

Protective effects of the carotenoids-rich alga *Dunaliella salina* against thioacetamide-induced liver fibrosis in rats

Dalia R. Hassan* and Salem A. Salem*

Home Economic Department, Faculty of Specific Education, Fayoum – University

*dr.dal.refaat@gmail.com

Received: November 19, 2021; accepted: December 22, 2021; Available online: December 27, 2021

ABSTRACT

Dunaliella salina is naturally occurring source of beta-carotene which acts as antioxidant and also, it is beneficial in the treatment of liver disorders. The goal of this research is to investigate the impact of *D. salina* algae as an antioxidant on liver fibrosis in rats. In the current investigation, male Albino rats (N= 30 rats) weighing (180-200g) were used. They were separated into five groups: (1) negative control; (2) Positive control which was treated with Thioacetamide (TAA); groups (3), (4), and (5) TAA induced liver fibroses rats fed 100, 200, and 300mg *D. salina* powder/kg diet. The experiment extended to 6 weeks. The following were tested in the serum samples of each group: aspartate aminotransferase "AST", alanine aminotransferase "ALT", alkaline phosphatase, total bilirubin, albumin, Malondialdehyde "MDA", Superoxide dismutase "SOD", and glutathione "GSH". The results showed that rats in group (2) with liver fibrosis had considerably higher levels of their serum AST, ALT, total bilirubin and MDA and were significantly lower in serum albumin, SOD, and GSH. On the other hand, treatment of the induced liver fibrosis with different doses of *D. salina* powder had a significant decline in the rat serum levels of AST, ALT, ALP, bilirubin, and MDA, as well as a significant rise in serum antioxidant SOD, GSH, and serum albumin. However, liver histological examination of rats with generated hepatic fibroses in groups (3, 4 & 5) revealed using *D. salina* at different doses can decrease liver injury, necrosis, and inflammatory cell infiltration. This was attributed to the presence of high levels of carotenoids (especially β -carotene) in *D. salina* which has protective activity opposed to TAA-induced hepatic fibrosis in rats.

Keywords: *D. salina*, liver, antioxidants, β -carotene, MDA, AST, ALT, SOD, GSH, histopathology.

INTRODUCTION

Algae have a high nutritional value with a wide range of bioactive compounds (Singh and Jialal, 2006). *Dunaliella salina* is a member of the Chlorophyceae family, which includes unicellular biflagellate green algae. It is safe to use as a food additive or as a protective and curative agent in a variety of ailments (El-Baz *et al.*, 2019). It is a naturally occurring source of β -carotene (Borowitzka, 2013; Borovkovet *al.*, 2020). The oral acute toxicity assessment for *D. salina* powder or extract revealed no fatalities or toxicity indications a maximum dosage of 5g/kg,

suggesting that it was safe (Farouk *et al.*, 2020). Under appropriate conditions, *D. salina* can accumulate the maximum amount of β - carotene, making this aquatic algae far more effective as an antioxidant because. It is regarded as a critical biomolecule in the treatment of atherosclerosis, and retinal degeneration (Bansal and Sapna, 2009; Xu and Harvey, 2019).The most common colors in nature are carotenoids, which include carotene, lycopene, lutein and zeaxanthin and these are powerful antioxidants in healthy human diets (Martin, 2007).

Liver fibrosis is a long-term condition which acts on the global population, and considered one of the leading morbidity and mortality causes (Schuppan and Kim, 2013). Also, it being the most common cause of death for 1.3 million around the world (Wong and Huang, 2018). Fibrosis develops at different speeds depending on the source of the liver disease. Cirrhosis is an advanced stage of liver fibrosis, it induces direct diverting of portal and arterial blood circulation into the hepatic (central veins), altering transfer between the liver subcarriers and the surrounding liver epithelium, i.e., hepatocytes (Detlef and Nezam, 2008).

Thioacetamide "TAA" is commonly used in the food, and many industries, like leather processing, laboratory, beverages, textile and motor fuel industries (Akhtar and Sheikh, 2013). It is recognized as a human carcinogen and as a liver toxic that necessitates the oxidative biosynthetic route in order to deactivate its hepatotoxic effect (Low *et al.*, 2004; Ghosh *et al.*, 2016). TAA causes glomerular necrosis in liver cells as well as elevations in plasma liver enzymes and bilirubin, resulting in acute liver injury, whereas prolonged exposure causes hepatic fibrosis, liver tumor development, and cytomegaly (Zarger *et al.*, 2017; Bashandy *et al.*, 2018). TAA treatment also causes structural alterations in renal corpuscles, including as glomerular's capsule and tubule deterioration (Omar, 2018). It damages DNA, induces oxidative stress, produces cytokines, and induces renal failure in rats (Zarger *et al.*, 2019). TAA bioactivation, results in the formation of thioacetamide S-oxide, which generates superoxide radicals and ROS (reactive oxygen species). These oxidative damage are subsequently distributed throughout the body's various organs (Ghosh *et al.*, 2016). TAA may be absorbed by skin or inhaled/ingested by humans (Zarger *et al.*, 2019).

The present study aims to evaluate the role of using *Dunaliella salina* powder as a protective agent in rats against TAA-induced liver fibrosis.

MATERIALS AND METHODS:

Materials:

Algae:

Fresh *Dunaliella salina* in freshwater was received from the National Research Center's plant, biochemistry division. BG11 media was used to extract and filter algae from water, which was then grown for two weeks in 2 L of medium. Following growth, the algal biomass was extracted and cultivated for an additional two weeks, then grown in a 17 L canning jar with 15 L of culture media under oxygenation, and the culture temperature was $20\pm 3^{\circ}\text{C}$. For cultivation, a 2500 lx continuous light was used.

Animals:

Thirty male adult rats weighed 180-200g were procured from Helwan Cairo's animal house and fed a regular laboratory diet while also having unrestricted access to tap water. The animals were kept in a climate-controlled environment with 12-hrs light/dark cycle facility and temperatures ranging from 22-25°C. All animals had been treated with kindness.

Kits:

Thioacetamide (TAA) kit was purchased from Sigma-Aldrich Co. Transaminase (AST), alanine transaminase (ALT), alkaline phosphatase (ALP), total bilirubin, malondialdehyde (MDA), superoxide dismutase (SOD), glutathione (GSH), and albumin kits were supplied by Biodiagnostic Co., Egypt.

β -carotene of *D. salina*:

Using an Agilent 5 prep-C18 Scalar column and an analytical HPLC system (5m, 150mm, 4.6mm). At a flow rate of 1.25ml/min, the following solvents were used: (A) acetone and (B)

Protective effects of the carotenoids-rich alga *Dunaliella salina* against thioacetamide-induced liver fibrosis in rats

methanol/water (9:1, v/v) containing 0.05% butylated hydroxytoluene) (BHT). A gradient of solvents (A and B) was used to separate β -carotene for 40 minutes, including 25 minutes at 80 to 20%, 10 minutes at 20%, and 5 minutes at 20-80%. A colorimetric detection was used to measure the segregated β -carotene.

Experimental design:

After a week on a baseline diet, all 30 rats were separated into five groups (each with six rats).

Group (1): fed a baseline diet and served as the negative control.

Group (2): TAA was administered to rats intraperitoneally (dose 200 mg/kg of rat weight) two days weekly for four weeks and fed a baseline diet for six weeks.

Group (3): Rats with induced liver fibrosis were daily given *D. salina* powder (100 mg/kg diet).

Group (4): Rats with induced liver fibroses were daily given *D. salina* powder (200 mg/kg diet).

Group (5): Rats with induced liver fibroses were daily given *D. salina* powder (300 mg/kg diet).

All rats were given light anaesthesia at the end of the trial, and blood samples were collected from the retro-orbital vessels. Blood samples were left to coagulate before being separated for 15 minutes at 3000 rpm to extract the serum.

Histopathological examination:

Pieces of liver from each group were preserved in formalin solution (10%), dehydrated using ascending grades of alcohol, imbedded in paraffin wax cutting

5 μ thick. Sections were hydrated using descending grades of alcohol, then distilled water and stained with hematoxylin and eosin (H&E).

Statistical Analysis

The statistical significance of standard deviation across groups was determined using analysis of variation in one direction (ANOVA). At ($P \leq 0.05$), the significance of mean differences is determined and the Least Significant Difference (LSD) test was used. For all analysis of data, SPSS software was employed (Version 16; SPSS Inc., Chicago., USA).

RESULTS AND DISCUSSION

The chemical composition of *Dunaliella salina* powder indicated that it contains proteins (22.84%), carbohydrates (32.5%), lipids (5.8%), ash (4.87%) and total carotenoid (14.19%) of its dry weight and that of β -carotene was 12.98%. Moisture represents 4.87% of dry weight of this alga. Abd El-Baky *et al.* (2007) found that *D. salina* accumulates carotenoids (15.2% of its dry weight) and β -carotene (12.6%), making it more beneficial in the prevention of liver fibrosis. Furthermore, Farouk *et al.*, (2020) stated that phytochemical analysis of *D. salina* bioactive extract revealed a considerable quantity of carotenoids, notably β -carotene, which has numerous benefits in liver illnesses. *D. salina*, which has a high concentration of carotenoids and xanthophylls, was demonstrated to be an effective source of antioxidants versus a wide range of oxidative stresses (HU *et al.*, 2008; Hsu *et al.*, 2008).

Table (1): Chemical composition of *Dunaliella salina* powder.

Moisture %	Protein %	Fat %	Carbohydrates %	Ash %	β -carotene %	Total carotenoid %
4.87	22.84	5.8	32.5	19.8	12.98	14.19

Carotenoids are made up of molecules such as β -carotene and xanthophylls like lutein and zeaxanthin, which are needed to protect cells from the damaging consequences of pollutants, light, air, and irritant pigments (Noeman, 1989). Carotenoids' ability to diminish excited sensitizer particles and superoxides is the fundamental system of action for this phenomenon. Furthermore, β -carotene prevents liposomes from superoxide, hydroxyl radical-induced lipid auto-oxidation, peroxidation and hydrolysis of lipids caused by Fe^{2+} -generated radicals (L^{\bullet} and $L^{OO\bullet}$) (Shinmoto, 1992.). Furthermore, degradation of the lipid hydroperoxide produces alkoxy (LO) and peroxy radicals (LOO). They ultimately produce a large number of carbonyls, which cause DNA deterioration and the development of cancer and ageing process

disorders (Shinmoto,1992.;Yavuz et al., 2014).

Table (2) shows that TAA rats in the positive group (2) exhibit a highly significant ($P \leq 0.01$) rise in ALT, AST, ALP, and bilirubin, respectively, and a non-significant drop in serum albumin when compared to the negative group (1). Except for albumin, all of the studied parameters of TAA-rat groups given 100, 200, and 300mg/kg diet of *D. salina* decreased considerably ($P \leq 0.01$). This was consistent with the findings of Afifi *et al.* (2016), who discovered that after hepatic cell injury, the liver enzymes AST and ALT are released into the bloodstream.

On the other hand, the group (2) of TAA-rats who received *D. salina* 100mg/kg diet showed non-significant in albumin when compared with positive control group.

Table (2) Mean blood ALT, AST, ALP, bilirubin, and albumin levels in TAA-rats fed meals containing varying doses of *D. salina* (Mean \pm S.E.).

	ALT U/L	AST U/L	ALP U/L	Bilirubin mg/dl	Albumin g/dl
Group (1): Negative control	42.12 \pm 1.21**	61.80 \pm 1.32**	53.10 \pm 1.29**	1.37 \pm 0.11**	4.53 \pm 0.21**
Group (2): Positive control	76.48 \pm 1.47	103.17 \pm 2.50	127.92 \pm 1.79	3.05 \pm 0.20	3.39 \pm .0.14
Group (3): TAA+<i>D. salina</i> 100mg/kg diet	54.87 \pm 1.73**	87.37 \pm 1.57**	109.08 \pm 3.37**	2.33 \pm 0.10**	3.76 \pm 0.07
Group (4): TAA+<i>D. salina</i> 200mg/kg diet	45.95 \pm 2.16**	83.17 \pm 1.10**	84.92 \pm 1.59**	1.62 \pm 0.10**	4.22 \pm 0.09**
Group (5): TAA+<i>D. salina</i> 300mg/kg diet	40.05 \pm 1.47**	78.57 \pm 0.95**	62.28 \pm 1.31**	1.46 \pm 0.12**	4.60 \pm 0.14**

*Significant differences from Positive group (2) at $P \leq 0.05$.

**Significant differences from Positive group (2) at $P \leq 0.01$.

In comparison to the negative group, TAA (200 mg/kg body weight) i.p. injection induced severe hepatic fibrosis as well as significant rises in serum levels of AST, ALT, ALP, and total bilirubin. This finding is consistent with the findings of Kuriakose and Kurup (2010), who discovered that animals administered paracetamol had substantial liver damage 24 hours later, as seen by considerably ($P \leq 0.05$) greater levels of hepato-specific AST, ALT, ALP, and bilirubin. However,

after treatment with *D. salina*, all of the previous measures were considerably ($P \leq 0.05$) decreased, suggesting that consuming 1000 mg/kg of this algae may be capable of causing liver cell regeneration and decreasing the leakage of these enzymes into the blood.

Albumin levels are low in chronic liver damage due to the liver cells' diminished ability to produce proteins, which were according to Zhao *et al.*, (2014). Furthermore, serum ALP and

Protective effects of the carotenoids-rich alga *Dunaliella salina* against thioacetamide-induced liver fibrosis in rats

bilirubin levels are linked to liver cell activity, whereas ALT is linked to liver disease and AST is linked to liver damage, myocardial infarction, and severe muscle injury.

Treatment with different doses of *D. salina* powder acted as a hepatoprotective in the current investigation, as evidenced by lowering all liver enzymes, particularly at high doses, which led in the equilibrium of the enzyme levels in the treated rats. These results are comparable to those of Madkour and Abdel-Daim (2013), who discovered that a 1000 mg/kg pretreatment with *D. salina* microalga restored hepatic enzyme levels in paracetamol-intoxicated rats.

Data in Table (3) show that, TAA induced highly significant decrease in SOD, GSH and TAC by percentage of 59.40%, 59.12% and 67.89%, respectively, and resulted in a highly considerable rise in MDA level by 145.11% as compared to the negative group. Also, it was noticed that there was a highly significant ($P \leq 0.01$) reduction in (MDA), and highly considerable rise in (SOD, GSH and TAC) in TAA rats administered 300mg/kg diet *D. salina* at group (5). But on the other hand, there was no substantial differences in both MDA and TAC levels in TAA rats given 100mg/kg diet *D. salina* at group (3) in comparison to the positive control group.

Table (3): The mean serum MDA, TAC, SOD, and serum GSH levels in TAA-rats fed meals containing varying doses of *D. salina* (mean + S.E.).

GROUPS	MDA nmol	TAC mmol/L	SOD U/L	GSH mg/dl
Group (1): Negative control	8.09 ± 0.67**	1.90 ± 0.11**	84.87 ± 1.88**	20.06 ± 0.75**
Group (2): Positive control	19.83 ± 0.93	0.61 ± 0.02	34.45 ± 2.11	8.20 ± 0.49
Group (3): TAA+ <i>D. salina</i> 100mg/kg diet	18.56 ± 0.55	0.76 ± 0.03	46.36 ± 1.50**	10.44 ± 0.61*
Group (4): TAA+ <i>D. salina</i> 200mg/kg diet	17.53 ± 0.55*	0.91 ± 0.04*	54.17 ± 1.70**	12.89 ± 0.78**
Group (5): TAA+ <i>D. salina</i> 300mg/kg diet	14.75 ± 0.63**	1.63 ± 0.15**	68.76 ± 2.36**	19.24 ± 1.02**

* Significant differences from Positive G (2) $P \leq 0.05$.

**Significant differences from Positive G(2) $P \leq 0.01$.

According to El-Baz *et al.* (2020), oxidative stress (OS) is an imbalance in the equilibrium between free radical generation and antioxidant defences that has been associated to the etiology of numerous liver disorders (Li *et al.*, 2015). OS also induces structural and functional changes in physiologically macromolecules like proteins, carbohydrates, and lipids, according to Sutti *et al.* (2014). RO are produced by the electron transport chain in mitochondria and peroxidases, largely in hepatocytes during metabolic or detoxifying activities (Murphy, 2009). To counteract ROS and mitigate their harm, living organisms have evolved complex antioxidant systems comprised of endogenous and food-derived antioxidants.

Antioxidant collaboration provides more resistance to oxygen radicals or nitrogen species. As a result, TAC might give more valuable biological information and must take into consideration the whole impact of all antioxidants found in plasma and bodily fluids (Kuriakose and Kurup, 2010).

The primary line of defence against free radicals caused by oxygen has been recognized as SODs (Jung *et al.*, 2011; Blackney *et al.*, 2014). Superoxide is a free radical with a negative charge was generated when oxygen receives a free electron (Hayyan *et al.*, 2016). SOD also plays an important role in the cellular antioxidant defence system. It eliminates superoxide (O_2^-) by converting it to H_2O_2 ,

which quickly convert to water by both (catalase and glutathione peroxidase) (GPx).

MDA is a byproduct of the lipid peroxidation process (Barriuso, 2013) and when the number of free radicals created surpasses the cell's ability to eliminate them, oxidative stress occurs. The increase in MDA and decrease in TAC and SOD levels in TAA-induced liver fibrosis rats implies to a rise in lipid peroxidation, which causes tissue necrosis and failure of antioxidant defense systems to prevent uncontrolled free radical generation. The *D. salina* therapy adequately reversed these changes, in addition, *D. alina* demonstrated antioxidant effects against TAA in the current study, which were mediated by an increase in GSH levels, which is an antioxidant and anti-carcinogenic tripeptide. Furthermore, this was linked to a reduction in MDA. The current result was consistent with Sukalingam and Ganesan (2018).

Histopathological results:

The microscopic anatomy of a normal rat liver (group 1) reveals the existence of normal hepatocytes, as well as central and portal veins (Fig. 1A). However, as seen in group (2), rats with TAA-induced hepatic fibroses had hepatocyte degradation, fibrosis, and mononuclear cell infiltration (Fig. 1B). In TAA-induced liver fibrosis rats fed 100mg/kg diet of *D. salina* (group 3); there was fine fatty alteration of hepatocytes and portal infiltration with inflammatory cells (Fig. 1C). In TAA-induced liver fibrosis rats fed a 200mg/kg

diet of *D. salina*, considerable multifocal macrovascular and microvascular steatosis in hepatocytes surrounding the central vein and minor hydropic degeneration of hepatocytes were detected (Fig. 1D) (group 3). TAA-induced liver fibrosis in rats given 300mg/kg *D. salina* diet (group 5) resulted in normal histological pattern of hepatocytes, modest perivascular fibrosis in the portal, and isolated regions of coagulative necrosis (Fig. 1E).

The current findings showed that *D. salina* provided antioxidant protection against TAA-induced fibrosis in the livers of the rats studied. TAA insult caused glomerulus deformation and congestion. This was demonstrated in rats from groups (3, 4, and 5) that were given 100, 200, and 300mg/kg diets of *D. salina* powder. As revealed by the biochemical studies, this can be explained by an increase in GSH levels and an inhibition of MDA levels. GSH includes a tripeptide that is both antioxidant and anticarcinogenic, which increases protection against oxidant-induced cell damage (Tsai *et al.*, 2012). Sukalingam and Ganesan's (2018) findings are compatible with the present findings. *D. salina* inhibited thioacetamide-induced inflammatory cell infiltration, indicating that it may have antihepatotoxic effects (Farouk *et al.*, 2020). Furthermore, the presence of β -carotene in *D. salina* can be related to the improvement in TAA-induced liver fibrosis group of rats (group 6). El-Baz *et al.*, (2020) found that β -carotene in *D. salina* reduced liver fibrosis in rats by lowering inflammatory mediators.

Protective effects of the carotenoids-rich alga *Dunaliella salina* against thioacetamide-induced liver fibrosis in rats

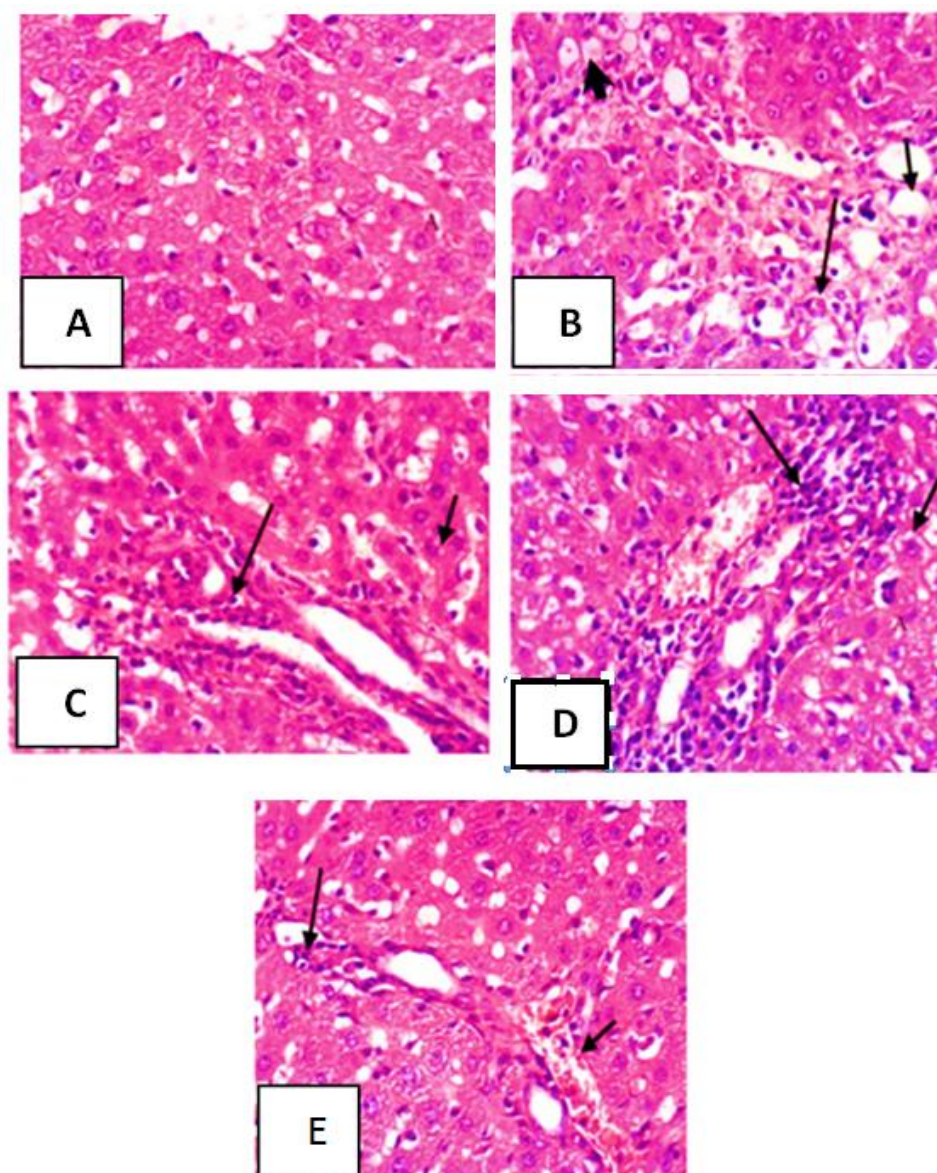


Figure (1A-E): Show microphotographs of sections of liver of rats from different groups stained with HE, with Magnification 400X .

(Fig 1-A): Negative G1, rats with normal histological pattern of liver hepatocytes, central and portal veins.

(Fig1-B):) Positive G2, rats with TAA induced liver fibroses showing degeneration of hepatocytes, presence of fibrosis and mononuclear cells infiltration (arrows).

(Fig 1-C): Group (3), rats with TAA induced liver fibroses and fed 100mg/kg diet of *D. salina* showing hepatocyte fatty changes and portal congestion with granulocytes (arrows) .

(Fig 1-D): rats with TAA induced liver fibroses and fed 200mg/kg diet of *D. salina* showing moderate multifocal macrovesicular and microvesicular steatosis in hepatocytes surrounding the central vein and slight hydropic degeneration of hepatocytes (arrow).

(Fig 1-E): rats with TAA induced liver fibroses and fed 300mg/kg diet of *D. salina* showing normal histological pattern of hepatocytes, mild perivascular fibrosis in the portal area and focal areas of coagulative necrosis (arrows).

Conclusion:

The inclusion of β -carotene in *D. salina* powder significantly reduced hepatic steatosis. These findings suggest that *D. salina* can be used as an effective and valuable medication in treatment of liver cirrhosis. Additional research and medical investigations are needed to assess the medicinal value of *D. salina* in anti-fibrotic rats and individuals with liver cirrhosis.

REFERENCES

- Afifi, N.A.; Ramadan, A.; El-Eraky, W.; Salama, A.A.A.; El-Fadaly, A.A. and Hassan, A. (2016). Quercetin protects against thioacetamide induced hepatotoxicity in rats through decreased oxidative stress biomarkers, the inflammatory cytokines; (TNF- α), (NF- κ B) and DNA fragmentation. *Der Pharma Chemica.*, 8:48-55.
- Bansal, M.P. and Sapna, J. (2009). Hypercholesterolemia induced oxidative stress is reduced in rats with diets enriched with supplement from *Dunaliella salina* algae. *Am. J. Biomed. Sci.*, 1(3): 196-204.
- Barriuso, B.; Astiasarán, I. and Ansorena, D. (2013): A review of analytical methods measuring lipid oxidation status in foods: A challenging task. *Eur. Food Res. Technol.*, 1–15.
- Bashandy, S.A.; Ebaid, H.; Moussa, S.A.A.; Alhazza, I.M.; Hassan, I.; Alaamer, A. and Al Tamimi, J. (2018). Potential effects of the combination of nicotinamide, vitamin B2 and vitamin C on oxidative-mediated hepatotoxicity induced by thioacetamide. *Lipids in health and Disease*, 17:1-9.
- Blackney, M.J.; Cox, R.; Shepherd, D. and Parker, J.D. (2014). Cloning and expression analysis of *Drosophila* extracellular Cu and Zn superoxide dismutase. *Biosci. Rep.*, 34:e00164.
- Borovkov, A.B.; Gudvilovich, I.N. and Avsiyan, A.L. (2020). Scale-up of *Dunaliella salina* cultivation: from strain selection to open ponds. *J. Appl. Phycol.*, 32:1545–1558
- Borowitzka, M.A. (2013). *Dunaliella*: biology, production, and markets. In: Richmond, A. and Hu, Q. (eds) *Handbook of Microalgal Culture*. Wiley, Chichester: 359–368
- Detlef, S. and Nezam, H.A. (2009). Liver Cirrhosis. *Lancet.*, 371(9615): 838–851.
- El-Baz, F.K.; Salama A.A.A. and Hussein R.A. (2020). *Dunaliella salina* microalgae oppose thioacetamide-induced hepatic fibrosis in rats. *Toxicol. Rep.*, 7: 36–45.
- Farouk K. El-Baz, aHanan F. Aly, b, and Abeer A.A. Salama (2019): Toxicity assessment of the green *Dunaliella salina* microalgae. *Toxicology Reports*. Vol: 6, 850-861.
- Ghosh, S.; Sarkar, A.; Bhattacharyya, S. and Sil, P.C. (2016). Silymarin protects mouse liver and kidney from thioacetamide induced toxicity by scavenging reactive oxygen species and activating PI3K-Akt pathway. *Frontiers in pharma*, 7:481.
- Harvey, P.J. and Ben-Amotz, A. (2020). Towards a sustainable *Dunaliella salina* microalgal biorefinery for 9-cis beta-carotene production. *Algal Res.*, 50:102002.
- Hayyan, M.; Hashim, M.A. and Alnashef, I.M. (2016). Superoxide Ion: Generation and Chemical Implications. *Chem. Rev.*, 116:3029–3085.
- Hsu, Y.W.; Tsai, C.F.; Chang, W.H.; Ho, Y.C.; Chen, W.K. and Lu, F.J. (2008). Protective effects of *Dunaliella salina* – A carotenoids-rich alga, against carbon tetrachloride-induced

Protective effects of the carotenoids-rich alga *Dunaliella salina* against thioacetamide-induced liver fibrosis in rats

- hepatotoxicity in mice. *Food Chem. Toxicol.*, 46:3311–7.
- Hu, C.C.; Lin, J.T.; Lu, F.J.; Chou, F.P. and Yang, A. (2008). Determination of carotenoids in *Dunaliella salina* cultivated in Taiwan and antioxidant capacity of the algal carotenoid extract. *Food Chem.*, 109(2):439-46.
- Jung, I.; Kim, T.Y. and Kim-Ha, J. (2011). Identification of *Drosophila* SOD3 and its protective role against phototoxic damage to cells. *FEBS Lett.*, 585:1973–1978
- Kuriakose, G.C. and Kurup, M.G. (2010). Antioxidant and hepatoprotective activity of *Aphanizomenon flosaquae* Linn against paracetamol intoxication in rats. *Indian J. Exp. Biol.*, 48:1123-30
- Li, S.; Tan, H.Y.; Wang, N.; Zhang, Z.J.; Lao, L.; Wong, C.W. and Feng, Y. (2015). The role of oxidative stress and antioxidants in liver diseases. *Int. J. Mol. Sci.*, 16:26087–26124
- Low, T.Y.; Leow, C.K.; Salto, T.M. and Chung, M.C. (2004). A proteomic analysis of thioacetamide-induced hepatotoxicity and cirrhosis in rat livers. *Proteomics*, 4:3960-3974.
- Martín, A.; Mattea, F.; Gutierrez, I.; Miguel, F. and Cocero, M.J. (2007). Co-precipitation of carotenoids and bio-polymers with the supercritical anti-solvent process. *J. Supercrit. Fluids*, 41(1):138–14.
- Murphy, M.P. (2009). How mitochondria produce reactive oxygen species. *Bioch. J.*; 417: 1–13.
- Norman I. Krinsky (1989). Antioxidant functions of carotenoids. *7*, (6): 617-635
- Omar, A.M.S. (2018). The potential protective influence of flaxseed oil against renal toxicity induced by thioacetamide in rats. *Saudi J. Biological Sci.*, 25:1696-1702.
- Schuppan, D. and Kim, Y.O. (2013). Evolving therapies for liver fibrosis. *J. Clin. Invest.*, 123 (5):1887–1901.
- Singh, U. and Jialal, I. (2006). Oxidative stress and Atherosclerosis. *Pathophysiology*, 6(13): 129-42.
- Shinmoto H., Dosako S., Nakajima, I. (1992). Antioxidant activity of bovine lactoferrin iron/ascorbate induced lipid peroxidation. *Biosci. Biotech. Biochem.*, 56:2079-2080.
- Sukalingam, K.; Ganesan, K. and Xu, B. (2018). Protective effect of aqueous extract from the leaves of *Justicia tranquebariensis* against thioacetamide-induced oxidative stress and hepatic fibrosis in rats. *Antioxidants* (Basel). *Antioxidants*, 7: 78
- Sutti, S.; Jindal, A.; Locatelli, I.; Vacchiano, M.; Gigliotti, L.; Bozzola, C. and Albano, E. (2014). Adaptive immune responses triggered by oxidative stress contribute to hepatic inflammation in NASH. *Hepatology*, 59:886–897
- Tsai, C.F.; Lu, F.J. and Hsu, Y.W. (2012). Protective effects of *Dunaliella salina* - a carotenoids-rich alga - against ultraviolet B-induced corneal oxidative damage in mice. *Mol. Vis.*, 18:1540-4.
- Wong, M.C.S. and Huang, J. (2018). The growing burden of liver cirrhosis: implications for preventive measures. *Hepatol. Int.*, 12(3):201–203.
- Xu, Y. and Harvey, P.J. (2019). Red Light control of β -carotene isomerisation to 9-cis β -carotene and carotenoid accumulation in *Dunaliella salina*. *Antioxidants*, 8:148-161
- Yavuz S.C.; Murat K. and Meltem, A.O. (2014). Biochemical composition and bioactivity screening of various extracts from *Dunaliella salina*, a green microalga. *EXCLI J.*, 13:679-690
- Zhao, Y.; Ma, X.; Wang, J.; He, X.; Hu, Y.; Zhang, P.; Wang, R.; Li, R.;

- Gong, M.; S. Luo, S. and Xia, O. (2014). Curcumin protects against CC14-induced liver fibrosis in rats by inhibiting HIF-1 alpha) through an ERK-dependent pathway. *Molecules*, 19:18767-80.
- Zargar, S.; Alonazi, M.; Rizwana, H. and Wani, T.A. (2019). Resveratrol reverses thioacetamide-induced renal assault with respect to oxidative stress, renal function, DNA damage, and cytokine release in Wistar rats..*Hindawi Oxidative Medicine and Cellular Longevity* Volume 2019, Article ID 1702959, 8 pages. <https://doi.org/10.1155/2019/1702959>
- Zargar, S.; Wani, T.A.; Alamro, A.A. and Ganaie, M.A. (2017). Amelioration of thioacetamide-induced liver toxicity in Wistar rats by rutin. *Int.J. Immuno. Pharma.*, 30:207-214

طحالب الدوناليليا سالينا تحمي من تليف الكبد الناجم عن مركب الثيوأسيتاميد في الجرذان

داليا رفعت حسن , سالم على سالم

قسم الاقتصاد المنزلي ، كلية التربية النوعية ، جامعة الفيوم

المستخلص

طحالب الدوناليليا هي مصدر طبيعي للبيتا كاروتين ولها تأثير علاجي في أمراض الكبدو تعرف بأنها مصدر جيد للبيتا كاروتين حيث يعتبر مضاد للأكسدة ، وعلى ذلك ، فان الهدف من هذه الدراسة هو دراسة من تأثير الطحالب كمضاد للأكسدة لعلاج التليف الناجم عن الثيوأسيتاميد المسبب لتليف الكبد في فئران التجارب. تم استخدام ثلاثين ذكور جرذ ألبينو وزنها (180-200 جم) في الدراسة الحالية. تم تقسيمهم إلى 5 مجموعات ؛ المجموعة (1) مجموعة السيطرة السلبية ؛ المجموعة (2) بمجموعة الثيوأسيتاميد) مجموعة التحكم الإيجابية) ؛ المجموعات (3 و 4 و 5) تغذت الفئران التي حققت بمادة الثيوأسيتاميد بـ 100 و 200 و 300 ملجم / كجم من مسحوق الدوناليليا سالينا اجريت التجربة لمدة 6 أسابيع تم قياس الأسبارتات أمينوترانسفيراز ، ألانين أمينوترانسفيراز ، الفوسفاتيز القلوي ، البيليروبين الكلي ، الألبومين ، مالون داي الدهيد ، سوبراوكسيد داي ميوتيز، والجلوتاثيون في سيرم الدم . وقد زادت بشكل ملحوظ كلا من معدل الأسبارتات أمينوترانسفيراز ، ألانين أمينوترانسفيراز، مالون داي الدهيد والبيليروبين . كما انخفضت بشكل ملحوظ كلا من سوبراوكسيد داي ميوتيز، والجلوتاثيون في سيرم الدم. وقد احدث مسحوق دوناليليا سالينا في مستويات مختلفة الهى انخفاض معنويا ملحوظا في مستويات الأسبارتات أمينوترانسفيراز ، ألانين أمينوترانسفيراز ، الفوسفاتيز القلوي ، البيليروبين بالاضافة الى انخفاض مستوي مالون داي الدهيد ، وعلى الجانب الاخر حدث ارتفاع معنوي في مضادات الأكسدة في السيرم (سوبراوكسيد داي ميوتيز، الجلوتاثيون) كما ارتفع مستوى الألبومين بالسيرم. كما أظهرت نتائج الهستوباثولوجيا أنسجة الكبد ان طحالب الدوناليليا ادت الى تقليل التليف ونخر الفصوص المركزية الكبدية و الخلايا الملتهبة الناتجة عن مادة الثيوأسيتاميد. أشارت النتائج إلى أن طحالب الدوناليليا تساعد بإجراءات وقائية ضد التليف الكبدي في الفئران. ختاماً كشف التحقيق الكيميائي النباتي عن وجود نسبة عالية من البيتتا كاروتين، وهي المسؤولة عن وقاية الكبد من التليف.