoic acid	38.56	0.28	C <sub>14</sub> H <sub>28</sub> O 2	228	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
4	1-Octadecene	39.38	0.30	$C_{18}H_{36}$	252	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
5	6,10,14-trimethyl-2- pentadecanone	40.40	0.26	C <sub>18</sub> H <sub>36</sub> O	268	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
6	(Z)-9-Hexadecenoic acid methyl ester	41.68	0.23	C <sub>17</sub> H <sub>32</sub> O 2	268	$\begin{array}{c} 268\ (3\%)\ [M]^{+}\ ,\ 55\ (100\%)\ [C_{4}H_{7}]^{+}\ ,\\ 69\ (67\%)\ [C_{5}H_{9}]^{+}\ ,\ 41\ (62\%)\ [C_{3}H_{5}]^{+},\\ 43\ (59\%)\ [C_{3}H_{7}]^{+}\ ,\ 83\ (58\%)\ [C_{6}H_{11}]^{+} \end{array}$		
7	Hexadecanoic acid	42.10	11.47	C <sub>17</sub> H <sub>34</sub> O	270	270 (12%) $[M]^+$ , 74 (100%)		

Table 1. Identified volatile constituents of hexane extract of *M. ciliata* using GC/MS technique.

	methyl ester			2		$[C_{3}H_{6}O_{2}]^{+}$ , 87 (78%) $[C_{4}H_{7}O_{2}]^{+}$ , 43
				2		$(26\%)$ $[C_3H_7]^+$ , 143 (25%)
						$[C_{8}H_{15}O_{2}]^{+}, 227 (24\%) [C_{14}H_{27}O_{2}]^{+}$
8						256 (30%) [M] <sup>+</sup> , 73 (100%)
		10.07	1.16	C <sub>16</sub> H <sub>32</sub> O	050	$[C_{3}H_{5}O_{2}]^{+}$ , 60 (74%) $[C_{2}H_{4}O_{2}]^{+}$ , 43
	n-Hexadecanoic acid	42.87	4.46	2	256	$(68\%) [C_3H_7]^+, 55 (61\%) [C_4H_7]^+, 57$
						$(61\%) [C_4H_9]^+$
9						$280(3\%) [M]^+$ , 83 (100%) $[C_6H_{11}]^+$ ,
	(E)-5-Eicosene	43.49	0.60	$C_{20}H_{40}$	280	55 (99%) $[C_4H_7]^+$ , 43 (99%) $[C_3H_7]^+$ ,
						97 (88%) $[C_7H_{13}]^+$ , 57 (88%) $[C_4H_9]^+$
10	1 <i>5</i> 41					284 (13%) [M] <sup>+,</sup> 74 (100%)
	15-methyl-	11.06	0.14	C <sub>18</sub> H <sub>36</sub> O	284	$[C_{3}H_{6}O_{2}]^{+}, 87 (76\%) [C_{4}H_{7}O_{2}]^{+}, 43$
	hexadecanoic acid	44.06	0.14	2	284	$(50\%) [C_3H_7]^+, 55 (49\%) [C_4H_7]^+, 41$
	methyl ester					$(39\%) [C_3H_5]^+$
11	(E,E)-9,12-			СЦО		$294 (13\%) [M]^{+.}$ , 67 (100%) $[C_5H_7]^+$ ,
	octadecadienoic acid	45.49	9.46	$C_{19}H_{34}O$	294	81 (95%) $[C_6H_9]^+$ ,95 (72%) $[C_7H_{11}]^+$ ,
	methyl ester			2		55 (60%) $[C_4H_7]^+$ , 79 (48%) $[C_6H_7]^+$
12						296 (7%) $[M]^+$ , 55 (100%) $[C_4H_7]^+$ ,
	(E)-9-octadecenoic	45.63	14.76	C <sub>19</sub> H <sub>36</sub> O	296	69 (70%) $[C_5H_9]^+$ , 74 (68%)
	acid methyl ester	45.05	14.70	2	270	$[C_3H_6O_2]^+$ , 41 (66%) $[C_3H_5]^+$ , 83
						$(61\%) [C_6 H_{11}]^+$
13						298 (17%) $[M]^+$ , 74 (100%)
		46.12	2.15	$C_{19}H_{38}O$	298	$[C_3H_6O_2]^+$ , 87 (80%) $[C_4H_7O_2]^+$ , 43
	Methyl stearate	40.12	2.15	2	270	$(31\%)$ $[C_3H_7]^+$ , 143 (29%)
						$[C_8H_{15}O_2]^+, 55 (27\%) [C_4H_7]^+$
14				C <sub>18</sub> H <sub>34</sub> O		282 (3%) $[M]^+$ , 55 (100%) $[C_4H_7]^+$ ,
	Oleic acid	46.65	22.49		282	$69 (74\%) [C_5 H_9]^+, 41 (70\%) [C_3 H_5]^+,$
				2		67 (60%) $[C_5H_7]^+$ , 83 (59%) $[C_6H_{11}]^+$
15						280 (3%) $[M]^{+}$ , 55 (100%) $[C_4H_7]^+$ ,
	(E)-9-Eicosene	47.71	0.90	$C_{20}H_{40}$	280	69 (83%) $[C_5H_9]^+$ , 41 (80%) $[C_3H_5]^+$ ,
		.,.,1	0.70	0201140	200	83 (76%) $[C_6H_{11}]^+$ , 97 (76%)
						$[C_7H_{13}]^+$
16	trans-13-	_		C <sub>18</sub> H <sub>34</sub> O	_	282 (3%) $[M]^+$ , 55 (100%) $[C_4H_7]^+$ ,
	Octadecenoic acid	50.53	0.35	2	282	69 (65%) $[C_5H_9]^+$ , 57 (62%) $[C_4H_9]^+$ ,
				2		$(41 (56\%) [C_3H_5]^+, 43 (56\%) [C_3H_7]^+$
17						326 (20%) [M] <sup>+</sup> , 74 (100%)
	Eicosanoic acid	51.26	0.50	$C_{21}H_{42}O$	326	$[C_3H_6O_2]^+$ , 87 (82%) $[C_4H_7O_2]^+$ , 43
	methyl ester			2		(43%) [C <sub>3</sub> H <sub>7</sub> ] <sup>+</sup> ,55 (40%) [C <sub>4</sub> H <sub>7</sub> ] <sup>+</sup> , 41
						$(56\%) [C_3H_5]^+$
	[]		Se	esquiterpen	es	
						216 (26%) $[M]^{+}$ , 83 (100%)
18	Ar-tumerone	36.70	2.61	$C_{15}H_{20}O$	216	$[C_5H_7O]^+$ , 119 (77%) $[C_9H_{11}]^+$ , 201
						(24%) [C <sub>14</sub> H <sub>17</sub> O] <sup>+</sup> , 132 (22%)
						$[C_{10}H_{12}]^+$
						218 (3%) $[M]^+$ , 120 (100%)
19		37.61	0.90	$C_{15}H_{22}O$	218	$[C_9H_{12}]^+$ , 83 (28%) $[C_5H_7O]^+$ , 105
	Curlone					(17%) [C <sub>8</sub> H <sub>9</sub> ] <sup>+</sup> , 91 (15%) [C <sub>7</sub> H <sub>7</sub> ] <sup>+</sup> , 55
				D'4		$(11\%) [C_4H_7]^+$
00		<b>50</b> 10		Diterpenes		
20	4,8,12,16-	52.10	0.32	$C_{21}H_{40}O$	324	$324  (2\%)  [M]^+,  99  (100\%)$

tetramethylheptadec	2	$[C_5H_7O_2]^+$ , 55 (38%) $[C_4H_7]^+$ , 43
an-4-olide		$(38\%) [C_3H_7]^+, 69 (30\%) [C_5H_9]^+, 57$
		$(25\%) [C_4 H_9]^+$

# Table 2. Identified volatile constituents of CH<sub>2</sub>Cl<sub>2</sub> extract of *M. ciliata* using GC/MS technique.

No	Common damage	RT	Area	Mol.	Mol.			
No.	Compound name	(min)	%	formula	Wt.	m/z (%)		
	Shikimates							
	2-Methoxy-4-					$150(100\%) [M]^+$ , $135(82\%) [C_8H_7O_2]^+$ ,		
22	vinylphenol	16.34	0.25	$C_9H_{10}O_2$	150	107 (28%) $[C_7H_7O]^+$ , 77 (22%) $[C_6H_5]^+$ ,		
						$51 (10\%) [C_4H_3]^+$		
00		10.05	0.00		1.64	164 (100%) $[M]^+$ , 149 (30%)		
23	trans-Isoeugenol	19.85	0.26	$C_{10}H_{12}O_2$	164	$ \begin{bmatrix} C_9H_9O_2 \end{bmatrix}^+, \ 91 \ (22\%) \ \begin{bmatrix} C_6H_3O \end{bmatrix}^+, \ 77 \\ (19\%) \ \begin{bmatrix} C_6H_5 \end{bmatrix}^+, 131 \ (18\%) \ \begin{bmatrix} C_9H_7O \end{bmatrix}^+ $		
						$180 (100\%) [M]^+, 165 (38\%)$		
			0.44	<b>a w a</b>	100	$[C_9H_9O_3]^+$ , 137 (24%) $[C_8H_9O_2]^+$ , 122		
24	4-Vinyl-syringol	22.51	0.41	$C_{10}H_{12}O_3$	180	$(12\%)$ $[C_7H_6O_2]^+$ , 181 (11%)		
						$[C_{10}H_{13}O_3]^+$		
	Phenol, 2,6-					194 (100%) [M] <sup>+.</sup> , 131 (82%)		
25	dimethoxy-4-(2-	25.53	0.29	$C_{11}H_{14}O_3$	194	$[C_9H_7O]^+$ , 167 (34%) $[C_9H_{11}O_3]^+$ , 163		
	propenyl)-					$(33\%) [C_9H_7O_3]^+, 103 (31\%) [C_7H_3O]^+$		
	(E)-2,6-Dimethoxy-					104 (100%) DVII <sup>+</sup> 170 (16%)		
26	4-(prop-1-en-1-	25.62	0.88		194	$\begin{array}{cccccccccccccccccccccccccccccccccccc$		
20	yl)phenol	23.02	0.00	$C_{11}H_{14}O_3$	174	$(14\%) [C_9H_7O]^+$ , 119 (13%) $[C_9H_{11}]^+$		
	A ((1E) 2							
	4-((1E)-3- Hydroxy-1-					180 (72%) [M] <sup>+</sup> , 137 (100%)		
27	propyl)-2-	26.44	0.42	$C_{10}H_{12}O_3$	180	$[C_8H_9O_2]^+$ , 124 (47%) $[C_7H_8O_2]^+$ , 91		
	methoxyphenol					$(32\%) [C_6H_3O]^+, 119 (23\%) [C_8H_7O]^+$		
	6-Hydroxy-4,4,7a-					196 (10%) [M] <sup>+</sup> , 111 (100%)		
• •	trimethyl-5,6,7,7a-			~ ~ ~ ~	10.5	$[C_6H_7O_2]^+$ , 178 (61%) $[C_{11}H_{14}O_2]^+$ ,		
28	tetrahydrobenzofur	27.06	1.07	$C_{11}H_{16}O_3$	196	140 (43%) $[C_9H_{16}O]^+$ , 135 (35%)		
	an-2(4H)-one					$[C_8H_7O_2]^+$ , 181 (34%) $[C_{10}H_{13}O_3]^+$		
	I		Acetoge	enins (Fat D	erivativ	es)		
			3			$282~(7\%)~[M]^{+},~57~(100\%)~[C_{4}H_{9}]^{+}~,$		
29	Eicosane	3.13	53.8	$C_{20}H_{42}$	282	71 (82%) $[C_5H_{11}]^+$ , 85 (69%)		
27		5.15	3	€20 <b>1 1</b> 42	202	$[C_6H_{13}]^+$ , 97 (40%) $[C_7H_{13}]^+$ , 55		
						(37%) [C <sub>4</sub> H <sub>7</sub> ] <sup>+</sup> 270 (12%) [M1 <sup>+</sup> , 74 (100%)]		
	Hexadecanoic acid,					$ \begin{array}{cccccccccccccccccccccccccccccccccccc$		
30	methyl ester	30.30	2.27	$C_{17}H_{34}O_2$	270	$[C_{3}\Pi_{6}O_{2}]$ , $37$ (75%) $[C_{4}\Pi_{7}O_{2}]$ , 143 (24%) $[C_{10}H_{23}]^{+}$ , 227 (24%)		
						$[C_{14}H_{27}O_2]^+$ , 55 (20%) $[C_4H_7]^+$		
	n-Hexadecanoic					256 (33%) [M] <sup>+</sup> , 73 (100%)		
31	acid	31.15	12.5	$C_{16}H_{32}O_2$	256	$[C_3H_5O_2]^+$ , 60 (73%) $[C_2H_4O_2]^+$ , 129		
51	uiu	51.15	4	C10H132C2	250	(59%) $[C_7H_{13}O_2]^+$ , 57 (57%) $[C_4H_9]^+$ ,		
						55 (51%) $[C_4H_7]^+$		

32	10-Octadecenoic acid, methyl ester	33.57	0.32	C <sub>19</sub> H <sub>36</sub> O <sub>2</sub>	296	$\begin{array}{c} 296\ (10\%)\ [M]^{+}\ ,\ 55\ (100\%)\ [C_{4}H_{7}]^{+}\ ,\\ 69\ (78\%)\ [C_{5}H_{9}]^{+}\ ,\ 83\ (73\%)\ [C_{6}H_{11}]^{+}\\ ,\ 84\ \ (60\%)\ \ [C_{6}H_{12}]^{+}\ ,\ 97\ \ (55\%)\\ [C_{7}H_{13}]^{+} \end{array}$
33	Heptadecanoic acid, 16-methyl-, methyl ester	34.06	0.50	C <sub>19</sub> H <sub>38</sub> O <sub>2</sub>	298	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
			F	Phenolic terp	ene	
34	Alliodorin	37.76	0.35	$C_{16}H_{20}O_3$	260	$\begin{array}{cccccccccccccccccccccccccccccccccccc$
				Triterpene	s	
35	Squalene	44.85	2.56	C <sub>30</sub> H <sub>50</sub>	410	$\begin{array}{c} 410\ (3\%)\ [M]^+,\ 69\ (100\%)\ [C_5H_9]^+,\ 81\\ (60\%)\ [C_6H_9]^+,\ 149\ (19\%)\ [C_{11}H_{17}]^+,\\ 137\ (18\%)\ [C_{10}H_{17}]^+,\ 95\ (18\%)\\ [C_7H_{11}]^+ \end{array}$
	Steroids					
36	Cholesta-3,5-diene	46.07	0.97	C <sub>27</sub> H <sub>44</sub>	368	$\begin{array}{cccccccccccccccccccccccccccccccccccc$

The major phyto-components of *M. ciliata* and its various biological activities obtained through the GC/MS study of hexane and methylene chloride fractions are presented in **Table 3**. The represented data revealed that both fractions are rich with bioactive volatile constituents which recommended phytopharmaceutical importance of the plant.

Predominant component	<b>Biological activity</b>					
	Acetogenins (Fat Derivatives)					
	- Antioxidant, Nematicide, Pesticide, Hypocholesterolemic,					
Hexadecanoic acid, methyl	Antiandrogenic flavor, Hemolytic, 5-Alpha reductase inhibitor [13].					
ester	- Antimicrobial activity [14].					
	- Antifungal activity [15].					
	- Antibacterial, antitumor, immunostimulant, chemopreventive and					
	lipoxygenase inhibitor [16].					
	- Antioxidant, Hemolytic, Nematicide, Pesticide,					
	Hypocholesterolemic, Lubricant [17]					
	- Alpha reductase and antipsychotic inhibitor [18].					
	- larvicidal activity against mosquitoes [19].					
	- Antifungal, flavor, potent antimicrobial agent, antimalarial pesticide					
	and antipsychotic [20].					
	- Cytotoxicity against human colorectal carcinoma cells (HCT-116)					
n-Hexadecanoic acid	[21] .					
	- Antifouling property [22].					
	- Control of human pathogens, pests, termites and maggots [23].					
	- Increasing proliferation of MSCs [24].					

Γ	
	- Considered as larvicide [25].
	- The leaf ethanolic extract of Centella asiatica showed significant
	inhibitory activity against <i>Mycobacterium tuberculosis</i> due to presence
	of major bioactive n-Hexadecanoic [26].
	- Repellent against Anopheles species and thus useful for malaria
	control [27].
	- Antiandrogenic [28].
	- Echo enhancement in sonographic doppler B-mode imaging [29].
9,12-Octadecadienoic acid,	-anti-inflammatory, antibacterial, hypocholesterolemic and
methyl ester, (E,E)	hepatoprotective activities, 11-octadecenoic acid, methyl ester for
	having antioxidant and antimicrobial properties [30].
9-Octadecenoic acid,	-antimicrobial, anti-inflammatory, antioxidant
methyl ester, (E)-	-hypocholesterolemic, cancer preventive, hepatoprotective,
-	antiarthritic, antihistimic [31].
Methyl stearate	-anti-inflammatory, intestinal lipid metabolism regulator,
<b>J</b>	antinociceptive, nematicidal, antioxidant, antimicrobial activity [32].
	- Mediterranean diet (rich in OA) consumption [33].
	- it has useful effects on cardiovascular disease, serum lipids and
Oleic Acid	special protecting effects against cancer [34].
olek Aciu	- <i>in vivo</i> antioxidative [35].
Eicosane	-it mimics the effect of indomethacin, reduces the secretion of
Elcosane	histamine, bradykinin, TXs, PGs, and LTs, inhibits the stimulation of
	cytokines such as IL-1b, TNF, IL-6, and interferon- $\alpha$ , interrelated with
	the cox enzyme [36].
<b>A AAAA</b>	Sesquiterpenes
Ar-tumerone	-antioxidant, anti-inflammatory, anticancer, antimicrobial,
	neuroprotective, cardioprotective and radioprotective effects [37].
	Shikimates
6-Hydroxy-4,4,7a-	-cytotoxic against MCF-7, antimicrobial, antiprofilative, anti-acetyl
trimethyl-5,6,7,7a-	cholinesterase activity, anti-inflammatory [38].
tetrahydrobenzofuran-	
2(4H)-one	
	Triterpenes
Squalene	critical for reducing free radical oxidative damage to the skin. Serum
	squalene originates partly from endogenous cholesterol synthesis and
	partly from dietary sources,
	especially in populations consuming large amounts of olive oil or
	shark liver [2]. The endogenous
	synthesis of squalene begins with the production of 3-hydroxy-3-
	methylglutaryl coenzyme A (HMG
	CoA). The initial reduction of HMG CoA (a niacin-dependent
	reaction) results in the formation of
	mevalonate [4].
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	methylglutaryl coenzyme A (HMG
	CoA). The initial reduction of HMG CoA (a niacin-dependent
	reaction) results in the formation of
	mevalonate [4].
	-critical for reducing free radical oxidative damage to the skin,
	antitumor properties [39].
	- emollient, skin hydration, antioxidant, it is used in cosmetic
	dermatology [40].
L	

Repeated chromatographic analysis of methylene chloride fraction using column chromatography and PTLC (silica gel, petroleum ether: ethyl acetate, 17:3) yielded compound (37,  $R_f = 0.33$ ). <sup>1</sup>H NMR and MS spectra proved that compound **37** is  $\beta$ -amyrin which was in agreement with those previously those established from *Aster yomena* [41].

# 4.1. Cytotoxic activity

The herbal medicines exert their multiple therapeutic and anticancer properties through inhibition of cancer activating enzymes and hormones, stimulation of DNA repair mechanism, promotion of protective enzymes production, induction of the antioxidant system, and enhancing the immune system [42].

The methanolic extract of *M. ciliata* as well as both fractions; hexane and  $CH_2Cl_2$  were screened for cytotoxic activity against HepG2 and MCF-7 cell lines using MTT-based cytotoxicity Inspect. After 48 h incubation period of the extracts, it was found that methylene chloride showed potent cytotoxicity against all cancer cell lines (HepG2, and MCF7) followed by the methanolic extract and the least one hexane fraction. The viability of cancer cell lines was considerably affected. The cell layer partially condensed, forming cell-free areas, and finally separated from the culture plate [43, 44].

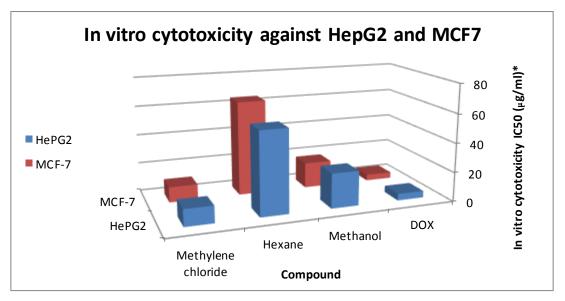


Fig (1): Cytotoxic activity against human cell lines of Moltkiopsis ciliata extracts

## 5. CONCLUSION

The study revealed that *Moltkiopsis ciliata* lipophilic extract contains various volatile constituents belonging to different classes, GC/MS analysis of both hexane and methylene chloride fractions has identified thirty-five compounds, the most predominant constituents have significant pharmaceutical importance, utilized as anticancer agents.

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## REFERENCES

- T. A. AL-Turki, and J. Thomas, "An account on the floral dimorphism and ecology of the genus Moltkiopsis I.M.Johnst. (Boraginaceae) in Saudi Arabia," *Turkish Journal of Botany*, vol. 34, no. 5, Jan. 2010, doi :10.3906/bot-0907-98.
- [2] A. M. Aldoweriej, K. B. Alharbi, E. M. A. Saeed and I. M. El-Ashmawy, "Antimicrobial activity of various extracts from some plants native to Alqassim Region, Saudi Arabia," *Journal of Food, Agriculture & Environment*, vol. 14, no. 1, Jan. 2016, doi: https://doi.org/10.1234/4.2016.1408.
- [3] S. A. Kawashty, H. M. Soliman, and N. A. Boquellah, "Chemical and biological characterize of some species from Mahdadh Dhahab region," *Journal of Innovations in Applied Pharmaceutical Sciences*, vol. 1, no. 3, sept. 2016.
- [4] J. M. Finefield, D. H. Sherman, M. Kreitman, and R. M. Williams, "Enantiomeric Natural Products: Occurrence and Biogenesis," *Angewandte Chemie International Edition*, a. vol. 51, no. 20, May. 2012, doi: 10.1002/anie.201107204.
- [5] I. F. Palici, E. Liktor-Busa, I. Zupkó, B. Touzard, and M. Chaieb, Edit Urbán6 and Judit Hohmann, " study of in vitro antimicrobial and antiproliferative activities of selected saharan plants, " *Acta*
- *Biologica Hungarica*, vol. 66, no. 4, Dec. 2015, doi: https://doi.org/10.1556/018.66.2015.4.3.
- [6] H. McNair and J. Miller, "Basic Gas Chromatography. 2rd ed. Hoboken," John Wiley & Sons, 2009.
- [7] B. Parthipan, M. G. T. Suky, and V. R. Mohan, "GC/MS Analysis of Phytocomponents in Pleiospermium alatum (Wall. ex Wight & Arn.) Swingle, (Rutaceae)," *Journal of Pharmacognosy and Phytochemistry*; vol. 4, no. 1, Feb. 2015, doi: https://doi.org/10.25004/IJPSDR.2016.080603.
- [8] R. Srivastava, M. Alok, and V. Amita, "GC/MS Analysis of phytocomponents in, pet ether fraction of Wrightia tinctoria seed," *Pharmacognosy Journal*, vol: 7, no. 4, May. 2015, doi:10.5530/pj.2015.4.7.

- [9] R. Prabakaran, B. Joseph, N. Pradeep, "Phyto medicinal compounds from Urginea indica Kunth: A synthetic drugs potential alternative," *British Journal of Pharmaceutical Research*, vol. 11, no. 5, Jan. 2016, doi: 10.9734/BJPR/2016/25216.
- [10] G. Ambrin, A. Mohammad, A. A. Abdulaziz, H. Abeer, F. A. Elsayed, and A. Altaf, "Conversion of Cytochrome P450 2D6 of human into a FRET-based tool for real-time monitoring of ajmalicine in Living cells," *Frontiers in Bioengineering and Biotechnology*, vol. 27, no. 7, Nov.2019, doi: 10.3389/fbioe.2019.00375.
- [11] P. T. Purushoth, P. Panneerselvam, R. Suresh, A. W. Clement, and S. Balasubramanian," GC-MS analysis of ethanolic extract of Canthium parviflorum Lamk leaf," *Journal of Applied Pharmaceutical Science*, vol. 3, no. 2, Feb. 2013, doi: 10.7324/JAPS.2013.30229.
- [12] N. Salah, N. J. Miller, G. Paganga, L. Tijburg, G. P. Bolwell, and C. Riceevans, "The antioxidant properties of theaflavins and their gallate esters radical scavengers or metal chelators?," *Archives* of *Biochemistry and Biophysics*, vol. 392, no. 1, Aug. 1995, https://doi.org/10.1016/0014-5793(96)00780-6.
- T. Sudha, S. Chidambarampillai, and V. Mohan, "GC/MS analysis of bioactive components of aerial parts of Fluggea leucopyrus Willd. (Euphorbiaceae)," *Journal of Applied Pharmaceutical Science*, vol. 3, no. 5, Nov. 2013, https://doi.org/10.1016/0014-5793(96)00780-6.
- [14] A. Akpuaka, M. Ekwenchi, D. Dashak, and A. Dildar," Biological activities of characterized isolates of n-hexane extract of Azadirachta indica A. Juss (Neem) leaves, "*Nature and Science*, vol. 11, no. 5, May. 2013, ISSN: 1545-0740.
- [15] D. Sharma, R. Rani, M. Chaturvedi, and J. P. Yadav, "Anibacterial efficacy and gas chromatographymass spectrometery analysis of bioactive compounds present in different extracts of Allium sativum," *Asian Journal of Pharmaceutical and Clinical Research*, vol. 11, no. 4, April. 2018, doi:10.22159/ajpcr.2018.v11i4.24053.
- [16] I. F. Begum, Mohankumar, R.; Jeevan, M. and Ramani K., "GC–MS analysis of bio-active molecules derived from Paracoccus pantotrophus FMR19 and the antimicrobial activity against bacterial pathogens and MDROs," *Indian journal of microbiology*, vol. 56, no. 4, Jul. 2016, doi: 10.1007/s12088-016-0609-1.
- [17] H. Takahashi, T. Sasaki, and M. Ito, "New flavonoids isolated from infected sugarbeet roots," *Bulletin of the Chemical Society of Japan*, vol. 60, no. 6, Jun. 1987, https://doi.org/10.1177/1934578X1501000675.
- [18] E. D. Vijisaral, and S. Arumugam, "GC-MS analysis of bioactive constituents of Indigofera suffruticosa leaves," *Journal of Chemical and Pharmaceutical Research*, vol. 6, no. 8, Dec. 2014, doi:10.20473/jkr.v1i2.3086.
- [19] A. Jeyasankar, and T. Chinnamani, "Larvicidal and pupicidal activities of Solonum pseudocapsicum fruits compunds against Aedes aegypti, Anopheles stephensi and Culex quinquefasciatus (Diptera: Culicidae)," *Journal of Infectious Diseases and Medical Microbiology*, vol. 2, no. 2, May. 2018.
- [20] D. Mathivanan, P. R. Gandhi, R. R. Mary, and S. Suseem, "Larvicidal and acaricidal efficacy of different solvent extracts of Andrographis echioides against blood-sucking parasites," *Physiological and Molecular Plant Pathology*, Vol. 101, no. 7, Mar. 2018, doi:10.1016/j.pmpp.2017.03.008.
- [21] L. Ravi, and K. Krishnan, "Cytotoxic potential of N-hexadecanoic acid extracted from Kigelia pinnata leaves," *Asian Journal of Cell Biology*, vol.12, no. 1, Feb. 2017, doi:10.3923/ajcb.2017.20.27.
- [22] A. Manilal, S. Sujith, B. Sabarathnam, G. Kiran, J. Selvin, C. Shakir, and A. Lipton, "Antifouling potentials of seaweeds collected from the southwest coast of India," *World Journal of Agricultural Sciences*, vol. 6, no. 3, Mar. 2010, doi: 10.3390/md15090265.
- [23] A. Manilal, S. Sujith, B. Sabarathnam, G. Kiran, J. Selvin, C. Shakir, and A. Lipton, "Biological activity of the red alga Laurencia brandenii," *Acta Botanica Croatica*, Vol. 70, no. 1, Apr. 2011.
- [24] D. F. Chen, X. Li, Z. Xu, X. Liu, S. H. Du, H. Li, J. H. Zhou, H. P. Zeng, and Z. C. Hua, "Hexadecanoic acid from Buzhong yiqi decoction induced proliferation of bone marrow mesenchymal stem cells," *Journal of medicinal food*, vol. 13, no. 4, Jan. 2010, doi: 10.1089/jmf.2009.1293.

- [25] B. R. Naik, G. S. Gowreeswari, Y. Singh, R. Satyavathi, S. Daravath, and P. R. Reddy, "Bio-synthesis of silver nanoparticles from leaf extract of Pongamia pinnata as an effective larvicide on dengue vector Aedes albopictus (Skuse) (Diptera: Culicidae)," *Advances in Entomology*, vol. 2, no. 2, Apr.2014, doi: 10.4236/ae.2014.22016.
- [26] M. Suresh, P. K. Rath, A. Panneerselvam, D. Dhanasekaran, and N. Thajuddin, "Anti-mycobacterial effect of leaf extract of Centella asiatica (Mackinlayaceae)," *Research Journal of Pharmacy and Technology*, vol. 3, no. 3, Jul. 2010, doi:10.1080/13880200490514177.
- [27] M. Ogunlesi, W. Okiei, E. Ofor, and A. E. Osibote," Analysis of the essential oil from the dried leaves of Euphorbia hirta Linn (Euphorbiaceae), a potential medication for asthma," *African Journal of Biotechnology*, vol. 8, no. 24, Dec. 2009, doi:10.4314/AJB.V8I24.68792.
- [28] S. Kumar, P. Samydurai, R. Ramakrishnan, and N. Nagarajan, "Gas chromatography and mass spectrometry analysis of bioactive constituents of Adiantum capillus-veneris L," *International Journal* of *Pharmacy and Pharmaceutical Sciences;* vol. 6, no. 4, Jan. 2014, doi: 10.32474/DDIPIJ.2021.03.000174.
- [29] P. Kushwaha, S. S. Yadav, V. Singh, and L. Dwivedi, "Phytochemical screening and GC-MS studies of the methanolic extract of Tridax procumbens," *International Journal of Pharmaceutical Sciences and Research*, vol. 10, no. 5, May. 2019, doi:10.13040/IJPSR.0975-8232.10(5).2492-96.
- [30] M. M. Rahman, S. H. Ahmad, M. T. M. Mohamed, M. Z. Ab Rahman, "Antimicrobial compounds from leaf extracts of Jatropha curcas, Psidium guajava, and Andrographis paniculata," *Scientific World Journal*, vol. 2014, no. 1, Aug. 2014, doi: 10.1155/2014/635240.
- [31] S. Sahreen, M. R. Khan, and R. A. Khan, "Evaluation of antioxidant activities of various solvent fruits," extracts of Carissa opaca Food Chem, vol. 6, no. 3, Mar. 2020, doi: 10.1016/j.heliyon.2020.e03637.
- [32] M. Adnan, Ch. M. N. Uddin, A. T. M. M. Kamal, M. O. K. Azad, A. Paul, S. B. Uddin, J. W. Barlow, M. O. Faruque, C. H. Park, D. H. Cho, "Investigation of the Biological Activities and Characterization of Bioactive Constituents of Ophiorrhiza rugosa var. prostrata (D.Don) & Mondal Leaves through In Vivo, In Vitro, and In SilicoApproaches, "*Molecules*, vol. 24, no.7, Mar. 2019, doi: 10.3390/molecules24071367.
- [33] C. Carrillo, M. D. Cavia, S. R.Alonso-Torre, "Antitumor effect of oleic acid; mechanisms of action; a review," *Nutricion Hospitalaria*, vol. 27, no. 6, Nov-Dec. 2016, doi: https://doi.org/10.1371/journal.pone.0157195.
- [34] A. Keys, A. Menotti, M. J. Karvonen, C. Aravanis, H. Blackburn, R. Buzina, et al., "The diet and 15year death rate in the seven countries study," *American Journal of Epidemiology*, vol. 124, no. 6, Dec. 1986, doi: 10.1093/oxfordjournals.aje.a114480.
- [35] Y. Wan, H. X. Li, G. M. Fu, X. Y. Chen, F. Chen, M. Y. Xie," The relationship of antioxidant components and antioxidant activity of sesame seed oil," *Journal of the Science of Food and Agriculture*, vol. 11, no. 6, May. 2015, doi: 10.1371/journal.pone.0157195.
- [36] P. N. Okechukwu, "Evaluation of anti-inflammatory, analgesic, antipyretic effect of eicosane, pentadecane, octacosane, and heneicosane," *Asian journal of pharamatheutical and clinical research*, vol. 13, no. 4, Apr. 2020, https://doi.org/10.22159/ajpcr.2020.v13i4.36196.
- [37] A. Amalraj, A. Pius, S. Gopi, and S. Gopi, "Biological activities of curcuminoids, other biomolecules from turmeric and their derivatives–A review," *Journal of traditional and complementary medicine*, vol. 7, no. 2, Jun. 2016, doi: 10.1016/j.jtcme.2016.05.005.
- [38] M. Dawra, Y. El Rayess, M. El Beyrouthy, N. Nehme, "Biological activities and chemical characterization of the Lebanese endemic plant Origanum ehrenbergii Boiss," *Flavour and Fragrance Journal*, vol. 36, no. 6, Dec. 2020, doi:10.1002/ffj.3646.
- [39] Z. Huang, Y. Lin, and J. Fang, "Biological and Pharmacological Activities of Squalene and Related Compounds: Potential Uses in Cosmetic Dermatology," *Natural library of medicine*, vol. 14, no. 1, Jan. 2009, doi: 10.3390/molecules14010540.

- [40] J. Huang, W. Zhou, W. Dong, A. M. Watson, Y. Hong, "Directed efficient, and versatile modifications of the Drosophila genome by genomic engineering," *Proceedings of the National Academy of Sciences* of the United States of America, vol. 106, no. 20, May. 2009, https://doi.org/10.1073/pnas.0900641106.
- [41] J. Qinglong, J. Hong-Guang, K. A. Ryun, and W. A. Eun-Rhan, "New Megastigmane Palmitate and a New Oleanane Triterpenoid from *Aster yomena* Makino," Helvetica Chimica Acta, vol. 95, no.8, Aug. 2012, https://doi.org/10.1002/hlca.201200035.
- [42] S. Shukla, and A. Mehta, "Anticancer potential of medicinal plants and their phytochemicals," *The Brazilian Journal of Botany*, vol. 38, no. 2, Jun. 2015, https://doi.org/10.1007/s40415-015-0135-0.
- [43] H. Liu, J. P. Dilger, and J. Lin, "Lidocaine Suppresses Viability and Migration of Human Breast Cancer Cells: TRPM7 as a Target for Some Breast Cancer Cell Lines," *Cancers*, vol. 13, May 2021, https://doi.org/10.3390/cancers13020234.
- [44] M. Ali, R. Iqbal, M. Safdar, "Antioxidant and antibacterial activities of Artemisia absinthium and Citrus paradisi extracts repress viability of aggressive liver cancer cell line," Molecular Biology Reports, vol. 48, Jan. 2022, https://doi.org/10.1007/s11033-021-06777-0.