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# Antimicrobial and antioxidant activities of phytic acid extracted from Wheat Bran

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#### Abstract

The objective of this study is to investigate the antimicrobial and antioxidant activities of pure phytic acid and phytate extracted from wheat bran. The obtained results showed that, Gram-positive bacteria had more sensitive for phytic acid than Gram- negative bacteria. Bacillus anthrakoid and Staphylococcus aureus were more sensitive at concentrations of pure phytate 5, 6 %. Inhibition effects in Escherichia coli O157:H7 and Staphylococcus aureus were 20.33 and 38.67 mm, respectively at 6% of pure phytate. Moreover, a moderate inhibition against fungi was observed. Besides the pure phytic acid and phytate extracted from wheat bran have an inhibitory effect on many types of microorganisms. The antioxidant activity of pure phytate increased with increasing the concentration, where, the scavenging activity was 22, 23 and 36% at concentration 1, 3, and 5 mM, respectively. The extracted phytate gave high scavenging activity that reached 90% if compared with pure phytic acid. Aqueous wheat bran extract gave a percentage of free radical scavenging activity reached 36.16%. From the obtained results it can be concluded that phytate extracted from wheat bran has good antimicrobial and antioxidant activity and could be use as natural preservatives.

#### **Keywords:**

Antimicrobial, Antioxidant, Phytic acid, phytate, wheat bran

#### **INTRODUCTION**

Phytic acid constitutes 1 to 5 % (w  $w^{-1}$ , dry weight basis) of most oilseeds, legumes, cereals, nuts, and pollen and represents from 50 to 80 % of the total phosphorus level in seeds. Phosphorus accumulation in developing seeds is higher than that needed for nominal cellular functions. Several plants use the excess phosphorus to synthesize phytic acid (Graf and Eaton, 1990 and Thavarajah et al., 2010). Phytic acid may effectively inhibit the growth of foodborne pathogens such as Escherichia coli, Staphylococcus aureus, Bacillus subtilis. Salmonella Typhimurium, and Clostridium perfringens (Zhou et al., 2018). Phytic acid has also enhanced the antibacterial effect of nisin against Listeria monocytogenes on cabbage and broccoli, significantly reducing bacteria (Bari et al., 2005). Phytic acid has(PA) been recognized as a potent antioxidant. It is a potent inhibitor of iron-catalyzed hydroxyl radical formation by chelating the iron required for the generation of hydroxyl radical via the Fenton-type reaction. That PA exhibited an antioxidant effect by suppressing iron (II)-enhanced hydroxyl radical formation induced by1-methyl-4phenylpyridinium ion (MPP+) in rat striatum. (Graf and Eaton, 1990; Rimbach and Pallauf, 1998 and Obata, 2003). Phytic acid and wheat bran as antioxidants inhibited metmyoglobin formation and stabilized red meat color. In addition, it inhibited lipid peroxidation and degradation of heme pigments caused by cooking and storage. Therefore, they may be useful as additives for meat processing to prevent the offflavor formation and extension of shelf life (Shatta et al., 2005). The phytic acid is an effective antioxidant as it has high chelating capacity of multivalent metal ions such as iron, zinc, and calcium. Phytic acid inactivates the iron's catalytic action and inhibits the hydroxyl radicals (OH) production by forming an iron chelate (Graf By binding with the six and Eaton, 1990). Fe<sup>3+</sup>coordination sites, phytic acid prevents the reaction between  $H_2O_2$  and chelated  $Fe^{3+}$  ion, hydroxyl radical avoiding iron formation. Different examinations have additionally demonstrated the capacity of phytic corrosive to repress lipid oxidation in meat-based foods (Lee and Hendricks, 1995 and Park et al., 2004). The utilization of phytic corrosive as a reasonable option for the meat industry to further develop industrialized meat items' security and quality, was additionally settled by Stodolak *et al.* (2007) while assessing phytic-corrosive containing arrangements as security extenders for a crude and cooked hamburger and pork meat held under refrigeration, it found that metmyoglobin creation was restrained within the sight of phytic corrosive in crude meat. Therefore, the aim of this study is to investigate the antimicrobial and antioxidant activities of pure phytic acid and phytate extracted from wheat bran.

### MATERIALS AND METHODS

#### Materials

Wheat bran obtained from the mills of Upper Egypt in Sohag Governorate during 2020. Chemicals used in this study were obtained from Pio Chem, Alpha Company, and Sigma – Aldrich (St. Lous, MO, USA).

#### Microorganisms

The tested microorganisms were summarized in Table (1).

No	Microorganisms	Source				
Gram- negative bacteria						
1	Escherichia coli O157:H7					
2	Klebsiella Pneumoniae	Botany Dept., Fac.of				
3	Proteus vulgaris					
4	Pseudomonas sp	Sci.,Assiut Univ				
Gram- positive bacteria						
1	Bacillus anthrakoid	Botany Dept., Fac.of				
2	Staphylococcus aureus	Sci Assiut Univ				
Fungi						
1	Aspergillus flavus	Botany Dept., Fac.of Sci.,				
2	Penicillium duclauxii	Fac.of Arche., Sohag Univ				
3	Trichoderma sp					

# Table (1): Tested microorganisms in the study.

#### Methods

#### **Preparation of phytate extract**

Phytic acid corrosive was removed from wheat bran as per the strategy portrayed by Canan *et al.* (2011): Briefly 20 grams of wheat bran was blended with 200 mL of 1.0 N HCL and shaken for one hour at room temperature on a stirrer, then its pH was acclimated to 6.2 utilizing 4.0 N NaOH and centrifuged at 3000 rpm for 10 min then the pH of the supernatant was acclimated to 8 with 1.5 M Na<sub>2</sub>CO<sub>3</sub> arrangement and stayed for 12 h at room temperature. After centrifugation at 3000 rpm for 10 min, the supernatant was tapped and the acquired pellet was resuspended in 1.0 N HCL. To eliminate foreign substances and protein denaturation, 10 mL formaldehyde and 0.5-gram celit were added. This suspension was then shaken for 2 h and stayed for 12 h at room temperature. The pre-arranged suspension was then gone through subjective Whatman channel paper No3 and the pH of the filtrate was acclimated to 7 by 1.5 M Na<sub>2</sub>CO<sub>3</sub>. At last, the framed pellet was recuperated through separating by subjective Whatman filter paper and dried at 60° C for one day.

#### Preparation of aqueous wheat bran extract:

Ten grams of each sample was extracted by stirring with 100 ml of distilled water for 1 hour at room temperature. After centrifugation and filtration through Whatman No. 1. This extract was used to perform the required analyzes.

#### Antibacterial activity

Antibacterial activity of phytic corrosive is not entirely set in stone by the agar dissemination strategy as per the NCCLS (1993).

#### Antifungal and Yeasts test

Antifungal and yeasts test were completed concurred to ELLOF (1998).

#### Determination of total antioxidant activity

The free radical scavenging activity of phytic acid and other antioxidants was measured with 2,2 diphenyl- 1- picrylhydrazyl (DPPH) free radical using the method described by Yen and Wu (1999). Sample solution (4 ml) was added to 1 ml, 0.2 mM DPPH in methanol. After reacting for 30 min in the dark, the absorbance was read at 547 nm by a spectrophotometer. Methanol (4 ml) was mixed with 1 ml DPPH and this served as the control. Radical scavenging activity (%) was calculated as follows:

Radical scavenging activity (%) = (1 - Absorbance of sample / Absorbance of control) x 100.

#### Statistical analysis

Data were analyzed by analysis of variance (ANOVA) using a completely randomized factorial design. Basic statistics and ANOVA were

performed to test the significance within replication and between treatments (MSTAT-C 1989). The (LSD) tests was used to determine the means at the level of 0.05%.

### **RESULTS AND DISCUSSION**

# Antimicrobial activity of phytate and wheat bran

The results shown in Table 2 indicate the antimicrobial activity of pure phytic acid (PA), phytate extract and wheat bran extract at different concentrations. Data showed that, Gram-positive bacteria was more sensitive for PA than Gramnegative bacteria. *Bacillus anthrakoid* was sensitive to pure phytic acid while, Staphylococcus aureus was more sensitive to pure phytate at concentrations 5, 6 %. Moreover, no inhibition against fungi was recorded. Besides, it could be noticed that microbial spectra were decreased with increasing the phytic acid concentrations. These results are in the line with those reported by Boukhris et al. (2020), they found that minimum inhibitory concentration of PA varied from 0.488 to 0.97 mg/ml for the Gram-positive bacteria that were tested, and was 0.244 mg/ml for the Gramnegative bacteria. Linear and general models were used to further explore the antibacterial effects of PA. The developed models were validated using experimental growth data for L. monocytogenes, S. aureus and S. Typhimurium. The antimicrobial activity of phytate extracted from wheat bran at concentration (5 %) was determined and inhibitation zones was recorded in same Table. Data showed that, Gram- positive bacteria was more sensitive than Gram- negative bacteria. Bacillus anthrakoid and Staphylococcus aureus were sensitive to concentration of phytate extract at 5 % concentration. Data showed that, the inhibition effects on Escherichia coli O157:H7, and Staphylococcus aureus were 20.33 and 38.67 mm, respectively for 6% concentration of pure phytate. Moreover, results revealed that a moderate inhibition against fungi was observed. The concentration of phytate extract 5 % had no inhibition effects on Trichoderma sp. Data revealed that the microbial spectra was decreased with increasing the concentrations of phytate extract and Gram- positive bacteria was more sensitive than Gram- negative bacteria. The antimicrobial

activity of aqueous wheat bran extract was also determined and the inhabitation zones illustrated in Table (2). From these date it can be conceded that, pure phytic acid and phytate extracted from wheat bran have an inhibitory effect on many types of microorganisms. The inhibition zones varied according to pure phytate the concentration degree and the highest inhibition zones were at a concentration 5% and 6%. The finding showed that effect of phytate extract on fungi, was moderate but was better than the effect of phytic acid, which did not show any effect on fungi.

# Antioxidant activity of phytic acid, phytate extract and wheat bran

Phytic acid, phytate extract and aqueous wheat bran extract were subjected to the antioxidant activity using 2,2-diphenyl-picrylhydrazyl (DPPH) radical scavenging assay. Results in Table 3 and Fig. 2 showed that, the free radical scavenging activity of phytic acid at (1, 3 and 5mM) concentrations were 22.23, 33.60 and 36, respectively. These means that the antioxidant activity of pure phytate increased with increasing in phytate concentration, these results are in agreement with those obtained by Skarin et al (2005) they reported that, phytic acid had low radical scavenging effect, which was in the range of 10.1 to 41.0%. Ahn et al. (2004) reported that the irradiation dose was positively correlated with the DPPH radical-scavenging effects. Phytic acid has antioxidant functions by virtue of forming a unique iron chelate and it suppresses the iron-

catalyzed oxidative reactions, serving as a potent antioxidant function in the preservation of seeds (Graf and Eaton, 1990). The same mechanism, dietary phytic acid may lower the incidence of colonic cancer and protect against other inflammatory bowel diseases. However, the free radical-scavenging activity of phytic acid has not yet been reported. A concentration effect of phytic acid was also observed, and radical-scavenging capacity increased with increasing concentration. Phytate extracted from wheat bran gave the free radical scavenging activity 16.70, 81 and 90.33% at concentrations 1, 3 and 5%, respectively. Phytate extract from wheat bran gave a high scavenging activity that reached 90% compared with pure phytic acid. On the other hand, aqueous wheat bran extract gave a free radical scavenging activity reached 36.16%. These results are in the line with those reported by Iqbal et al. (2007), they found that the remaining amount as percentage of DPPH radical at 5 min after initiation of the reaction were 24, 30, 38,43, and 48% for five wheat varieties bran. Free radical and radical cation scavenging activity were comparable to previous findings for wheat bran of different varieties from USA (Yu et al., 2003). Li et al. (2007) reported that the DPPH scavenging activity of purple wheat bran extract was 63.17%. From this result it can be concluded that pure phytic acid as well as phytate extracted from wheat bran have an important role as natural antioxidants that can be used to preserving many food items.

**Table (2)** Diameter of inhibition zones (mm) of phytic acid, phytate extract, and wheat bran extract against some selected microorganisms:

Concentration (mg/ml)	Diameter of inhibition zones (mm)					
Microorganism	PA 1%	PA 3%	PA 5%	PA 6%	PE 5%	WBE
Escherichia coli 0157:H7	N.d	N.d	12.67	20.33	20.33	16.33
Klebsiella Pneumoniae	N.d	N.d	30.67	33.00	15.67	N.d
Proteus vulgaris	N.d	N.d	N.d	N.d	19.67	N.d
Pseudomonas sp	N.d	N.d	30.67	35.00	17.00	N.d
Bacillus anthrakoid	24.33	24.00	31.67	20.33	17.33	N.d
Staphylococcus aureus	N.d	N.d	31.00	38.67	23.33	20.67
Aspergillus flavus	N.d	N.d	N.d	N.d	20.00	22.00
Penicillium duclauxii	N.d	N.d	N.d	N.d	35.00	49.33
Trichoderma sp	N.d	N.d	N.d	N.d	N.d	23.67
LSD 5%	4.29	3.03	3.59	9.36	10.52	3.20

PA=phytic acid, PE= phytate extract, WBE= wheat bran extract, N.d= Not detected

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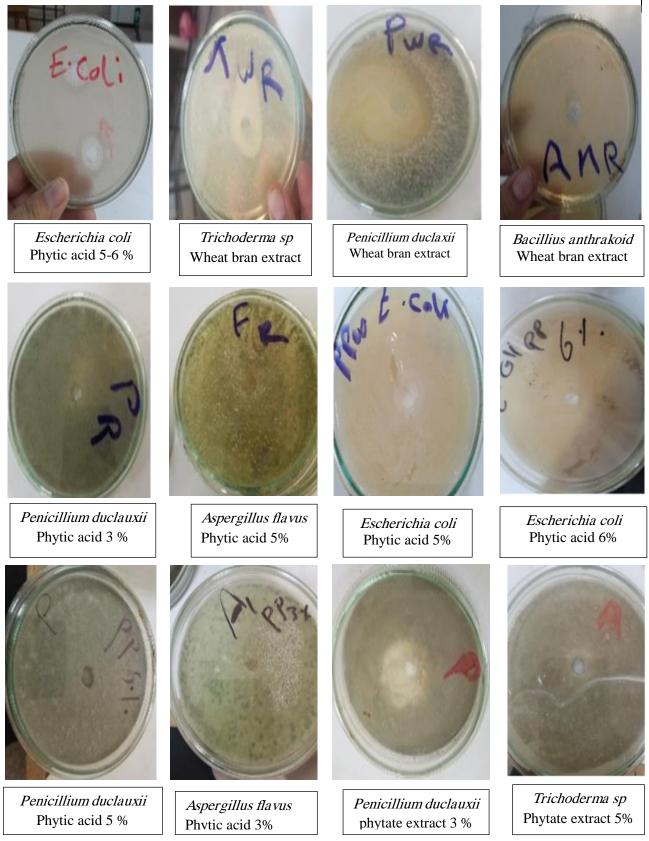
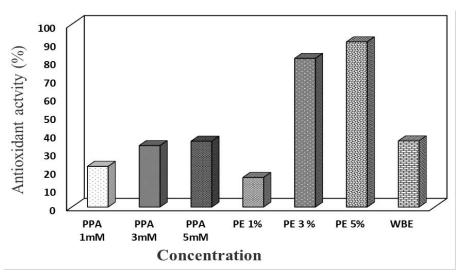


Fig (1): Antimicrobial activity of phytate against certain types of microorganisms.

Samples	Antioxidant activity			
Pure phytic acid 1mM	22.23			
Pure phytic acid 3mM	33.60			
Pure phytic acid 5mM	36.00			
Phytate extract 1%	16.17			
Phytate extract 3%	81.30			
Phytate extract 5%	90.33			
Wheat bran	36.13			
LSD 5%	3.28			

Table (3): Antioxidant activity of phytic acid, phytate extract, and wheat bran.



**Fig** (2): Antioxidant activity of phytic acid, phytate extract, and wheat bran Extract.  $PPA_1$ = pure phytic acid 1mM,  $PPA_3$  = pure phytic acid 3mM,  $PPA_5$  = pure phytic acid 5mM,  $PE_1$  = phytate extract 1%,  $PE_3$  = phytate extract 3%,  $PE_5$  = phytate extract 5%, WBE= wheat bran extract

# CONCLUSION

The obtained results showed that, Gram-positive bacteria had more sensitive for phytic acid than Gram- negative bacteria. A moderate inhibition against fungi was observed. The results indicated that pure phytic acid and phytate extracted from wheat bran have an inhibitory effect on many types of microorganisms. Phytate extracted from wheat bran gave a high scavenging activity that reached 90% compared with pure phytic acid. Aqueous wheat bran extract gave a percentage of free radical scavenging activity reached 36.16%. It can be concluded that phytate extracted from wheat bran have antimicrobial and antioxidant activity with the possibility of using them as natural preservatives

# REFERENCES

- Ahn, H. J., Kim, J. H., Jo, C., Kim, M. J., & Byun, M. W. (2004). Comparison of irradiated phytic acid and other antioxidants for antioxidant activity. *Food Chemistry*, 88(2), 173-178.
- Bari, M. L., Ukuku, D. O., Kawasaki, T., Inatsu, Y., Isshiki, K., & Kawamoto, S. (2005).
  Combined efficacy of nisin and pediocin with sodium lactate, citric acid, phytic acid, and potassium sorbate and EDTA in reducing the Listeria monocytogenes population of inoculated fresh-cut produce. *Journal of food protection*, 68(7), 1381-1387.
- Boukhris, I., Smaoui, S., Ennouri, K., Morjene, N., Farhat-Khemakhem, A., Blibech, M., ... & Chouayekh, H. (2020). Towards understanding the antagonistic activity of phytic acid against

common foodborne bacterial pathogens using a general linear model. *PloS one*, *15*(4), e0231397.

- Canan, C., Cruz, F. T. L., Delaroza, F., Casagrande, R., Sarmento, C. P. M., Shimokomaki, M., & Ida, E. I. (2011). Studies on the extraction and purification of phytic acid from rice bran. *Journal of food composition and analysis*, 24(7), 1057-1063.
- Eloff, J. N. (1998). A sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria. *Planta medica*, 64(08), 711-713.
- Graf, E., & Eaton, J. W. (1990). Antioxidant functions of phytic acid. *Free Radical Biology and Medicine*, 8(1), 61-69.
- Iqbal, S., Bhanger, M. I., & Anwar, F. (2007). Antioxidant properties and components of bran extracts from selected wheat varieties commercially available in Pakistan. *LWT-Food Science and technology*, *40*(2), 361-367.
- Lee, B. J., & Hendricks, D. G. (1995). Phytic acid protective effect against beef round muscle lipid peroxidation. *Journal of Food Science*, 60(2), 241-244.Li, W., Pickard, M. D., & Beta, T. (2007). Effect of thermal processing on antioxidant properties of purple wheat bran. *Food chemistry*, 104(3), 1080-1086.
- MSTAT-C (1989). A micromputer program for the design, management, and analysis of agronomic research experiments, ver.1.2 Michigan State University, East Lansing, MI48824.
- NCCLS (1993). Performance standards for antimicrobial Disc susceptibility Test. Approved Standards NCLLS Publication M2-A5, Villanova, PA, USA.
- Obata, T. (2003). Phytic acid suppresses 1-methyl-4-phenylpyridinium ion-induced hydroxyl radical generation in rat striatum. *Brain research*, 978(1-2), 241-244.
- Rimbach, G., & Pallauf, J. (1998). Phytic acid inhibits free radical formation in vitro but does not affect liver oxidant or antioxidant status in growing rats. *The Journal of nutrition*, *128*(11), 1950-1955.
- Shakrin, N.M.; Saad, N.; Esa,N.M.(2005).Phytic acid extracted from rice bran as natural antioxidant, Abstracts of the 20<sup>th</sup> Scientific Conference of the Nutrition Society of

Malaysia , Youth and nutrition, the future of the nation , Kuala Lumpur, March 24-25.

- Shatta, A.A.; Gab-Alla,A.A.and El-Shami.A.M. (2005). The effect of phytic acid as antioxidant in meat products. *Egyptain Journal of Food Science*, *33*(2), 87–99.
- Stodolak, B., Starzyńska, A., Czyszczoń, M., & Żyła, K. (2007). The effect of phytic acid on oxidative stability of raw and cooked meat. *Food Chemistry*, 101(3), 1041-1045.
- Park, H. R., Ahn, H. J., Kim, J. H., Yook, H. S., Kim, S., Lee, C. H., & Byun, M. W. (2004). Effects of irradiated phytic acid on antioxidation and color stability in meat models. *Journal of agricultural and food chemistry*, 52(9), 2572-2576.
- Thavarajah, P., See, C. T., & Vandenberg, A. (2010). Phytic acid and Fe and Zn concentration in lentil (Lens culinaris L.) seeds is influenced by temperature during seed filling period. *Food Chemistry*, *122*(1), 254-259.
- Yen, G. C., & Wu, J. Y. (1999). Antioxidant and radical scavenging properties of extracts from Ganoderma tsugae. *Food chemistry*, 65(3), 375-379.
- Yu, L., Perret, J., Harris, M., Wilson, J., & Haley, S. (2003). Antioxidant properties of bran extracts from "Akron" wheat grown at different locations. *Journal of Agricultural and Food Chemistry*, 51(6), 1566-1570.
- Zhou, Q. I., Zhao, Y. U., Dang, H., Tang, Y., & Zhang, B. (2019). Antibacterial effects of phytic acid against foodborne pathogens and investigation of its mode of action. *Journal of food protection*, 82(5), 826-833.

الملخص العربي

# النشاط المضاد للأكسدة والميكروبات لحمض الفيتك المستخلص من ردة القمح

محمد عبد الحميد سرور، أبو الحمد السيد مهني، السيد على محمود و أسماء جمال أحمد. قسم علوم الأغذية والتغذية، كلية الزرعة جامعة سوهاج،سوهاج، مصر.

الهدف من هذه الدراسة هو التعرف على النشاط المضاد للميكروبات والأكسدة لحامض الفيتك النقى والمستخلص من ردة القمح. حيث أظهرت النتائج أن البكتيريا الموجبة الجرام كانت أكثر حساسية لحامض الفيتك من البكتريا السالبة لجرام. وكانت بكتريا Bacillus anthrakoid د Staphylococcus اکثر حساسية من لحمض الفيتك النقى عند تركيزات 5، 6٪. وكانت تأثيرات التثبيط بكتريا القولون Escherichia coli O157: H7 والمكورات العنقودية الذهبية 20.33 و38.67 مم على التوالي بتركيز 6٪ من الفيتات النقي. علاوة على ذلك، أظهرت النتائج وجود تثبيط معتدل للفطريات. اضافة إلى ذلك وجد أن حامض الفيتك النقى والفيتات المستخلصة من ردة القمح لهما تأثير مثبط على العديد من الكائنات الحية الدقيقة. زاد النشاط المضاد للأكسدة للفيتات النقية بزيادة تركيز الفيتات حيث كان النشاط المضاد اللأكسدة 22 و23 و36٪ بتركيزات 1 و3 و5 مل مول على التوالي. أعطى حامض الفيتك المستخلص من ردة القمح نشاط مضاد للأكسدة عالى وصل إلى 90٪ مقارنة بحامض الفيتك النقى قد أعطى مستخلص ردة القمح نسبة نشاط مضاد الأكسدة وصلت 36.16٪. من النتائج المتحصل عليها تتضح أن الفيتات المستخلصة من ردة القمح لهاً نشاط مضاد للميكر وبات والأكسدة و يمكن استخدامها كمواد حافظة طبيعية.