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Genetic Diversity Among some Parental Varieties of *Vicia faba* L. Using ISSR and SCoT Molecular Marker Techniques

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ABSTRACT



This study was executed to evaluate molecular variations among 13 genotypes of *Vicia faba* from Egypt and Kurdistan of Iraq for use in crop breeding programs in Kurdistan. The genetic relationships have been detected among these genotypes using ISSR and SCoT markers. Ten and twelve primers had successfully generated reproducible polymorphic amplicons that were suitable for studying the genetic variability between studied genotypes. Out of 225 clear amplicons detected, (77 for ISSR and 92 for SCoT were polymorphic) with 70.71 and 70.03 polymorphism %, respectively. SCoT markers provided more discriminating data and were more informative with high UM%, DI, and Rp values (37.0, 0.23, and 3.81, respectively) compared with ISSR markers (25.7, 0.21, and 3.50, respectively). Cluster analysis and PCo analysis based on ISSR-SCoT combined data succeeded in the description of genetic diversity and heterogeneity within studied genotypes. Hence, setting the studied genotypes into four major groups. The first group included check variety only while the second group included sakha1, sakha3 and sakha4, the third group included only Giza843, and the last group contained all other genotypes, including all studied cultivars from Kurdistan. Crosses between divergent genotypes out of these groups may lead to the development of effective breeding strategies in the future. Therefore, we recommend more studies on the genetic improvement of these cultivars and their inclusion in breeding programs in Kurdistan of Iraq due to the significant genetic differences between them.

Keywords: Genetic diversity among of Vicia faba L. - SCoT and ISSR.

INTRODUCTION

The faba bean (Vicia faba L. 2n=12) is considered to be one of the most important pulse crops produced in Iraq. The small-seeded Vicia faba major (broad bean), the large-seeded Vicia faba minor field or (horse bean), and the medium-seeded type (Vicia faba) equine are the three different varieties of faba beans. In actuality, the only difference between the three types of seed size is that they all belong to the same species Hanelt and Mettin (1989). The edible seed of faba beans, a resource with great economic value that complements cereal-based diets with substantial amounts of protein, makes it an ideal crop for rotation with cereals. The ultimate objective of plant breeders in order to sustain high agricultural productivity is the production of varieties with high yield potential. A novel cultivar should not only have a high yield potential but also consistent performance and extensive adaptability to a variety of settings. Given that genotypes reply to environmental change in different ways, the genotype environment (G \times E) interaction is crucial for faba bean breeders. Strong G×E interactions for quantitative traits like seed yield can dramatically reduce the benefits of selecting superior genotypes for improved cultivar development (Fathi et al., 2013). Evaluating the stability of performance and breadth of adaptability for cultivars being chosen for a wide range of settings has become more crucial. When genotypes are tested in various conditions, number of stability metrics have been proposed to define yield stability, with varying degrees of success (Dawod and Al-Layla, 2008).

On the other hand, the assessment of genetic variation is a crucial component of genetic studies, biodiversity research, germplasm characterization, and choosing favorable genotypes in plant breeding programs. Where it is becoming known that significant levels of genetic variation are not expressed in the phenotype. Recently, technological developments in molecular genetics have made it possible to quantify genetic diversity at the DNA level by detecting different molecular markers (Abid *et al.*, 2015). One of the most important advantages of molecular markers is that they are not affected by plant developmental stages or environmental factors (Abdel-Razzak *et al.*, 2012). Therefore, molecular marker techniques are used in

order to assess and utilize genetic variation in favor of a breeding program. Where, marker-assisted selection (MAS) is expected to change the strategies used for faba bean breeding, as it has in many other crop species. Already, different types of molecular marker techniques have been employed to assess the genetic variations among V. faba genotypes such as RFLP, RAPD, AFLP, TRAP, and SSAP (Abid et al., 2015). Intersimple sequence repeat (ISSR) molecular marker technique was also utilized in order to explore the genetic diversity of V. faba genotypes (Terzopoulos and Bebeli, 2018; Asfaw et al., 2018). Recently, Collard and Mackill (2009) outlined a quick and innovative DNA marker technology called start codon targeted (SCoT) polymorphism. This marker was developed based on the short-conserved region flanking the Adenine-Thymine-Guanine (ATG) start codon in plant genes. SCoT markers are like RAPD and ISSR molecular marker techniques because a single primer is used as forward and reverse. Moreover, as a PCR-based gene target technique, SCoT analysis has low cost and is effective to use (Bhattacharyya et al., 2013).

Many studies found that SCoT might be more effective than other dominant DNA molecular markers like RAPD and ISSR because it is gene-targeted (Gupta *et al.*, 2018). Also, SCoT is superior to these markers in higher

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polymorphism and better marker resolvability (Gorji *et al.*, 2011). Moreover, SCoT can generate co-dominant markers caused by insertions and deletions, as well as they can generate dominant markers caused by sequence variations like RAPD and ISSR (Aswathy *et al.*, 2017). However, using ISSR and SCoT markers together gives very effective, reliable, and more superior results in genetic diversity study than the use of single markers (Mao *et al.*, 2018).

Therefore, the objective of this study was to assess molecular variations among 13 faba bean genotypes from Egypt and Kurdistan using ISSR and SCoT molecular marker techniques to test the efficiency of these techniques for genotype identification and marker-assisted selection. Hence, then help in managing the germplasm, improving present **Table 1. List of cultivars used in the study**. breeding methods, and introducing new cultivars in breeding programs in Kurdistan.

MATERIALS AND METHODS

This research was conducted during summer season 2021 at a private laboratory in Egypt (Creative Egyptian Biotechnologists; Giza, Egypt) to evaluate molecular variation among 13 genotypes of faba bean.

Genetic materials

The genetic materials used in this study were seeds of thirteen varieties of *Vicia faba* (Table 1) shows the pedigree of these genotypes and contains information such as origin, earliness, foliar diseases reaction, Orobanche reaction, drought-tolerant, seed color and seed size.

	Cultivars	Abbreviation	Origin	Pedigree	Earliness	Foliar Diseases reaction	Orobanche reaction	Drought tolerant	Seed color	Seed size
1.	Sakha 1	Sa1	FCRI, EGYPT	Giza 716 x 620/283/85	Early	Resistant	Susceptible	Susceptible	light brown	Medium
2.	Sakha 3	Sa3	FCRI, EGYPT	Promising line 716/402/2001 derived from cross716 (Giza461 X 503/453/83)	Late	Resistant	Susceptible	Susceptible	light brown	Medium
3.	Sakha 4	Sa4	FCRI, EGYPT	Sakha 1 X Giza 3	Early sowing	Resistant	Susceptible	Susceptible	light brown	Medium
4.	Nubaria 3	Nu3	FCRI, EGYPT	Selected from Ahnacia line	Late	Resistant	Susceptible	tolerant	light brown	Medium
5.	Nubaria 4	Nu4	FCRI, EGYPT	-	Late	Resistant	Susceptible	tolerant	light brown	Medium
6.	Nubaria 5	Nu5	FCRI, EGYPT	-	Late	Resistant	Susceptible	tolerant	light brown	Medium
7.	Giza 716	Gi716	FCRI, EGYPT	461\842\83x503\453\83	Early	Resistant	Susceptible	tolerant	light brown	large
8.	Giza 843	Gi843	FCRI, EGYPT	561/2076/85 X 461/845/83	Late	Moderately resistant	Resistant	Susceptible	light brown	Medium
9.	Misr3	Mi3	FCRI, EGYPT	L667 x (Cairo 241 x Giza 461)	Moderate	Moderately resistant	Resistant	Susceptible	light brown	Medium
10.	FLIP17- 066FB	R117	IRAQ	Selection for heat from shambat	-	-	-	-	-	Medium
11.	FLIP17- 071FB	R179	IRAQ	\$88135-3-2-1	-	-	-	-	-	Medium
12.	FLIP19- 218FB	F19	IRAQ	S88094-8-1	-	-	-	-	-	Medium
13.	Check variety	CV	IRAQ	Local variety	Moderate	Moderately resistant	known	known	brown	Medium

Molecular SCoT and ISSR analysis DNA Extraction

2-Start Codon Target Translation (SCoT).

Thirteen cultivars of *Vicia faba* were collected separately then DNA extraction was performed using DNeasy plant Mini Kit (Bio Basic).

Polymerase chain reaction (PCR)

In the molecular analysis of thirteen types of Vicia faba, genomic DNA was employed as a template for PCR amplification using 10 ISSR primers and 12 SCoT primers. Primers for ISSRs purchased from Operon Technology in Alameda, California, the consensus sequence used in Joshi *et al.* (1997) and Sawant *et al.* (1999). Research, on the other hand, was used to create SCoT primers, which were purchased from Biobasic Com. The ISSR and SCoT methods' amplification reactions were carried out in accordance with the instructions provided by Fathi *et al.* (2013) and Xiong *et al.* (2011), respectively.

1- Inter Simple Sequence Repeat (ISSR)

The DNA amplifications were performed in an automated thermal cycle (model Techno 512) programmed for one cycle at 94° C for 4 min followed by 40 cycles of 1 min at 94° C, 1 min at 57° C, and 2 min at 72° C. the reaction was finally stored at 72° C for 10 min the reaction was finally stored at 4 °C.

The DNA amplifications were performed in an automated thermal cycle (model Techno 512) programmed for one cycle at 94° C for 4 min followed by 40 cycles of 1 min at 94° C, 1 min at 55° C, and 2 min at 72°C. The reaction was finally stored at 72° C for 10 min the reaction was finally stored at 4 °C.

Gel electrophoresis

Amplified products on a 1.5% agarose gel with ethidium and 100 bp to 1.5 kb ladder markers, were separated. The run took place in a tiny submarine gel Bio-Rad for about 30 minutes at 100V.

Gel reading and analysis

DNA GelAnalyzer3 software was employed to evaluate banding pattern images after they were taken with the Bio-1D Gel Documentation System. This program rated clear amplicons as present (1) or absent (0) for each primer and input the results as a binary data matrix. According to Adhikari *et al.*, (2015) DNA-profiles for ISSR and SCoT methods were produced from this matrix.

Polymorphic Information Content (PIC) and DI (Diversity Index) were calculated according to, PIC = 1-p2-q2, DI=average PIC value Gorji *et al.*, (2011) where p is frequency of present amplicon and q is frequency of absent amplicon.

The ability of each primer to differentiate among the studied was also evaluated using binary data and the resolving power (Rp) value calculated as described by Prevost and Wilkinson (1999) using the formula:

$$Rp = \sum I_{a} = \sum 1 - [2 \times (|0.5 - p|)]$$

Where, ¹a (amplicon informativeness) was calculated for each amplicon scored individually by the primer, p being the ratio of studied lines containing theI amplicon.

Comparative analysis

According to binary data matrix, similarity coefficients were calculated by Dice coefficient Nei and Li, (1979). Agglomerative hierarchical clustering (AHC) analysis (Cluster analysis) derived from Unweight pair-group average (UPGMA) method and Principal Coordinate (PCo) analysis were performed using XLSTAT 2019.2.2 software according to Hamada *et al.*, (2019). Where PCo analysis is used for classification of genotypes and in fact, it is a supplementary method for cluster analysis Singh and Chaudhary, (1985).

RESULTS AND DISCUSSION

Molecular variation assessment using ISSR technique.

Molecular banding patterns and DNA profiles estimated from ISSR technique were shown in Figs. 1 and 2.



Fig. 1. Banding patterns of ISSR -PCR products for 13 Vicia faba L. cultivars produced with ten primers. L, ladder (0.1 :1.5 kb).

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Fig. 2. DNA-profile representation of ISSR fingerprints of 13 Vicia faba L. cultivars based on 9666 amplicons 21 of them were marker loci according to Adhikari *et al.* (2015).

These Figures clearly demonstrated that there were differences among patterns of *Vicia faba* genotypes. This was accomplished by using ten primers targeted 96 scoreable amplicons with product sizes ranging between 174 and 1955 bp.

Molecular data obtained from analyzing ISSR banding patterns and recorded in Table 2 showed that polymorphism levels varied from one primer to the other and ranged from 0.0 to 100%. Where the 14A primer targeted the highest number of amplicons (21) with the highest value of Rp (9.394) and polymorphism percentage and PIC were 95.24% and 0.309. Demonstrating that the 14A primer showed a high level of discrimination and informativeness in the identification when compared to other primers.

On the other hand, Table 3 showed the similarity matrix based on ISSR-technique data where the lowest values of similarity coefficient were 0.585 between Giza843 and check variety, followed by 0.628 between Giza716 and check variety. While the highest values of similarity coefficient were 0.931 between Misr3 and FLIP17-071FB followed by 0.914 between Nubaria3 and Nubaria5.

Table 2. Molecular d	ata estimated from banding patterns of ISSR technique.
Primer	Amplicons

	Comment	Mal	lan]	Polymorph	ic	-	Poly	morphism	Pol	ymorphic	Resolving
Name	$(5' \rightarrow 3')$	size i	cular range	Monomorphi	c Without	Unique	Unique	Total	·	%	ina	(PIC)	power Rn
	(8 7 0)	Sinc	unge		unique	(+)	(-)					(110)	T.P
14A	(CT)8TG	174:	1867	1	15	2	3	21		95.24		0.309	9.394
44B	(CT)8GC	191:	1236	4	5	2	1	12		70.50		0.162	2.618
49-A	(CT) ₆ AG	342:	1809	1	2	0	2	5		80.00		0.169	1.694
49-B	(CA)6GG	267:	1569	0	10	2	1	13		100.00		0.324	6.006
HB-09	(GT) ₆ GC	359:	1955	3	3	0	0	6		50.00		0.152	1.386
HB-11	(GT) ₆ CC	212	:410	2	0	0	0	2		0.00		0.000	0.000
HB-12	(CAC) ₃ GC	237:	1190	0	7	1	1	9		100.00		0.318	4.158
HB-13	(GAG) ₃ GC	275:	1340	2	4	0	4	10		80.00		0.170	2.310
HB-14	(CTC) ₃ GC	186	: 692	2	3	1	0	6		66.70		0.220	2.464
HB-15	(GTG) ₃ GC	283:	1601	4	7	1	0	12		66.67		0.268	4.928
Table 3	3. Similarity	y matr	ix for 1	13 Vicia faba	L. cultiva	rs based o	n ISSR-t	echniqu	ue dat	a			
Cultiva	rs	Sa 1	Sa 3	Sa 4	Nu 3 N	u4 Nu	15 Gi'	716 Ĝ	i 843	Mi 3	R 117	R179	F19
Sa 3	C).843											
Sa 4	0).846	0.843										
Nu 3	C).790	0.803	0.807									
Nu 4	C).748	0.793	0.797	0.899								
Nu 5	C).720	0.748	0.752	0.914 0.	903							
Gi 716	C).689	0.767	0.770	0.818 0.	894 0.8	367						
Gi 843	C).673	0.648	6 0.710	0.754 0.	794 0.7	766 0.7	52					
Mi 3	C).746	0.742	0.762	0.865 0.	883 0.8	371 0.8	33 0	.806				
R 117	C).721	0.697	0.793	0.810 0.	800 0.8	303 0.7	'91 0	0.772	0.857			
R179	C).721	0.750	0.754	0.891 0.	851 0.8	367 0.8	314 0).784	0.931	0.853		
F19	C	0.701	0.730	0.735	0.848 0.	853 0.8	355 0.8	300 0).783	0.863	0.839	0.889	
CV	0).699	0.752	0.680	0.695 0.	<u>672</u> 0.6	545 0. 6	5 <u>28</u> 0).585	0.720	0.727	0.694	0.690

Comparative analysis for data of molecular markers techniques enables the determination of proximity or distance between genotypes through displays of genotypes in clusters (Ozturk et al., 2022). For this purpose, cluster analysis was

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performed among thirteen cultivars of faba bean, a dendrogram for the genetic relationship among cultivars were carried out as Fig. 3 which separated them into four major groups at a similarity coefficient = 0.799. The first group included (check variety) only while the second group included (sakha3, sakha1 and sakha4), the third group included only (Giza83), the last group divided into (four) subgroups, the first subgroup included (FLIP17-071FB, Misr3) and (FLIP19-218FB) only, the second subgroup (Nubaria5, Nubaria3, Nubaria4), the third one is (Giza716) and the last one is (FLIP17-066FB).



Fig. 3. UPGMA clustering dendrogram for 13 *Vicia faba* L. cultivars based on ISSR molecular data using Nei and Lis similarity coefficient.

Legend: TL represents truncated line at the coefficient of similarity =0.799.

Molecular variation assessment using SCoT technique.

In the same vein, Figs. 4 and 5 displayed various molecular banding patterns and DNA profiles conducted but using the SCoT molecular marker technique. This was accomplished by targeting 129 scoreable amplicons using 12 primers with product sizes between 116 and 1635 bp.

Table 4 shows molecular data obtained from analyzing SCoT banding patterns. Polymorphism levels varied amongst primers, with the SCoT-7 primer targeting the highest number of amplicons (17) with a polymorphism percent of 94.10% and a PIC of 0.357 in comparison to other primers. While the ScoT-5 primer targeted the fewest amplicons and furthermore demonstrated the lowest RP value of 1.386, polymorphism%, and PIC, which were 60.00% and 0.139, respectively. This demonstrates the SCoT-7 primer's high-informative and discriminatory capabilities in the discovery of genetic diversity.

Table 5 showed the similarity matrix based on SCoTtechnique the lowest value of similarity coefficient was 0.680 between Sakha3 and check variety, followed by 0.693 between Sakha1 and FLIP17-066FB. While the highest value of similarity coefficient was 0.900 between Giza716 and Misr3 followed by 0.893 between Sakha4 and Nubaria3.

Cluster analysis was performed among the thirteen cultivars of faba bean using SCoT molecular data. A dendrogram obtained from this analysis was illustrated in Fig. 6 which separated all studied cultivars into four groups at a similarity coefficient =0.795. The first group included (Sakha1, Sakha3) while the second group included FLIP17-071FB, FLIP19-218FB, FLIP17-066FB and check variety. The third group included (Giza843) only, while the last included Sakha4, Nubaria3, Misr3, Giza716, Nubaria4 and Nubaria5.



Fig. 4. Banding patterns of SCoT-PCR products for 13 *Vicia faba* L. cultivars produced with 12 primers. L, 1.5 kb



Fig. 5. DNA-profile representation of ScoT fingerprints of 13 *Vicia faba* L. cultivars based on 129 amplicons 21 of them were marker loci according to Adhikari *et al.*, (2015).

Table 4. Molecular data estimated from banding patterns of SCoT technique.

Primer			A	nplicons					Polymorphic	Decolving
Name	Sequence	Molecular	Manamahia	Po	lymorphi	c T	Tetal	Polymorphism	index	power
Iname	$(5^{\prime} \rightarrow 3^{\circ})$	range	Monomorphic	unique	Unique (+)	Unique (-)	Total	70	(PIC)	[°] Rp
SCoT-2	CAACAATGGCTA CCACCC	115:1386	4	6	4	0	14	71.43	0.233	5.236
SCoT-5	CAACAATGGCTA CCACGA	202: 624	2	2	0	1	5	60.00	0.139	1.386
SCoT-6	CAACAATGGCTA CCACGC	199:1119	1	4	1	4	10	90.00	0.216	2.772
SCoT-7	CAACAATGGCTA CCACGG	197:1635	1	15	1	0	17	94.10	0.357	8.932
SCoT-8	CAACAATGGCTA CCACGT	214:1283	6	2	0	1	9	33.33	0.100	1.386
SCoT-9	CAACAATGGCTA CCAGCA	239:1372	5	1	2	1	9	44.44	0.095	1.078
SCoT-10	CAACAATGGCTA CCAGCC	236:1477	5	5	1	1	12	58.33	0.166	2.618
SCoT-11	AAGCAATGGCTA CCACCA	277:1107	3	5	2	0	10	70.00	0.232	3.388
SCoT-12	ACGACATGGCTA CCAACG	213:1064	2	6	0	0	8	75.00	0.337	4.312
SCoT-13	ACGACATGGCG ACCATCG	224:1542	5	10	0	1	16	68.80	0.272	6.776
SCoT-14	ACGACATGGCG ACCACGC	214:1438	1	9	1	0	11	99.90	0.319	5.082
SCoT-15	ACGACATGGCG ACCGCGA	185:794	2	6	0	0	8	75.00	0.255	2.772

The underlines of ATG codon in the primer sequence were fixed

Table 5. Similarity matrix for 13 Vicia faba L. cultivars based on SCoT-technique data.												
Cultivars	Sa 1	Sa 3	Sa 4	Nu 3	Nu 4	Nu 5	Gi 716	Gi 843	Mi 3	R 117	R179	F19
Sa 3	0.809											
Sa 4	0.803	0.813										
Nu 3	0.797	0.783	0.893									
Nu 4	0.716	0.768	0.802	0.786								
Nu 5	0.797	0.782	0.849	0.855	0.859							
Gi 716	0.764	0.775	0.864	0.881	0.874	0.860						
Gi 843	0.711	0.737	0.762	0.793	0.767	0.780	0.821					
Mi 3	0.758	0.744	0.844	0.840	0.865	0.852	0.900	0.837				
R 117	0.693	0.719	0.722	0.741	0.725	0.764	0.757	0.758	0.809			
R179	0.761	0.759	0.805	0.800	0.824	0.824	0.828	0.807	0.865	0.826		
F19	0.748	0.775	0.772	0.755	0.765	0.753	0.823	0.773	0.815	0.781	0.833	
CV	0.736	0.680	0.761	0.817	0.714	0.755	0.748	0.723	0.778	0.782	0.795	0.814





Legend: TL represents truncated lineat the coefficient of similarity =0.795.

Resulted data comparison of discriminating capacity between ISSR and SCoT Molecular markers techniques in Table (6) were recorded. The total number of scorable ISSR amplicons was 96 with an average of 9.6 amplicons/primer, with a product size ranged from 174 and 1955 bp. While the total number of scorable SCoT amplicons was 129 with an average of 10.8 amplicons/primer, with a product size ranged from 115 and 1635 bp. Also, through better discrimination capabilities compared with ISSR, SCoT primers targeted generating 92 polymorphic amplicons with an average of 7.7/primer and 21 unique markers with an average of 1.8/primer. While ISSR primers except HB-11 targeted generating 77 polymorphic amplicons with an average of 7.7/primer and 21 unique markers with an average of 7.7/primer and 21 unique markers with an average of 7.7/primer. This discrimination capability for the SCoT technique was confirmed by UM%, DI%, and Rp% values which were 37.0, 0.23 and 3.81, respectively compared with ISSR which were 25.7%, 0.21 and 3.50, respectively. This indicates the high discriminatory potential of using SCoT primers compared with ISSR primers. Where SCoT markers were more discriminating, provided more informative data. Also, confirms that it can be relying on the SCoT techique to evaluate the genetic diversity among the faba bean cultivars better than ISSR markers. More importantly SCoT marker is generated from the functional region of the genome. So genetic analyses such as genetic diversity genotype identification, construction of linkage maps and QTL mapping using this marker would be more useful Hajibarat *et al.* (2015).

This result agrees with Abdel-Hameed et al. (2020); Bashandy et al. (2020); Ola-Ahmed and Abd EL-Aziz (2021) in Grape, Gorji et al. (2011) in potato and Etminan et al. (2016) in durum wheat. They found that the SCoT marker was more informative and effective than the ISSR marker to estimate the genetic diversity and perform fingerprinting in these plants. While this result disagrees with Ramadan et al. (2019), who found that the ISSR marker is more discriminating and provides more informative data than SCoT in fennel cultivars. While Baghizadeha and Dehghan (2018) and other researchers recommended that it is preferable to use these molecular marker techniques in combination with each other for distinctive fingerprinting. Also indicated that cluster analysis based on ISSR and SCoT data obviously discriminated among the Iranian pistachio cultivars, this was confirmed by Abd EL-Aziz et al. (2019), who showed that the combined data of ISSR and SCoT molecular marker techniques were suitable and more informative for assessing the genetic relationships and genetic diversity among apricot strains.

Table 6. Comparison of discriminating capacity between ISSR and SCoT Molecular markers techn
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Technique	MS –	SA		PA		IUA		D 0/	TIM0/	DI	Dn
recinique		Total	Mean	Total	Mean	Total	Mean	1 /0	UIVI /0	DI	кр
ISSR	174: 1955	96	9.6	77	7.7	21	2.1	70.7	25.7	0.21	3.5
ScoT	115: 1635	129	10.8	92	7.7	21	1.8	70	37	0.23	3.81

MS: Molecular size; SA: Scorable Amplicons; PA: Polymorphic Amplicons; TUA: Total unique amplicons (+&-); P%: Polymorphism %; UM%: Unique Marker %; DI: Diversity Index; Rp: Resolving power,

In Table 7 on combined data of both ISSR and SCoTtechnique the lowest value of similarity coefficient was 0.667 between Giza843 and check variety, followed by 0.695 between Sakha1 and Giza843. While the highest value of similarity coefficient was 0.894 between Misr3 and FLIP17-071FB followed by 0.883 between Nubaria4 and Giza716.

Fable 7. Similarity matrix for 13 Vicia faba L. cultivars based on combined data of ISSR and SCoT technique.													
Cultivars	Sa 1	Sa 3	Sa 4	Nu 3	Nu 4	Nu 5	Gi 716	Gi 843	Mi 3	R 117	R179	F19	
Sa 3	0.823												
Sa 4	0.820	0.824											
Nu 3	0.794	0.791	0.858										
Nu 4	0.731	0.779	0.800	0.837									
Nu 5	0.763	0.767	0.808	0.882	0.879								
Gi 716	0.731	0.771	0.826	0.854	0.883	0.863							
Gi 843	0.695	0.700	0.742	0.777	0.779	0.774	0.792						
Mi 3	0.753	0.743	0.810	0.851	0.873	0.861	0.870	0.824					
R 117	0.705	0.710	0.750	0.770	0.759	0.781	0.772	0.764	0.830				
R179	0.744	0.755	0.784	0.840	0.837	0.843	0.822	0.797	0.894	0.838			
F19	0.727	0.755	0.756	0.797	0.807	0.801	0.812	0.778	0.837	0.807	0.859		
CV	0.721	0.710	0.729	0.766	0.696	0.707	0.697	0.667	0.753	0.759	0.752	0.759	

The results of the-detected and characteristic ISSR and SCoT molecular markers of faba bean genotypes used in this investigation are mentioned in Table (8). All studied genotypes had positive and negative unique amplicons except-Nubaria3, Misr3 and FLIP17-071FB. These markers were 21 positive unique and 21 negative unique amplicons. Where Nubaria5 was characterized by the highest number of positive unique amplicons (6), while Sakha1 was characterized by the highest number of negative unique markers which was also 6 amplicons. However, the Check variety was characterized by the highest number of total unique markers with a total of 9 amplicons (4 positives and 5 negatives).

 Table 8. Detected and characteristic ISSR and SCoT

 molecular markers of the studied genotypes

	Unique markers										
Inbred	D	Molecular	T	N	umber	•					
lines	Primer	size	Type –	+	-	All					
	49-A	342, 486	-								
C - 1	SCoT-6	1017,1119	-	0	6	~					
Sal	SCoT-9	827	-	0	6	6					
	SCoT-10	236	-								
a 2	SCoT-5	263	-	1	1	2					
5a 5	SCoT-9	1372	+	1	1	2					
Sa 4	SCoT-7	1447	+	1	0	1					
Nu 3	_	-		0	0	0					
NL- 4	SCoT-10	693	+	2	0	2					
INU 4	SCoT-11	328,774	+	3	0	3					
	14-A	1224	+								
NJ. 5	49-B	1095	+	6	0	6					
INU 5	HB-12	692	+	0	0	0					
	SCoT-2	242,283,1252	+								
C: 716	14-A	775	-	1	1	C					
GI / 10	49-B	340	+	1	1	Z					
	HB-13 4	409,483,810,1130	-								
	SCoT-2	115	+								
Gi 843	SCoT-6	328	-	3	5	8					
	SCoT-9	301	+								
	SCoT-14	342	+								
Mi 3	-	-		0	0	0					
D 117	49-B	942	-	1	1	r					
К 117	SCoT-6	494	+	1	1	2					
R179	-	-		0	0	0					
	HB-12	304	-								
F19	HB-14	692	+	1	2	3					
	SCoT-13	224	-								
	14-A	354,876	-								
	14-A	368	+								
	44-B	669	-								
CV	44-B	282,733	+	4	5	9					
	HB-15	373	+								
	SCoT-6	377	-								
	SCoT-8	1090	-								

Comparative analysis based on ISSR and SCoT combined molecular data

Comparative analysis of genotypes based on combined molecular data of more than one molecular technique enables the determination of proximity or distance between genotypes with more accurately and clearly (Abd EL-Aziz et al., 2019). A dendrogram for the genetic relationship among genotypes was carried out as Fig. 7 which separated them into four major groups at a similarity coefficient =0.790. The first group included check variety only while the second group included sakha1, sakha3 and sakha4, the third group included only Giza83. The last group was divided into two subgroups, the first subgroup included (FLIP17-066FB) only, and the second subgroup includes the other genotypes and was divided into two sub-subgroups. The first sub-subgroup included only FLIP19-218FB, and the second sub-subgroup included six genotypes were Misr3, FLIP17-071FB, Nubaria4, Giza716, Nubaria3, and Nubaria5.



Fig. 7. UPGMA clustering dendrogram for 13 *Vicia faba* L. cultivars based on ISSR-SCoT combined data using Nei and Lis similarity coefficient.

Legend: TL represents truncated line at the coefficient of similarity = 0.790 using ISSR-SCoT combined data.

The PCO in Fig. 8 results reveal that the first three axes represent only 31,47% of the total variability, of these the two first coordinates accumulated 16,54% of variability and the first 14.93%. Moreover, in this analysis four main groups and an independent one, comprising check variety are observed. Scattering diagram ISSR and SCoT data were subjected to a principal component analysis (PCo) in order to obtain an alternative view of the relationships between the varieties in Fig. 7. In the two pco (F1 and F2), in general all the accessions of faba bean were classified into four groups of the plot from pco. The scales of pco scattering diagram constructed from the data, four groups are distinguishable from the dendrogram which was confirmed also by pco diagram, group one included 3 genotypes (Giza843, Misr3 and Nubaria4), group two included 4 genotypes (FLIP17-066FB, FLIP17-071FB, FLIP19-218FB and check variety), group three included 2 genotypes (Giza716 and Nubaria5) finally group four included 4 genotypes (Nubaria3, Sakha4, Sakha1 and Sakha3).



Fig. 8. PCo scattering diagram for 13 Vicia faba L. cultivars based on dice dissimilarity index for combined data.

CONCLUSION

Two molecular markers ISSR and SCoT were used to analyze the relationship among thirteen cultivars of *Vicia faba L*. It was discovered that the SCoT markers are effective than ISSR for differentiating and identifying the cultivars of faba bean under study. In summary, SCoT markers successfully evaluated the genetic relationships among the *Vicia faba* genotypes used and generated a high level of RP. The 13 genotypes were clustered into four groups using the UPGMA dendrogram, crosses between divergent genotypes out of these groups may lead to the development of effective breeding strategies in the future. The results of this study will aid in managing the germplasm, improving present breeding methods, and introducing new cultivars.

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REFERENCES

- Abd El-Aziz, M. H.; S. Y. Mohamed and Hadeer E. Magwaid (2019). Molecular and phytochemical assessment for some seedy strains of Alamar apricot rootstock under salinity stress. Egyptian Journal of Basic and Applied Sciences, 6 (1): 173– 186.
- Abdel-Hameed, U. K.; K. Abdelaziz and Nahla El-Sherif (2020). Genetic diversity of grapevine (vitis vinifera l.) Cultivars in almadinah al-munawara based on molecular markers and morphological traits. Bangladesh J. Plant Taxon. 27(1): 113– 127.

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- Abdel-Razzak, H.S.; Alfrmawy, A.M.; Ibrahim, H.M.; El-Hanafy, A.A. (2012). Genetic Diversity in Faba Bean (*Vicia faba* L.) Using Inter-Simple Sequence Repeat (ISSR) Markers and Protein Analysis. Life Sci. J., 9, 497–503.
- Abid, G.; Mingeot, D.; Udupa, S.M.; Muhovski, Y.; Watillon, B; et al. (2015). Genetic Relationship and Diversity Analysis of Faba Bean (*Vicia faba* L. Var. Minor) Genetic Resources Using Morphological and Microsatellite Molecular Markers. Plant Mol. Biol. Rep., 33, 1755–1767.
- Adhikari,S.; S. Saha; T. K. bandyopadhyay and P. Ghosh (2015). Efficiency of ISSR marker for characterization of cymbopogon gemplasm and their suitability in molecular barcoding. Plant systematic and Evalouation, 301:439-450.
- Asfaw, B.M.; Dagne, K.;Wakayo, G.K.; Kemal, S.A.; Muleta, K.T. (2018). Genetic Diversity Study of Ethiopian Faba Bean (Vicia faba L.) Varieties Based on Phenotypic Traits and Inter Simple Sequence Repeat (ISSR) Markers. Afr. J. Biotechnol., 17, 433–446.
- Aswathy, L.; R.S. Jisha and V. H. Masand (2017). Computational strategies to antimalarial thiazine alkaloid lead compounds based on an Australian marine sponge *Plakortislita*. J Biomol Struct Dyn., 35(11): 2407-2429.
- Baghizadeha, A. And E. Dehghanb (2018). Efficacy of SCoT and ISSR markers in assessment of genetic diversity in some Iranian pistachio (Pistaciavera L.) cultivars. Pistachio and Health Journal, 1(1): 37-43
- Bashandy, T.; S. Kamel and H. Ferweez (2020). Evaluation of Yield, Fruit Quality and Molecular Diversity for Three Grape Cultivars under New Valley Conditions. J of Agricultural Chemistry and Biotechnology, Mansoura Univ.,11 (7): 229-233.
- Bhattacharyya, P.; Kumaria, S.; Kumar, S.; Tandon, P. (2013). Start Codon Targeted (SCoT) Marker Reveals Genetic Diversity of Dendrobium Nobile Lindl., an Endangered Medicinal Orchid Species. Gene, 529, 21–26.
- Collard, B.C.Y. and Mackill, D.J. (2009). Start Codon Targeted (SCoT) Polymorphism: A Simple, Novel DNA Marker Technique for Generating Gene-Targeted Markers in Plants. Plant Mol. Biol. Rep., 27, 86–93
- Dawod, Kh. M. and M. J. Al-Layla (2008). Genotype-environment instruction in Durum Wheat.
- Etminan, A.; A. Pour-Aboughadareh; R. Mohammadi; A. Ahmadi-Rad; A Noori; Z. Mahdavian and Z. Moradi (2016). Applicability of start codon targeted (SCoT) and inter-simple sequence repeat (ISSR) markers for genetic diversity analysis in durum wheat genotypes. Biotechnology & Equipment, 30 (6):1075-1081.
- Fathi, M.A.; SH. M. Hussein and S.Y. Mohamed (2013). Horticultural and molecular genetic evaluation of some peach selected strains cultivated under kalubiah governorate conditions, 9(1s):12-23.
- Gorji, A. M.; P. Poczai; Z. Polgar and J. Taller (2011). Efficiency of Arbitrarily Amplified Dominant Markers (SCoT, ISSR and RAPD) for Diagnostic Fingerprinting in Tetraploid Potato. Am. J. Pot. Res., 88:226–237.

- Gupta, D.D.; Hui, P.K. and Tag, H. (2018). Genotypic variation in Acmella paniculata across different phytogeographical ranges of Northeast India inferred through ISSR & SCoT based markers. J. Appl. Res. Med. Aromat. Plants, 11:3–11.
- Hajibarat, Z; A, Saidi; Z. Hajibarat and R. Talebi (2015). Characterization of genetic diversity in chickpea using ISSR markers, Start Codon Targeted polymorphism (SCoT) and conserved DN- Derived polymorphism (CDDP). Physiology and Molecular Biology of plants, 21 (3):365-373.
- Hamada, M.S.; M. H. Abd El-Aziz and Norhan M. Sharshera (2019). Generation mean analysis for some selected crosses based on molecular distances estimated using ISSR and RAPD techniques in squash 9th International Conference for Sustainable Agricultural Development 4-6 March, Fayoum J. Agric. Res, & Dev., 33 (1) 495- 510.
- Hanelt P, Mettin D (1989). Biosystematics of the genus Vicia L. (Leguminosae). Annu. Rev. Ecol. Syst. 20:199–223.
- Joshi, C. P.; H. Zhou; X. Huang and V. L.Chiang (1997). Context sequences of translation initiation codon in plants. Plant Mol. Biol., 35: 993–1001.
- Mao, R.; Xia, P.; J. Liu; X. Li; R. Han; F. Liu; H. Zhao and Z. Liang (2018). Genetic diversity and population structure assessment of Chinese Senna obtusifolia L. by molecular markers and morphological traits of seed. Acta Physiol. Plant. 40.
- Nei, M. and W. H. Li (1979). Mathematical model for studing genetic variation in terms of restriction endonucleases. Proc. Natl. Acad .Sci. USA, 76: 5269- 5273.
- Ola A. Ahmed and M. H. Abd EL-Aziz (2021). Description and Evaluation of some Newly Introduced Grape Cultivars Under Egyptian Conditions. Journal of Agricultural Chemistry and Biotechnology, 12(7):127-136, 2021.
- Ozturk, H.I.; V. Donderalp; H. Bulut and R. Korkut (2022). Morphological and molecular characterization of some pumpkin (*Cucurbita pepo* L.) genotypes collected from Erzincan province of Turkey. Sci Rep. 26;12(1):6814-6824.
- Prevost, A. and M.J. Wilkinson (1999). A new system of comparing PCR primersapplied to ISSR fingerprinting of potato cultivars. *TAG Theoretical and Applied Genetics*, 98:107-112.
- Ramadan, Walaa A.; R. M. Shoaib, Rania T. Ali and N. S. Abdel-Samea (2019). Assessment of genetic diversity among some fennel cultivars (Foeniculum vulgare Mill.) by ISSR and SCoT Markers. African J. Biol. Sci., 15 (1): 219-234.
- Sawant, S.V.; P. K. Singhl; S. K. Gupta; R. Madnala and R. Tuli (1999).Conserved nucleotide sequences in highly expressed genes in plants. J. Genet., 78:123-131.
- Singh, R. K and B. D. Chaudhary (1985). Biometrical methods in quantitative genetic analysis. Kalyani Publishers, New Delhi. pp. 102-157.
- Terzopoulos, P.J.; Bebeli, P.J. (2018). Genetic Diversity Analysis of Mediterranean Faba Bean (*Vicia faba* L.) with ISSR Markers. Field Crop. Res., 108, 39–44.
- Xiong, F.Q.; R.C. Zhong; Z.Q.Han and J.Jiang (2011). Start codon targeted polymorphism for evaluation of functional genetic variation and relationships in cultivated peanut (*ArachishypogaeaL.*) genotypes. *Mol. Biol. Rep*, 38: 3487-3494.

التنوع الوراثي بين بعض الأصناف الأبوبة للفول البلدي باستخدام تقنيات العلامات الجزيئية ISSR و SCoT

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الملخص

نفذت هذه الدراسة لتقيم الاختلافات الوراثية الجزيئية بين 13 صنف أو تركيب وراثى من الفول البلدى من مصر وكردستان العراق لاستخدامها في برامج تربية المحاصيل في كردستان. وتم الكشف عن العلاقات الوراثية بين هذه التراكيب باستخدام كل من التقية الجزيئية SCOT و SCOT حيث نجحت عشرة واثتي عشر من البلائات على الترتيب في ابتاج متضخمات (أمبليكونات) متباينة قابلة للتكرار وكانت مناسبة لدراسة التباين الوراثي بين التراكيب الوراثية تحت العراق (عنه بين هذه التراكيب باستخدام كل من التقية الجزيئية SCOT و SCOT حيث نجحت عشرة واثتي عشر من البلائات على الترتيب في ابتاج متضخمات (أمبليكونات) متباينة قابلة للتكرار وكانت مناسبة لدراسة التباين الوراثى بين التراكيب الوراثية تحت الدراسة. وتم الكشف عن 225 متضخم (77 لـ SCR و 29 لـSOT) بنسبة تباين 70.71 و 70.03 و 70.03 و 70.03 و 70.03 و 70.03 و 70.05 و 70.03 و 20.05 على التوالي) أعلى مقارنة بتقنية SSR توعرفترة منيزية (70.05 و 20.03 و 3.81 على التوالي) أعلى مقارنة بتقنية ISSR (70.05 على التوالي) العلى مقارنة بتقنية ISSR (تراكيب الوراثية وعبرا لتنوع عدم التقلي) أعلى مقارنة بتقنية SSR عن 20.05 (عليان الوراثي العرابية) على التراك على التوالي) أعلى مقارنة بتقنية SSR على التوالي. واظهرت التتلج ان تقنية SOT كن أكثر تميزًا بنسبة علامات منفرة على اليتاك المحاصيل في (70.05 على التوالي) أعلى مقارنة بتقنية SSR على التراكيب الوراثية المعنوبية الأسلسى بالإعتماد على البيات المدموم و 20.05 على التوالي) أعلى مقارنة بتقنية SSR (70.05 و 20.05 على التوالي) أعلى مقارنة بنقية SSR التراكيب الور اثية المدروسة إلى أربع مجموعات رئيسية المجموعة الأولى تضمنت الصنف المحلي التنائية تضمنت التراكيب الوراثية المدروسة إلى أربع مجموعات رئيسية المجموعة الأولى تضمنت الصنف المحرفي التوالي في الوراثية المتراكيب الوراثية المدروسة إلى أربع مجموعات منيسية المجموع التراكيب الوراثية الأخرى ما في نفل معربيات المحول عن عربينا مالمي على التوالي في المدروسة إلى ألمن من عمن عمليات الصنف المحلي الخبيرة من كردستان وم التراكيب الوراثي المدروسة إلى التي المدروسة إلى ألتي معمليات الموراثي المدروسة المدروسة على ألمن المدوسة عمرة المدروسة المدروسة المدروسة المدروسة معربيان معملي المرائية الكبير مع ماليت المدروسة المدروسة ومالية تضمنعة ميزة معرات مالمد ما