

## Assessment of larvicidal efficiency of nanoemulsion from *Pimpinella anisum* L. essential oil on *Culex pipiens* L. (Diptera: Culicidae)

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### ABSTRACT

In the current study, the nanoemulsion prepared from the essential oil of *Pimpinella anisum* was evaluated as an efficient larvicide against the insect disease vector *Culex pipiens*. The essential oils were distilled from seeds of *P. anisum* by steam distillation, then the nanoemulsion was prepared using the ultrasonication method. The mean droplet size of the nanoemulsion was 116 nm with a polydispersity index of 0.21. The efficacy of nanoemulsion was assessed against the third instar larvae through several concentrations of 30, 35, 40, 45 and 50 ppm. The mortality of larvae was recorded 24h after exposure. The LC<sub>50</sub> and LC<sub>90</sub> for the nanoemulsion were 40.1 and 55.1 ppm while the LC<sub>50</sub> and LC<sub>90</sub> for the bulk emulsion were 44.9 and 69.2 ppm, respectively related to *P. anisum* essential oil. The Larvicidal activity of the formulated nanoemulsion was more toxic than that of the bulk emulsion. The findings of the present study revealed that the nanoemulsion of *P. anisum* EO can be used as a candidate in the integrated management programs to control *C. pipiens*.

### INTRODUCTION

*Culex* mosquitoes are significant vectors of many diseases as they transmit a number of arboviruses including encephalitis, Rift Valley fever and West Nile viruses (Murugan *et al.*, 2012). *C. pipiens* L. is the most abundant mosquito in Egypt, it is found in urban and suburban regions. Chemical insecticides are not recommended nowadays due to several reasons such as the development of insect resistant and the resulted health problems (Bigoga *et al.*, 2013). In addition, these synthetic compounds leave residues that accumulate in the environment causing pollution such as water and soil pollution (Scott *et al.* 2013; Shah *et al.*, 2015). Insecticides with botanical origin may provide efficient and suitable alternatives (Pavela & Govindarajan, 2017) Most botanicals are safer than synthetic insecticides and less harmful to non-target organisms. Essential oils (EOs), also named volatile oils, are natural volatile compounds obtained from parts of aromatic plants such as roots, stems, leaves, flowers, fruits and seeds according to the type of plant (Aktar *et al.*, 2009). EOs are widely used in several industries including food applications, pharmaceutical, perfumery, medicine, sanitary, and cosmetic products (Azmy,

2021b). EOs were proved to be efficient Larvicides against mosquito (Cheng *et al.*, 2009 and Ghosh *et al.*, 2012). To prevent vaporization of the volatile compounds and to reserve biological activity of the EO, the EO should be formulated as nanoemulsion (Bakkali *et al.*, 2008; Osanlo *et al.*, 2017).

EO Nanoemulsions are emulsions with droplet size in the nano-range (McClements, 2011); this formulation improves solubility of oils (Magdassi *et al.*, 2013). Many EOs nanoemulsions were reported as effective larvicides such as nanemulsions of eucalyptus, rosemary, orange (Sugumar *et al.*, 2014; Duarte *et al.*, 2015, Azmy *et al.*, 2019; Azmy *et al.*, 2021 a). *P. anisum* belongs to family Apiaceae, the EO of this plant can be extracted from the seeds, the major component of *P. anisum* EO is trans-anethole (Orav *et al.*, 2008), the insecticidal activity of this compound was reported against *C. pipiens* (Zoubiri *et al.*, 2014; El Zayyat *et al.*, 2017). From this prospective, the present study intended to evaluate the larvicidal efficiency of nanoemulsion of *P. anisum* EO against third instar larvae of *C. pipiens* under laboratory conditions.

## MATERIALS AND METHODS

### **Mosquito larvae:**

Mosquito larvae were obtained from the Egyptian Research Institute of Medical Entomology. Mass rearing of larvae was under optimum humidity ( $75 \pm 3\%$ ) temperature ( $25 \pm 2^\circ\text{C}$ ), and 16 L:8 D photoperiod.

### **Oil extraction:**

Essential oil extraction from seeds of *P. anisum* (Apiaceae) was done through Hydro- distillation using a Clevenger-type apparatus according to Angioni *et al.* (2006).

### **Nanoemulsion formulation:**

The emulsion was prepared using EO, Tween 20 and distilled water according to Duarte *et al.* (2015). To get the nanoemulsion, the emulsion was subjected for 30 minutes according to Anjali *et al.* (2010) to Sonicator 30 kHz frequency and 750 W power output (Ultrasonics, USA/ digital ultrasonic cleaner cd 4830) at the Electron Microscope unit in faculty of Science, Ain Shams University. The part of emulsion which was not subjected to sonication was considered to be the bulk emulsion.

### **Characterization of nanoemulsion**

#### **Droplet Size Distribution & poly dispersity index PDI:**

Measurement of the Droplet size was done using dynamic light scattering technique according to Sugumar *et al.* (2014) at Egyptian Petroleum Research Institute using particle size analyzer (Malvern-UK, 4700).

#### **Stability:**

Thermodynamic stability of the Nanoemulsion was evaluated via storing it at  $4^\circ\text{C}$  and  $25^\circ\text{C}$  for a month. In addition, it was centrifuged at 10,000 rpm for 30 min, and then was observed for any cracking, phase separation or creaming according to Ghosh *et al.* (2013).

**Bioassay tests:**

Bioassay was performed on the third instar larvae of *C. pipiens*, the larvae were treated with different concentrations of nanoemulsion according to the standard WHO (2005) protocol.

Five concentrations of the nanoemulsion and bulk emulsion 30, 35, 40, 45, and 50 ppm were used. Three replicates of twenty larvae were used for each treatment. For the control, the same concentrations were used but with the surfactant only. Mortality was recorded after 24 hours of treatment. Lethal concentration were determined at the 95% confidence level using probit analysis, the percentages of larval mortality were calculated for each concentration of the nanoemulsion.

**RESULTS****Charazterization of nanoemulsion****Droplet size:**

The droplet size distribution of the nanoemulsion is shown in Fig. 1, with the peak at 116 nm as shown in Fig. 2, the value of the PDI equal to 0.21.

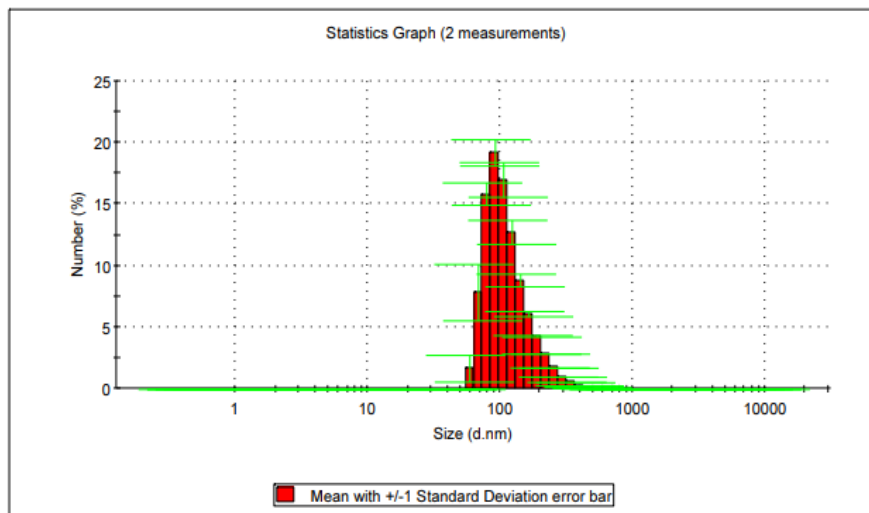


Fig. 1: Droplet size distribution of the nanoemulsion droplets by dynamic light scattering.

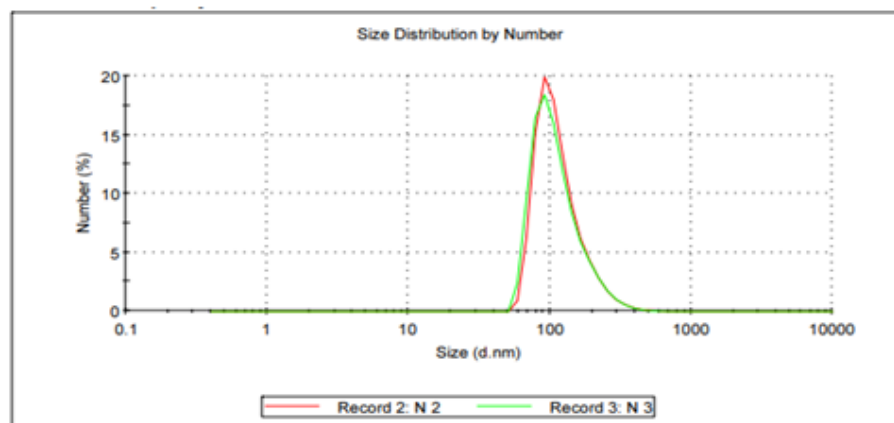


Fig. 2: Droplet size distribution.

### Stability:

There was no observed sign of instability of the nanoemulsion, including creaming, phase separation or cracking, it was stable after centrifugation for 30 min at 10,000 rpm and also it was stable when stored for a month at 4 °C and 25 °C .

### Bioassay

The toxic effect of the nanoemulsion on the 3<sup>rd</sup> larval instar of *C. pipiens* was significantly increased with the increase of concentration ( $P < 0.05$ ). Regression analysis showed a concentration-dependent correlation of the nanoemulsion with mortality of larvae. Table 1 shows Mortalities percentage of the larvae caused by bulk emulsion and nanoemulsion after 24 h of treatment.

Table 1: Mortality percentages of nanoemulsion *P. anisum* EO against 3<sup>rd</sup> instar larvae of *C. pipiens* after 24 hrs.

Nanoemulsiom		Bulk emulsion	
Concentration ppm	Mortalities % (Mean ±SE)	Concentration Ppm	Mortalities % (Mean ±SE)
30	17±0.5 a	30	12±0.5 a
35	23± 1.9 b	35	20±1 b
40	52± 1.1 c	40	40± 1c
45	68.1±1. d	45	50± 0.2d
50	83±0.1 e	50	62±0.2e

Fig. 3 and 4 Show the LC<sub>50</sub> of the nanoemulsion and bulk emulsion on the regression line of probit mortality . The nanoemulsion is more effective than the bulk emulsion regarding the LC<sub>50</sub> and LC<sub>25</sub> values.

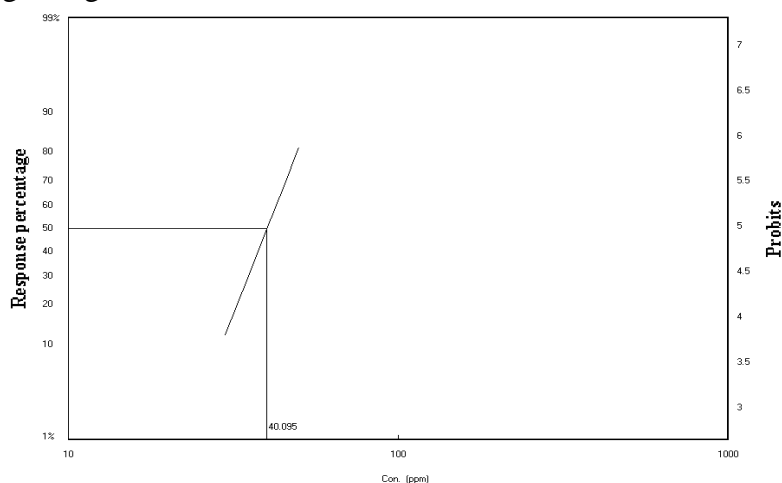


Fig. 3: Regression line of probit mortality of *C. pipiens* larvae against the log concentrations of the nanoemulsion of *P. anisum* EO.

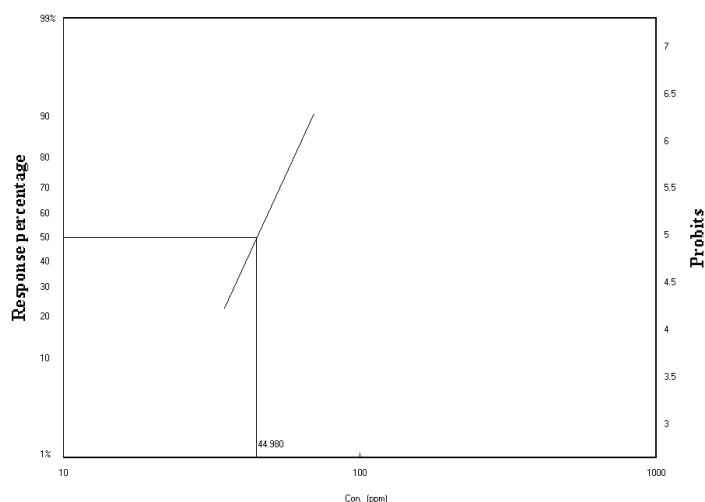


Fig. 4: Regression line of probit mortality of *C. pipiens* larvae against the log concentrations of the bulk emulsion of *P. anisum* EO.

The lethal concentration values of nanoemulsion and bulk emulsion calculated from the regression line of probit mortality of *C. pipiens* larvae against the log concentrations are shown in Table 2.

Table 2: The LC<sub>25</sub>, LC<sub>50</sub> and LC<sub>90</sub> values of nanoemulsion and bulk emulsion of *P. anisum* EO against 3<sup>rd</sup> instar larvae of *C. pipiens* after 24 hrs.

Lethal concentration	Nanoemulsion Concentration Ppm	Bulk emulsion Concentration Ppm
LC <sub>25</sub>	33.9	35.8
LC <sub>50</sub>	40.1	44.9
LC <sub>75</sub>	47.3	56.4
LC <sub>90</sub>	55.1	69.2
LC <sub>99</sub>	71.3	98.4

## DISCUSSION

Insecticides from botanical sources such as EOs have been reported as safe effective substitutes to synthetic insecticides as they are recognized being environmentally friendly (Sugumar *et al.*, 2014; Azmy *et al.*, 2021c). On the other hand, EOs have poor water solubility, while larvicides used to control mosquito larvae should be soluble in water because they are aquatic organisms. Therefore, EOs should be formulated as nanoemulsions to overcome this natural obstacle (Duarte *et al.*, 2015).

The larvicidal activity of the nanoemulsion of *P. anisum* against the *C. pipiens* larvae in this study may be due to trans-anethole which is the major component of the EO, it was reported to have insecticidal property (Abdel-Baki *et al.*, 2021). Our results agree with several studies on the nanoemulsions based on EOs as effective insecticides (Anjali *et al.*, 2011; Ghosh *et al.*, 2013; Sugumar *et al.*, 2014; Duarte *et al.*, 2015).

*al.*, 2015, Azmy *et al.*, 2019; Azmy *et al.*, 2021a). In addition, the results of this research come in contact with the research that studied the toxic effects of *P. anisum* EO on the different life stages of *Cx. quinqua-fasciatus* (Pavela & Govindarajan, 2017), Knio *et al.*, 2008 reported that *P. anisum* seed EO had toxic effect on larvae of *Ochlerotatus caspius*. Furthermore, It was reported that *P. anisum* Eo resulted in complete mortality to *Tribolium castaneum* and *Ephestia kuehniella* adults after 24 h after treatment with 64 µl/L air concentration (Mikhael, 2001). Our findings could be attributed to the penetration of EO to the body of the larvae causing neural toxicity to their nervous system and inhibition of the normal biological processes (Chantawee & Soonwera, 2018). Nanoemulsions based on EO have various advantages including stability, long shelf life (Mishra *et al.*, 2017). The uniformity and stability of the droplet size in the nanoemulsion can be indicated by PDI value is an indication of (Anjali *et al.*, 2010), the low value of PDI of the prepared nanoemulsion in this study indicates the high uniformity of droplet size and the long-term stability. Stabilization of the nanoemulsion is because of the presence of the surfactant as it provides a mechanical barrier to prevent the droplets from accumulation. The high efficiency of the nanemulsion may be due to the small size of the droplets, which increases the surface area and facilitates the penetration of the effective component into the larvae according to Anjali *et al.* (2010). The botanical molecules are able to interact with enzymes, hormones and bind to membranes and cellular components and thus interfere with biochemical pathways of the mosquito (Powell, 2009 and Fallatah, 2010). Further investigation is needed to reveal the mode of action of this larvicide and its effect on the different biological aspects of the treated larvae.

## CONCLUSION

The present research reported that natural products of botanical origin such as the nanoemulsion of *P. anisum* EO can be used as low-cost safe alternative to humans and the environment for controlling the disease-vector *C. pipiens*.

## ETHICAL APPROVAL:

This research paper was approved by the research ethics committee from Faculty of Science, Ain Shams University (ASU-SCI/ENTO/2023/3/1).

## REFERENCES

- Abdel-Baki, A. A. S.; Aboelhadid, S. M.; Sokmen, A.; Al- Quraishy, S., Hassan, A. O. and Kamel, A. A.** (2021). Larvicidal and pupicidal activities of *Foeniculum vulgare* essential oil, trans- anethole and fenchone against house fly *Musca domestica* and their inhibitory effect on acetylcholinesterase. *Entomol. Res.*, 51(11): 568-577.
- Aktar, W.; Sengupta, D. and Chowdhury, A.** (2009). Impact of pesticides use in agriculture: their benefits and hazards. *Interdisciplinary toxicol.*, 2(1): 1-12.
- Anjali, C.; Khan, S.; Goshen, K. and Magdassi, S.** (2010). Formulation of water-dispersible nanopermethrin for larvicidal applications. *Ecotoxicol. and Env. Safety.*, 73: 1932-1936.

- Anjali, C. ; Sharma, Y., Mukherjee A. and Chandrasekaran N.** (2011). Neem oil (*Azadirachta indica*) nanoemulsion - a potent larvicidal agent against *Culex quinquefasciatus*. Wiley Online Library.
- Azmy, R. M.; El Gohary, E. G.; Mahmoud, D.; Salem, D.; Abdou, M. and Salama, M.** (2019). Assessment of larvicidal activity of nanoemulsion from *Citrus sinensis* essential oil on *Culex pipiens* L.(Diptera: Culicidae). Egyptian J. of Aquatic Biol. and Fisheries, 23(3): 61-67.
- Azmy, R. M.; El Gohary, E. G. E.; Salem, D. A.; Abdou, M. A.; Salama, M. S. and Mahmoud, D. M.** (2021a). Biochemical and histopathological effect of the essential oil of *Citrus sinensis* (L.) Osbeck on larvae of *Culex pipiens* Linnaeus, 1758 (Diptera: Culicidae). Aquatic Insects, 42(1), 78-90.
- Azmy, R. M.** (2021b). Nanoformulated Materials from Citrus Wastes. In Waste Recycling Technologies for Nanomaterials Manufacturing (pp. 649-669). Springer, Cham.
- Azmy, R. M. et al.** (2021c). Evaluation of the larvicidal activity of nanoemulsion from *Citrus aurantifolia* (Christm) Swingle peel on *Culex pipiens* L.(Diptera: Culicidae) and the induced morphological aberrations. Egyptian Journal of Aquatic Biol. and Fisheries, 25(3): 421-434.
- Bakkali, F.; Averbeck, S.; Averbeck, D. and Idaomar, M.** (2008) Biological effects of essential oils - a review. Food Chem Toxicol, 46(2): 446–475.
- Bigoga, J.; Saahkem, P.; dindeng, S.; Ngondi, J. and Negue, M.** (2013). Larvicidal and Repellent Potential of *Chenopodium ambrosiodes* Linn Essential oil against *Anopheles gambiae* Giles (Diptera: Culicidae). The open Entomolgy journal, 7:16-22.
- Chantawee, A. and Soonwera, M.** (2018). Larvicidal, pupicidal and oviposition deterrent activities of essential oils from Umbelliferae plants against house fly *Musca domestica*. Asian Pacific Journal of Tropical Medicine, 11(11): 621.
- Cheng, S.; Huang, C.; Chen, Y.; Yu, J.; Chen, W. and Chang, S.** (2009). Chemical compositions and larvicidal activities of leaf essential oils from two eucalyptus species. Bioresource Tech., 48: 4868 - 4873.
- Duarte, J.; Amado, J.; Oliveira, A.; Cruz, R.; Ferreira, A.; Souto, R.; Falcão, D.; Carvalh,o J. and Fernandes, C.** (2015). Evaluation of larvicidal activity of a nanoemulsion of *Rosmarinus officinalis* essential oil. Rev. Bras., 25(2):189–192.
- El Zayyat, E. A.; Soliman, M. I.; Elleboudy, N. A., and Ofaa, S. E.** (2017). Bioefficacy of some Egyptian aromatic plants on *Culex pipiens* (Diptera: Culicidae) adults and larvae. J. of Arthropod-Borne Diseases, 11(1), 147.
- Fallatah, S.** (2010). Histopathological Effects of Fenugreek (*Trigonella foenumgraceum*) extracts on the larvae of the Mosquito *Culex Quinquefasciatus* Agr. Tech. and Biol.Sciences, 5(2): 123-130.
- Ghosh, A.; Choudhary, N.; and Chandra, G.** (2012). Plant extracts as potential mosquito larvicides. The Indian J.of Med. Res.,135(5): 581-598.
- Ghosh, V.; Mukherjee, A.; and Chandrasekaran, N.** (2013). Formulation and characterization of plant essential oil based Nanoemulsion: evaluation of its Larvicidal activity against *Aedes egypti*. Asian J. Chem., 25:S321- S323.

- Knio, K. M.; Usta, J.; Dagher, S.; Zournajian, H. and Kreydiyyeh, S.** (2008). Larvicidal activity of essential oils extracted from commonly used herbs in Lebanon against the seaside mosquito, *Ochlerotatus caspius*. *Bioresource tech.*, 99(4), 763-768.
- Magdassi, S.; Mukherjee, A. and Chandrasekaran, N.** (2013). Distinctive effects of nano-sized permethrin in the environment. *Environmental Science and Pollution Research*, 20: 2593-2602.
- McClements, D.** (2011). Edible nanoemulsions: fabrication, properties, and functional performance. *Soft Matter*, 7(6): 2297-316.
- Mikhael, A. A.** (2011). Potential of some volatile oils in protecting packages of irradiated wheat flour against *Ephesia kuehniella* and *Tribolium castaneum*. *J. of Stored Products Res.*, 47(4): 357-364.
- Mishra, P.; Tyagi, B.; Chandrasekaran, N. and Mukherjee, A.** (2017). Biological nanopesticides: a greener approach towards the mosquito vector control *Environ Sci. Pollut. Res.*, 1–13.
- Murugan, K.; Kumar, P.; Kovendan, D.; Subrmaniam, J. and Hwang, J.** (2012). Larvicidal, pupicidal, repellent and adulticidal of *Citrus sinensis* orange peel extract against *Anopheles stephensi*, *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). *Parasitol. Res.*, 111(4):1757-1769.
- Orav, A.; Raal, A. and Arak, E.** (2008). Essential oil composition of *Pimpinella anisum* L. fruits from various European countries. *Nat. P. Res.*, 22(3): 227-232.
- Osanlo, M.; Amani, A.; Sereshti, H.; Abai, M.; Esmaeili, F. and Sedaghat, M.** (2017). Preparation and optimization nanoemulsion of Tarragon (*Artemisia dracunculus*) essential oil as effective herbal larvicide against *Anopheles stephensi*. *Ind Crop. Prod.*, 109: 214–219.
- Pavela, R. and Govindarajan, M.** (2017). The essential oil from *Zanthoxylum monophyllum* a potential mosquito larvicide with low toxicity to the non-target fish *Gambusia affinis*. *J. of Pest Sci.*, 90(1): 369-378.
- Powell, R.** (2009). Plant seeds as sources of potential industrial chemicals, pharmaceuticals, and pest control agents. *J. Nat. Prod.*, 72(3): 516-23.
- Scott, J. G.; Leichter, C. A.; Rinkevich, F. D. et al.** (2013) Insecticide resistance in house flies from the United States: Resistance levels and frequency of pyrethroid resistance alleles. *Pesticide Biochem. and Physiol.*, 107: 377–384.
- Shah, R. M.; Abbas, N. and Shad, S. A.** (2015) Assessment of resistance risk in *Musca domestica* L. (Diptera: Muscidae) to methoxyfenozide. *Acta Tropica* 149: 32–37.
- Sugumar, S.; Clarke, S.; Nirmala, M.; Tyagi, B.; Mukherjee, A.; and Chandrasekaran, N.** (2014). Nanoemulsion of eucalyptus oil and its larvicidal activity against *Culex quinquefasciatus*. *Bull. Entomol. Res.*, 104 (03): 393–402.
- WHO.** Guidelines for laboratory and field testing of mosquito larvicides, 2005.
- Zoubiri, S.; Baaliouamer, A.; Seba, N. and Chamouni, N.** (2014) Chemical composition and larvicidal activity of Algerian *Foeniculum vulgare* seed essential oil. *Arabian J. of Chem.*, 7: 480–485.