



Pathological Aspects of Rabbits Hepatotoxicity and Fibrosis Experimentally Induced by Thioacetamide and The Role of Silymarin as Anti fibrotic Effects



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THIS study was designed to estimate the pathological aspects of hepatotoxicity and fibrosis of the liver using Thioacetamide (TAA) and the hepatoprotective effects of Silymarin in rabbits via biochemical tests and histopathological changes. A total of 24 male rabbits were used, randomly separated into 4 groups: Group I worked as a negative control, group II treated with Silymarin only by oral administration, group III was given IP injection TAA only, and finally Group IV was given TAA followed by Silymarin through oral route. Results revealed that rabbits treated with TAA had indicated a significant rise in total serum bilirubin, alkaline phosphatase, aspartate transaminase, and alanine transaminase levels. Histopathologically, liver sections showed severe centrilobular, periportal, hepatocellular vacuolation and necrosis respectively. Cholangitis with severe collagen fibres deposition was detected in the portal area accompanied by biliary duct epithelium hyperplasia and ductal fibrosis. The treatment with Silymarin followed by TAA exposure leads to enhancement of both biological parameters and pathological features of the liver in Group IV. Finally, the present study concludes that Silymarin has a hepatoprotective and anti-fibrotic activity mechanism.

Keywords: Histopathology, Liver, Fibrosis, TAA, Silymarin.

Introduction

Hepatotoxicity is one developing case because of the high use of compounds and environmental contact with xenobiotic substances [1,2]. The liver has multifunctions in metabolism, detoxification, secretion, and storage [3]. Liver fibrosis develops as a result of a large aggregate of scar tissues, which occurs when the liver starts to restore and replace damaged cells [4, 5]. It is a changeable reaction to any acute or chronic hepatocellular damage that reflects a balance between hepatic repair and fibrosis development [6]. Many disorders can lead to fibrosis, especially when the damage continues repeatedly or persistently, sometimes occurring after just one injury. When severe inflammation-induced as a result of acute hepatitis, the liver usually repairs itself by regeneration of new hepatocytes

and attaching them to a network of connective tissue [7]. When a repetitive liver injury occurs, necrotic hepatocytes will be substituted by newly regenerated cells. If the liver injury is chronic, a failure of replacement and regeneration of hepatocytes with rich extracellular matrix and collagen fibres will be seen [8, 9].

Thioacetamide (TAA) is well recognized as a hepatotoxic substance [10]. It is an organo-sulfur compound, which is frequently doing as a fungicide for controlling the decay of the fruit, also, in the leather industry, it works as an organic solvent [11]. So, exposure to TAA has a toxic effect through skin contact and/or inhalation [12, 13]. TAA is used as a model to induce both hepatotoxicities and to prompt fibrosis in different lab animals. TAA induces hepatotoxicity through oxidative stress as a

consequence of inflammatory reactions, though TAA is metabolized by cytochrome P450 to thioacetamide-sulfoxide (TASO) [14]. There are slightly experimental studies of the rabbit as a model for TAA-induced liver injury, acute hepatotoxicity and chronic hepatitis and fibrosis.

Silymarin a milk thistle component (*SilybumMarianum*) has antioxidant (free radicals scavenging), hepato-protective, hepatocyte regeneration, anti-neoplastic, anti-inflammatory, membrane stabilizing, antifibrotic, and immune moderating characteristics [15]. There is a slightly experimental investigation of TAA induce liver injury in the rabbit as a model study of acute hepatotoxicity and fibrosis.

Material and Methods

Chemicals

TAA was purchased from Sigma (AVONCHEM, UK). The dose of TAA was calculated from a pilot study. A Silymarin dose was obtained from previous studies [16, 17].

Animals

Our research was permitted by the Ethical Committee of the Mosul University of the College of Veterinary medicine.

Experimental protocol

A total of 24 male rabbits were separated into main (4) groups (6 each), the groups are: Control group (group I), and Group II, rabbits were given Silymarinorally dissolved in saline200mg/kg B.W. daily for 5 weeks. In group III, rabbits were orally treated with TAA (I/P) at a dose of 150 mg/kg B.W. twice weekly for 8 weeks. Group

IV, animals receiving TAA for 8 weeks followed by Silymarinorally dissolved in saline200mg/kg B.W. daily for 5 weeks. Toward the end of this investigation, samples of blood are taken for serum tests and liver samples are stored in a formalin solution of 10% neutral buffer for histopathological examination [18].

Assessment of hepatotoxicity

Hepatotoxicity was measured by calculating the total serum proteins (TSP), total serum bilirubin (TSB), alkaline phosphatase (ALP), and alanine aminotransferase (ALT), and aspartate aminotransferase (AST) levels in serum. Blood samples were collected from all groups, and collected serum was evaluated by spectrophotometer via a diagnostic kit (BIOLABO /French). The numbers were investigated via the statistical program SPSS, and the mean and standard error were calculated using the ANOVA test (one-way analysis of variance), the significant difference for all tests was at the level of significance of $P \leq 0.05$. Liver samples were kept in a neutral buffered formalin (10%) for the tissue slide preparation and histopathological examination.

Results

The results of the biochemical tests are summarized in (Table 1). Levels of liver enzymes in group III were significantly elevated, and there was an increase in the level of (TSB, ALP, AST, and ALT). In the TAA-treated rabbits, in contrast, to control and other groups ($p < 0.05$), while in group IV, which represent the treatment with Silymarin after TAA-intra peritoneal injection in

TABLE 1. Liver function test results in the studied groups

GROUPS	TSP	TSB	ALP/ALK	AST/GOT	ALT/GPT
CONTROL	7.34 ± 0.47 ^a	0.69 ± 0.16 ^a	34.48 ± 4.49 ^a	16.58 ± 1.63 ^a	8.88 ± 0.87 ^a
SILYMARIN	7.48 ± 0.53 ^a	0.73 ± 0.29 ^a	30.14 ± 0.57 ^a	18.52 ± 1.47 ^a	6.48 ± 1.63 ^{ab}
TAA-	6.6 ± 0.27 ^a	2.02 ± 0.35 ^b	49.36 ± 3.17 ^b	25.92 ± 1.21 ^b	11.38 ± 1.07 ^{ab}
TAA-	7.02 ± 0.48 ^a	1.2 ± 0.22 ^{ab}	39.86 ± 4.95 ^{ab}	19.38 ± 0.99 ^a	9.06 ± 0.58 ^a
SILYMARIN					

Data expression as Mean ± Standard error SE

The difference letters mean there are significant differences between groups at $p \leq 0.05$

rabbits revealed significantly ($p < 0.05$) decreased these enzymes related to TAA group ($p < 0.05$).

The histopathological assessment of liver sections demonstrates many pathological changes as a result of treatment with each TAA and in combination together (groups III and IV) respectively. Data revealed no pathological changes not only in the control group but also in group II (Silymarin). The deviation in the histology of liver sections in the TAA-treated group a severe centrilobular and periportal hepatocellular vacuolation and necrosis were

seen, and dilatation and congestion of central veins are noticed as well. Hepatic cells in the affected lobule were less eosinophilia. Cholangitis is with severe collagen fibres deposition in the portal area with biliary duct epithelium hyperplasia, in addition to the priductular fibrosis and mononuclear inflammatory cells, infiltrate the portal triad. The portal fibrosis extends towards the Neighboring portal areas and inside the hepatic lobules. Liver sections from group IV displayed minor changes in the histopathological picture in comparison to TAA alone. This group displayed minor portal fibrosis with moderate

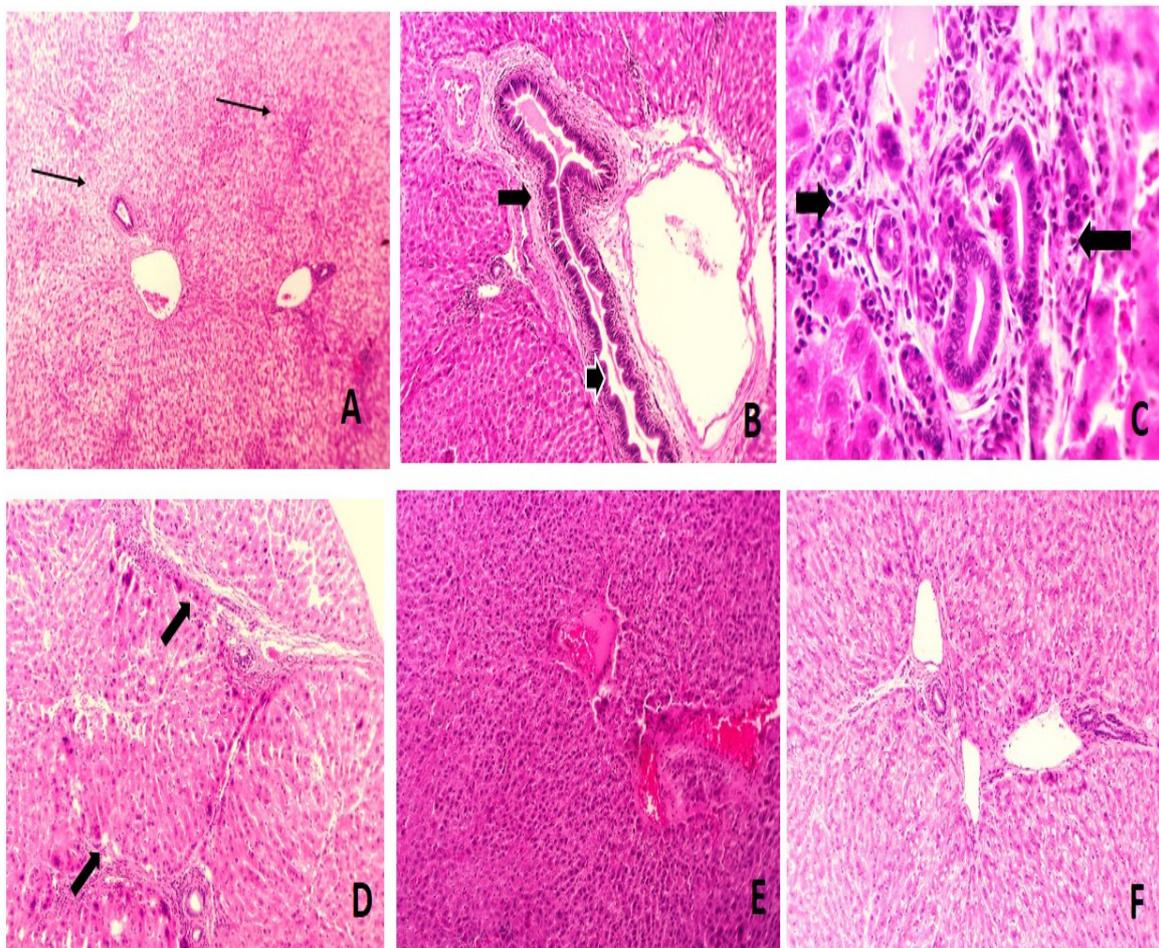


Fig.1. Liver of Male rabbits treated with TAA - silymarin. (A) dilation of central veins with severe centrilobular and periportal hepatocellular vacuolation and necrosis (Arrow) (H & E stain, 10x). (B) Severe collagen fibres deposition in the portal area (Arrow) with biliary duct epithelium hyperplasia with priductular fibrosis (Arrow head) (H & E stain, 10x). (C) mononuclear inflammatory cells infiltration in the portal triad (Arrow) (H & E stain 40x). (D) portal fibrosis extends towards the neighbouring portal areas and inside the hepatic lobules (Arrow) (H & E stain 10 x). (E & F) group of TAA followed by Silymarin displayed mild portal fibrosis with inflammatory cell accumulation in the portal area. Hepatocytes show less vacuolation and necrosis (H & E stain 10 x).

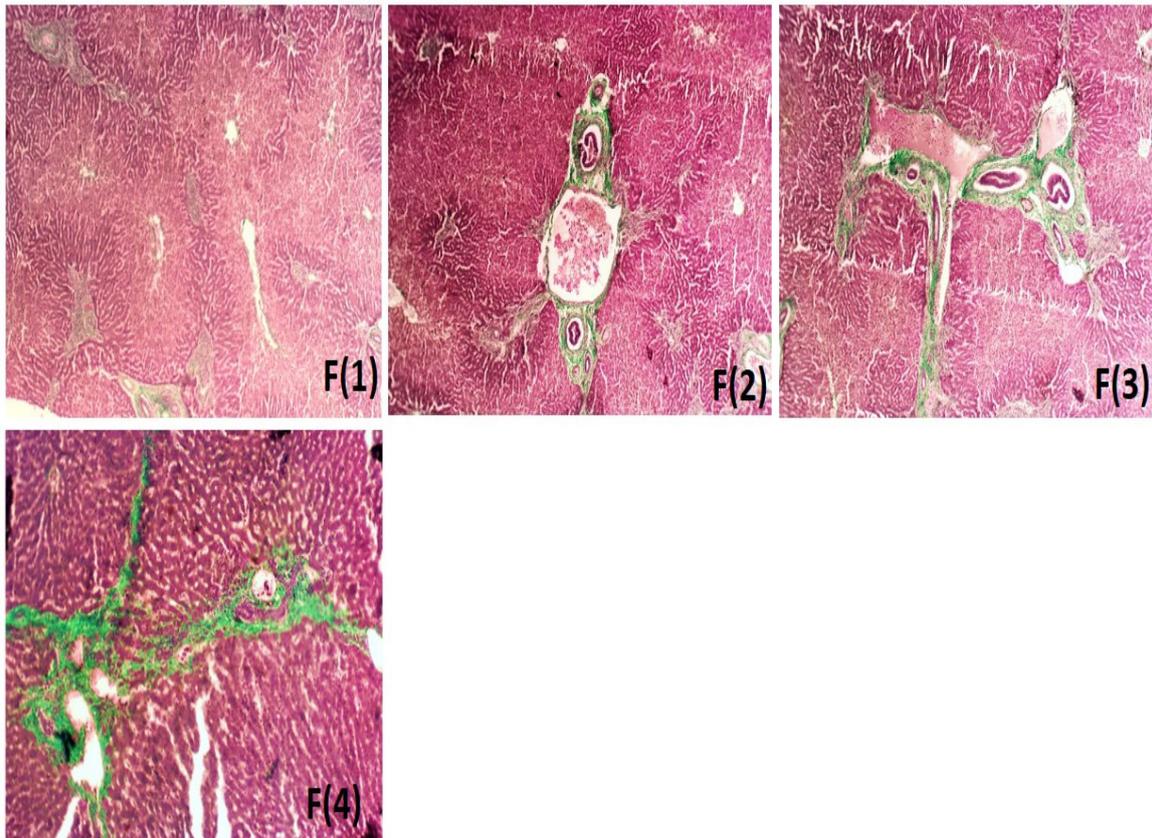


Fig. 2. Liver of Male rabbits (group IV) treated with TAA-silymarin. Histopathological staging of liver fibrosis. F(1) Fibrosis development to some portal triad with small fibrous septa. F(2) Fibrosis spreading out of portal areas with moderate fibrous septa. F (3) Fibrosis enlargement in portal areas with a marked portal to portal connecting as well as a portal to central fibrous septa. F(4) Fibrosis spreading out of portal triad with an obvious portal to portal connecting besides portal to central fibrous septa. Liver of Male rabbits (group III) treated with TAA (Masson's trichrome stain 10 x).

inflammatory cell accumulation in the portal area. Liver cells illustrate less vacuolation and necrosis. (Fig. 1 and 2).

Discussion

The liver is the essential glandular vital organ in the body and has numerous roles in regulating biological processes [19]. This study focused on the follow-up of TAA to induce hepatotoxicity and fibrosis, whereas the pilot study was used to determine the chronic dose in rabbits. TAA is used as a model to cause hepatotoxicity and induced fibrosis [20]. The objective of our experiment was to identify the properties of Silymarin against fibrosis in TAA-model and hepatic damage in rabbits. Results of this study declared that Silymarin caused a significant improvement in the numbers of liver enzymes and histopathological pictures of liver section

and fibrosis recovery. Additionally, the results presented a significant increase in the numbers of TSB, ALP, ALT and AST enzymes in the serum of rabbits treated with TAA compared to the control group and this confirm by many types of research [21].

Various researchers indicated that chronic use of TAA causes severe injuries in the liver, ranging from severe necrosis to cirrhosis. TAA is metabolized by cytochrome P450 to thioacetamide-sulfoxide (TASO) [22] which is an effective free radical that binds to macromolecules in hepatocytes and triggers changes in the permeability of cell wall and Ca^{+2} influx. This disturbance of Ca^{+2} levels in the cell leads to inhibits the activity of mitochondria, leading to hepatocytes vacuolation and necrosis [23, 24].

The generation of ROS is well-known to have a significant role in hepatotoxicity [25, 26]. Cytochrome P450 is the most source in the induction of oxidative stress in the liver, as well as the role of TAA metabolites in reactive oxygen species (ROS) formation at a large level, leading to impairment of the antioxidant defence mechanism and cellular components, leading to peroxidation of lipid and DNA damage [27, 28]. Liver fibrosis is highlighted by the increased deposition of both collagen fibres and extracellular matrix. Triggered hepatic stellate cells (HSCs) cells, fibroblasts from portal triad, and myofibroblasts of bone marrow origin have been recognized as the main collagen-producing cells in the damaged liver [29]. These cells are activated by cytokines and growth factors like TGF- β 1, and angiotensin II [30, 31].

The hepatoprotective characteristics of Silymarin are suggested to its anti-inflammatory, anti-oxidative, anti-fibrotic, cellular regeneration and immunomodulatory properties [32]. Several studies tested the role of silymarin against liver inflammation and fibrosis in many animal models. The anti-inflammatory effects of Silymarin are via inhibition of neutrophils migration to the site of inflammation and thus decreased the inflammatory mediators and cytokines as well as elimination of the radicals' species and preventing lipid peroxidation [33,34]. The anti-fibrotic effects of Silymarin are commonly due to its capability to prevent the activation and transformation of HSCs into myofibroblasts [35,36], which is responsible for collagen deposition by inhibition of fibrogenic pathways of the cytoskeletal formation, pro-fibrogenic collagen, and through depressed-regulates TGF- β 1 mRNA, which stops NF- κ B activation and prevents the stimulation of HSCs [37,38].

Conclusions

This study concludes that Silymarin is a hepatoprotective and antifibrotic medical plant in curing hepatotoxicity and liver fibrosis induced by TAA through the biochemical examination and histopathology of liver sections.

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Conflict of Interest

The investigator announces there is no conflict of interest exists.

References

1. Stephen Robert, J. M., Peddha, M. S. and Srivastava, A. K. Effect of Silymarin and Quercetin in a Miniaturized Scaffold in Wistar Rats against Non-alcoholic Fatty Liver Disease. *ACS Omega*, **6**(32),20735–20745(2021).doi.org/10.1021/acsomega.1c00555.
2. Younossi, Z., Tacke, F., Arrese, M., Chander Sharma, B., Mostafa, I., Bugianesi, E., Wai-Sun Wong, V., Yilmaz, Y., George, J., Fan, J. and Vos, M. B. Global Perspectives on Nonalcoholic Fatty Liver Disease and Nonalcoholic Steatohepatitis. *Hepatology (Baltimore, Md.)*, **69**(6), 2672–2682(2019). doi.org/10.1002/hep.30251.
3. Acharya, P., Chouhan, K., Weiskirchen, S. and Weiskirchen, R. Cellular Mechanisms of Liver Fibrosis. *Frontiers in Pharmacology*, **12**, 671640(2021).doi.org/10.3389/fphar.2021.671640.
4. Nallagangula, K. S., Nagaraj, S. K., Venkataswamy, L. and Chandrappa, M. Liver fibrosis: a compilation on the biomarkers status and their significance during disease progression. *Future Science OA.*, **4**(1), FSO250 (2017). doi.org/10.4155/fsoa-2017-0083.
5. Roehlen, N., Crouchet, E. and Baumert, T. F. Liver Fibrosis: Mechanistic Concepts and Therapeutic Perspectives. *Cells*, **9**(4), 875(2020). doi.org/10.3390/cells9040875.
6. Mohammed, S. A. and Abu-Deif, E.E. Animal Model for hepatic fibrosis and Cirrhosis. *Cytol. Histol. Int. J.*, **2**(1),1-4(2018).doi.org/10.23880/chij-16000105.
7. Bao, Y. L., Wang, L., Pan, H. T., Zhang, T. R., Chen, Y. H., Xu, S. J., Mao, X. L. and Li, S. W. Animal and Organoid Models of Liver Fibrosis. *Frontiers in Physiology*, **12**, 666138(2021). doi.org/10.3389/fphys.2021.666138.
8. Delire, B., Stärkel, P. and Leclercq, I. Animal Models for Fibrotic Liver Diseases: What We Have, What We Need, and What Is under Development. *Journal of Clinical and Translational Hepatology*, **3**(1),53–66(2015). doi.org/10.14218/JCTH.2014.00035.

9. Nevzorova, Y. A., Boyer-Diaz, Z., Cubero, F. J. and Gracia-Sancho, J. Animal models for liver disease - A practical approach for translational research. *Journal of Hepatology*, **73**(2), 423–440(2020). doi.org/10.1016/j.jhep.2020.04.011.
10. Akhta, T. and Sheikh, N. An overview of thioacetamide-induced hepatotoxicity. *Toxin. Rev.*, **32**(3), 43–46(2013).
11. Zargar, S., Wani, T. A., Alamro, A. A. and Ganaie, M. A. Amelioration of thioacetamide-induced liver toxicity in Wistar rats by rutin. *International Journal of Immunopathology and Pharmacology*, **30**(3), 207–214(2017). doi.org/10.1177/0394632017714175.
12. Koblihová, E., Mrázová, I., Vernerová, Z. and Ryska, M. Acute Liver Failure Induced by Thioacetamide: Selection of Optimal Dosage in Wistar and Lewis Rats. *Physiol. Res.*, **63**(4), 491–503(2014). doi.org/10.33549/physiolres.932690.
13. Schyman, P., Printz, R. L., Estes, S. K., O'Brien, T. P., Shiota, M. and Wallqvist, A. Assessing Chemical-Induced Liver Injury *In Vivo* From *In Vitro* Gene Expression Data in the Rat: The Case of Thioacetamide Toxicity. *Frontiers in Genetics*, **10**, 1233(2019). doi.org/10.3389/fgene.2019.01233.
14. Gillessen, A. and Schmidt, H. H. Silymarin as Supportive Treatment in Liver Diseases: A Narrative Review. *Advances in Therapy*, **37**(4), 1279–1301(2020).
15. Hellerbr, C., Schattenberg, J. M., Peterburs, P.H., Lechner, A. and Brignoli, R. The potential of silymarin for the treatment of hepatic disorders. *Clinical Phytoscience*, **2** (1), 1–14(2017). doi.org/10.1186/s40816-016-0019-2.
16. Nada, S., Gowif, A. M., El-Denshary, E. S., Salama, A., Khalil, M. and Ahmed, K. Protective Effect of Grape Seed Extract and/or Silymarin Against Thioacetamide-induced Hepatic Fibrosis in Rats. *J. Liver*, **4**(2), 1–7(2015). doi.org/10.4172/2167-0889.1000178.
17. Tighe, S.P., Akhtar, D., Iqbal, U. and Ahmed, A. Chronic Liver Disease and Silymarin: A Biochemical and Clinical Review. *J. Clin. Transl. Hepatol.*, **8**(4), 454–458(2020).
18. Luna, L.G. Manual of histologic staining methods of the armed forces institute of pathology. 3rd ed., New York: McGraw-Hill, pp.28–38 (1968).
19. Ghanim, A., Younis, N. S. and Metwaly, H. A. Vanillin augments liver regeneration effectively in Thioacetamide induced liver fibrosis rat model. *Life Sciences*, **286**, 120036(2021). doi.org/10.1016/j.lfs.2021.120036
20. Jafar, S. N. and Mawlood, K. A. The Protective Effects Of Extract Of Punica Granatum L Peel And Piper Longum Fruit Against Hepatotoxicity Induced By Thioacetamide In Male Albino Rats. *Annals of Tropical Medicine and Public Health*, **23**(7), 1–10(2020). doi.org/10.36295/ASRO.2020.23739.
21. Emam, M. A., Farouk, S. M. and Abdo, M. The Ameliorative Potential of Probiotics and/or Silymarin on Thioacetamide Induced Hepatotoxicity in Rats: Histological and Immunohistochemical Study. *International Journal of Morphology*, **36**(2), 661–669(2018). doi.org/10.4067/s0717-95022018000200661.
22. Đurašević, S., Pejić, S., Grigorov, I., Nikolić, G., Mitić-Ćulafić, D., Dragičević, M. and Đorđević, J. Effects of C60 Fullerene on Thioacetamide-Induced Rat Liver Toxicity and Gut Microbiome Changes. *Antioxidants (Basel)*, **10**(6), 911(2021). https://doi.org/10.3390/antiox10060911.
23. Robb-Gaspers, L.D. and Thomas, A.P. Coordination of Ca²⁺ signalling by intercellular propagation of Ca²⁺ waves in the intact liver. *The Journal of Biological Chemistry*, **270**(14), 8102–8107(1995). doi.org/10.1074/jbc.270.14.8102.
24. Yahya, S., Shalaby, R., Mannaa, F., Abdel-Wahhab, K., Mohamed, N., Shabana, M. and Elwakeel, S. Hepatoprotective Effects of Chitosan on Thioacetamide-Induced Liver Toxicity in Male Albino Rats. *Biointerface Research in Applied Chemistry*, **11**(6), 14490–14505(2021). doi.org/10.33263/BRIAC116.1449014505.
25. Schyman, P., Printz, R. L., Estes, S. K., Boyd, K. L., Shiota, M. and Wallqvist, A. Identification of the Toxicity Pathways Associated With Thioacetamide-Induced Injuries in Rat Liver and Kidney. *Front Pharmacol.*, **9**, 1272(2018) doi.org/10.3389/fphar.2018.01272.
26. Pellicano, A., Ramachandran, P., Iredale, J.P. and Fallowfield, J.A. Liver fibrosis and repair: immune regulation of wound healing in a solid organ. *Nat. Rev. Immunol.*, **14**(3), 181–194(2014). doi.org/10.1038/nri3623.

27. Ghosh, Sh., Sarkar, A., Bhattacharyya, S. and Sil, P. C. Silymarin Protects Mouse Liver and Kidney from Thioacetamide Induced Toxicity by Scavenging Reactive Oxygen Species and Activating PI3K-Akt Pathway. *Front Pharmacol.*, **7**, 481 (2016). doi.org/10.3389/fphar.2016.00481.
28. Amirtharaj, G. J., Natarajan, S. K., Pulimood, A., Balasubramanian, K. A., Venkatraman, A. and Ramachandran, A. Role of oxygen free radicals, nitric oxide and mitochondria in mediating cardiac alterations during liver cirrhosis induced by thioacetamide. *Cardiovasc. Toxicol.*, **17**(2), 175-184 (2016). doi.org/10.1007/s12012-016-9371-1
29. Hempel, F., Roderfeld, M., Savai, R., Sydykov, A., Irungbam, K. and Schermuly, R. Depletion of Bone Marrow-Derived Fibrocytes Attenuates TAA-Induced Liver Fibrosis in Mice. *Cells*, **8** (10), 1210 (2019). doi.org/10.3390/cells8101210.
30. AL-zuwainy, S. J. Improved Anti-Inflammatory Effect of Silymarin in Rats Induced Liver Carcinogenesis. *JUBPAS.*, **26**(4), 348-355 (2018).
31. Brenner, D. A. Molecular Pathogenesis of Liver Fibrosis. *Trans. Am. Clin. Climatol. Assoc.*, **120**, 361-368 (2009). PMID: 19768189.
32. Camini, F.C. and Costa, D.C. Silymarin: not just another antioxidant. *J. Basic Clin. Physiol. Pharmacol.*, **31**(4), j/bcphp (2020). doi.org/10.1515/jbcpp-2019-0206.
33. Anthony, K.P. and Saleh, M.A. Free radical scavenging and antioxidant activities of silymarin components. *Antioxidants*, **2**(4), 398-407 (2013). doi.org/10.3390/antiox2040398.
34. Wu, J.P., Tsai, C.C., Yeh, Y.L., Lin, Y.M., Lin, C.C., Day, C.H., Shen, C.Y., Padma, V.V., Pan, L.F. and Huang, C.Y. Retracted: Silymarin Accelerates Liver Regeneration after Partial Hepatectomy. *Evid Based Complement Alternat. Med.*, **2575273** (2017). doi.org/10.1155/2015/603529.
35. Kalopitas, G., Antza, C., Doundoulakis, I., Siargkas, A., Kouroumalis, E., Germanidis, G., Samara, M. and Chourdakis, M. Impact of Silymarin in individuals with nonalcoholic fatty liver disease: A systematic review and meta-analysis. *Nutrition*, **83**, 111092 (2021). doi.org/10.1016/j.nut.2020.111092.
36. MacDonald-Ramos, K., Michán, L., Martínez-Ibarra, A. and Cerbón, M. Silymarin is an ally against insulin resistance: A review. *Ann. Hepatol.*, **23**, 100255 (2021). doi.org/10.1016/j.aohep.2020.08.072.
37. Di Sario, A., Bendia, E., Taffetani, S., Omenetti, A., Candelaresi, C., Marzioni, M., De Minicis, S. and Benedetti, A. Hepatoprotective and antifibrotic effect of a new silybin-phosphatidylcholine-Vitamin E complex in rats. *Dig. Liver Dis.*, **37**(11), 869-76 (2005). doi.org/10.1016/j.dld.2005.05.011.
38. Jia, J. D., Bauer, M., Cho, J. J., Ruehl, M., Milani, S., Boigk, G., Riecken, E. O. and Schuppan, D. Antifibrotic effect of silymarin in rat secondary biliary fibrosis is mediated by downregulation of procollagen alpha1(I) and TIMP-1. *Journal of Hepatology*, **35**(3), 392-398 (2001). doi.org/10.1016/s0168-8278(01)00148-9.

الجوانب المرضية لتسمم وتليف الكبد في الارانب المحدث تجريبياً عن طريق الثايواسيتامايد ودور السليمارين كمضاد للتليف

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صممت هذه الدراسة لتقييم الجوانب المرضية للسمية الكبدية والتليف الكبدية باستخدام مادة الثايواسيتامايد والتأثيرات الوقائية للسليمارين في الأرانب عن طريق استخدام الاختبارات الكيموحياتية ودراسة التغيرات المرضية النسجية. تم استخدام ٢٤ من ذكور الأرانب قسمت بشكل عشوائي إلى ٤ مجموعات: المجموعة الأولى مجموعة سيطرة ، المجموعة الثانية عوملت بالسليمارين فقط عن طريق التجريع بالفم ، المجموعة الثالثة عوملت بمادة الثايواسيتامايد عن طريق الحقن داخل الخلب والمجموعة الرابعة عوملت بمادة الثايواسيتامايد والسليمارين. أظهرت النتائج أن الأرانب التي عوملت بالثايواسيتامايد أظهرت ارتفاعاً معنوياً في مستويات كل من البيلوروبين الكلي وخميرة الفوسفاتيز القلوي (ALP) وخميرة ناقلة أمين الأسبارتات (AST) وخميرة اقلية أمين الألانين (ALT) ، أما نتائج الفحص المجهرى ، أظهرت مقاطع الكبد وجود تفجى وتنخر الخلايا الكبدية في مركز الفصيص الكبدى والخلايا الكبدية حول البوابة الكبدية ووجود التهاب في القنويات الصفراوية مع ترسب لألياف الكولاجين في منطقة البوابة الكبدية فضلاً عن فرط تنسج في ظهارة القناة الصفراوية وتليفها. سبب استخدام السليمارين بعد المعاملة بالثايواسيتامايد إلى التحسن في كل من نتائج الاختبارات الكيموحياتية فضلاً عن التغيرات المرضية النسجية للكبد في المجموعة الرابعة. أخيراً ، خلصت الدراسة الحالية إلى أن السليمارين لديه خصائص في حماية الكبد ويعد كمضاد للتليف.

الكلمات الدالة: التشريح المرضي ، الكبد ، التليف ، الثايواسيتامايد ، السليمارين.