Original Article

Ameliorative Effects of Vitamin C and Vanillin against Potassium Bromate Toxicity in Male Albino Rats: Biochemical, Hematological and Histological Study

Sohir M. Farag, Nahed A. Omar and Hekmat L. El-Gammal

Department of Zoology, Faculty of Science, Damietta University, New Damietta, Damietta, Egypt

ABSTRACT

Introduction: Potassium bromate (KBrO₃) is a halogen that is widely existing in the environment and used mainly as a food additive. KBrO₃ causes severe damage to many tissues through ROS generation. Exogenous antioxidants are used to protect against and/or treat the KBrO₃ toxicity.

Aim of the Work: To examine the treatment efficiency of vitamin C (Vit C) or/and vanillin against KBrO₃-induced biochemical, hematological, oxidative stress and hepatic and renal histological alterations in male albino rats.

Materials and Methods: Forty five adult male albino rats were categorized into 9 groups equally; Control, Vit C (30mg/kg), Vanillin (100mg/kg), Vit C + Vanillin, KBrO₃ (25mg/kg), KBrO3 withdrawal, KBrO₃ + Vit C, KBrO₃ + Vanillin and KBrO₃ + Vit C + Vanillin. Rats administrated KBrO₃, vitamin C and vanillin orally and daily for 10 weeks. Rats were sacrificed, blood and tissues were gathered and handled for biochemical, hematological and histological studies.

Results: KBrO₃ oral administration increased the serum levels of alanine amino transferase (ALT), aspartate amino transferase (AST), gamma glutamyl transferase (γ -GT), malondialdehyde (MDA), uric acid and creatinine and significantly decreased the levels of total protein (TP), super oxide dismutase (SOD), WBCs, RBCs, platelets and hemoglobin (Hb). KBrO₃ induced hepatic central vein congestion, hepatocytes necrosis, cellular infiltration, atrophy of the glomeruli, renal tubular epithelial cells necrosis and degeneration and infiltration of inflammatory cells. Combined vitamin C or/and vanillin administration attenuated the KBrO3-induced dysfunction by decreasing ALT, AST, γ -GT, MDA uric acid and creatinine levels' and elevating TP, SOD, WBCs, RBCs, platelets and Hb levels' and by improving the liver and kidney architectural lesions.

Conclusion: Vitamin C and Vanillin ameliorates the hematological markers, liver and kidney functional and histological alterations induced by KBrO3 through their antioxidant properties.

Received: 19 February 2023, Accepted: 10 May 2023

Key Words: Antioxidant, curcumin nanoparticles, radiation, reactive oxygen species, tongue.

Corresponding Author: Sohir M. Farag, MSc, Department of Zoology, Faculty of Science, Damietta University, New

Damietta, Damietta, Egypt, Tel.: +20 10 17502081, E-mail: sohirmohammed@du.edu.eg

ISSN: 1110-0559, Vol. 47, No. 2

INTRODUCTION

Potassium bromate (KBrO₃) is a halogen generated from disinfecting water by using chlorine and ozone^[1]. Mostly, bromate is used in bread-baking processes, cosmetic industries, cheese production and beer making^[2,3,4]. The major cause of in *vivo* toxicity of KBrO₃ is oxygen reactive species (ROS) aggregation and oxidative stress^[1,5]. This prompted oxidative stress leads to tissues' damage through modifications in the cellular critical molecules^[6]. KBrO₃ was reported to induce tissues' toxicity and tumors in several animals and human^[7].

Several antioxidants have a protecting or/and treating adequacy against KBrO₃-induced toxicity. Vitamin C (Vit C) is a highly effective water-dissolvable antioxidant. Vit C does not synthesized in human, so its sources are vegetables and fruits. Vit C is desired for collagen formation, maintaining healthy skin, gums and blood

vessels, inorganic iron absorption, plasma cholesterol level reduction, and immune system inducment^[8]. It protects the essential biological macromolecules from oxidative stress damage^[9].

Natural products of medicinal plants possess hepatoprotective, antioxidant, chemotherapeutic, and anti-inflammatory properties[10]. Vanillin is a polyphenol compound isolated from Vanilla planifolia pod and bean. It is vastly utilized as a relish material in food, drinks, pharmaceutical products and perfumes[11]. possess several biological properties; antioxidant, antimutagenic anti-inflammatory, and antitumor properties[12,13,14] and these properties are related to its dynamic ingredients^[15]. Therefore, our study was designated to estimate Vit C or/and vanillin ability in treating KBrO, toxicity through evaluating the biochemical, oxidative and anti-oxidative, hematological markers and histological variations in adult male albino rats.

Personal non-commercial use only. EJH copyright © 2024. All rights served

DOI: 10.21608/ejh.2023.191687.1853

MATERIALS AND METHODS

Study design

Forty five adult male albino rats, their weights range from 130 to 150 gm were housed in the animal house of Faculty of Science, Damietta University, New Damietta under the standard conditions in neat fully aired plastic crates and had a loose access to typical diet and water throughout the experiment. All animals' proceedings were performed based on the National Institute of Health guide for the care and use of laboratory animals, and the protocol of the study was confirmed by our institute (5/3/1/3/2).

Rats were classified into 9 groups; each one had 5 rats and received one or more of KBrO₃, Vit C and vanillin orally and daily for 10 weeks.

Group1 (control): Rats served as a control group.

Group 2 (Vit C): Rats obtained 30mg/kg of Vit C^[16].

Group 3 (Vanillin): Rats obtained 100 mg/kg of vanillin^[17].

Group 4 (Vit C +Vanillin): Rats obtained a mixture of Vit C and vanillin.

Group 5 (KBrO₂): Rats obtained 25 mg/kg of KBrO₂^[18].

Group 6 (KBrO₃ withdrawal): Rats obtained 25 mg/kg of KBrO₃ then KBrO₃ was withdrawn for 2 additional weeks

Group 7 (KBrO₃+Vit C): Rats obtained 25mg/kg of KBrO₃, and then Vit C (30mg/kg) was administrated 2 hr later.

Group 8 (KBrO₃+Vanillin): Rats obtained 25mg/kg of KBrO₃, and then vanillin (100mg/kg) was administrated 2 hr later.

Group 9 (KBrO₃+Vit C+Vanillin): Rats received 25mg/kg of KBrO₃, and then a mixture of Vit C and vanillin was administrated 2 hr later.

Chemicals

The chemicals used in the present study are: KBrO₃ (Sigma Chemicals, Sigma, St Louis, MO), absolute ethanol and Xylene (Supelco Inc- Sigma-Aldrich, USA), vitamin C (Windmill vitamin C-1000mg, USA), Vanillin (powder, ReagentPlus®, Sigma-Aldrich USA), Eosine yellow (Alpha Chemika, India), Hematoxylin (solution, Techno Pharmchem, India) and PBS (liquid, ThermoFisher, USA).

Specimens' preparation

By the experiment end, rats were sacrificed after anesthetization by 50 mg/kg ketamine and 5 mg/kg xylazine intraperitoneally^[19] and blood from heart were directly piled up. For rating hepatic and renal function parameters' values, the blood was let to curd at 37°c for 10 min, centrifuged for 10 min at 3000rpm at room temperature then serum was gathered and preserved at -20°c for further use. Blood aliquots were added to EDTA containing tubes

and used immediately for the hematological assessment. Liver homogenates were prepared by homogenizing liver samples in 5-10 ml cold buffer per gram tissue then centrifuged for 15 min at 18000g (4°C) to estimate oxidative stress markers.

For the histological checking, liver and kidney samples' were taken and fixed in 10% buffered formalin and handled to paraffin blocks. Then 5µm sections were stained with hematoxylin/eosin (H&E)^[20].

Biochemical assessment of hepatic function parameters

Important biochemical parameters including serum ALT, AST, γ -GT and TP were rated using GPT (ALT)-LQ (SPINREACT, Spain), GOT (AST)-LQ kit (SPINREACT, Spain), (γ -GT) kit (BioSystems, Spain) and total protein kit (SPINREACT, Spain), respectively.

Estimation of some oxidative stress markers

The liver supernatant of homogenate was used to measure spectrophotometrically the MDA and SOD levels using MDA and SOD kits (Biodiagnostic, Egypt), respectively.

Biochemical assessment of renal function parameters

Serum was used to measure uric acid and creatinine using Uric acid-LQ kit (SPINREACT, Spain) and CREATININE-J kit (SPINREACT, Spain), respectively.

Assessment of hematological parameters

The blood samples collected into a tube containing EDTA were used to preform complete blood count (CBC) using automated hematology analyzer (KT6400, Genrui Biotech, China).

Statistical analysis

For each measured parameter; the statistical significance of the differences among groups was determined using One-Way Analysis of Variance (ANOVA) followed by Post Hoc tests. When P < 0.05, it is considered significant.

RESULTS

Liver biochemical parameters

No remarkable discrepancy in serum ALT (P=0.857, 1.000, 0.998), AST (P= 0.998, 1.000, 1.000), γ -GT (P=1.000, 1.000, 1.000), and TP (P=0.461, 0.973, 1.000) levels' (Figure 1) between control group and Vit C, vanillin, and Vit C and vanillin-treated groups, respectively. In KBrO₃-treated group, the serum ALT level was increased but not significantly (P=0.339), the AST and γ -GT levels' were notably increased (P=0.000 and 0.004), respectively, and the TP level was crucially declined (P=0.000) comparing to control (Figure 1). When comparing KBrO₃-treated and withdrawn group with the KBrO₃-treated group, the serum ALT and AST levels' were decreased but not significantly (P= 0.539 and 0.098), respectively, γ -GT was appreciably decreased (P= 0.037) and the

TP level was grow up but not significantly (P= 0.990) (Figure 1). Treated-rats with KBrO₃ and Vit C revealed no significant (P= 0.941 and 0.956) reduce in ALT and γ-GT levels', respectively, while the AST level was decreased significantly (P= 0.000) and the TP level was increased crucially (P= 0.000). Treatment with KBrO₃ and vanillin expressed no significant decrease (P= 0.655 and 0.584) in ALT and γ-GT values', respectively, a notable decrease in AST (P= 0.000) and a significant boost (P= 0.000) in the TP level (Figure 1). Combined treatment with KBrO₃, Vit C and vanillin caused significant (P= 0.391 and 0.499) diminution in ALT and γ-GT, respectively, a significant low in the AST (P= 0.000) and serious increase the TP level (P= 0.000) (Figure 1).

Oxidative and antioxidant parameters

No significant modifications (*P*=1.000 for each) in MDA level between control and Vit C, vanillin and Vit C and vanillin treated groups was appeared (Figure 2A). Compared with control, MDA level exposed noteworthy increase (*P*=0.000) in KBrO₃-treated group (Figure 2A). MDA levels' were significantly lessened (*P*=0.001, 0.000, 0.000, 0.000), respectively, in KBrO₃-treated and withdrawn, KBrO₃ and Vit C, KBrO₃ and vanillin and KBrO₃ and Vit C and vanillin-treated groups compared to KBrO₃-treated group (Figure 2A).

The SOD level as antioxidant activity marker, did not significantly (P=0.050, 0.658 and 0.861), respectively, differ between the control and Vit C, vanillin and Vit C and vanillin-treated groups (Figure 2B). In the KBrO₃-treated group, the SOD efficiency was significantly declined (P=0.000). While in KBrO₃-treated and withdrawn, KBrO₃ and Vit C, KBrO₃ and vanillin and KBrO₃ and Vit C and vanillin - treated groups, SOD value was markedly rised (P=0.001, 0.000, 0.000, 0.000), respectively, matched with the KBrO₃-treated group (Figure 2B).

Biochemical parameters

There was no notably change in the levels of uric acid (P=1.000 for each) and creatinine (P=0.919, 1.000, 0.974), respectively, (Figures 2C,D) between the control and Vit C, vanillin, and Vit C and vanillin-treated groups. In KBrO₃-treated group, uric acid was significantly enhanced (P=0.000), while the creatinine level was increased but not significantly (P=0.662) (Figures 2C,D). Uric acid level in KBrO, -treated and withdrawn, KBrO, and Vit C, KBrO, and vanillin, KBrO, and Vit C and vanillin treated groups, was significantly lowered (P=0.010, 0.014, 0.000, 0.000), respectively (Figure 2 C). Compared to KBrO₃-treated group, creatinine was diminished significantly (P=0.002)in KBrO, and Vit C-treated rats, and decreased but not significantly (P=0.183, 0.458, 0.223,), respectively, in KBrO₃ treated and withdrawn, KBrO₃ and vanillin, KBrO₃ and Vit C and vanillin treated groups (Figure 2 D).

Hematological parameters

No significant variances in the WBCs (P=1.000, 0.998, 0.956), RBCs (P= 0.933, 0.999, 1.000), platelet counts

(P=1.000, 1.000, 1.000) and Hb (P=0.812, 0.932, 0.736)(Figure 3) between the control and Vit C, vanillin, and Vit C and vanillin treated groups, respectively, were detected. In KBrO₂-treated group, WBCs, RBCs, platelet counts and Hb concentration were significantly declined (P= 0.008, 0.031, 0.014, 0.000), respectively compared with control (Figure 3). In contrast to KBrO, treated group, the WBCs, RBCs and platelet counts were increased but not significantly (P= 0.980, 0.311, 0.477), respectively, while Hb concentration was markedly increased (P= 0.002) in KBrO, treated and withdrawn group (Figure 3). KBrO, and Vit C treated rats revealed no significant (P=0.586, 0.722and 0.395) increase in WBCs, RBCs and platelet counts, respectively, while the Hb concentration was significantly raised (P= 0.014). Treatment with KBrO₃ and vanillin increased but not significantly (P= 0.442 and 0.056) WBCs and RBCs, respectively, while the platelet count and Hb concentration was significantly mounted up (P= 0.030 and 0.000), respectively, (Figure 3). Treatment with KBrO₂ and Vit C and vanillin increased WBCs, RBCs and platelet count but not significantly (P=0.107, 0.505 and 0.055), respectively, while the Hb level was significantly enhanced (P=0.000) (Figure 3).

Histological results of liver

Checking H&E stained liver sections of control, Vit C, Vanillin and Vit C and vanillin treated rat groups showed similar histological structure which showed normal hepatic architecture. Hepatocytes were coordinated in cords raying out from central veins to the portal areas and isolated by sinusoids. Hepatocytes have vesicular nuclei at the center with eosinophilic cytoplasm. Some cells have binucleated nuclei. Blood sinusoids contain Kupffer cells (Figures 4A-D). Sections of KBrO₂-treated group expressed hepatic architectural abnormalities with congested and expanded central vein, sinusoids were infiltrated with inflammatory cells and sinusoidal spaces were filled with blood. Apparent increases in kupffer cells number were detected. Some necrotic hepatocytes and other with vacuolated cytoplasm were observed (Figures 4E,F). In KBrO₃ treated and withdrawn, KBrO₃ and Vit C, KBrO₃ and vanillin and KBrO3 and Vit C and vanillin treated groups; the hepatic architecture restored its normality with some congested sinusoids and some dilated sinusoids with slight kupffer cells' presence. Hepatocytes are close to normal with normal euchromatic nuclei (Figures 4G-J).

Histological results of kidney

Checking H&E stained kidney sections of control, Vit C, Vanillin and Vit C and vanillin treated rat groups revealed normal renal cortex architecture. Typical glomeruli framed by Bowman's capsule and identical renal tubules were seen (Figures 5A-D). Rats treated with KBrO₃ showed atrophy of the glomeruli, glomerular tuft's congestion and expansion of the Bowman's space. Necrosis, degeneration and loss of the lining epithelium of the epithelial cells of the kidney tubules were observed. Also interstitial hemorrhage and interstitial cellular infiltration were observed (Figures

5E,F). In KBrO₃ treated and withdrawn and KBrO₃ and Vit C treated groups, renal Malpighian corpuscles and tubules restored their normal structure but some degenerative tubules, interstitial hemorrhage and interstitial infiltration

were observed (Figures 5G,H). KBrO₃ and vanillin and KBrO₃ and Vit C and vanillin treated groups showed close to normal renal architecture (Figures 5I,J).

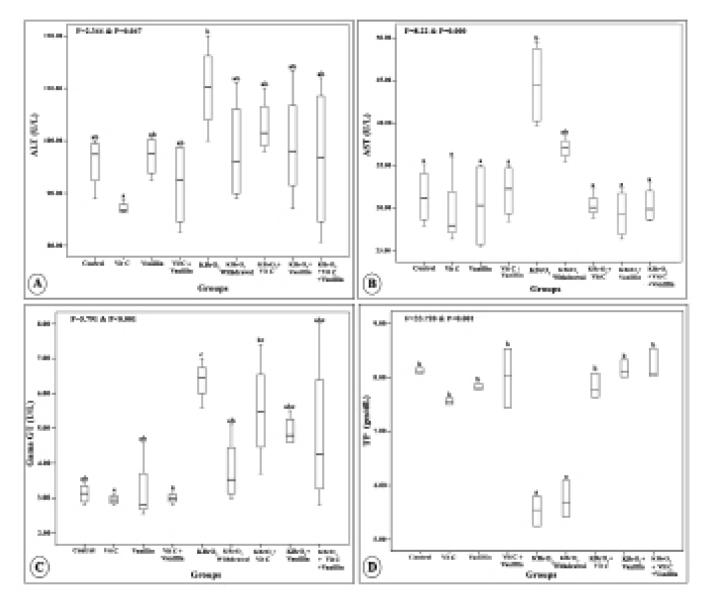


Fig. 1: Serum levels of liver function parameters in different rat groups. A: ALT, B: AST, C: γ -GT and D: TP. Bars represent standard errors. Values with same letter indicate no significant variation. Significant variation is represented by different letters.

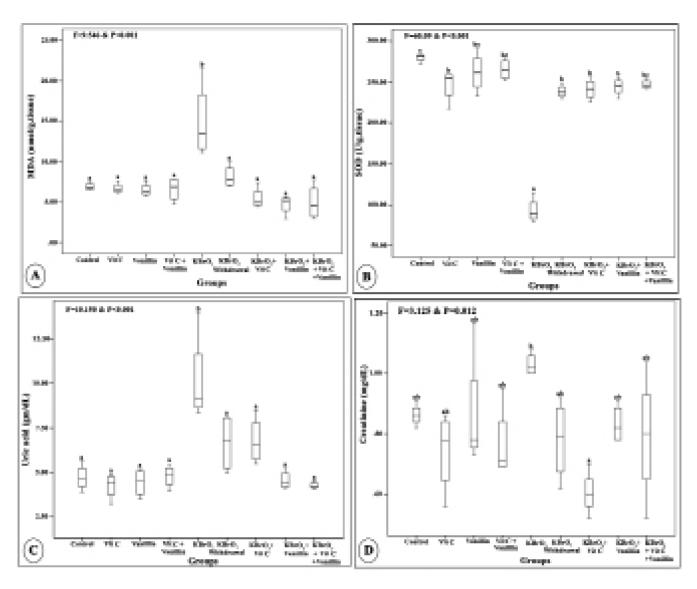


Fig. 2: Levels of some oxidative, antioxidant, and kidney function parameter in different rat groups. A: MDA, B: SOD, C: uric acid and D: creatinine. Bars represent standard errors. Values with same letter indicate no significant variation. Significant variation is represented by different letters.

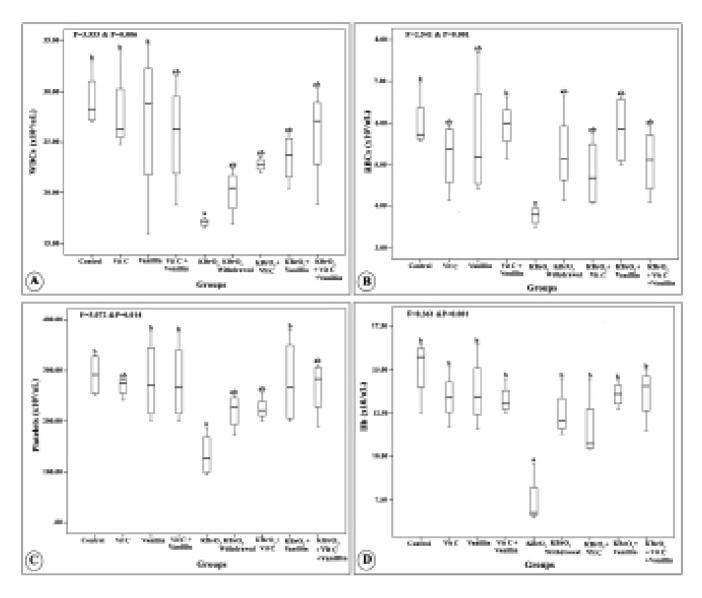


Fig. 3: Levels of hematological parameters in different rat groups. A: WBCs count, B: RBCs count, C: platelets count and D: Hb concentration. Bars represent standard errors. Values with same letter indicate no significant variation. Significant variation is represented by different letters.

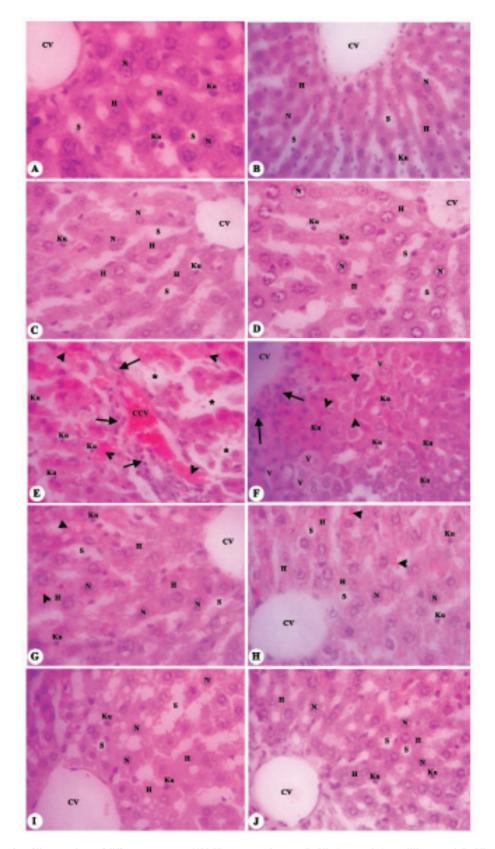


Fig. 4: Photomicrographs of liver sections of different rat groups (400 X). A: control group, B: Vit C-treated, C: vanillin-treated, D: Vit C and vanillin-treated, E&F: KBrO3-treated, G: KBrO3-treated and withdrawn, H: KBrO3 and Vit C-treated, I: KBrO3 and vanillin-treated, J: KBrO3 and Vit C and vanillin-treated. CV: central vein, CCV: congested central vein, H: hepatocyte, N: nucleus, S: sinusoid, Ku: kupffer cells, arrow: infiltration, arrow head: blood congestion, V: vacuolated cytoplasm, *: necrosis

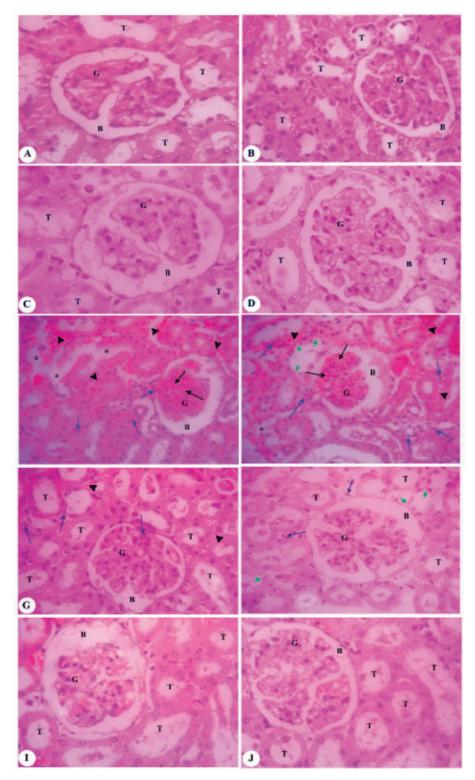


Fig. 5: Photomicrographs of kidney sections of different rat groups (400 X). A: control group, B: Vit C-treated, C: vanillin-treated, D: Vit C and vanillin-treated, E&F: KBrO3-treated, G: KBrO3-treated and withdrawn, H: KBrO3 and Vit C-treated, I: KBrO3 and vanillin-treated, J: KBrO3 and Vit C and vanillin-treated. G: glomeruli, B: Bowman's space, T: tubules, black arrow: glomerular tuft congestion, blue arrow: interstitial infiltration, green arrow: loss of the lining epithelium, arrow head: interstitial hemorrhage, *: necrosis.

DISCUSSION

Metabolism of certain drugs and toxic chemicals releases free radicals. Oxidative stress derived from disparity between the free radicals production and antioxidants disturbing cell functionality leading to tissue injury and various pathological states^[21].

KBrO₃ is widely used in bread making, pharmaceuticals and cosmotic industries^[4,22]. KBrO₃ biotransformation has virulent effects in *vivo* by outputting ROS, which desolate cellular homeostasis and tissues' architectural perfection^[23,24].

Hepatic injury manipulates the hepatocytes' cell membrane constancy triggering seepage of the intracellular enzymes into circulation [25]. Clinically, the injury of the liver is investigated through measuring the biochemical parameters levels [26]. The high serum AST and ALT activities are often used as hepatic injury markers [21]. In recent study, KBrO $_3$ caused significant augmentation in AST and γ -GT levels, while ALT level was increased but not significantly and a crucial lowering in the TP level. The current outcomes are in harmony with previous studies [27-29].

KBrO₃ is a potent oxidant that promotes lipid peroxidation whose waste products damage cellular macromolecules. MDA is indicator for assessing peroxidation of lipids. The increased concentration of MDA and decreased antioxidants' levels trigger tissue spoil and failure of cell's antioxidants to hinder excess free radicals formation^[29,30]. A significant elevation in the concentration of MDA and a notable reduction in SOD level in KBrO₃-treated group were observed in our study. The current results are compatible with the former studies^[29,31-35] which revealed excess MDA and paucity of hepatic antioxidants (SOD, CAT, GPx, GST and GSH) in KBrO₃, treated animals.

Kidney is from the main and primary target organs of toxic substances^[36]. KBrO₃ is highly toxic and hurtful to kidneys, brain and spinal cord^[24]. It leads to renal tissue damage and haemolysis^[37]. The renal functionality is indicated by appreciating creatinine, urea and uric acid levels^[24,38,39]. Remarkable elevation in the serum concentration of uric acid and elevation but not significantly in creatinine in the current KBrO₃ treated group indicating reduction in glomerular filtration rate and impairment of kidney. These results are coincidence with the prior studies results^[24,36,40,41].

KBrO₃ intake induces hematological changes. KBrO₃ caused a noteworthy drop in the count of WBCs, RBCs and platelets and in the Hb concentration in the ongoing study. Our results agree with the previous results of [1,42-46]. The decreased leucocytes and platelets counts could be owed to mutations in in these cells' DNA which resulted from oxidative stress induced by KBrO₃ [47], hematopoiesis disturbance [43] and bone marrow suppression especially depression of megacaryocyte [48]. KBrO₃ toxic effects on bone marrow and hematopoietic organs lead to inability

of them to release normal RBCs in the circulation^[49,50] and also KBrO₃-induced oxidative stress damages RBC's membrane and impairs its deformability^[44]. ROS oxidize ferrous ions to ferric causing methemoglobin formation and this resulted in reduction in Hb concentration^[51]. Also KBrO₃ causes deficiency in iron, folic acid and vitamin B12 levels^[1] which are desired for normal haematopoiesis, erythropoiesis and functional formation of haemoglobin and myoglobin^[52,53].

In the existing study, KBrO, administration caused damage and deformity of the hepatic architecture. Central vein congestion, hepatic degeneration with inflammatory infiltration and congested sinusoids were observed. Cellular necrosis and vacuolated hepatocytes were also recorded. More activated Kupffer cells with obvious raise in their number were manifested. The histological changes detected in the ongoing study are similar to that recorded in [28,29,54] studies. Histological abnormalities in the kidney were detected in the KBrO,-treated rats. Atrophy of the glomeruli with expansion of the Bowman's space, congestion of the glomerular tuft and renal tubules necrosis and loss of the lining epithelium were observed. Interstitial hemorrhage and interstitial inflammatory cells infiltration were recorded. The histological observations recorded in our study are consistent with^[41,55,56]. Accumulation of ROS and toxins damages the mitochondria and lysosomes leading to proteolytic and apoptotic enzymes release into the cytoplasm that causes the cellular destruction and appearance of these histological changes^[56].

Oxidative stress results from excess formation and incomplete elimination of reactive species from cells. To counteract the ROS adverse effects; cells have several defense mechanisms including antioxidative enzymes and small molecules such as Vit C and E, etc. Extra exogenous antioxidants are required to boost the cellular defense system in illness^[57].

Vit C has the potency to effectively rake ROS, inhibits free radicals generation, protects bio-membranes from peroxide damage, reacts with superoxide directly and seeks for homeostasis^[58-60]. In the ongoing study, KBrO₃ and Vit C administrated rats revealed drop in ALT, γ-GT and AST levels' and notably increased TP level. A serious decrease in MDA and a remarkable elevation in SOD were recorded in KBrO, plus Vit C treated rats. These results are consistent with [45,29,61] results. A significant relief in uric acid and creatinine levels' was recorded. Also treatment with Vit C improved the hematopoiesis indicated by the accretion in the WBCs, RBCs and platelets count and significant elevation in the Hb concentration. Vit C treatment improved the massive histological modulations induced by KBrO₃. The liver showed a reasonable level of integrity and normality except for congestion of some blood sinusoids. The damage of the kidney was also disappeared and the renal tissue returned to normal structure except for the interstitial infiltration and loss of lining epithelium of some renal tubules. This is in concord with the prior studies of [29,62-64]. These Vit C improvements are related to its ability as antioxidant to combat free radical mechanism of KBrO₃.

Vanillin shows antioxidant, anticancer, antimicrobial, anti-inflammatory, hepatoprotective and cardioprotective activities [65]. Vanillin administration to KBrO $_3$ -treated rats in our study improved biochemical parameters levels by lowering the ALT, γ -GT and AST levels and significantly increasing the TP level. Also MDA was remarkably decreased and SOD was markedly raised. Our results are in endorsement with the former studies [14,66-68].

Vanillin is a powerful chemopreventive factor against KBrO₃-mediated renal injury^[67]. In the existing study, vanillin administration to KBrO₃-treated rats exposed a notable reduce in uric acid levels, decrease but not significant in creatinine level, enhance in the level of WBCs, RBCs, platelets count and Hb concentration. The current changes in the hematological parameters are in harmonization with^[1] study.

Vanillin markedly improved KBrO₃ associated histopathological modulations in the ongoing study. Both liver and kidney tissues of rats treated with vanillin restored their normal structure and pathological alterations resulted from administration of KBrO₃ were faded. This is in harmony with the previous work of [6,14,44]. The hepatic, renal, hematological and histological improvements associated with vanillin administration illustrates its antioxidant activity, free radicals scavenging ability and its ability to stabilize cellular membranes. The vanillin antioxidant ability can be related to hydroxyl group (OH–) presence linked to aromatic ring that hemolytic fragments the O–H bond [69,70].

Combined administration with Vit C and vanillin to the KBrO₃ treated rats improved the biochemical and hematological parameters in our study. This improvement was illustrated by the drop in ALT, γ-GT and AST levels', noteworthy increase in TP level, significant dwindling in MDA level, remarkable rise in SOD level, significant lessening in uric acid level, decrease but not significant in the creatinine level, the elevation in the count of RBCs, WBCs and platelets and outstanding increase in the Hb concentration. Combination of Vit C and vanillin also ameliorated KBrO₃ toxicity and upgraded the histological variations in the hepatic and renal architecture.

The existing study showed that vanillin's ability and activity in ameliorating the toxicity of KBrO₃ is more than that of Vit C. The study of [53] approved the stronger activity of vanillin than that of Vit C in the 2,20 -azinobis 3-ethylbenzothiazoline-6-sulfonic acid (ABTS_b)-scavenging and considerable activity in the 1-diphenylpicrylhydrazyl (DPPH) radical-assay. This ability of vanillin result of the existing of functional groups such as phenolic, ether and aldehyde moieties [71,72].

CONCLUSION

Our study exposed that KBrO₃ induced hepatic, renal injury and hematological alterations by ROS generation

and oxidative stress initiation. Severe histopathological modulations in the liver and kidney were related to KBrO₃ administration. It was proved that the treatment with Vit C and/or vanillin and with both vit C and vanillin together alleviated biochemical changes and hematological changes induced by KBrO₃, attenuated the hepatic oxidative injury by preventing oxidation of lipids and enhancing the defense mechanisms and markedly ameliorated the pathological variations in the liver and kidney.

CONFLICT OF INTERESTS

There are no conflicts of interest.

REFERENCES

- Saad HB, Amara IB, Krayem N, Boudawara T, Kallel C, Zeghal KM and Hakim A: Ameliorative effects of vanillin on potassium bromate induces bone and blood disorders in *vivo*. Cellular and Molecular Biology. (2015) 61(7): 12-22.
- IARC: Some Naturally Occurring and Synthetic Food Components, Furocoumarins and Ultraviolet Radiation, in: Monographs on the Evaluation of the Carcinogenic Risk of Chemicals to Humans. Lyon, France. (1986).
- 3. Cunningham DK and Anderson JA: Decomposition of bromate in fermenting and nonfermenting doughs. Cereal Chem. (1956) 33: 290-299.
- 4. Ahmad MK, Khan AA, Ali SN and Mahmood R: Chemoprotective effect of taurine on potassium bromate-induced DNA damage, DNA-protein cross linking and oxidative stress in rat intestine. PLoS One. (2015) 10(3): e0119137.
- Zhang X, Bull RJ, Fisher J, Cotruvo JA and Cummings BS: The synergistic effect of sodium chlorite and bromochloroacetic acid on BrO3-induced renal cell death. Toxicology. (2011) 289(2-3): 151-159.
- Saad HB, Driss D, Chaabouni SE, Boudawara T, Zeghal KM, Hakim A and Amara IB: Vanillin mitigates potassium bromate-induced molecular, biochemical and histopathological changes in the kidney of adult mice. Chemico-Biological Interactions. (2016 a) 252: 102-113.
- 7. Khan RA, Khan MR and Sahreen S: Protective effects of rutin against potassium bromate induced nephrotoxicity in rats. BMC Complement Altern Med. (2012) 12: 204 DOI:10.1186/1472-6882-12-204.
- 8. Rekha C, Poornima G, Manasa M, Abhipsa V, Devi JP, Kumar HTV and Kekuda TRP: Ascorbic acid, total phenol content and antioxidant activity of fresh juices of four ripe and unripe citrus fruits. Chem Sci Trans. (2012) 1(2): 303–10.
- 9. Tawfik MS and Al Badr N: Adverse effect of monosodium glutamate on liver and kidney functions in adult rats and potential protective effect of vitamin C and E. Food Nutr Sci. (2012) 3(5): 651-659.

- Rahman MA, Mossa JS, Al-Said MS, Al-Yahya MA: Medicinal plant diversity in the flora of Saudi Arabia
 1: A report on seven plant families. Fitoterapia. (2004)
 75 (2) 149-161.
- 11. Ho K, Yazan LS, Ismail N and Ismail M: Apoptosis and cell cycle arrest of human colorectal cancer cell line HT-29 induced by vanillin. Cancer Epidemiol. (2009) 33(2): 155-160.
- 12. Lee Y, Kwon J, Khang G and Lee D: Reduction of inflammatory responses and enhancement of extracellular matrix formation by vanillin-incorporated poly (lactic-co-glycolic acid) scaffolds. Tissue Eng Part A. (2012) 18(19-20): 19671-978.
- 13. Pedroso LS, Fávero GM, de Camargo LEA, Mainardes RM and Khalil NM: Effect of the o-methylcatechols apocynin, curcumin and vanillin on the cytotoxicity activity of tamoxifen. J Enzyme Inhib Med Chem. (2013) 28(4): 734-740.
- 14. Sefi M, Elwej A, Chaâbane M, Bejaoui S, Marrekchi R, Jamoussi K, Gouiaa N, Boudawara-Sellemi T, El Cafsi M, Zeghal N and Soudani N: Beneficial role of vanillin, a polyphenolic flavoring agent, on manebinduced oxidative stress, DNA damage, and liver histological changes in Swiss albino mice. Human & experimental toxicology. (2019) 38(6): 619-631.
- 15. Gupta S and Sharma B: Pharmacological benefits of agomelatine and vanillin in experimental model of Huntington's disease. Pharmacol Biochem Behav. (2014) 122: 122-135.
- 16. Oliveira CP, da Costa Gayotto LC, Tatai C, Nina BID, Lima ES, Abdalla DS, Lopasso FP, Laurindo FRM and Carrilho F: Vitamin C and vitamin E in prevention of nonalcoholic fatty liver disease (NAFLD) in choline deficient diet fed rats. Nutrition journal. (2003) 2: 1-5.
- 17. Al Asmari A, Al Shahrani H, Al Masri N, Al Faraidi A, Elfaki I and Arshaduddin M: Vanillin abrogates ethanol induced gastric injury in rats via modulation of gastric secretion, oxidative stress and inflammation. Toxicology reports. (2016) 3: 105-113.
- 18. Olajide JE, Akanji MA and Daikwo MA: Modulation of enzyme activities following the coadministration of potassium bromate and chloroquine in selected tissues and serum of albino rats. Animal Research International. (2016) 13(1): 2359-2367.
- Struck MB, Andrutis KA, Ramirez HE, Battles AH.
 2011. Effect of a Short-term Fast on Ketamine– Xylazine Anesthesia in Rats. J Am Assoc Lab Anim Sci 50:344–348.
- 20. Bancroft JD and Layton C: The Hematoxylins and Eosin. In: Suvarna SK, Layton C and Bancroft JD. Bancroft's Theory and Practice of Histological Techniques. 8th ed., Elsevier, Philadelphia. (2019) pp: 126.

- 21. Sabiu S, Wudil AM and Sunmonu TO: Combined administration of Telfaira occidentalis and Vernonia amygdalina leaf powders ameliorates garlic-induced hepatotoxicity in Wistar rats. Pharmacologia. (2014) 5(5): 191-198.
- 22. Oloyede OB and Sunmonu TO: Potassium bromate content of selected bread samples in Ilorin, Central Nigeria and its effect on some enzymes of rat liver and kidney. Food Chem Toxicol. (2009) 47: 2067-2070.
- 23. Tahir M, Rehman MU, Lateef A, Khan R, Khan AQ, Qamar W, Ali F, O'Hamiza O and Sultana S: Diosmin protects against ethanol-induced hepatic injury via alleviation of inflammation and regulation of TNF-α and NF-κB activation. Alcohol. (2013) 47(2): 131-139.
- 24. Altoom NG, Ajarem J, Allam AA, Maodaa SN and Abdel-Maksoud MA: Deleterious effects of potassium bromate administration on renal and hepatic tissues of Swiss mice. Saudi journal of biological sciences. (2018) 25(2): 278-284.
- 25. Solter PF: Clinical pathology approaches to hepatic injury. Toxicol Pathol. (2005) 33(1): 9-16.
- 26. Ramaiah SK: A toxicologist guide to the diagnostic inter-pretation of hepatic biochemical parameters. Food Chem Toxicol. (2007) 45(9): 1551-1557.
- 27. Omer R, Abuelgasim AI and Elmahdi B: Effect of potassium bromate on liver and blood constituents of Wistar albino rats. American Journal of Food Technology. (2008) 3(5): 310-314.
- 28. Dimkpa U, Ukoha U, Anyabolu E, Uchefuna R, Anikeh L, Oji O, Besong E and Emenjo O: Hepatotoxic Effects of Potassium Bromate on Adult Wistar Rats. J Biol Agri Healthcare. (2013) 3: 111-115.
- 29. Bayomy NA, Soliman GM and Abdelaziz EZ: Effect of potassium bromate on the liver of adult male albino rat and a possible protective role of vitamin C: histological, immunohistochemical, and biochemical study. The anatomical record. (2016) 299(9): 1256-1269.
- 30. Almaaty AHA, Hendam BM, Althobaiti F, Fayad E and Abd El-Aziz YM: Evaluation of the hepatoprotective and antioxidant effects of Tegillarca granosa flesh body extract against potassium bromide toxicity via targeting the histomorphometry, chromosomal and expressions of TGF-β1, VEGF and COX-2 genes in rats. Biocell. (2022) 46(1): 219-234.
- 31. Waffa SA and Farida AA: Effect of consumption of kiwi fruit on potassium bromate induced oxidative stress in rats. Australian Journal of Basic and Applied Sciences. (2012) 6: 519-524.
- 32. Kujawska M, Ignatowicz E, Ewertowska M, Adamska T, Markowski J, Liebert JJ: Attenuation of KBrO₃ induced renal and hepatic toxicity by cloudy apple juice in rat. Phytotherapy Research. (2013) 27: 1214–1219.

- 33. Wahba HMA and Ibrahim TAA: Protective effect of flaxseed oil and vitamin E on potassium bromate-induced oxidative stress in male rat. International Journal of Current Microbiology and Applied Sciences. (2013) 2(9): 299-309.
- 34. De Angelo AB, George MH, Kilburn SR, Moore TM, Wolf DC: Carcinogenicity of potassium bromate administered in the drinking water to male B6C3F1Mice and F344/N rats. Toxicologic Pathology. (2016) 26: 587-594.
- 35. Al-Anazi KM, Al-Mareed AA, Farah MA, Ali MA, Hailan WA and Al-Hemaid FM: Protective Effect of Capparis spinosa Extract against Potassium Bromate Induced Oxidative Stress and Genotoxicity in Mice. Evid Based Complement Alternat Med. (2021) 2021: Doi:10.1155/2021/8875238.
- 36. Ahmad MK, Khan AA and Mahmood R: Taurine ameliorates potassium bromate-induced kidney damage in rats. Amino Acids. (2013) 45(5): 1109-1121.
- 37. Robert IA and William BC: Carcinogenicity of potassium bromate in rabbit. Biol Edu. (1996) 34: 114-120.
- 38. Al-Qarawi AA, Abdel-Rahman H, Mousa HM, Ali BH and El-Mougy SA: Nephroprotective action of Phoenix dactylifera. in gentamicininduced nephrotoxicity. Pharmaceutical Biology. (2008) 46(4): 227-230.
- 39. Nabavi SF, Nabavi SM, Habtemariam S, Moghaddam AH, Sureda A, Jafari M and Latifi AM: Hepatoprotective effect of gallic acid isolated from Peltiphyllum peltatum against sodium fluorideinduced oxidative stress. Industrial Crops and Products. (2013) 44: 50-55.
- 40. Ahmad MK, Naqshbandi A, Fareed M and Mahmood R: Oral administration of a nephrotoxic dose of potassium bromate, a food additive, alters renal redox and metabolic status and inhibits brush border membrane enzymes in rats. Food Chem. (2012) 134(2): 980-985.
- 41. Abdel-Latif AS, Abu-Risha SE, Bakr SM, El-Kholy WM and El-Sawi MR: Potassium bromate-induced nephrotoxicity and potential curative role of metformin loaded on gold nanoparticles. Science Progress. (2021) 104(3): Doi:10.1177/00368504211033703.
- 42. Achukwu PU, Ufelle SA, Ukaejiofo EO, Ejezie FE, Nwachukwu DN, Nwagha UI, Nworie WC and Anyaehie USB: The effect of potassium bromate on some haematological parameters of wistar rats. Niger J Physiol Sci. (2009) 24(1): 59-61.
- 43. DHbare AJ and Dale PG: Potassium bromate induced a hematological alteration in European rabbit. The Journal of Zoology Studies. (2017) 4(3): 1-5.

- 44. Mohamed EAK and Saddek EA: The protective effect of taurine and/or vanillin against renal, testicular, and hematological alterations induced by potassium bromate toxicity in rats. The Journal of Basic and Applied Zoology. (2019) 80(1): 1-11.
- 45. Shehab ZA and Ghadhban RF: Effect of Potassium Bromate on Some Hematological and Biochemical Parameters and Protective Role of Vitamin C on Laboratory Rats (Rattus_Rattus). Annals of the Romanian Society for Cell Biology. (2021) 25(2): 669-674.
- 46. Radwan S, El-Wessemy A, Abdel-Aziz B and Abdel-Baky E: Study the protective effect of vitamin E against potassium bromate toxicity on some hematological, renal, and hepatic functions in male rats. Bull. Pharm. Sci., Assiut University. (2022) 45(2): 903-913.
- 47. Parsons JL and Chipman JK: The role of glutathione in DNA damage by potassium bromate in *vitro*. Mutagenesis. (2000) 15(4): 311-316.
- 48. Hoffbrand AV, Petit JE and Moss PAH: Essential Haematology. 4th ed., Blackwell, Oxford. (2004) pp: 252-253.
- 49. Hussein MA and Kata FS. Some hematological and biochemical effects of potassium permanganate (KMnO4) on female mice (Mus musculus L.). Journal Basrah Researchs (Science). (2008) 34(3): 9-13.
- 50. El-Boshy ME, Risha EF, Abdelhamid FM, Mubarak MS and Hadda TB. Protective effects of selenium against cadmium induced hematological disturbances, immunosuppressive, oxidative stress and hepatorenal damage in rats. J Trace Elem Med Biol. (2015) 29: 104-110.
- 51. Unzai S, Eich R, Shibayama N, Olson JS and Morimoto H: Rate constant for O2 and CO bonding to the β and α subunits within the R and T states of human hemoglobin. J Biol Chem. (1998) 273(36): 23150-9.
- 52. Fenech M: The role of folic acid and Vitamin B12 in genomic stability of human cells. Mutat Res. (2001) 475(1-2): 57-67.
- 53. Tai A, Sawano T and Yazama F: Antioxidant properties of ethyl vanillin in *vitro* and in *vivo*. Bioscience, biotechnology, and biochemistry. (2011) 75(12): 2346-2350.
- 54. Oyewo OO, Onyije FM, Awoniran PO: Hepatotoxic effect of potassium bromate on the liver of wistar rats. J Morphol Sci. (2013) 30: 107-114.
- 55. Ali BH, Za'abi MA, Karaca T, Al Suleimani Y, Al Balushi KA, Manoj P, Ashique M and Nemmar A: Potassium bromate-induced kidney damage in rats and the effect of gum acacia thereon. Am J Transl Res. (2018) 10(1): 126-137.

- 56. Elgendy HA and Bayomy NA: Effect of rosmarinic acid on potassium bromate induced renal cortical oxidative stress and apoptosis in adult male albino rat. Eur J Anat. (2020) 24(2): 89-98.
- 57. Poljsak B, Šuput D and Milisav I: Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants. Oxid Med Cell Longev. (2013) 2013: 956792. DOI:10.1155/2013/956792
- 58. Sminorff N and Wheeler GL: Ascorbic acid in plants biosynthesis and function. Crit Rev Biochem Mol Biol. (2000) 19: 267-290.
- 59. Shigeoka S, Ishikawa T, Tamoi M, Miyagawa Y, Takeda T, Yabuta Y, Yoshimura K: Regulation and function of ascorbate peroxidase isoenzymes. J Exp Bot. (2002) 53: 1305-1319.
- 60. Giampieri F, Alvarez-Suarez JM, Mazzoni L, Forbes-Hernandez TY, Gasparrini M, Gonzalez-Paramas AM, Santos-Buelga C, Quiles JL, Bompadre S, Mezzetti B and Battino M: Polyphenol-rich strawberry extract protects human dermal fibroblasts against hydrogen peroxide oxidative damage and improves mitochondrial functionality. Molecules. (2014) 19(6): 7798–816.
- 61. Umemura T, Kitamura Y, Kanki K, Maruyama S, Okazaki K, Imazawa T, Nishimura T, Hasegawa R, Nishikawa A and Hirose M: Dose-related changes of oxidative stress and cell proliferation in kidneys of male and female F344 rats exposed to potassium bromate. Cancer Sci. (2004) 95(5): 393-398.
- 62. Rokaya HAS, Abulyazid I, Thanaa MSB: Protective effect of cape gooseberry fruit against mutagenicity of potassium bromate in mice. J Am Sci. (2012) 8: 22-29.
- 63. Sabiu S, Sunmonu TO, Ajani EO and Ajiboye TO: Combined administration of silymarin and vitamin C stalls acetaminophen-mediated hepatic oxidative insults in Wistar rats. Revista Brasileira de Farmacognosia. (2015) 25: 29-34.
- 64. Kashef S, Abd-El-Hafez A, Sarhan N and Shal A: Effect of Vitamin C on Histological Changes Induced by Potassium Bromate on the Renal Cortex of Adult Male Albino Rat. (2014), Thesis.

- 65. Olatunde A, Mohammed A, Ibrahim MA, Tajuddeen N and Shuaibu MN: Vanillin: A food additive with multiple biological activities. European Journal of Medicinal Chemistry Reports. (2022) 5: 100055. https://doi.org/10.1016/j.ejmcr.2022.100055.
- 66. Makni M, Chtourou Y, Fetoui H, Garoui EM, Boudawara T and Zeghal N: Evaluation of the antioxidant, anti-inflammatory and hepatoprotective properties of vanillin in carbon tetrachloride-treated rats. European journal of pharmacology. (2011) 668(1-2): 133-139.
- 67. 67. Saad H, Driss D, Amara IB, Boudawara O, Boudawara T, Chaabouni SE, Zeghal KM and Hakim A: Altered hepatic m RNA expression of immune response-associated DNA damage in mice liver induced by potassium bromate: Protective role of vanillin. Environmental Toxicology. (2016 b) 31(12): 1796-1807.
- 68. Saad HB, Kammoun I, Boudawara O, Hakim A and Amara IB: Preventive effect of vanillin on lipid peroxides and antioxidants in potassium bromate-induced cardiotoxicity in adult mice: Biochemical and histopathological evidences. Journal of Pharmacognosy and Phytochemistry. (2017) 6(4): 1379-1383.
- 69. Dimitrios B: Sources of natural phenolic antioxidants. Trends Food Sci Technol. (2006) 17(9): 505-512.
- 70. Machado KCM, Oliveira GLS, de Sousa ÉBV, Costa LHF, Machado KC, de Sousa DP, Satyal P and de Freitas RM: Spectroscopic studies on the in *vitro* antioxidant capacity of isopentyl ferulate. Chem Biol Interact. (2015) 225: 47-53.
- 71. Takebayashi J, Asano R, Nakae Y, Saito M, Gohda E, Yamamoto I, Tai A: 2- O-a- D-glucopyranosyl-L-ascorbic acid scavenges 1, 1-diphenyl-2-picrylhydrazyl radicals via a covalent adduct formation. Biosci Biotechnol Biochem. (2007) 71(3): 754-760.
- 72. Saito S and Kawabata J: A novel oxidative dimer from protocatechuic esters: contribution to the total radical scavenging ability of protocatechuic esters. Biosci Biotechnol Biochem. (2008) 72(7): 1877-1880.

الملخص العربي

التأثيرات التحسينية لفيتامين سي و الفانيلين ضد سمية برومات البوتاسيوم في ذكور التأثيرات الجرذان البيضاء: دراسة فسيولوجية و نسيجية

سهير محمد فرج، ناهد أحمد عمر، حكمت لطفى الجمال

قسم علم الحيوان، كلية العلوم، جامعة دمياط، دمياط الجديدة، دمياط، مصر

المقدمة: برومات البوتاسيوم هي هالوجين موجود علي نطاق واسع في البيئة و يستخدم بصورة أساسية كمضاف غذائي. تسبب برومات البوتاسيوم ضررا بالغا للعديد من الأنسجة من خلال تكوين أنواع الأكسجين النشطة (ROS). تستخدم مضادات الأكسدة الخارجية للحماية من أو لعلاج سمية برومات البوتاسيوم.

الهدف من العمل: التحقق من الكفاءة العلاجية لفيتامين سي, أو/و الفانيلين ضد التغيرات الكيميائية الحيوية ،الدموية، التأكسدية والتركيبية الكبدية و الكلوية في ذكور الجرذان البيضاء.

المواد و الطرق: تم تقسيم 63 من ذكور الجرذان البيضاء إلى 9 مجموعات متساوية: المجموعة الضابطة ، فيتامين سي (73) مجم/كجم), فانيلين (70) مجم/كجم), فيتامين سي (70) مجم/كجم), انسحاب برومات البوتاسيوم (70) مجم/كجم), انسحاب برومات البوتاسيوم (70) البوتاسيوم (70) مجم/كجم) البوتاسيوم (70) مجمركجم البوتاسيوم (70) مجمركجم البوتاسيوم البوتاسيوم

alanine amino transferase (ALT), gamma glutamyl transferase (γ -GT), malondialdehyde aspartate amino transferase (AST), gamma glutamyl transferase (γ -GT), malondialdehyde TP)), حمض اليوريك uric acid و الكرياتينين uric acid و قلل بشكل ملحوظ مستويات البروتين الكلي ((MDA)), حمض اليوريك Super oxide dismutase (SOD), عدد خلايا الدم البيضاء (WBCs), عدد كرات الدم الحمراء (RBCs), و عدد الصفائح الدموية (platelets) و الهيموجلوبين (Hb). سببت برومات البوتاسيوم احتقان الوريد المركزي الكبدي، غدر الخلايا الكبدية، الارتشاح الخلوي، ضمور الكبيبات، تأكل و نخر الخلايا الطلائية الأنبوبية الكلوية و تسرب الخلايا الالتهابية. أدى تناول فيتامين سي أو الفانيلين أو كليهما معًا إلى تخفيف الخلل الوظيفي الناجم عن برومات البوتاسيوم عن طريق خفض مستويات AST و AST و γ -GT و حمض اليوريك acid, MDA و الكرياتينين و رفع مستويات البروتين الكلي و SOD و عدد خلايا الدم البيضاء WBCs, عدد كرات الدم الحمراء RBCs, الصفائح الدموية و الهيموجلوبين و عن طريق تحسين التغيرات التركيبية في الكبد و الكلية.

الإستنتاج: يعمل فيتامين سي و الفانيلين على تحسين الدلائل الدموية، و التغيرات الوظيفية و النسيجية للكبد و الكلى المستحثة ببرومات البوتاسيوم من خلال خصائصهما المضادة للأكسدة.