

Clinicopathological Studies on Neem and Ginger Effects as Feed Additives in Normal and *E. coli* Infected Weaned Rabbits

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Abstract

The present study was performed to investigate the clinicopathological effects of neem and ginger as feed additives. One hundred and twenty weaned White New Zealand rabbits were divided into 6 equal groups. Group (1) kept as control. Groups (2 and 5) received ration contained neem leaves daily (5% of diet). Groups (3 and 6) received ration contained ginger powder daily (2% of diet). Groups (4, 5 and 6) were experimentally infected by *E. coli* (O103 once orally with a dose of 3 ml of suspension containing 3×10^7 CFU at the end of the 4th week of experiment. The results revealed normal parameters in none infected as well as neem and ginger treated groups (1, 2 and 3). However a significant decrease in the serum total protein, albumin, globulin and catalase levels (CAT) on the 1st, 3rd and 15th day PI was observed in infected non treated animals (Group 4). On the other hand a significant increase in alanine aminotransferase (ALT), aspartate aminotransferase (AST), alkaline phosphatase (ALP), malondialdehyde level (MDA), phagocytic percentage and phagocytic index were observed in the same group. In Groups 5 and 6; animals showed a significant increase in total protein, albumin, globulin, CAT and a significant decrease in the other parameters comparing with the infected group. It could be concluded that both neem and ginger can be used as feed additives in rabbit ration to enhance hepatic, renal and antioxidant activities beside cell mediated immunity. Moreover, Neem was better than ginger in amelioration of the harmful effects of *E. coli* infection.

Keywords: Antibacterial, Immunostimulant, Neem, Ginger, Antioxidant

Introduction

There is a worldwide trend towards the use of natural additives in food as spices and herbal medicine. Neem and ginger are important members of herbal medicine [1,2]. Both of them have compounds of many biological activities as hepato and renal protective effects in addition to anti-oxidant and immunostimulant properties [2,3].

The effect of dietary neem leaves meal on serum biochemical parameters was investigated by Obikaonu *et al.* [4]. The AST, ALT and ALP levels were decreasing by increasing neem leaves percent in diet comparing with the control. Mukherjee *et al.* [5] found that the administration of oral mixture of leaves and seed extract at 10% concentration for 6 weeks in rodent caused

specific activation of T-lymphocytes and phagocytic cells. Biswas *et al.* [1] found an increase in catalase activity (CAT) while malondialdehyde level (MDA) was decreased. Lebda *et al.* [6] studied the effect of different ginger treatments on hepatic oxidative stressed male New Zealand rabbits. The results revealed that AST, ALT, ALP and urea were decreased with increased creatinine level comparing with control. Onu and Aja [7] recorded a significantly decreased MDA level with increased CAT activity using ginger (0.25% of diet) on weaned rabbits as supplement for 10 weeks. Oral ginger administration as feed additives stimulated phagocytic activity [8].

Escherichia coli is one of the most important etiological agents of enteritis and losses in rabbit industry [9]. The most pathogenic

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serotypes inducing high mortalities are (O109:H2), with neonatal diarrhea, (O103:H2) weaning diarrhea, O123 and O132 [10, 11].

This work aimed to study hepatic and renal protective effects as well as antioxidants and immunological effects of neem leaves and ginger rhizomes on normal and *E. coli* experimentally infected rabbits.

Material and Methods

Experimental Design

One hundred and twenty weaned white New Zealand rabbits apparently healthy were obtained from a private farm near Menia Elkameh, Sharkia Governorate. The basal and balanced growing ration was given to animals. The rabbits were divided into 6 equal groups. Group 1 was kept as control. Groups (2 and 5) received neem leaves powder as 5% of diet [12]. Groups (3 and 6) received ginger powder 2% of diet daily for 6 weeks [6]. Groups (4, 5 and 6) were experimentally infected rabbits by *E. coli* with of O103 strain once orally with a dose of 3 ml of suspension containing 3×10^7 CFU [13] at the end of the 4th week of experiment. The animals were observed for clinical examination daily along the experiment.

Feed additives

Neem, fresh matured neem leaves were harvested from garden in El-Asher Men Ramadan City. The samples were identified in Botany Department, Faculty of Science, Zagazig University. The leaves were cleaned, dried in shaded place and powdered. The powder was added to ration as 5% of diet [12].

Ginger rhizomes were collected from local markets as dry rhizomes then were ground well till became fine powder and added to ration as 2 % of diet [6].

Pathogenic *E. coli* O103 was obtained from Animal Health Research Institute, Dokki, Giza, Egypt and was used for infection.

Blood samples

Blood samples were collected from ear vein on the 1st, 3rd and 15th day PI. The first sample was collected on heparin for phagocytic activity determination [13] and the second one was

collected for serum biochemical analysis AST, ALT [14] ALP [15] total protein [16] albumin [17] globulin was calculated, urea [18] and creatinine [19].

Antioxidant activities

Catalase activities and MDA levels were calorimetrically determined using Kits of Bio Diagnostics [20].

Phagocytic activity

The Phagocytic percent and index were evaluated according to Wilkinson [21]. The phagocytic percent is the total number of phagocytic cells at any stage of phagocytosis in 100 mononuclear phagocytic cells. Phagocytic index (PI) is the number of *C. Albicans* ingested by 100 phagocytic cells.

Statistical analysis

The obtained data were statistically analyzed by F-test [22] using MSTAT-C computer program.

Results and Discussion

Neem and ginger are important members of herbal medicine [1, 2]. Both of them have biological activities as hepato and renal protective in addition to anti-oxidant and immunostimulant properties [2, 3].

The use of neem or ginger as feed additives in rabbit ration with a percent of 5% and 2% respectively were investigated for their beneficial impact on rabbit biochemical and immunological properties as well as their antibacterial efficacy.

Our results revealed that Groups (2 and 3) treated non infected animals showed no significant changes in AST, ALT and ALP hepatic enzymes all over the experimental periods compared to control group (Tables 1, 2). This may be due to that neem leaves meal or ginger are safe and has no deleterious effect on the liver and these results confirmed histopathologically where there is no pathological lesions in the hepatic tissues. These results are in agreement with Haque *et al.* [23] and Ogbuewu *et al.* [12] who stated that AST and ALT were non significantly changed in neem and ginger treated groups comparing with the control.

Table (1): Effect of Neem and Ginger on hepatic enzymes and proteinogram in infected and normal rabbits through 15 days PI (mean \pm SE)

Hepatic enzymes									
Parameter	AST (U/L)			ALT (U/L)			ALP (U/L)		
Group \ Time	1 st day	3 rd Day	15 th day	1 st day	3 rd day	15 th day	1 st day	3 rd day	15 th day
	PI	PI	PI	PI	PI	PI	PI	PI	PI
GP (1)	40.20	40.40	41.00	40.60	42.00	44.00	35.00	35.80	40.00
Control	$\pm 0.374^d$	$\pm 0.509^d$	$\pm 0.447^d$	$\pm 0.24^d$	$\pm 0.31^c$	$\pm 0.44^c$	$\pm 0.44^{de}$	$\pm 0.37^{de}$	$\pm 0.54^d$
GP (2) Neem	39.60	39.40	38.00	41.60	41.00	40.00	33.20	34.20	37.40
	$\pm 0.509^d$	$\pm 1.122^d$	$\pm 0.836^d$	$\pm 0.50^d$	$\pm 0.37^c$	$\pm 0.44^c$	$\pm 2.37^d$	$\pm 0.37^{de}$	$\pm 0.24^d$
GP (3)	40.40	39.20	39.40	38.20	40.40	41.20	35.40	35.00	39.00
Ginger	$\pm 0.244^d$	$\pm 0.37^d$	$\pm 0.244^d$	$\pm 0.37^d$	$\pm 0.40^c$	$\pm 0.37^c$	$\pm 0.24^{de}$	$\pm 0.31^{de}$	$\pm 0.31^d$
GP (4) E. coli	67.40	90.60	60.60	58.60	80.00	60.20	57.20	88.60	60.20
	$\pm 1.029^a$	$\pm 0.871^a$	$\pm 1.691^a$	$\pm 1.02^a$	$\pm 0.89^a$	$\pm 1.28^a$	$\pm 1.11^c$	$\pm 0.60^a$	$\pm 1.31^c$
GP (5) Neem + E. coli	50.60	68.20	44.20	49.00	64.40	48.50	48.20	76.20	49.40
	$\pm 0.40^c$	$\pm 0.86^c$	$\pm 0.969^c$	$\pm 0.44^b$	$\pm 1.12^b$	$\pm 0.50^b$	$\pm 8.06^d$	$\pm 0.58^b$	$\pm 1.4^d$
GP (6)	55.60	74.80	50.20	45.80	60.20	50.10	52.4	78.2	51.6
Ginger + E.coli	$\pm 1.50^b$	$\pm 0.91^b$	$\pm 0.583^b$	$\pm 1.8^c$	$\pm 1.28^b$	$\pm 1.31^b$	$\pm 206^{cd}$	$\pm 0.91^b$	$\pm 0.24^{cd}$
F-test	*	*	*	*	*	*	*	*	*

Proteinogram									
Parameter	Total protein (g/dL)			Albumin (g/dL)			Globulins (g/dL)		
Group \ Time	1 st day	3 rd day	15 th day	1 st day	3 rd day	15 th day	1 st day	3 rd day	15 th day
	PI	PI	day PI	PI	PI	PI	PI	PI	PI
GP (1)	6.60	6.62	6.60	3.34	3.34	3.34	3.26	3.28	3.26
Control	± 0.04	$\pm 0.05^a$	$\pm 0.06^a$	± 0.02	$\pm 0.02^a$	$\pm 0.02^a$	± 0.024	$\pm 0.03^a$	$\pm 0.04^a$
GP (2) Neem	6.58	6.62	6.76	3.34	3.34	3.40	3.24	3.28	3.36
	± 0.04	$\pm 0.03^a$	$\pm 0.02^a$	± 0.024	$\pm 0.02^a$	$\pm 0.04^a$	± 0.024	$\pm 0.02^a$	$\pm 0.02^a$
GP (3)	6.58	6.56	6.70	3.32	3.32	3.36	3.26	3.24	3.32
Ginger	± 0.03	$\pm 0.04^a$	$\pm 0.04^a$	± 0.024	$\pm 0.02^a$	$\pm 0.02^a$	± 0.05	$\pm 0.02^{ab}$	$\pm 0.03^a$
GP (4) E. coli	6.36	4.76	5.82	3.30	2.70	2.98	3.06	2.06	2.86
	± 0.15	$\pm 0.07^d$	$\pm 0.09^c$	± 0.15	$\pm 0.10^c$	$\pm 0.03^b$	± 0.18	$\pm 0.07^c$	$\pm 0.12^b$
GP (5) Neem + E. coli	6.70	6.30	6.76	3.36	2.96	3.48	3.34	3.34	3.28
	± 0.07	$\pm 0.15^b$	$\pm 0.06^a$	± 0.02	$\pm 0.02^b$	$\pm 0.04^a$	± 0.07	$\pm 0.16^a$	$\pm 0.03^a$
GP (6)	6.72	5.86	6.40	3.46	2.88	3.30	3.26	2.98	3.10
Ginger +E. coli	± 0.07	$\pm 0.07^b$	$\pm 0.08^a$	± 0.08	$\pm 0.08^b$	$\pm 0.08^a$	± 0.12	$\pm 0.13^b$	$\pm 0.03^a$
F-test	N.S	*	*	N.S.	*	*	N.S.	*	*

N.S.: non significant

*: Significant at 0.05 probability PI: post infection

However, The increase in serum ALT, AST and ALP activities was observed in group (4) which may be attributed to the damaging effect of E. coli (lipopolysaccharide) on the liver and increased cell membrane permeability [24-25]. This confirmed pathologically (Figure 1A and B) which showed severe congestion of the hepatic blood vessels, hydropic degeneration and coagulative necrosis of the hepatocytes with pyknotic and karyolytic nuclei. The significant decrease in the hepatic enzymes in groups (5 and 6)

comparing with group (4) may be due to administration of neem and ginger which contain hepatoprotective and antibacterial active principals [26,27]. This was confirmed pathologically (Figure 1C) and was manifested by as reduced degenerative changes in hepatocytes. Ginger has hepatoprotective effect and repair the damaged hepatic tissue [28,29]. Decreasing in proteinogram in group (4) may be due to E coli effects on liver and kidney [9]. Serum total protein, albumin and globulin levels were increased in groups (5

and 6) on the (3rd and 15th) day post infection comparing with infected group [29,30].

A significant increase in urea and creatinine (Table 2) was found in group (4). *E. coli* infection (lipopolysaccharide) caused damage in renal tissue [9,25,31]. This was confirmed pathologically (Figure 1D) with multiple cystic dilatations of renal tubules with hyaline and cellular casts. Renal protective and antibacterial effects of neem and ginger ameliorated the elevation in urea and creatinine levels in groups 5 and 6. Similar results were obtained previously by Iebda *et al.* [6] and Ezz-Din *et al.* [32]. Also this may be due to polyphenols and flavonoids present in ginger that remove waste products from plasma [6].

In respect to antioxidant activity, Groups (2 and 3) showed a significant increase in CAT and a significant decrease in MDA on the 1st, 3rd and 15th day PI (Table 3). It could be explained by the antioxidant effects of neem that may increase the synthesis of antioxidant molecules as mentioned previously by Dhillon *et al.* [33]. Also, this may be due to ginger contains antioxidants that act as a free radical scavenger. A significant decrease in CAT and increase MDA was recorded in group (4) which may be attributed to lipopolysaccharide [34,35]. Groups (5 and 6) showed a significant increase in CAT and decrease in MDA level in comparison to infected group. This may be attributed to scavenging effects of neem [33, 36] and ginger [37] among free radicals and so relieving oxidative stress of *E. coli* infection.

Groups (2, 3, 5 and 6) showed a significant increase in phagocytic percent and index all over the experimental period (Table 4). This may be due to neem increase nonspecific immune response [36,38]. Ginger stimulated phagocytic activity as described by Ajith *et al.*, and Mallikarjuna *et al.* [39,40]. Phagocytic percent and index were significantly increased in group (4). This may be referred to immune response against *E. coli* as explained by Eisa and Alam [41, 42]. Leukocytosis, neutrophilia and monocytosis were found in groups 4, 5, and 6 on 1st day PI. Neutrophilia and monocytosis were still present on 3rd day. Lymphopenia was found in the same groups on the 1st and 3rd day PI. These changes may be due to the acute defense mechanism against bacterial infection as recorded by Edrees *et al.*, and El-Boushy *et al.* [9,25]. A significant decrease in TLC was found in group 4 on the 3rd and 15th day PI. The changes in leukogram may be due to immunodepressant effect of *E. coli* infection as stated by many authors [31,41,43,44].

Conclusion

It could be concluded that neem (5% of diet) and ginger (2% of diet) enhanced hepatic, renal and antioxidant activities of rabbits. Also, they acted as immune stimulant and ameliorated the harmful effects of *E. coli* infection. Moreover, neem effects were stronger than ginger effects.

Table (2): Effect of Neem and Ginger on Serum urea and creatinine in infected and normal rabbits through 15 days PI (mean ± SE)

Parameter	Time	Serum urea (mg/dL)			Serum creatinine (mg/dL)		
		1 st day PI	3 rd day PI	15 th day PI	1 st day PI	3 rd day PI	15 th day PI
GP (1) Control		17.30 ± 0.104 ^b	17.17 ± 0.100 ^d	17.63 ± 0.163 ^c	1.26 ± 0.014 ^b	1.24 ± 0.009 ^d	1.27 ± 0.011 ^d
GP (2) Neem		17.16 ± 0.094 ^b	17.68 ± 0.554 ^d	17.39 ± 0.140 ^c	1.25 ± 0.005 ^b	1.27 ± 0.006 ^{cd}	1.28 ± 0.0096 ^{cd}
GP (3) Ginger		17.36 ± 0.066 ^b	17.30 ± 0.074 ^d	17.88 ± 0.112 ^c	1.28 ± 0.008 ^b	1.30 ± 0.007 ^{cd}	1.30 ± 0.009 ^{cd}
GP (4) E. coli		19.0 ± 0.447 ^a	26.80 ± 0.622 ^a	24.60 ± 0.484 ^a	1.46 ± 0.050 ^a	1.88 ± 0.014 ^a	1.42 ± 0.031 ^a
GP (5) Neem + E. coli		17.39 ± 0.191 ^b	19.75 ± 0.139 ^c	17.40 ± 0.155 ^c	1.25 ± 0.007 ^b	1.67 ± 0.021 ^b	1.34 ± 0.008 ^b
GP (6) Ginger + E. coli		17.88 ± 0.333 ^b	21.89 ± 0.150 ^b	18.78 ± 0.092 ^{bc}	1.28 ± 0.008 ^b	1.70 ± 0.020 ^b	1.37 ± 0.010 ^b
F-test		*	*	*	*	*	*

*=significant at 0.05 probability. PI: post infection

Table (3): Effect of Neem and Ginger on antioxidant activities of liver tissues of infected and normal rabbits Through 15 days PI (mean ± SE)

Parameter	Time	Catalase (Mu/mg)			MDA (nmol/mg)		
		1 st day PI	3 rd day PI	15 th Day PI	1 st day PI	3 rd day PI	15 th Day PI
GP (1) Control		1.36 ± 0.058 ^b	1.36 ± 0.066 ^b	1.37 ± 0.059 ^b	1.53 ± 0.008 ^d	1.53 ± 0.007 ^d	1.42 ± 0.007 ^b
GP (2) Neem		1.53 ± 0.056 ^a	1.60 ± 0.046 ^a	1.80 ± 0.038 ^a	1.35 ± 0.006 ^e	1.30 ± 0.005 ^f	1.00 ± 0.20 ^c
GP (3) Ginger		1.49 ± 0.032 ^a	1.50 ± 0.023 ^a	1.70 ± 0.049 ^a	1.40 ± 0.005 ^f	1.35 ± 0.006 ^f	1.10 ± 0.007 ^c
GP (4) E. coli		0.78 ± 0.048 ^d	0.71 ± 0.031 ^d	1.206 ± 0.062 ^c	1.93 ± 0.020 ^a	2.20 ± 0.36 ^a	1.78 ± 0.008 ^a
GP (5) Neem + E. coli		1.13 ± 0.050 ^c	1.178 ± 0.018 ^c	1.35 ± 0.024 ^b	1.75 ± 0.012 ^{bc}	1.90 ± 0.003 ^{bc}	1.44 ± 0.006 ^b
GP (6) Ginger + E. coli		1.10 ± 0.007 ^c	1.134 ± 0.017 ^c	1.33 ± 0.024 ^b	1.81 ± 0.06 ^b	1.73 ± 0.006 ^{bc}	1.45 ± 0.010 ^b
F-test		*	*	*	*	*	*

*=significant at 0.05 probability. PI: post infection

Table (4): Effect of Neem and Ginger on phagocytic % and phagocytic index in infected and normal rabbits through 15 days PI (mean ± SE)

Parameter	Time	Phagocytic %			Phagocytic index		
		1 st day PI	3 rd day PI	15 th day PI	1 st day PI	3 rd day PI	15 th day PI
GP (1) Control		63.04 ± 0.09 ^f	62.12 ± 0.14 ^d	64.18 ± 0.177 ^b	2.38 ± 0.037 ^e	2.46 ± 0.050 ^e	2.46 ± 0.050 ^c
GP (2) Neem		70.44 ± 0.15 ^{cd}	70.26 ± 0.18 ^c	73.24 ± 0.172 ^a	3.06 ± 0.092 ^c	3.09 ± 0.060 ^c	3.55 ± 0.063 ^a
GP (3) Ginger		65.92 ± 0.23 ^e	70.94 ± 0.21 ^c	71.18 ± .139 ^a	2.56 ± 0.050 ^d	2.66 ± 0.092 ^d	2.74 ± 0.067 ^b
GP (4) <i>E. coli</i>		80.60 ± 0.68 ^b	74.34 ± 0.22 ^{ab}	62.70 ± 0.234 ^c	2.86 ± 0.050 ^c	3.46 ± 0.074 ^b	2.40 ± 0.141 ^b
GP (5) N + <i>E. coli</i>		86.66 ± 0.20 ^a	75.10 ± 0.24 ^a	64.10 ± 0.264 ^b	4.62 ± 0.073 ^a	3.82 ± 0.058 ^a	3.02 ± 0.058 ^a
GP (6) G + <i>E. coli</i>		83.64 ± 0.22 ^b	74.44 ± 0.24 ^a	63.14 ± 0.163 ^b	4.34 ± 0.050 ^b	3.58 ± 0.037 ^b	2.69 ± 0.124 ^b
F-test		*	*	*	*	*	*

C: Control, N: Neem, G: Ginger

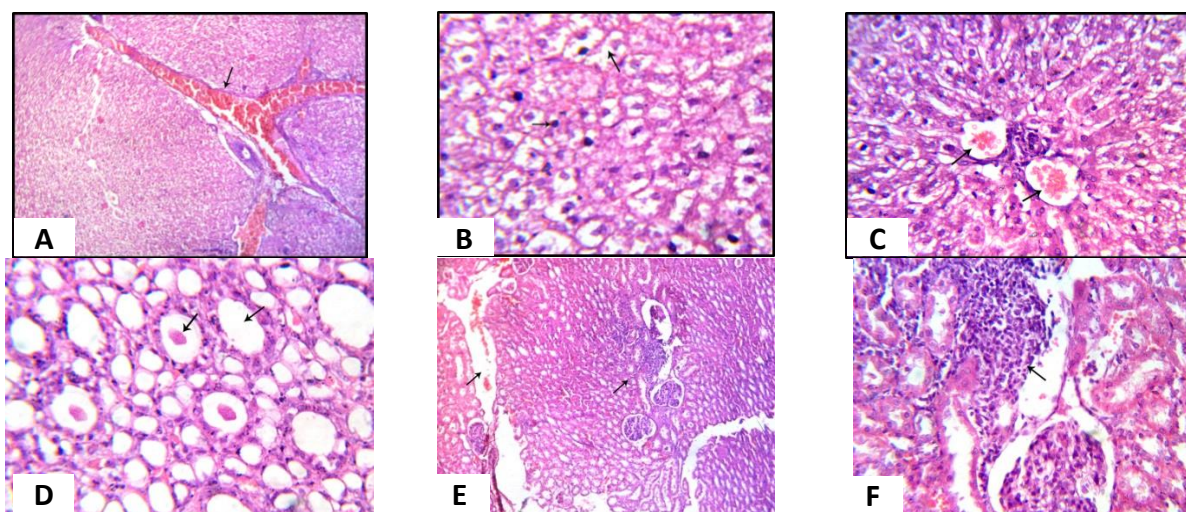


Figure 1: A) Liver (Group 4), 3 days PI showing severe congestion of the hepatic blood vessels (H&Ex130); B) Liver (Group 4), 3 days PI showing severe hydropic degeneration and coagulative necrosis of the hepatocytes with pyknotic and karyolytic nuclei (H&E x520); C) Liver (Group 5), on the 3rd day PI, showing congestion of the hepatic blood vessels together with mononuclear leukocytic cells infiltration (H&E x520); D) Kidney (Group 4), 3 days PI showing many cystic dilatations of some renal tubules with hyaline and cellular casts inside it (H&E x520); E) Kidney (Group 5), 3 days PI showing mild congestion of renal blood vessels in the renal cortex together with mononuclear leukocytic cell infiltration (H&E x300); F) Kidney (Group 6), 3 days PI showing mononuclear leukocytic cells infiltrating the renal cortex (H&E x 520).

Conflict of interest

None of the authors have any conflict of interest to declare.

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References

[1] Biswas, K.; Chattopadhyaya, I.; Banerjee, R.K. and Bandyopadhyay, U. (2002):

Biological activities and medicinal properties of Neem (*Azadirachta indica*). *Current Science J.*, 82 (11): 1336-1345.

[2] Ahmad, N.; Suhaniza, S.; or, A.; Nor, A.M.; Noor, A.A. and Yasmin, A.M. (2006): Effects of ginger extract on antioxidant status of hepatocarcinoma induced. *Malaysian Bioc. and Mol. Bio. J.*, 14 (1): 7-12.

[3] Joshi, R.N.; Parakh, S.R. and Sampada, B. (2006): Neem: A Tree for Solving Global Problems. *Pharma Info. Net.*

<http://www.pharmainfo.net/reviews/neem-tree-solving-global-problems>.

- [4] Obikaonu HO, Okoli IC, Opara MN, Okoro VMO., Ogbuewu IP, Etuk EB and Udedibie ABJ (2012): Haematological and serum biochemical indices of starter broilers fed leaf meal of neem. *Agric. Tech. J.*, 8 (1): 71-79
- [5] Mukherjee S, Garg S and Talwar GP (1999): Early post implantation contraceptive effects of a purified fraction of neem seeds given orally in rats: possible mechanisms involved. *Ethanopharmacol. J.*, 67 (3): 287-296.
- [6] Lebda, M.A.; Taha, N.M.; Korshom, M.A.; Mandour, A.E.A. and El-Morshedy, A.M. (2012): Biochemical effect of ginger on some blood and liver parameters in male New Zealand rabbits. *Journal of Animal and Feed Research*, 2 (2): 197-202.
- [7] Onu PN and Aja PM (2011): Growth performance and hematological indices of weaned rabbits fed garlic and ginger supplemented diets. *F. Inter. J.*, 1 (1): 51-59.
- [8] Chakraborty B and Sengupta M (2012): Boosting of non specific host response by aromatic spices turmeric and ginger in immunocompromised mice. *Cell immunol.* 280 (1): 92-100.
- [9] Edrees Nariman, M.M.; Hashim, M.A. and Alam, R.T. (2008): Clinicopathological studies on the effect of some bacterial infection in rabbits. *Zag. Vet. J.*, 36 (2): 199-206.
- [10] Peeters JE, Geeroms R and Orskov F (1988): Biotypes, serotypes and pathogenicity of attaching and effacing enteropathogenic *Escherichia coli* strains isolated from diarrhetic commercial rabbits. *Infect. Immun. J.*, 56: 1442-1448.
- [11] Camguilhem R and Milon A (1989): Biotypes and serogroups of *Escherichia coli* involved in intestinal infections of weaned rabbits. *Clin. Microbiol. J.*, 27: 743-747.
- [12] Ogbuewu, I.P.; Kadurumba, O.E.; Okoli, I.C. and Iloeje, M.U. (2010): Evaluation of toxicological effects of leaf meal of an ethanomedicinal plant-neem on blood chemistry of puberal chinchilla rabbit does. *Rep. Opi. J.*, 2 (2):29-34.
- [13] Feldman, B.F.; Zinkl, J.G. and Jain, N.C. (2000): *Schalm's Veterinary Hematology*. 5th ed., Lippincott Williams and Wilkins .A Wolters Company. Philadelphia, Baltimore, New York, London, Buenos Aires, Hong Kong, Sydney, Tokyo.
- [14] Reitman, S. and Frankel, S. (1957): Colorimetric method for the determination of serum transaminase activity. *Am. J. Clin. Pathol.*, 28: 56-58.
- [15] Tietz NW (1995): *Clinical Guide to Laboratory Tests*. 3rded Philadelphia; W.B. Saunders.
- [16] Grant, G.H.; Sliverman, L.M. and Christenson, R.H. (1987): *Amino Acids and Protein in: Fundamental of Clinical Chemistry*. 3rd ed. Philadelphia, W.B. Saunders Company.
- [17] Doumas, B.T.; Bayso, D.D.; Carler, R.J.; Peler, T. and Schaffer, R. (1981): Determination of serum albumin. *Clin. Chem.*, 27:1632
- [18] Henry, T.J. (1974): Determination of serum creatinine *Clin. Chem. Principles and Techniques*. 2nd ed. Harper and Row publishers New York.
- [19] Tietz, N.W. (1995): *Clinical Guide to Laboratory Tests*, 3rd ed Philadelphia; W.B. Saunders.
- [20] Packer, L and Glazer, AN (1990): *Method in Enzymology*. Vol. 186 Part B, Academic Press Inc. New York.
- [21] Wilkinson, P.C. (1976): Recognition and response in mononuclear and granular phagocytes. *Clin. Exp. Immunol.*, 25 (3): 355-366.
- [22] Tamhane, A.C. and Dunlop, D.D. (2000): *Statistics and Data Analysis from Elementary to Intermediate*. Upper Saddle River, USA

- [23] Haque, E.; Mandal, I.; Pal S. and Baral, R. (2006): Prophylactic dose of neem leaf preparation restricting murine tumor growth in non-toxic hematostimulatory and immunostimulatory. *Immunopharmacol. Immunotoxicol.*, 28 (1): 33-50
- [24] Smith H and Pearce JH (1972): Microbial pathogenicity in man and animals. Cited in: Eisa A.M. (1998): Clinicopathological studies on some anti-diarrheal drugs in rabbits. M.V.Sc., Thesis (Clinical pathology) Faculty of Veterinary Medicine, Zagazig University.
- [25] El-Boushy, M.E.; Ramdan, T.M. and Hala, N.I. (2005): Hematological, biochemical and pathological studies on colibacillosis in rabbits. 4th Int. Sci. Conf., Mansoura University
- [26] Chattopadhyay, R.R.; Bandy, O. and Padhyay, A. (1992): Effect of *Azadirachta indica* leaf extract on normal and paracetamol treated rats. *Bio. Res. Afr. J.*, 8:101-104.
- [27] Koul, A.; Mukherjee, N. and Gangar, S.C. (2006a): Inhibitory effects of *Azadirachta indica* on DMBA-induced skin carcinogenesis in Balb/c mice. *Mol. Cell Biochem.*, 283(1-2):47-55.
- [28] Al-Attar, A.M. and Zari, T.A. (2007): Modulatory Effects of Ginger and clove oils on physiological Responses in streptozotocin-induced Diabetic Rats. *Internet. J. Pharmacol.*, 3 (1): 34-40.
- [29] Koshalk, M.Y.; Edrees, N.E.; El-Nabtity, S.M. and Abd El-Latief, S.A. (2008): Hepatoprotective Effects of some Plants. 9th Vet. Med. Zag. Conference, 20-22 August, Port-Said.
- [30] Aruway, A.; Maigandi, SA.; Malami, BS and Daneji, A.I. (2011): Hematological and biochemical parameters of udd lambs fed graded levels of alkali-treated neem kernel cake. *Nigerian Journal of Basic and Applied Science*, 19 (2): 277-284.
- [31] Badr, S.M.A. (2007): Clinicopathological studies on the effect of levamisole in rabbits. M.V. Sc., Thesis (Clinical Pathology), Faculty of Veterinary Medicine, Zagazig University
- [32] Ezz-Din Doaa.; Gabry, M.S.; Farrag, A.R.H. and Abdel Moneim, A.E. (2011): Physiological and histological impact of neem leaves extract in a rat model of cisplatin-induced hepato and nephrotoxicity. *Journal of Medicinal Plants Research*, 5 (23): 5499-5506.
- [33] Dkhill, M.A.; Ahmed, E. Abdel Moneim, and Saleh, Al-Quraishy (2012): Antioxidant, hepatoprotective, and ameliorative effects of *Azadirachta indica* on *Eimeria papillata* induced infection in mice. *Journal of Medicinal Plants Research*, 6 (20): 3640-3647.
- [34] Yazar, E.; Col R.; Konyalioglu, S.; Birdane, Y.O.; Elmas, M. and Bas, A.L. (2004): Effects of vitamin E and prednisolone on biochemical and hematological parameters in endotoxaemic New Zealand white rabbits. *Revue Med. Vet.*, 155 (11): 538-542.
- [35] Keskin, E.; Oztekin, E.; Col, R.; Sivrikaya, A.K. and Yazar, U.E. (2005): Effect of Pentoxifylline on antioxidant status of healthy and endotoxaemic New Zealand white rabbits. *Acta. Vet. Brno.*, 74: 17-21.
- [36] Dasgupta, T.; Banerjee, S.; Yadava, P.K. and Rao, A.R. (2004): Chemopreventive potential of *Azadirachta indica* (Neem) leaf extract in murine. *Ethnopharmacol. J.*, 92 (1): 23-36.
- [37] Manju, V. and Nalini, N. (2005): Effect of ginger on bacterial enzymes in 1,2-dimethylhydrazine induced experimental colon carcinogenesis. *Eur. J. Cancer Prev.*, 15 (5): 377-83.
- [38] Ray, A.; Banerjee, BD and Sen, P (1996): Modulation of humoral and cell mediated immune responses by *Azadirachta indica* in mice. *Indian J. Exp. Biol.*, 34 (7): 698-701
- [39] Ajith, T.A.; Nivitha, V. and Usha, S. (2007): *Zingiber officinale* Rose alone and in combination with tocopherol protect the

- kidney against cisplatin-induced acute renal failure. Ed. Chem. Toxicol., J. 45 (6): 921-927.
- [40] Mallikarjuna, K.; Chetan, S.P.; Reddy, S.K. and Rajendra, W. (2008): Ethanol toxicity: Rehapitation of hepatic antioxidant defense system with dietary ginger. *Fitoterapia*, 79 (3): 174-178.
- [41] Eisa, A.M. (1998): Clinicopathological studies on some antidiarrheal drugs in rabbits. M.V.Sc., Thesis (Clinical Pathology) Faculty of Veterinary Medicine, Zagazig University.
- [42] Alam, Rasha T. (2008): Clinicopathological studies on the effect of some bacterial infection in rabbits. M.V. Sc., Thesis (Clinical Pathology), Faculty of Veterinary Medicine, Zagazig University.
- [43] El-Deep, M.A.; Metwally, A.M. and Galal, G.E. (2006): The impact of botanical extract, capsicum oil supplementation and their interactions on the productive performance of broiler chicks. XII, EPC, Verona, Italy.
- [44] Verma, S.K.; Singh, J. and Khamesra, A. (1994): Effect of ginger on platelet aggregation in man. *Indian J Med Res* 98: 240- 242

الملخص العربي

دراسات باثولوجية اكلينيكية على تأثير النيم والزنجبيل في الأرانب السليمة والمعدية بالميكروب القولوني (بعد الفطام)

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اجريت هذه الدراسة لمعرفة تأثير النيم والزنجبيل على الأرانب السليمة والمعدية بالميكروب القولوني. اجرى هذا البحث على مائة وعشرون ارنب عمر شهر وتم تقسيمهم الى ٦ مجموعات متساوية وكانت مدة التجربة ٦ اسابيع. تم اعطاؤهم العليقة الاساسية مضاف اليها مسحوق اوراق النيم الجافة بنسبة ٥٪ من وزن العليقة للمجموعتين (٢ و ٥) و بودرة الزنجبيل بنسبة ٢٪ من وزن العليقة للمجموعتين (٣ و ٦) يوميا طوال فترة التجربة. تمت عدوى تجريبية بالميكروب القولوني (*E. coli* O103) عن طريق الفم جرعة واحدة (3×10^7 CFU) عند نهاية الاسبوع الرابع فى المجموعات (٤ و ٦). اوضحت النتائج ان المجموعتين (٣،٢) كانت جميع قياساتهما مماثلة للمجموعة الضابطة (١) طوال فترة التجربة. اظهرت نتائج المجموعة الرابعة وجود نقص معنوى فى البروتين الكلى والزرلال والجلوبيولين و معدل نشاط انزيم الكاتاليز. بينما وجد زيادة معنوية فى معدل انزيمات الكبد ومستوى البولين والكرياتينين و مستوى المالونداي الدهايد والنشاط التلعمى والقدرة القاتلة للخلايا وحيدة النواة. وأدى تناول النيم والزنجبيل الى تحسن نتائج المجموعتين (٥ و ٦) فى جميع القياسات مقارنة بالمجموعة الرابعة واقتربت من الطبيعى فى اليوم الخامس عشر. وجد ارتفاع معنوى فى عدد كرات الدم البيضاء نتيجة لزيادة العدد النوعى لخلايا النيتروفيل فى المجموعة ٤ فى اليوم الأول بينما استمر فى لليوم الثالث بعد العدوى فى المجموعتين ٥،٦. انخفض عدد كرات الدم البيضاء نتيجة لانخفاض عدد خلايا الليمفوسيت فى المجموعة ٤. انخفض عدد خلايا الليمفوسيت فى المجموعات ٤،٥،٦ فى اليومين الثالث والخامس عشر. وجد ارتفاع معنوى فى عدد كرات الدم البيضاء نتيجة لزيادة العدد النوعى لخلايا النيتروفيل فى المجموعة ٤ فى اليوم الأول بينما استمر فى لليوم الثالث بعد العدوى فى المجموعتين ٥،٦. انخفض عدد كرات الدم البيضاء نتيجة لانخفاض عدد خلايا الليمفوسيت فى المجموعة ٤. انخفض عدد خلايا الليمفوسيت فى المجموعات ٤،٥،٦ فى اليومين الثالث والخامس عشر. مما سبق يتضح ان اضافة النيم بنسبة (٥٪) والزنجبيل بنسبة (٢٪) الى العليقة له تأثير مضاد للميكروب القولوني كما أدى الى وتحسين كفاءة الكبد والكلى ومضاد للأكسدة ورفع الحالة المناعية. وكان النيم أقوى تأثيرا من الزنجبيل.