# A new species, first record and synonymy of eriophyid mites (Acari, Prostigmata) from Egyptian weeds

## Ashraf S. Elhalawany

Fruit Tree Mite Department, Plant Protection Research Institute, Agricultural Research Centre, Dokki, Giza, Egypt, E-mail: dr\_ashraf\_said@yahoo.com, ashrafelhalawany@arc.sci.eg, ORCID <u>https://orcid.org/0000-0001-5195-3942</u>

#### ABSTRACT

Eriophyoids have a significant economic impact as weed pests, particularly when it comes to the malformations they can occasionally cause, such as rusting, bronzing, leaf rolling, erineum, bud galls, stunting, blisters, leaf galls, damaged seeds, and mosaic virus disease. Six eriophyoid mite species collected from Egyptian weeds are described. *Aceria newplucheae* **sp. nov.** is described as a vagrant on the leaf axil of *Pluchea dioscoridis* (L.) DC. (Compositae). A complementary description to the first-recorded species, *Aceria chenopodia* Xue, Sadeghi & Hong infesting *Chenopodium album* L. and *Chenopodium murale* L. (Amaranthaceae) in Egypt was done based on females, males and immature stages. Three eriophyoid mite species have been synonymized: *Aceria noxia* Flechtmann & Tassi is a junior synonym of *Aceria lividus* Elhalawany; *Aceria mosalahi* Lewandowski & Elsayed is a junior synonym of *Aceria dioscoridis* (Soliman & Abou-Awad); and *Aceria aegyptiacus* (Soliman & Abou-Awad) is a junior synonym of *Aceria tosichella* Keifer. Furthermore, *Vittacus plucheae* Abou-Awad & Nasr, a vagrant causing rusting on *P. dioscoridis*, is designated as a junior synonym of *Neooxycenus plucheae* Abou-Awad and re-described herein.

**Keywords:** Systematics, biological control, *Aceria*, *Neooxycenus*, *Pluchea dioscoridis*, Amaranthaceae, Poaceae.

# **INTRODUCTION**

Eriophyoidea are an economically significant group due to the direct harm they can inflict on their hosts, their ability to spread dangerous plant diseases, and the potential for using them as biological weed control agents (Lindquist et al. 1996). Almost 80% of reports are about a single host species, 95% are about a single host genus, and 99% are about a single host family (Skoracka et al. 2010).

The weed *Pluchea dioscoridis* (L.) DC. (Compositae = Asteraceae) is a significant wild evergreen shrub. The plant can be found growing near water canals, roads, and marshes. It is most common in the Nile region, the Western Desert Oases, the Mediterranean coastal strip, the Eastern Desert, and Sinai Peninsula, Egypt (Boulos 2002). The genus *Amaranthus* is found all over the world, with approximately 65 species in tropical, subtropical, and warm climates (Boulos 1999). This weed is a major threat to a variety of agricultural crops in addition to vegetables and orchards. It is an annual spread made up of long-lived seeds that are mostly

propagated by wind and water, but also by machines (Zaki 2000).

Keifer (1969a) established the genus *Vittacus* based on the species *Vittacus mansoni*. This genus is distinguished by its fusiform body, gnathosoma that projects downward; flat dorsal opisthosoma, and semiannuli that appear as transverse thickened bands, forming a slight furrow, especially caudally; each band forms subdorsal ridges laterally. There are currently nine species of *Vittacus* known worldwide, only two species of which have previously been recorded from Egypt, *Vittacus plucheae* Abou-Awad & Nasr, 1986 on *P. dioscoridis* and *Vittacus bougainvilleae* (Keifer, 1959) on *Bougainvillea glabra* Choisy (Nyctaginaceae) (Guo et al. 2015; Elhalawany 2018).

Abou-Awad (1981) established the genus *Neooxycenus* based on *Neooxycenus plucheae* on *P. dioscoridis* and described it as having a fusiform, flattened body divided into broad dorsal semiannuli and narrow microtuberculate ventral semiannuli. The prodorsal shield is rounded and broader, with a prominent anterior lobe; the posterior portion is narrow and appears as a projection covering the anterior 2–3 dorsal

semiannuli; scapular tubercles are widely separated and located near posterolateral margins of the shield; subdorsal ridges bordering the furrow are somewhat prominent. So far, three *Neooxycenus* species are known worldwide, with only one species, *N. plucheae* previously recorded from Egypt.

In addition, approximately 30 eriophyoid species have been recorded on amaranthaceous plants; 21 of which are from the genus *Aceria*, with only one, *Aceria lividus* Elhalawany, 2018, being recorded on *Amaranthus lividus* L. (Elhalawany 2018; Lewandowski et al. 2021, Ripka and Takacs 2021; Amrine and de Lillo personal communication 2022).

More than 1,044 species have been assigned to the genus *Aceria* Keifer, 1944 with approximately 51 of them being found in Egypt to date. With this study, Egyptian eriophyoid fauna increased to 121 species from 37 genera (Zaher 1984; Elhalawany 2012, 2015, 2018; Elhalawany and Ueckermann 2015, 2018, 2022; Elhalawany et al. 2014, 2018, 2019a, b, 2020, 2021, 2022; Halawa 2015).

Thus, the aim of this work is to describe a new eriophyid species, *Aceria newplucheae* **sp. nov.** on *P. dioscoridis*, as well as to complete the descriptions of the female, male and immature stages of a new record, *Aceria chenopodia*. Moreover, four eriophyoid mite species have been synonymized based on new samples collected in Egypt.

## MATERIALS AND METHODS

Live eriophyoid mites were collected from plant samples in four governorates: Qalyubia, Gharbia, Fayoum and Giza, Egypt. Specimens were cleared in Keifer's solution for two hr at 40°C and mounted on glass microscope slides in Keifer's F-medium (Amrine and Manson 1996). Mounted specimens were examined using a phase contrast (Carl Zeiss Nr, Germany) research microscope. According to de Lillo et al. (2010), line drawings were created by hand using a drawing tube attached to the phase contrast microscope. Generic classification is based on Amrine et al. (2003), and descriptions of all genera published after 2003. Morphological terminology mostly followed Lindquist (1996). All measurements are in micrometers (µm), and are taken with the software computer program (compuEye) (Bakr 2005). The range of the specimens (i.e., holotype and paratypes) is listed in parentheses after the measurements of the female holotype. Only the measurement ranges for males and immature stages are provided.

The type materials of the new species have been deposited at the mite reference collection of Fruit Tree Mites Department, Plant Protection Research Institute, Agricultural Research Centre, Dokki, Giza governorate, Egypt (PPRI-ARC). Furthermore, some paratypes have been deposited at the mite reference collection of the Egyptian Society of Acarology Museum (ESAM), Zoology and Agricultural Nematology Department, the Faculty of Agriculture, Cairo governorate, University. Giza Egypt; the Arthropod/Mite Collection of the Department of Entomology, Nanjing Agricultural University, Jiangsu Province (NJAU), China; and the mite collection of Department of Plant, Soil and Food Sciences (Di.S.S.P.A.), University of Bari Aldo Moro, Italy (UNIBA, formerly indicated as UBI by Zhang 2018).

The following abbreviations were used according to Amrine et al. (2003): AD. Anterodorsal view of mite; CGF. Coxi-genital region of female; D. Dorsal view of mite; DN. Dorsal view of nymph; em. Empodium; GM. Genital region of male; IG. Internal female genitalia; LO. Lateral view of semiannuli; L1. Leg I; L2. Leg II; LM. Lateral view of mite; VN. Ventral view of nymph.

#### **RESULTS AND DISCUSSION**

Eriophyidae Nalepa, 1898 Subfamily Eriophyinae Nalepa, 1898 Tribe Aceriini Amrine & Stasny, 1994 Genus *Aceria* Keifer, 1944 *Aceria newplucheae* sp. nov. (Figures 1–3, Table 1).

# **Description**

*Female* (n=16) body vermiform, 230 (170–238) long including gnathosoma, 45 (45-47) wide, 47 (40-54) thick; whitish in life. Gnathosoma 20 (18-20) long, projecting obliquely downwards, basal seta ep 2, antapical seta d 4 (3–4), cheliceral stylets 16 (15-16) long. Prodorsal shield 24 (21-25) long with short frontal lobe acuminate 3 (3–4), 30 (25 - 30)wide; subtriangular; prodorsal shield ornamentation generally faint, with dart-shaped mark of the faint median line and short admedians on posterior 1/4 of prodorsal shield and transverse lines extending laterally curved ahead of scapular tubercles, also admedians connected to two cells lateral on each

side of prodorsal shield, sub-median line absent. Scapular tubercles on rear shield margin, 17 (16-19) apart, setae sc 19 (19-22), projecting posteriorly. Coxigenital area smooth, with three genitalia. semiannuli between coxae and prosternal apodeme present 4 (4–5); anterolateral setae on coxisternum I *lb* 7 (7–8), 9 (9–10) apart; proximal setae on coxisternum I la 19 (14–19), 11 (10–11) apart; proximal setae on coxisternum II 2a 27 (21–28), 21 (20–22) apart. Leg I 25 (25– 27), femur 7 (7–8), basiventral femoral seta by 9 (9-10); genu 4 (3-4), antaxial genul seta l'' 21 (20-23); tibia 3 (3-4), paraxial tibial seta l' 4 (3-4), setae located  $\frac{1}{2}$  from dorsal base; tarsus 6 (6– 7); tarsal empodium simple em 5 (5-6), 4-rayed, tarsal solenidion  $\omega$  slightly tapered, 8 (7–9), paraxial fastigial tarsal seta ft' 15 (14–16), antaxial fastigial tarsal seta ft'' 18 (18–20), tarsal setae u' 1–2. Leg II 24 (23–25), femur 8 (8–9), seta bv 10 (9–11); genu 3, seta l'' 9 (8–10); tibia 3 (3-4); tarsus 6 (5-6); tarsal em simple 4 (4-5), 4rayed,  $\omega$  slightly tapered 8 (7–9), seta ft' 8 (7–9), seta ft'' 22 (20–22), tarsal seta u' 1–2. Opisthosoma with 52 (50-56) dorso-ventral semiannuli, subequal; dorsal semiannuli with oval-elongate microtubercles on rear annular margins, 8-14 caudal dorsal semiannuli without microtubercles; ventrally with oval microtubercles on rear annular margins, the last  $5^{\text{th}}$  ventral microtubercles linear. Lateral setae c217 (15–20), 43 (43–44) apart, on annulus 7 (7–8) from coxae II; ventral setae, d 38 (35-48), 30 (28–31) apart, on annulus 19 (19–21); ventral setae, e 10 (8-10), 27 (25-27) apart, on annulus 30 (30-32); ventral setae, III f 22 (20-26), 19 (18–20) apart, on  $5^{\text{th}}$  annulus from rear. Setae *h1* 4 (4–5); setae h2 55 (52–60). External genitalia 12 (12–13), 20 (18–21) wide, coverflap with ten longitudinal ridges in a single row, proximal setae on coxisternum III 3a, 10 (8-10), 18 (17-19) apart.

*Male* (n=4). Similar to female. Body vermiform, 130–188 long including gnathosoma, 30–41 wide and 38–40 thick; whitish in life. **Gnathosoma** 18–19, cheliceral stylets 14–15 long, setae ep 2–3, setae d 3–4. **Prodorsal shield** pattern similar to that of female, 21–22 long including frontal lobe, 25–27 wide; Scapular tubercles near the rear shield margin, 18–19 apart, setae sc 14–15, projecting diagonal posteriorly. **Coxigenital area** smooth, prosternal apodeme present 4–5; setae *1b* 5–6, 9–10 apart; setae *1a* 13–19, 10–11 apart; setae *2a* 20–22, 20–22 apart. **Leg I** 22–23, femur

8-9, seta bv 6-8; genu 3-4, seta l'' 17-20; tibia 3–4, seta l' 2–3; tarsus 5–6; empodium simple *em* 4–5, 4-rayed, tarsal solenidion  $\omega$  6–7 slight tapered, setae ft'13-15, setae ft'' 17-20, setae u'1-2. Leg II 20-21, femur 5-6, seta bv 6-8; genu 3-4, seta l'' 6-8; tibia 3; tarsus 4-5; em 4-5, 4rayed,  $\omega$  7–8 tapered, seta ft' 5–7, seta ft" 15–16, seta u' 1–2. **Opisthosoma** with 45–46 dorsal semiannuli and 50-53 ventral semiannuli, microtubercles shape similar to that of female. Lateral setae c2 14-16, 37-39 apart, on annulus 8 from coxae II; ventral setae d 26–28, 28–30 apart, on annulus 18-19; setae e 8-9, 25-26 apart, on annulus 27–28: setae f 17–18, 18–19 apart, on  $5^{\text{th}}$  annulus from rear. Setae h1 4–6; setae h2 40-46. External genitalia 12-13 long, 16-18 wide, surface below eugenital setae with granules, setae 3a 5-6, 14-15 apart.

Nymph (n=3). Body vermiform, 145–160 long including gnathosoma, 30-41 wide and 38-40 thick. Gnathosoma 14-15, cheliceral stylets 12-13 long, seta ep 1-2, seta d 3-4. Prodorsal shield subtriangular, smooth, 21–22 long, 25–27 wide; scapular tubercles on rear shield margin, 18–19 apart, setae sc 14–15, directed to the rear. **Coxigenital area** smooth; seta *1b* 4–5, 8–9 apart; seta 1a 11-13, 8-9 apart; seta 2a 15-16, 16-17 apart; seta 3a 6-7, 5-6 apart. Leg I 16-17, femur 4-5, seta by 4-5; genu 2-3, seta l'' 11-12; tibia 2–3, seta l' 2; tarsus 3; empodium simple em 4, 3rayed, tarsal solenidion  $\omega$  5 tapered, seta ft' 9–10, seta ft" 11–12, seta u' 1–2. Leg II 15–16, femur 3-4, seta by 4-5; genu 2-3, seta l'' 7-9; tibia 2-3; tarsus 3–4; em 3–4, 3-rayed,  $\omega$  4–5 tapered, seta ft' 7–8, seta ft'' 12–13, seta u' 1–2. **Opisthosoma** with 50-52 dorsal semiannuli and 44-46 ventral semiannuli, with linear elongate microtubercles situated on rear margin of dorsal semiannuli, pointed on ventral semiannuli, linear elongated on last four ventral semiannuli. Lateral setae c28–9, 35–37 apart, on annulus 8 from coxae II; ventral setae d 16–20, 33–34 apart, on annulus 17-18; setae e 5-6, 14-15 apart, on annulus 25-26; setae f 10–12, 18–19 apart, on  $4^{\text{th}}$  annulus from rear. Setae *h1* 1–2; setae *h2* 15–20.

*Larva* (n=3). Body vermiform, 130–145 long including gnathosoma and 38–40 wide. **Gnathosoma** 13–14, cheliceral stylets 11–12 long, seta *ep* 1–2, seta *d* 3–4. **Prodorsal shield** subtriangular, smooth, 16–17 long, 25–26 wide. Tubercles *sc* on rear shield margin, 14–15 apart, setae *sc* 9–10, directed to the rear.

Characters	<i>Aceria alfierii</i> (Dçbski,	Aceria dioscoridis †	Aceria dioscoridis 8	A. dioscoridis Current study	Aceria newplucheae
	1919) †	atoseoriais *	utoseottuis 3	Current Study	sp. nov.
Body length	180	217.5-232.5	214 (166–250)	215(180-243)	230 (170–238)
Body width	41	41.3–45	46 (37–51)	52(47-60)	45 (45–47)
body Thickness	-	-	46 (37–51)	48 (36–50)	47 (40–54)
Gnathosoma Length	_	19	15 (15–18)	18 (16–20)	20 (18–20)
Cheliceral stylets Length	_	-	11 (10–14)	12 (12–16)	16 (15–16)
Prodorsal shield Length	-	-	20 (20-25)	26 (24–30)	24 (21–25)
Prodorsal shield width	-	-	32 (30–36)	34 (32–39)	30 (25–30)
Seta sc length	15.6	19	19 (18–22),	19 (17–21)	19 (19–22)
Seta sc apart	_	12.5	12 (10–14)	20 (18-22)	17 (16–19)
Leg I length	37.4+ <i>em</i>	22.5	24 (19–24)	26 (22–30)	25 (25-27)
Leg II length	36.9 + em	21	19 (17–21)	24 (20–28)	24 (23–25)
Number of Empodial rays	4	4	4	4	4
Tarsal solenidion $\omega$	tapered	Tapered	Tapered	Tapered	Tapered
Setae 1b length	6.5	_	10 (9–12)	8 (8–9)	7 (7–8)
Tubercles 1b apart	_	_	6 (5–7)	9 (6–9)	9 (9–10)
Setae 1a length	16.1	_	26 (20-30)	20 (18–22)	19 (14–19)
Tubercles <i>1a</i> apart	_	_	5 (5-7)	9(9–11)	11 (10–11)
Setae 2a length	26.9	35	35 (31–39)	30 (25-30)	27 (21–28)
Tubercles 2 <i>a</i> apart	_	_	16 (15–18)	24 (22–24)	21 (20–22)
Coxae I & II surface	smooth	Smooth	Smooth	Smooth	Smooth
Dorsal semiannuli	<u>(</u> )		50 (54 64)		51 (50 50)
number	60	66	59 (54–64)	<u>55 (54–58)</u>	51 (50–52)
Ventral semiannuli	<b>C</b> 0				
number	60	66	56 (52–58)	57 (55-60)	55 (52–56)
Dorsal semiannuli shape	elongate	large spine	sharp conical	large spine	oval-elongate
Ventral semiannuli shape	elongate	large spine	sharp conical	spine	Oval
Setae <i>c2</i> length	13.1	17	18 (17–23)	19 (17–21)	17 (15–20)
On annulus	_	10	7 (7–8)	8 (7–8)	7 (7–8)
Setae <i>d</i> length	28.7	39	46 (42–50)	34 (30-46)	38 (35-48)
On annulus	_	26	18 (18–20)	19 (18–20)	19 (19–21)
Setae <i>e</i> length	6	11	12 (10–16)	12 (11–16)	10 (8–10)
On annulus	_	38	33 (29–34)	33 (32–43)	30 (30–32)
Setae <i>f</i> length	15	20	20 (17–24)	20 (18–22)	22 (20–26)
On annulus	_	5 <sup>th</sup>	5 <sup>th</sup>	5 <sup>th</sup>	5 <sup>th</sup>
Setae <i>h1</i> length	3.6	3.75	5 (5-7)	5 (5-6)	4 (4–5)
Setae <i>h2</i> length	41.3	52	53 (50-70)	40 (38–50)	55 (52-60)
Female genitalia length	16.4	14	11 (9–12)	12 (12–14)	12 (12–13)
Female genitalia width	_	19	22	19 (19–24)	20(18-21)
Ridges number	б	7	6 (5-9)	8 (8–10)	10
Setae <i>3a</i> length	10.1	11	15 (14–19)	12(12-14)	10 (8–10)
Relation to host	gall	Gall	Gall	Gall	vagrant on the leaf axil

Table 1. Differences between Aceria species associated with Pluchea dioscoridis in Egypt

<sup>†</sup>After Sayed (1946), <sup>‡</sup> after Soliman and Abou-Awad (1977), § after Lewandowski et al. (2021)

**Coxigenital area** smooth; seta *1b* 3–4, 8–9 apart; seta *1a* 9–10, 9–10 apart; seta *2a* 15–17, 17–18 apart; seta *3a* 3, 6–7 apart. **Leg I** 13–15, femur 3–4, seta *bv* 4–5; genu 2, seta *l''* 8–9; tibia 2, setae *l'* 1–2; tarsus 2–3; empodium *em* 3–4 simple, 3-rayed, tarsal solenidion  $\omega$  4–5 tapered, setae *ft'* 9–10, setae *ft''* 11–12, setae *u'* 1–2. **Leg II** 11–13, femur 3–4, seta *bv* 3–4; genu 2, seta *l''* 3–4; tibia 2–3; tarsus 2–3; *em* 3, 3-rayed,  $\omega$  4–5

tapered, seta ft' 5–7, seta ft'' 9–11, seta u' 1–2. **Opisthosoma** with 45–47 dorsal semiannuli and 40–42 ventral semiannuli, with pointed microtubercles on semiannuli margin. Lateral setae c2 6–7, 30–32 apart, on annulus eight from coxae II; ventral setae d 11–12, 26–27 apart, on annulus 16–17; setae e 4–5, 13 apart, on annulus 25; setae f 10–12, 17 apart, on 4<sup>th</sup> annulus from rear. Setae h1 1–2; setae h2 14–15.



**Figure 1.** Line drawings of *Aceria newplucheae* **sp. nov.** Scale bar: 10 μm for D, CGF, GM, IG; 10 μm for L1, L2; 2.5 μm for em.



Figure 2. Line drawings of Aceria newplucheae sp. nov. Scale bar: 10 µm for DN, VN, DL, VL.



**Figure 3.** Phase microphotograph of *Aceria newplucheae* **sp. nov.**: A. Dorsal shield of female B. Coxi-genital region of female; C. Legs; D. Internal female genitalia; E. Empodium F. Coxi-genital region of male. Scale bar: 10 μm for A, B, C, F, 20 μm for D and E.

55

**Host plant.** *Pluchea dioscoridis* (L.) DC. (Compositae).

**Relation to the host plant.** Vagrant on the leaf axil, no damage was observed.

Type Locality. El-Sad village, Qalyubia governorate  $(30^{\circ}15'50.46''N)$ , 31°14'51.85"E), Egypt 30 September, 2020 and 20 September 2021. governorate (30°1'8.21"N, Giza 31°12'28.04"E), Egypt, 20 October, 2020. Kafr governorate (30°41'56.00"E, Ibri. Gharbia 31°10'49.07"N), Egypt, 10 November, 2022; coll. A.S. Elhalawany.

**Type material.** Holotype female on a slide (slide no. EGYErio118.1), deposited at (PPRI-ARC), Egypt. Paratypes: 20 females, five males, five nymphs and four larvae on nine slides (EGYErio118.2 –118.10), deposited at (PPRI-ARC), Egypt. Four paratype females on two slides (EGYErio118.11 –118.12), deposited at (ESAM), Egypt. Two paratype females on two slides (EgPID05-06), deposited at (UNIBA), Italy. Four paratype females on two slides (NJAUAcariE20.1-20.2), deposited at (NJAU) China.

**Etymology.** The species name "*newplucheae*", is a composition of *new*, and + *plucheae*, referring to the specific name of the host plant.

Differential diagnosis. The new species collected from leaf axil of *P. dioscoridis* in Egypt was compared with all Aceria species found on Pluchea dioscoridis, it is similar to Aceria alfierii (Dcbski, 1919) and Aceria dioscoridis (Soliman & Abou-Awad, 1977) collected from galls on stem and leaves of P. dioscoridis from Egypt. The former species differs from the new species by having prodorsal shield smooth, with short curved line at rear third between tubercles (with dart-shaped mark of the faint median line, short admedians on posterior 1/4 of prodorsal shield, and transverse lines extending laterally curved ahead of scapular tubercles in A. newplucheae sp. **nov.**); length of setae *sc* 15.6 (19–22 in *A*. newplucheae sp. nov.), setae c2 13.1 (15-20 in A. newplucheae sp. nov.), setae d 28.7 (35–48 in A. newplucheae sp. nov.), setae e 6 (8–10 in A. newplucheae sp. nov.); number of dorsal semiannuli 60 (50-52 in A. newplucheae sp. nov.); and number of ridges on genital coverflap six (ten in A. newplucheae sp. nov.); furthermore A. alfierii caused galls on the leaves, whereas A. *newplucheae* **sp. nov. is a** vagrant on the leaf axil without causing damage (Table 1). Aceria dioscoridis differs by having incomplete median line at middle third, faint; admedian lines divergent to the rear, curved in the rear part, forming U-shape (with dart-shaped mark of the faint median line, short admedians on posterior <sup>1</sup>/<sub>4</sub> of the prodorsal shield, and transverse lines extending laterally curved ahead of scapular tubercles; submedian lines I and II present only in the rear part of the shield parallel to admedian lines and curved inwards just before the rear edge of the shield in *A. newplucheae* **sp. nov.**); with large spine (conical) microtubercles (ovalelongate microtubercles in *A. newplucheae* **sp. nov.**) (Table 1).

# Aceria dioscoridis (Soliman & Abou-Awad, 1977)

*Eriophyes dioscoridis* Soliman & Abou-Awad, 1977: 673.

Eriophyes dioscoridis — Davis et al. 1982: 125.

Aceria dioscoridis.—Zaher et al. 1978: 42; Amrine and Stasny 1994: 42; Chandrapatya et al. 2016: 63; Chandrapatya et al. 2017: 11.

Aceria mosalahi Lewandowski & Elsayed in Lewandowski et al. 2021: 1401 (**new** synonymy).

(Figures 4, 5, 6a, Table 1)

**Relation to the host plant.** The mite was found in gall on the upper and lower surfaces of the leaves and stem (Figure 6a).

**Type host.** *Pluchea dioscoridis* (L.) DC. (Compositae).

**Type locality.** Egypt (Soliman and Abou-Awad 1977).

Distribution in Egypt. All over Egypt.

**Geographic distribution.** Egypt; Thailand (Chandrapatya et al. 2017).

Materials examined. Twenty females and five males on five slides (slides no. EGYErio82.1-82.5) from P. dioscoridis, El-Sad village, (30°15'50.46"N, Oalyubia governorate 31°14'51.85"E), Egypt, 30 September 2020 and 20 September 2021, deposited at (PPRI-ARC), Egypt. Eight females and two males on two slides (slides no. EGYErio82.6-82.7), Kafr Ibri, Gharbia (30°41'56.00"E, governorate 31°10'49.07"N), 10 November, 2021, 10 February 2022, deposited at (ESAM), Egypt. Five females on two slides (slides no. EGYErio82.8-82.9), Fayoum governorate (29°20'0.48"N, 31°42'18.23"E), 20 March, 2021, deposited at (PPRI-ARC), Egypt. Two slides (slides no. EgPlD07-08), deposited at (UNIBA), Italy; coll. by A.S. Elhalawany.

56



**Figure 4.** Line drawings of *Aceria dioscoridis* (Soliman & Abou-Awad, 1977)**:** Scale bar: 10 μm for AD, CGF, CGM, IG; 5 μm for L1, L2; 2.5 μm for em.





**Figure 5.** Phase microphotograph of *Aceria dioscoridis*: A. Ventral view of female B. Dorsal view of female; C. Legs; D. Internal genitalia; E. Empodium. Scale bar: 10 μm for A, B; 20 μm for C, D and E.



Figure 6. Galls induced by *Aceria dioscoridis* on leaves and stem of *Pluchea dioscoridis* (A); *Chenopodium murale* L. (B); *Chenopodium album* L. (C); *Amaranthus lividus* L. (D).

## Remarks

Aceria dioscoridis was originally described by Soliman and Abou-Awad (1977) in Egypt based on adult female. Lewandowski et al. 2021 described female, male and immature stages of this mite species under the name Aceria mosalahi Lewandowski & Elsayed inducing galls on the leaves and stems of Chenopodiastrum murale (L.) (=Chenopodium murale L.) (Amaranthaceae) Figure 6b, but the host plant is incorrectly identified as shown in Lewandowski et al. 2021 (page 1400: Figure 1a). *Pluchea dioscoridis* is the correct host plant (Figure 6). The female morphometrics match the original description by Soliman and Abou-Awad (1977) (Table 1).

Aceria chenopodia Xue, Sadeghi & Hong, 2009: 461.(Figures 7–8, Table 2)

Female (n=10) body vermiform, 250 (198-280) long including gnathosoma, 40 (37-49) wide, 45 (41-45) thick; light yellow in life. Gnathosoma 22 (20 - 24)long, projecting obliquely downwards, basal setae ep 3 (2-3), antapical setae d 7 (7–8), cheliceral stylets 20 (18–22) long. Prodorsal shield 30 (26-30) long with short frontal lobe acuminate, 35 (32–39) wide; sub-triangular; prodorsal shield ornamentation with median line complete, broken medially; admedian and first submedian lines complete, gently diverging to rear, and parallel, second submedian lines incomplete on posterior third of prodorsal shield ahead of scapular tubercles; lateral lines complete parallel to lateral shield; numerous granules in area between median and admedian lines, and lateral sides of prodorsal shield. Scapular tubercles on rear shield margin, 27 (26–29) apart, setae sc 48 (39–50), projecting posteriorly. Coxigenital area with minor granules, with 6 (6–7) semiannuli between coxae and genitalia, prosternal apodeme present 6 (5-7); setae 1b 8 (7-9), 12 (11-12) apart; setae 1a 21 (19–25), 9 (9–10) apart; setae 2a 45 (31–46), 23 (22–24) apart. Leg I 35 (31–35), femur10 (8– 10), basiventral femural seta bv 10 (10-12); genu 5 (5–6), antaxial genual seta l'' 25 (22–26); tibia 7 (7–8), paraxial tibial seta l' 7 (6–8), seta located <sup>1</sup>/<sub>4</sub> from dorsal base; tarsus 7 (6–8); empodium em simple 6 (6–7), 5-rayed, simple, tarsal  $\omega$  slightly tapered, 8 (8–9), tarsal seta ft' 15 (14–17), tarsal seta ft" 22 (20–23), tarsal seta u' 5–6. Leg II 29 (29–30), femur 7 (7–8), seta bv 10 (8–10); genu 5 (4-5), seta l'' 10 (9-10); tibia 5 (5-6); tarsus 7 (7-8); tarsal *em* simple 6 (6–7), 5-rayed,  $\omega$  8 (7– 9) slightly tapered, seta ft' 9 (9–11), seta ft'' 23 (22-26), tarsal seta u' 5–6. **Opisthosoma** dorsally with 67 (65-70) semiannuli, with elliptical microtubercles on rear annular margins, last 9-10 semiannuli with pointed microtubercles; ventrally with 65 (58-65) semiannuli, with elliptical to round microtubercles on rear annular margins, the last 9-10<sup>th</sup> ventral microtubercles linear. Lateral setae c2 50 (37-50), 39 (38-42) apart, on annulus 10 (9–10) from coxae II; ventral setae d55 (40–55), 36 (35–38) apart, on annulus 23 (22– 23); ventral setae e 40 (25–40), 21 (20–23) apart, on annulus 42 (41–43); ventral setae f 34 (25– complete, gently diverging to rear, and parallel, short line between admedian and submedian lines 36), 20 (20–21) apart, on 6<sup>th</sup> annulus from rear. Setae h1 7 (7–8), setae h2 57 (52–70). **External** genitalia 16 (14–18) long and 24 (23–25) wide, coverflap with ten longitudinal ridges in a single row plus three transverse lines at the base, proximal setae on coxisternum III 3a, 40 (36– 45), 19 (18–19) apart. **Internal genitalia** spermathecae ovoid, oriented posterolaterad; spermathecal tubes relatively short; transverse genital apodeme trapezoidal, distally folded.

(n=10). Similar to female. Body Male vermiform, 160-220 long including gnathosoma, 32–35 wide and 42–45 thick; light yellow in life. Gnathosoma 20-24, cheliceral stylets 18-20 long, setae ep 3-4, setae d 5-6. Prodorsal shield shape and patterns similar to those of female, 26-29 long including frontal lobe, 27-29 wide; scapular tubercles on the rear shield margin, 21-22 apart, setae sc 29-40, projecting diagonal Coxigenital area with minor posteriorly. granules, prosternal apodeme present 5-6; setae *1b* 7–8, 11–12 apart; setae *1a* 20–23, 8–10 apart; setae 2a 28-32, 18-22 apart. Leg I 30-32, femur 9–10, seta by 7–10; genu 5–6, seta l'' 22–25; tibia 6-7, seta l' 6-7; tarsus 7-8; tarsal empodium em simple 6–7, 5-rayed, tarsal solenidia  $\omega$  distally slight tapered 8–9, seta ft'13–15, seta ft" 19–21, seta u' 3–4. Leg II 27–28, femur 8–9, setae bv 7– 9; genu 4–5, setae l'' 9–10; tibia 5–6; tarsus 5–6; tarsal empodium em simple 6-7, 5-rayed, tarsal solenidia  $\omega$  distally slight tapered 8–10, setae ft' 9–10, setae ft'' 20–22, setae u' 3–4. Opisthosoma dorsally with 65-70 semiannuli; ventrally with 60-62 semiannuli, microtubercles shape similar to those of female. Lateral setae  $c^2$  30–33, 30–33 apart, on annulus ten from coxae II; ventral setae d 44-60, 30-31 apart, on annulus 22-23; setae e 35–40, 22–23 apart, on annulus 38–39; setae f22–35, 16–17 apart, on 6<sup>th</sup> annulus from rear. Setae h2 46–50; setae h1 5–6. External genitalia 13–15 long and 18–21 wide, with granules, setae *3a* 35–38, 18–19 apart.

*Nymph* (n=5). Body vermiform, 150–170 long and 36–40 wide. **Gnathosoma** 22–23, curved downward, setae *ep* 1–2, *d* 5–6; cheliceral stylets 15–17 long. **Prodorsal shield** sub-circular, 26– 27, including frontal lobe 2–3 over the gnathosomal base 30–35 wide, prodorsal shield ornamentation with median line complete, broken at middle; admedian and submedian lines at base of prodorsal shield, lateral line incomplete, on posterior third of prodorsal shield; numerous granules in area between median and admedian lines, and lateral sides of prodorsal shield. Tubercles *sc* on rear shield margin, 21–22 apart; *sc* 29–40. **Coxisternal plates** smooth, *1b* 4–5, 8–9 apart; *1a* 12–13, 6–7 apart; *2a* 19–22, 18–19 apart; *3a* 6–7, 5–6 apart. **Leg I** 19–22; femur 4–5, *bv* 6–7; genu 3–4, *l''* 13–15; tibia 3–4, *l'* 4–5; tarsus 4–5, *ft'* 13–15, *ft''* 18–20, setae *u'* 2– 3; tarsal  $\omega$  5–6; *em* 4–5, simple, 5-rayed. **Leg II** 16–18; femur 4–5, *bv* 7–8; genu 3–4, *l''* 6–8; tibia 3–4; tarsus 4–5, *ft'* 7–9, *ft''* 18–20, setae *u'* 2–3;  $\omega$ 5–6; *em* 4–5, simple, 5-rayed. **Opisthosoma** with 50-54 dorsal semiannuli, with elliptical microtubercles situated on rear margin of each annulus, 48-53 ventral semiannuli with minute round microtubercles, situated on rear margin of each annulus; elongated on the posterior semiannuli. Setae c2 27-30, 35-37 apart, on ten ventral annuli; setae d 31-43, 21-22 apart, on 19-20 ventral annuli; setae e 24-26, 14-15 apart, on 28-29 ventral semiannuli; setae f 20-22, 15-16 apart, on 5<sup>th</sup> annulus from rear. Setae h1 2–3; *h2* 40–58.

**Table 2.** Measurements of females and males of *Aceria chenopodia* associated with *C. album* and *C. murale* in Egypt and Iran, the measurement precedes the mite corresponding range for paratypes (given in parentheses).

	From Iran	From Egypt			
Characters	Females	Females	Males	Nymphs	Larvae
	n= 8	n= 10	n= 5	n=5	n= 3
Body length	255 (255-262)	250 (198-280)	160-220	150-170	110-130
Body width	53 (52–56)	40 (37–49)	32–35	36–40	27-30
Gnathosoma length	20 (20-23)	22 (20-24)	20-24	22–23	13–14
Cheliceral stylets length	17 (16–17)	20 (18-22)	18-20	15-17	12-13
Prodorsal shield length	28 (28-30)	30 (26–30)	26-29	26-27	20-21
Prodorsal shield width	42 (40-43)	35 (32–39)	27-29	30–35	24-25
Seta sc length	58 (56-63)	48 (39–50)	29-40	35-37	12-14
Distance between sc	22 (22–23)	27 (26–29)	21-22	18–19	11-12
Leg I length	40 (38–41)	35 (31–35)	30-32	19–22	19–20
Leg II length	36 (35–38)	29 (29–30)	27-28	16–18	15–17
Number of empodial rays	6	5	5	5	4
Seta 1b length	12 (12–13)	8 (7–9)	7–8	4–5	4–5
Distance between 1b	12 (12–13)	12 (11–12)	11-12	8–9	6–7
Seta <i>1a</i> length	31 (31–35)	21 (19–25)	20-23	12–13	12-13
Distance between 1a	10 (9–10)	9 (9–10)	8-10	6–7	4–5
Seta 2a length	45 (43–48)	45 (31–46)	28-32	19–22	16–18
Distance between 2a	23 (23–24)	23 (22–24)	18-22	18–20	13–14
Seta 3a length	33 (33–34)	40 (36–45)	35–38	6–7	3–4
Dorsal semiannuli number	65 (60–66)	67 (65–70)	65-70	50–54	49–52
Ventral semiannuli number	62 (58–64)	65 (58–65)	60–62	48–53	38–40
Seta <i>c2</i> length	50 (50-53)	50 (37–50)	30–33	27-30	19–25
Seta <i>c2</i> on semiannuli	10 (10–11)	10 (9–10)	10	10	9–10
Seta <i>d</i> length	53 (46–53)	55 (40–55)	44-60	31–43	17-20
Seta d on semiannuli	22 (21–22)	23 (22–23)	22-23	19–20	17–18
Seta <i>e</i> length	52 (52–56)	40 (25–40)	35–40	24–26	9–12
Seta <i>e</i> on semiannuli	39 (39–39)	42 (41–42)	38–39	28–29	25-26
Seta $f$ length	33 (31–33)	34 (25–36)	22-35	20-22	13–15
Seta $f$ on semiannuli	$6^{\text{th}}$	$6^{\text{th}}$	$6^{th}$	$5^{\rm th}$	$4^{\text{th}}$
Seta <i>h1</i> length	7 (7–8)	7 (7–8)	5–6	2–3	2–3
Seta h2 length	83 (80-89)	60 (52–70)	46–50	40–58	30–33
Genital coverflap length	14 (13–15)	16 (14–18)	13–15	_	—
Genital coverflap width	24 (21–25)	24 (23–25)	18–21	_	—
Longitudinal ridges	8	10	—	_	_



**Figure 7.** Line drawings of *Aceria chenopodia* Xue, Sadeghi & Hong, 2009: Scale bar: 10 μm for AD, Al, CGF, CGM, IG, PM; 5 μm for L1, LO, L2; 2.5 μm for em.



**Figure 8.** Line drawings of *Aceria chenopodia* Xue, Sadeghi & Hong, 2009: Scale bar: 10 μm for DN, VN, DL, VL.

*Larva* (n=3). Body vermiform, 110–130 long and 27 - 30wide. Gnathosoma 13–14 curved downward, setae ep 1–2, d 3–4, cheliceral stylets 12-13 long. Prodorsal shield sub-circular, 20-21 long and 24-25 wide; with incomplete median line broken at middle, first sub-median short at base 1/4 between scapular tubercles, second submedian lines complete sub-parallel to admedian lines; granules present on lateral sides of prodorsal shield. Tubercles sc near the rear shield margin, 11-12 apart; setae sc 12-14 directed anteriorly. Coxisternal plates smooth, setae 1b 4-5, 6-7 apart; *1a* 12-13, 4-5 apart; *2a* 16-18, 13-14 apart. Setae 3a 3-4, 6-7 apart. Leg I 1920; femur 4–5, bv 4–5; genu 3–4, l'' 8–9; tibia 3– 4, setae l' 5–6; tarsus 3–4, ft' 9–10, ft'' 13–15;  $\omega$ 5–6; em 4–5, simple, 4-rayed. **Leg II** 15–17; femur 4–5, bv 5–6; genu 2–3, l'' 5–7; tibia 2–3; tarsus 3–4, ft' 6–8, ft'' 13–15;  $\omega$  5–6; em 4–5, 4rayed. **Opisthosoma** dorsally with 49–52 semiannuli, ventrally with 38–40 semiannuli, with minute round microtubercles situated on rear margin of each annulus. Setae c2 19–25, 24– 25 apart, on 9–10 ventral semiannuli; setae d 17– 20, 17–18 apart, on 17–18 ventral semiannuli; setae e 9–12, 10–11 apart, on 25–26 ventral semiannuli; setae f 13–14, 13–14 apart, on 4<sup>th</sup> annulus from rear; h1 2–3; h2 30–33. Host plants in Egypt. Chenopodium album L. (Figure 6c) and Chenopodium murale L (Figure 6b). (Amaranthaceae).

**Distribution.** Iran and Egypt.

Relation to the host plant. Vagrant on buds and flowers, no damage symptom to the host was observed.

Type Locality. Razavi Khorasan Province, Iran.

(slide no. EGYErio122.1), from C. album, Toukh Egypt, 15 October, 2015 and 2021, deposited at distinct, Qalyubia governorate (30°17'20.02"N; (PPRI, ARC), Egypt. Ten females and two males October, 31°14'51.85"E), Egypt, 30 deposited at (PPRI-ARC), Egypt. Twenty females, with the same previous data, deposited at five males, six nymph and four larvae on five (ESAM), Egypt. Two slides (slides no. EgAV01slides (slide no. EGYErio122.2-122.6), from C. 02), deposited at (UNIBA), Italy. Two slides murale, deposited at (PPRI-ARC), Egypt. Four (slides no. NJAUAcariE19.1-19.2), deposited at females and two males on two slides from C. (NJAU) China, coll. by A.S. Elhalawany. *murale*, with the same previous data, deposited at (ESAM), Egypt; coll. A.S. Elhalawany.

#### **Remarks.**

This is the first record of A. chenopodia females and males in Egypt. Xue et al. (2009) described this species based on adult female from Iran. Except for the lengths of the following characters, the morphometry of the female resembles the original description: gnathosoma, body width, prodorsal shield width, cheliceral stylets length, lengths of setae c2 and legs, and number of ridges on genital coverflap. Setae sc, 1b, 1a, e, and h2 are slightly longer in Iranian specimens than Egyptian specimens; genital coverflap with ten longitudinal striae in Egyptian specimens (eight in Iranian specimens); and tarsal empodium in Iranian specimens is 6-rayed (5-rayed in Egyptian specimens) (Table 2).

# Aceria lividus Elhalawany, in Elhalawany et al., **2018 (Figures 6d, 9, Table 3)**

Aceria lividus Elhalawany, in Elhalawany et al., 2018: 18.

Aceria noxia Flechtmann & Tassi, 2020: 312, figures 1–6 (new synonym).

Type data. Amaranthus lividus L.

(Amaranthaceae).

Geographic distribution. Egypt, Brazil.

Host plant in Egypt. Amaranthus lividus L. and A. viridis L. (Amaranthaceae).

Relation to host. Vagrant on buds and flowers causes stunted, deformed flowers (Figure 6d).

Remarks. Elhalawany et al. (2018) reported this species from Egypt. Following the description of Aceria noxia Flechtmann & Tassi, 2020, the authorof the current study contacted Prof. Carlos Flechtmann to inquire whether he was aware of the

description of Aceria lividus on Amaranthus lividus in Egypt (Elhalawany et al. 2018). The Egyptian specimens slightly differ from the Brazilian specimens by small differences in the length of setae *sc* and *c2* (Table 3).

Material examined. Forty females, ten males, six nymphs, and five larvae paratypes on 20 slides (slide no. EGYErio62.1-62.20), Qalyubia Material examined. Single female on a slide governorate (30°15'50.46"N, 31°14'51.85"E), 2020, on two slides (slides no. EGYErio62.21-62.22),

> Table 3. Measurements of females of Aceria noxia and Aceria lividus associated with Amaranthus lividus and Amaranthus viridis from Brazil and Egypt.

	Acoria noria	Aceria	
Characters	Erom Drozil	lividus	
	From Brazil	From Egypt	
Dody low oth	224 (160-	200 (173-	
Body length	227)	213)	
Body width	61 (50–62)	52(50-62)	
Gnathosoma length	—	25 (25–28)	
Cheliceral stylets length	8 (8–16)	17 (16–18)	
Prodorsal shield length	31 (28–33)	34 (33–35)	
Prodorsal shield width	49 (43–49)	45 (42–50)	
Seta sc length	18 (18–23)	26 (24–27)	
Distance between sc	26 (20-26)	30 (26–30)	
Leg I length	38 (40–33)	33 (30–35)	
Leg II length	_	29(28-32)	
Number of Empodial rays	5	6	
Seta 1b length	8 (8–10)	7 (7–9)	
Seta 1a length	27 (18–27)	23 (22–25)	
Seta 2a length	40 (40-45)	27 (25–33)	
Seta 3a length	18 (13–18)	17 (15–18)	
Dorsal semiannuli	02(76,02)	65 (62 69)	
number	95 (70-95)	03 (03–08)	
Ventral semiannuli	71 (61 76)	65 (63 68)	
number	/1 (01-70)	03 (03–08)	
Seta <i>c2</i> length	40 (28–40)	25 (25–27)	
Seta d length	57 (42–58)	55 (53–60)	
Seta <i>e</i> length	14 (11–15)	11 (9–12)	
Seta f length	27 (26–28)	25 (24–30)	
Seta <i>h1</i> length	4	3 (3–4)	
Seta <i>h2</i> length	77 (65–77)	65 (64–70)	
Genital coverflap length	15 (15–18	16 (15–18)	
Genital coverflap width	21 (21–24)	23 (22–25)	
Longitudinal ridges	16 (14–16)	12-14	



**Figure 9.** Line drawings of *Aceria lividus* Elhalawany, 2018: Scale bar: 10 μm for AD, AL, CGF, GM, IG; 5 μm for L1, L2; 2.5 μm for em.

#### Aceria tosichella Keifer, 1969

Aceria tosichella Keifer, 1969a: 2. Eriophyes tulipae Keifer, 1938: 185.

Aceria tulipae.—Keifer 1944: 22, 1954: 123.

*Aceria tosichella.*—Keifer et al.1982: 160; Amrine and Stasny 1994: 92; Navia et al. 2006: 189; Skoracka et al. 2005: 64; Xue et al. 2011:143; Denizhan et al. 2015: 30.

Aceria aegyptiacus Soliman & Abou-Awad, 1977 (new synonymy).

Figure (10 A-E)

**Type data.** *Triticum sativum* Lam. (Poaceae); Zemun-Beograd, Serbia.

**Geographic distribution.** Africotropical, Antarctic, Australian, Indomalayan, Nearctic, Neotropical,

Palaearctic (Denizhan et al. 2015).

Host plant in Egypt. *Bromus catharticus* Vahl (Poaceae).

**Relation to host.** The mites live at the leaf bases, under the leaf surface, they cause the leaves to curl.

**Remarks.** Aceria aegyptiacus was described by Soliman and Abou-Awad (1977), based on a single female collected from litter and weeds in orange

In the present study A. tosichella was collected on four nymphs and six larvae on ten slides (slides the weed, Bromus catharticus also in citrus no. EGYErio12.1-12.10) from B. catharticus, in Qalyubia governorate, Egypt. Vahl, orchards at Comparing the morphological characters of both A. Qalyubia governorate, Egypt, 23 February 2021, aegyptiacus and A. tosichella, no consistent deposited at (PPRI-ARC), Egypt, coll. A.S. differences were found. Thus, Aceria aegyptiacus is Elhalawany. Ten females and two males on two a junior synonym of Aceria tosichella.

Egypt.

orchards at Luxor and Qena governorates, Egypt. Material examined. Twenty females, five males, Kaha (30°17'20.1"N 31°12'45.2"E), slides (slides no. EGYErio12.11-12.12), with the same previous data, deposited at (ESAM), Egypt. Distribution in Egypt. Qalyubia governorate, Five females on two slides (slides no. NJAUAcariE5.1-5.2), deposited at (NJAU), China.



Figure 10. Phase microphotograph of Aceria tosichella Keifer, 1969: A. prodorsal shield of female B. Coxi-genital region of female; C. Genital male; D. Internal genitalia; E. Empodium. Scale bar: 10 µm for A, B, C; 20 µm for D and E.

# Subfamily Phyllocoptinae Nalepa, 1892 Tribe Anthocoptini Amrine & Stasny, 1994 *Neooxycenus plucheae* Abou-Awad, 1981

*Neooxycenus plucheae* Abou-Awad, 1981: 368, figure 1.

*Vittacus plucheae* Abou-Awad & Nasr, 1986: 159 figure 1 (**new synonym**).

(Figures 11–14, Table 4).

# **Redescription**

*Female* (n=15) body fusiform, 194 (183–216) long including gnathosoma, 65 (61-69) wide, and light 70 (65 - 72)thick; yellow in life. Gnathosoma 28 (26 - 31)long. projecting obliquely downwards, basal setae ep 2 (2–3), antapical setae d 6 (5–6), cheliceral stylets 16 (16–17) long. Prodorsal shield 75 (64–75) long, including round frontal lobe 7 (6-8), 65 (65-70) wide; oval; prodorsal ornamentation obscure and with variations, two slight shoulders anteriorly, middle portion broad, posterior portion narrow and appears as a projection covering the first three dorsal semiannuli; median and submedian lines absent, admedian lines incomplete meeting and forming flask shape, sometimes with short transverse line at middle between admedian lines forming hexagonal shape at middle of the shield (Figure 14). Scapular tubercles on rear shield margin widely separated, 50 (45-50) apart, setae sc 8 (7–9), directed posteriad divergently. Coxigenital area smooth, with 4 (3-4)semiannuli between coxae and genitalia, sternal line forked; anterolateral setae on coxisternum I 1b 8 (7-8), 13 (13-14) apart; proximal setae on coxisternum I 1a 23 (21-25), 11 (11-12) apart; proximal setae on coxisternum II 2a 36 (32–41), 28 (27-31) apart. Leg I 31 (30-33), femur 9 (8-10), basiventral femural seta bv 9 (9-11); genu 6 (5-6), antaxial genual seta l'' 19 (18-20); tibia 6 (6-7), seta l' 3 (3-4), seta located in middle of segment; tarsus 7 (6-8); empodium em simple 6 (5–6), 4-rayed, simple, tarsal  $\omega$  slightly knobbed, 8 (7–9), tarsal seta ft' 18 (17–19), tarsal seta ft''22 (20–23), tarsal seta u' 2–3. Leg II 29 (27–30), femur 8 (8–9), seta by 9 (9–11); genu 5 (4–5), seta l'' 10 (6–10); tibia 6 (5–6); tarsus 7 (7–8); tarsal em simple 6 (5–6), 4-rayed,  $\omega$  8 (7–8) slightly knobbed, seta ft' 7 (5-8), seta ft" 20 (20-22), tarsal seta u' 2–3. Opisthosoma with two subdorsal ridges, prominent, begin near scapular tubercles and ending before setae f. Dorsally with (24–26) semiannuli, smooth, last 3–4 25 semiannuli with pointed microtubercles; ventrally with 70 (63–71) semiannuli, with round microtubercles on rear annular margins, the last  $13-17^{\text{th}}$  ventral semiannuli with microtubercles linear. Lateral setae c2 9 (8–10), 55 (54–60) apart, on annulus 10 (10–14) from coxae II; ventral setae d 32 (30–35), 32 (31–34) apart, on annulus 22 (22–26); ventral setae e 14 (14–15), 14 (14–15) apart, on annulus 38 (38–42); ventral setae III f 22 (22–23), 20 (20–21) apart, on 5<sup>th</sup> annulus from rear. Setae h1 3 (no range), setae h2 40 (35–45). **External genitalia** 14 (13–16), 25 (24–26) wide, coverflap with 10 (9–10) longitudinal ridges in a single row plus three transverse lines at the base, proximal setae on coxisternum III 3a 15 (12–15), 12 (12–14) apart.

Male (n=3). Similar to female. Body fusiform, 155–170 long including gnathosoma, 55–58 wide, and 63-67 thick; light yellow in life. Gnathosoma 25–29, cheliceral stylets 15–16 long, setae ep 2–3, setae d 4–5. Prodorsal shield shape and patterns similar to those of female, 60-65 long including frontal lobe, 50-55 wide; scapular tubercles on the rear shield margin, 40-45 apart, setae sc 7–8, projecting diagonal posteriorly. Coxigenital area smooth, sternal line forked; setae 1b 6-7, 11-12 apart; setae 1a 22-23, 9-10 apart; setae 2a 32-35, 27-29 apart. Leg I 24–26, femur 8–9, seta by 8–9; genu 4–5, seta l'' 18-19; tibia 5-6, seta l' 3-4; tarsus 5-6; tarsal empodium em simple 5-6, 4-rayed, tarsal solenidia  $\omega$  distally knobbed 7–8, seta ft'15–18, seta ft" 19–21, seta u' 2–3. Leg II 24–25, femur 7-8, seta bv 8-9; genu 4-5, seta l'' 8-9; tibia 5-6; tarsus 5-6; tarsal empodium em simple 5-6, 4rayed, tarsal solenidia  $\omega$  distally knobbed 7–8, seta ft' 5-8, seta ft" 19-21, setae u' 2-3.

**Opisthosoma** similar to female, dorsally with 24–25 semiannuli; ventrally with 50–55 semiannuli, microtubercles shape similar to those of female. Lateral setae c2 8–9, 50–55 apart, on semiannuli 10–11 from coxae II; ventral setae d 30–31, 29–30 apart, on semiannuli 19–20; setae e 13–14, 12–13 apart, on semiannuli 31–32; setae f 20–21, 18–19 apart, on 5<sup>th</sup> annulus from rear. Setae h1 3; setae h2 28–30. **External genitalia** 12–13 long and 18–19 wide, with granules, setae 3a 13–15, 13–15 apart.



**Figure 11.** Line drawings of *Neooxycenus plucheae* Abou-Awad, 1981**:** Scale bar: 10 μm for D, CGF, GM, IG, LM; 5 μm for L1, L2; 2.5 μm for em.



**Figure 12.** Line drawings of *Neooxycenus plucheae* Abou-Awad, 1981: Scale bar: 10 μm for DN, VN, DL, VL.



**Figure 13.** Phase microphotograph of *Neooxycenus plucheae* Abou-Awad, 1981: A. Dorsal view of female B. Ventral view of female; C. Ventral view of male; D. Internal genitalia; E. Empodium. Scale bar: 10 μm for A, B, C; 20 μm for D.

Nymph (n=4). Body fusiform, 140–150 long and 49–50 wide. Gnathosoma 17–18, curved downward, setae ep 1–2, d 3–4, cheliceral stylets 12-15 long. Prodorsal shield sub-circular, 54-66, including frontal lobe 3–4 over the gnathosomal base 50-54 wide. prodorsal ornamentation obscure and with hexagonal shape at middle of the shield. Tubercles sc on rear shield margin, 31-39 apart; sc 6-8. Coxisternal plates smooth, 1b 5-6, 13-14 apart; 1a 11-13, 8-9 apart; 2a 22-23, 28-30 apart; 3a 7-9, 10-12 apart. Leg I 22-24; femur 6-7, bv 7-8; genu 3-4, l'' 13-15; tibia 3-4, l' 2-4; tarsus 5-6, ft' 10-12, ft" 16–18, seta u' 1–2; tarsal  $\omega$  5–6; em 4–5, simple, 3-rayed. Leg II 19–20; femur 5–6, bv 7– 8; genu 3, l'' 6-7; tibia 3-4; tarsus 4-5, ft' 6-7, ft" 15–17, seta u' 1–2;  $\omega$  5–6; em 4–5, simple, 3-**Opisthosoma** with 25 - 26dorsal rayed. semiannuli, with elliptical microtubercles situated on rear margin of each annulus, 45-48 ventral semiannuli with pointed microtubercles, situated on rear margin of each annulus; elongated on the posterior semiannuli. Setae c26-7, 42-44 apart, on ten ventral annuli; setae d 15–16, 25–26 apart, on 22–23 ventral semiannuli; setae e 10-11, 14-15 apart, on 31-32 ventral semiannuli; setae f 16–17, 18–20 apart, on  $4^{\text{th}}$ annulus from rear. Setae h1 1-2; h2 30-33.

Larva (n=3). Body fusiform, 125–135 long and 40-45 wide. Gnathosoma 15–16, curved downward, setae ep 1–2, d 3–4, cheliceral stylets 12-13 long. Prodorsal shield sub-circular, 54-58, including frontal lobe 3–4 over the gnathosomal base 48–55 wide, prodorsal ornamentation obscure and with hexagonal shape at middle of the shield. Tubercles sc on rear shield margin, 25-27 apart; sc 6-7. Coxisternal plates smooth, 1b 3-4, 10-11 apart; 1a 8-9, 6-7 apart; 2a 17-18, 15-16 apart; 3a 5-6, 7-8 apart. Leg I 19–20; femur 5, bv 4–5; genu 2–3, l'' 10– 11; tibia 2–3, l' 2–3; tarsus 4, ft' 10–11, ft" 13–14, seta u' 1–2; tarsal  $\omega$  4–5 knobbed; em 4, simple, 3-rayed. Leg II 16–17; femur 5, bv 4–5; genu 2, l" 5; tibia 3; tarsus 4, ft' 6–7, ft" 13–14, seta u' 1– 2;  $\omega$  4–5; *em* 4, simple, 3-rayed. **Opisthosoma** with 23-24 dorsal semiannuli, with elliptical

microtubercles situated on rear margin of each annulus, 37–41 ventral semiannuli with pointed microtubercles, situated on rear margin of each annulus; elongated on the posterior semiannuli. Setae c2 4–5, 32–33 apart, on 8–9 ventral annulus; setae d 10–12, 19–20 apart, on 18–19 ventral annulus; setae e 7–8, 10–11 apart, on 27–28 ventral annulus; setae f 10–11, 11–12 apart, on 4<sup>th</sup> annulus from rear. Setae h1 1–2; h2 16–20.

**Host plant.** *Pluchea dioscoridis* (L.) DC. (Compositae).

**Distribution in Egypt.** Qalyubia, Gharbia, Kafr El-Sheikh.

**Relation to the host plant.** Vagrant on the lower leaf surface, no damage symptom to the host was observed.

Material examined. Twenty-two females, four males, three nymphs and two larvae on ten slides (slide no. EGYErio83.1-83.10), from Р. dioscoridis, Qalyubia governorate (30°15'50.46"N, 31°14'51.85"E), Egypt, 30 September, 2020 and 20 September, 2021, deposited at (PPRI-ARC), Egypt. Four females and one male on two slides (slide no. EGYErio83.11-83.12), Gharbia governorate (30°41'56.00"E, 31°10'49.07"N), 10 November, 2021, 10 February, 2022, deposited at (ESAM), Egypt. Two slides (slides no. EgPlD03-04), deposited at (UNIBA), Italy; coll. by A.S. Elhalawany.

#### Remarks.

described Abou-Awad (1981)Neooxycenus plucheae from only females collected on Pluchea dioscoridis. Abou-Awad and Nasr (1986) described the same species as Vittacus plucheae based on the variations in the design of the prodorsal shield. Females, males and immature stages of N. plucheae were collected in the current study. After examining many samples from different localities in Egypt, the morphometry of the female matches the original description by Abou-Awad (1981) (Table 4). Therefore, the current research suggests Vittacus plucheae is a junior synonym of Neooxycenus plucheae.

Characters	Vittacus	Neooxycenus	N. plucheae	N. plucheae
	plucheae †	plucheae ‡	female (15)	male (3)
Body length	145–183	123–163	194 (183–216)	155–170
Body width	53–59	55-63	65 (61–69)	55–58
Body thickness	_	_	70 (65–72)	63–67
Gnathosoma length	19	25	28 (26–31)	25–29
Cheliceral stylets length	15	_	16 (16–17)	15–16
Prodorsal shield length	55	63	75 (64–75)	60–65
Prodorsal shield width	53	54	65 (65–70)	50–55
Setae <i>sc</i> length	6	7.5	8 (7–9)	7–8
Setae sc apart	33	36	50 (45-50)	40–45
Leg I length	24	29	31 (30–33)	24–26
Leg II length	21	27	27 (27–32)	24–25
Number of empodial rays	4	4	4	4
Tarsal solenidion $\omega$	knobbed	Knobbed	Knobbed	Knobbed
Setae <i>1b</i> length	_	—	8 (7–8)	6–7
Tubercles 1b apart	_	_	13 (13–14)	11–12
Setae 1a length	_	—	23 (21–25)	22–23
Tubercles 1a length	_	—	11 (11–12)	9–10
Setae 2a length	25	43	36 (32–41)	32–35
Tubercles 2a apart	_	_	28 (27-31)	27–29
Coxae I &II surface	Smooth	Smooth	Smooth	Smooth
Dorsal semiannuli	22	24	25(24,26)	24 25
number	25	24	23 (24-20)	24-23
Ventral semiannuli	67	19	70 (62, 71)	50 55
number	07	40	/0 (03-/1)	30-33
Dorsal semiannuli shape	Smooth	Smooth	Smooth	Smooth
Ventral semiannuli shape	rounded	Rounded	Rounded	Rounded
Setae <i>c2</i> length	8	7.5	9 (8–10)	8–9
On annulus	13	7	10 (10–14)	10–11
Setae d length	25	30	32 (30–35)	30–31
On annulus	27	16	22 (22–26)	19–20
Setae <i>e</i> length	13	16	14 (14–15)	13–14
On annulus	45	29	38 (38–42)	31–32
Setae f length	13	20	22 (22–23)	20-21
On annulus	5 <sup>th</sup>	5 <sup>th</sup>	5 <sup>th</sup>	5 <sup>th</sup>
Setae <i>h1</i> length	3	3	3	3
Setae <i>h2</i> length	32	43	40 (35–45)	30-32
Female genitalia length	14	14	14 (13–16)	12–13
Female genitalia width	17	19	25 (24–26)	18–19
Ridges no.	10	8	10 (9–10)	_
Setae 3a length	9	9	15 (12–15)	13–15

**Table 4.** Differences between Vittacus plucheae and Neooxycenus plucheae on Pluchea dioscoridis in<br/>Egypt

†After Abou-Awad and Nasr 1986, ‡ after Abou-Awad 1981.



**Figure 14.** Phase microphotograph of *Neooxycenus plucheae* Abou-Awad, 1981: Variant prodorsal shield. Scale bar: 10 μm.

#### REFERENCES

- Abou-Awad BA. 1981. Some eriophyoid mites from Egypt with descriptions of two new species (Acari: Eriophyoidea). *Acarologia*, 22 (4), 367–372.
- Abou-Awad BA, Nasr AK. 1986. An eriophyid mite, Vittacus plucheae sp. n. (Acari: Eriophyoidea: Eriophyidae), on Pluchea dioscoridis L. (Compositae) from Egypt. Acarologia, 27 (2), 159–161.
- Amrine JWJr, Manson DCM. 1996. Preparation, Mounting and Descriptive Study of Eriophyoid Mites. In: Lindquist EE, Sabelis MW, Bruin J (Eds), Eriophyoid Mites. Their Biology, Natural Enemies and Control. World Crop Pests, 6, Amsterdam, The Netherlands: Elsevier Science Publishers, p. 383–396. DOI:10.1016/S1572–4379(96)80023–6
- Amrine JWJr, Stasny TA. 1994. Catalog of the Eriophyoidea (Acari: Prostigmata) of the World. Michigan: Indira Publishing House, 798 pp.
- Amrine JWJr, Stasny TA, Flechtmann CHW.
   2003. Revised Keys to the World Genera of the Eriophyoidea (Acari: Prostigmata).
   Michigan: Indira Publishing House, 244 pp.
- Bakr EM. 2005. A new software for measuring leaf area, and area damaged by *Tetranychus urticae* Koch. *Journal of Applied Entomology*, 129 (3), 173–175. DOI:10.1111/j.1439-0418.2005.00948
- Boulos L. 1999. *Flora of Egypt*. Al–Hadara Publication, Cairo, 419 pp.
- Boulos L. 2002. *Flora of Egypt*. Volume 3: Verbenaceae - Compositae. Al-Hadara Publishing, Cairo, 189 pp.
- Chandrapatya A, Konvipasruang P, Amrine JrJW. 2016. Present status of eriophyoid mites in Thailand. *Journal of Acarological Society of Japan*, 25 (S1), 81–104.
- Chandrapatya A, Konvipasruang P, Amrine JrJW. 2017. Catalog of Thai Eriophyoidea (Acari: Prostigmata) with Illustrations and Keys to Genera of Thai Mites. Printing Office, Extension and Training Office, Kasetsart University, 526 pp.

- Dçbski B. 1919. Nouvelles additions à ma liste des cédicies [sic] d'Egypte. *Bulletin Society Entomology of Egypt*, 3, 65–70.
- de Lillo E, Craemer C, Amrine JWJr, Nuzzaci G. 2010. Recommended procedures and techniques for morphological studies of Eriophyoidea (Acari: Prostigmata). *Experimental & Applied Acarology*, 51, 283–307. DOI:10.1007/s10493-009-9311-x
- Davis R, Flechtmann CHW, Boczek JH, Barké HE. 1982. *Catalogue of Eriophyid mites* (*Acari: Eriophyoidea*). Warsaw Agricultural University Press, Warsaw, Poland: 1–254.
- Denizhan E, Monfreda R, de Lillo E, Çobanogðlu S. 2015. Eriophyoid mite fauna (Acari: Trombidiformes: Eriophyoidea) of Turkey: new species, new distribution reports and an updated catalogue. *Zootaxa*, 3991 (1), 1–63. DOI:10.11646/zootaxa.3991.1.
- Elhalawany AS. 2012. Survey of eriophyid mites on some fruit trees, with re-descriptions of two newly recorded species and a checklist of eriophyid mites in Egypt (Acari: Eriophyoidea). Egyptian Academic Journal of Biological Sciences, A. Entomology, 5 (2), 205–216.
  DOI:10.21608/EABSA.2012.4833
- Elhalawany AS. 2015. Description of one new species and two first records of eriophyid mites (Prostigmata: Eriophyidae) on grasses in Egypt. 5<sup>th</sup> international conference of Plant Protection Research Institute. Hurghada, Egypt. 3–6 May 2015. *Egyptian Journal of Agricultural Research*, 93 (1a), 41–59.
- Elhalawany AS. 2018. A new species, new synonymy and a new record of eriophyoid mites (Acari: Eriophyidae), from Egypt. *ACARINES: Journal of the Egyptian Society of Acarology*, 12, 7–16. DOI:10.21608/ajesa.2008.164281
- Elhalawany AS, Ueckermann EA. 2015. Four new Aceria species (Acari: Trombidiformes: Eriophyidae) on Acacia nilotica from Egypt. International Journal of Acarology, 41, 272–282. DOI:10.1080/01647954.2015.1035320
- Elhalawany AS, Ueckermann EA. 2018. Three new Aceria species (Acari: Trombidi-

formes: Eriophyidae) associated with the invasive weed *Imperata cylindrical* (L.) (Poaceae) from Egypt. *International Journal of Acarology*, 44, 7–20. DOI:10.1080/01647954.2017.1402955

- Elhalawany AS, Ueckermann EA. 2022. Description of three new species and a new record of eriophyid mites (Acari: Eriophyoidea) from Egypt. *Systematic & Applied Acarology*, 27 (6), 1000–1019. DOI:10.11158/saa.27.6.3
- Elhalawany AS, Amer AI, Mesbah AE. 2019a. Redescription and illustration of eight eriophyoid mites (Acari: Prostigmata: Eriophyoidea) with emphasis of their host plants from family Moraceae in Egypt. Egyptian Journal of Plant Protection Research Institute, 2 (1), 22–48.
- Elhalawany AS, Amrine JWJr, Ueckermann E. 2019b. Description of five new species (Acari: Trombidiformes: Eriophyidae: Phyllocoptinae: Anthocoptini) associated with the weed *Imperata cylindrical* (Poaceae) from Egypt. *Systematic & Applied Acarology*, 24 (5), 742–770. DOI:10.11158/saa.24.5.3
- Elhalawany AS, Amrine JWJr, Ueckermann E. 2021. A new species and new record of eriophyoid mites (Trombidiformes: Eriophyoidea) from mango in Egypt with a note on the population dynamics of four eriophyoid species. *ACARINES: Journal of the Egyptian Society of Acarology*, 15, 1– 22. DOI:10.21608/AJESA.2021.240501
- Elhalawany AS, El-Sayed KM, Amer AI. 2018. A new species and record of *Aceria* (Acari: Prostigmata: Eriophyoidea) on weeds from Egypt. *ACARINES: Journal of the Egyptian Society of Acarology*, 12, 17–26. DOI:10.21608/ajesa.2008.164283
- Elhalawany AS, Mohamed AA, Ueckermann EA. 2022. Two new species and complementary descriptions of four new records of family Eriophyidae (Acari: Trombidiformes) in Egypt. *Systematic & Applied Acarology*, 27 (4), 670–696. DOI:10.11158/saa.27.4.5
- Elhalawany AS, Sanad AS, Xue X-F. 2014. Four new records of eriophyids and associated phytoseiids from Egypt. ACARINES: Journal of the Egyptian Society of Acarology, 8, 1–8.

DOI:10.21608/ajesa.2014.163832

- Elhalawany AS, Xue X-F, Amrine, JWJr. 2020. Five new eriophyid mite species from Egypt (Acari: Eriophyidae) associated with the weeds of the family Poaceae. *Systematic & Applied Acarology*, 25 (2), 379–408. DOI:10.11158/saa.25.2.13
- Flechtmann CHW, Tassi AD. 2020. An Aceria species (Prostigmata, Eriophyidae) from Amaranthus in Brazil. Persian Journal of Acarology, 9 (4), 311–319. DOI:10.22073/pja.v9i4.63707
- Guo, J.F., Sadeghi, H., Gol, A. and Xue, X.F. 2015. A new species of the genus *Vittacus* Keifer (Acari: Eriophyidae) from Iran. *Persian Journal of Acarology*, 4(1), 57–63.
- Halawa AM. 2015. New species and new record of the general *Aceria* Keifer and *Calepitrimerus* Keifer (Prostigmata: Acari: Eriophyidae) from Egypt. *Egyptian Academic Journal of Biological Sciences*, 8 (3), 43–48.

DOI:10.21608/EAJBSA.2015.12867

- Keifer HH. 1938. Eriophyid studies I. Bulletin of California Department of Agriculture, 27, 181–206.
- Keifer HH. 1944. Eriophyid studies XIV. Bulletin of the California Department of Agriculture, 33, 18–38
- Keifer HH. 1954. Eriophyid studies XXII. Bulletin of California Department of Agriculture, 43, 121–131.
- Keifer HH. 1959. Eriophyid Series XXVIII. Occasional Papers. *California Department* of Agriculture, 2: 1–20.
- Keifer HH. 1969a. Eriophyid studies C-3. Agricultural Research Service, United State Department of Agriculture, 1–20.
- Keifer HH. 1969b. Eriophyid studies C-3. Agricultural Research Service, United State Department of Agriculture, 1–24.
- Keifer HH, Baker EW, Kono T, Delfinado M, Styer WE. 1982. An Illustrated Guide to Plant Abnormalities Caused by Eriophyid Mites in North America. USDA, ARS, Agricultural Handbook, 573, 1–178 pp.
- Lewandowski M, Abo-Mostafa AM, Druciarek T, Elsayed AK. 2021. Two new species of

Aceria (Acariformes: Eriophyoidea) associated with Amaranthaceae in Egypt. Systematic & Applied Acarology, 26(8), 1399–1414. DOI:10.11158/saa.26.8.1Lindquist EE. 1996. External Anatomy and Notation of Structures In: Lindquist EE, Sabelis MW, Bruin J, (Eds), Eriophyoid Mites. Their Biology, Natural Enemies and Control. Amsterdam: Elsevier, World Crop Pests, 6, p. 3–31.

DOI:10.1016/S1572-4379(96)80003-0

- Nalepa A. 1892. Neue Arten der Gattung Phytoptus Duj. und Cecidophyes Nal. Denkschriften der kaiserlichen.Akademie der Wissenschaften. Mathematisch-Naturwissenschaften, Wien, 59, 525–540.
- Nalepa A. 1898. Zur Kenntniss der Gattung TrimerusNal. Zoologische Jahrbücher, 11 (5), 405–411.
- Navia D, Truol G, Mendonça RS, Sagadin M. 2006. Aceria tosichella (Keifer, 1969) (Acari: Eriophyidae) from Wheat Streak Mosaic Virus-infected wheat plants in Argentina. International Journal of Acarology, 32 (2), 189–193. DOI:10.1080/01647950608684460
- Ripka G, Takacs A. 2021. Description of a new eriophyid species from Hungary. *Folia Entomologica Hungarica Rovartani Közlemények*, 82, 109–120.

DOI:10.17112/FoliaEntHung.2021.82.109

- Sayed MT. 1946. Three new eriophyid mites from Egypt (Acarina, Eriophyidae). Bulletin Society Fouad l<sup>er</sup> Entomology, 30, 149–154.
- Skoracka A, Lewandowski M, Boczek J. 2005. A Catalogue of Eriophyoid Mites (Acari: Eriophyoidea) of Poland. Catalogus faunae Poloniae (N.S.), No. 1. Natura optima dux Foundation, Museum and Institute of Zoology, Polish Academy of Sciences, Warszawa, 199 pp.

Skoracka A, Smith L, Oldfield G, Cristofaro M, Amrine JW Jr. 2010. Host–plant specificity and specialization in eriophyoid mites and their importance for the use of eriophyoid mites as biocontrol agents of weeds. *Experimental & Applied Acarology*, 51 (1– 3), 93–113.

DOI:10.1007/s10493-009-9323-6

- Soliman ZR, Abou-Awad A. 1977. Five new species of the genus *Eriophyes* in the A.R.E. (Acarina: Eriophyoidea: Eriophyidae). *Acarologia*, 19 (4), 668–677.
- Xue X-F, Sadeghi H, Hong X-Y. 2009. Eriophyoid mites (Acari: Eriophyoidea) from Iran, with descriptions of three new species, one new record and a checklist, *International Journal of Acarology*, 35 (6), 461–483.

DOI:10.1080/01647950903427618

- Xue X-F, Sadeghi H, Hong X-Y, Sinaie, S. 2011. Nine eriophyoid mite species from Iran (Acari, Eriophyidae). *ZooKeys*, 143, 23–45. DOI:10.3897/zookeys.143.2162
- Zaher MA. 1984. Survey and Ecological Studies on Phytophagous, Predaceous and Soil Mites in Egypt. 1. Phytophagous Mites in Egypt (Nile Valley and Delta). PI 480 Programme U.S.A., Project No. EG-ARS-30, Grant No. FG-Eg-139, 228 pp.
- Zaher MA, Soliman ZR, Rasmy AH, Abou-Awad BA. 1978. Eriophyoid mites of Ègypt. IV Conf. Control. NRG. Cairo: 815–817.
- Zaki MA. 2000. *Identification and Control of Important Weeds in Egypt*. Al-Ahram Commercial Press - Kalyoub, Egypt, 266 pp.
- Zhang Z-Q. 2018. Repositories for mite and tick specimens: acronyms and their nomenclature. *Systematic & Applied Acarology*, 23, 2432–2446. DOI:10.11158/saa.23.12.12