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Enhancing the Nutritional Value of Corn Flakes by Adding Quinoa Flour

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Abstract

The effect of replacing yellow corn flour (YCF) by quinoa flour (QF) at different levels (20, 40, 60, 80, and 100%) on the nutritional values, phenolics, flavonoids compounds, antioxidant activity, color quality, and sensory evaluation of produced corn-quinoa flakes was investigated. Results revealed that substituting yellow corn flour with different levels of QF caused a significant increase (P < 0.05) in crude protein, ether extract, crude fiber and ash content, while carbohydrates content decreased. Also, the contents of minerals (Ca, P, Mg, K, Fe, and Zn) and essential amino acids (Histidine, Lysine, Isoleucine, Threonine, Methionine+Cystein and Phenylalanine+Tyrosine) were contained increased gradually by increasing replacement levels of OF from 20% to 100% in all tested samples compared with control. In this concern, the results indicated that the incorporation of QF in YCF led to a significant increase (P < 0.05) in the total content of phenolics, flavonoids compounds, and antioxidant activity of prepared corn-quinoa flakes by increasing of OF ratio in the flakes formulation. Color values of produced corn-quinoa flakes indicated that L^* and b^* values of produced flakes decreased gradually with increasing the substitution amount from 20 to 100% QF compared with the control sample. While a^* values increased significantly with increasing the replacement levels of QF compared with the control. Sensory evaluation tests showed that up to 40% of YCF could be replaced with QF in produced flakes and still more acceptable for consumers than the control sample prepared from 100% YCF. Thus, it can be suggested that combining OF in YCF led to strengthening and improving the national values of produced flakes.

Keywords: Chemical analysis, Corn flakes, Nutritional value, Quinoa flour, Sensory evaluation.

1.Introduction

Cereal grains are a wide foodstuff consumed in most countries and provide most of the caloric energy, proteins, minerals, and vitamins for the world population (1). Ready-to-eat (RTE) breakfast cereals have become a staple established on breakfast tables almost all over the world and are defined as "a food made from processed grains such as corn, wheat, oat, and other grains usually with added flavor and fortifying ingredients for human consumption". The global edible flakes market was valued at USD 17.43 billion in 2019 and is expected to grow significantly (2). Breakfast cereals are important sources

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of minerals, vitamins, antioxidants, and phytochemicals (3 and 4).

Corn flakes are the most popular breakfast cereals in the daily diet for children and adolescents. Mize or corn (*Zea mays* L.) is the major raw material used to manufacture corn flakes. It has a good source of minerals, vitamins, and some phytochemical components (5). On the other side, corn proteins have poor nutritional value for humans because they have limited amounts of some essential amino acids such as lysine and tryptophan which average (about 2%) are less than one-half of the recommended concentration for human nutrition (6 and 7).

(Chenopodium Quinoa quinoa, Willd.) is a pseudo cereal belonging to (Chenopodiaceae). Quinoa seeds have been consumed to make breakfast cereal or ground to flour to produce toasted and baked goods such as; cookies, biscuits, bread, noodles, and flakes (8). Quinoa seeds can be considered a very healthy, nutritious cereal compared to commonly consumed cereals such as wheat, barley, and corn (9). It has a rich source of protein, essential full amino acids, all of vitamins/minerals, essential fatty acids, dietary fiber, other beneficial compounds, and a low glycemic index. It can be used in breakfast flakes, provides energy, and aids in weight loss (10, 11, and 12). The amino acid value is higher than wheat and corn, and a more incredible amount of lysine makes quinoa superior to other cereals (13).

Ouinoa-based foods play а beneficial role in reducing childhood malnutrition (14); reducing risk of developing cardiovascular disease (15); diabetes (16); anti-obesity (17); people with anemia (18); individuals with lactose intolerance (19); women prone to osteoporosis (20) and celiac disease (21) due to its properties including a high nutritional value, therapeutic features, and gluten-free content.

The present research aimed to assess the proximate chemical composition, amino acids, minerals content and some phytochemicals of corn flakes supplemented with quinoa flour as a good source of essential amino acids and other nutrients as assessing the color quality and sensory properties of products.

2. Materials and Methods

2.1. Materials

Yellow corn flour was purchased from the Egyptian-Italian Company for maize products (Maiza), 10th of Ramadan City, Sharquia, Egypt. Quinoa seeds were obtained from Field Crops Research Institute, Agriculture Research Center, Giza, Egypt. All chemicals used in this study for analysis were of analytical grade and purchased from Delta Aromatic International Company, Giza, Egypt.

2.2. Methods

2.2.1. Preparation of quinoa flour (QF)

Quinoa seeds were cleaned from dust, broken seeds, and other foreign matters. The seeds were washed many times with tap water in a tank equipped with a mixer until there was no more foam in the washing water to remove saponins. Afterward, the seeds were rinsed and dried in an electric oven at $50\pm2^{\circ}$ C for 12 hr. The quinoa seeds were milled to a fine powder in an electric grinder of stainless steel using a laboratory disc mill and sifted through a 60 mesh to obtain quinoa flour (22).

2.2. 2. Preparation of Corn-Quinoa flour blends:

Six blends were prepared by mixing yellow corn flour (YCF) with quinoa flour

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(QF) using an electric blender in the percentage ratio as presented in Table (1).

Table (1): Formulation of corn/quinoa nour biends				
Treatments	Flour blends			
T1 (Control)	100% YCF			
T2	80% YCF+ 20% QF			
T3	60% YCF+ 40% QF			
T4	40% YCF+ 60% QF			
T5	20% YCF+ 80% QF			
T6	100% QF			

 Table (1): Formulation of corn/quinoa flour blends:

2.2.3. Preparation of corn-quinoa flakes

Flakes samples (control and cornquinoa flakes) were prepared according to the method of Leusner (23). Flour blends were mixed with water until the dough formed (moisture content 22 to 35%); flakes dough that had been formed was then flattened using a roller (thickness 0.38 – 0.63 mm). After that, it was cut into flakes (3.5-6.5 g) and dried using an electric oven at $210\pm2^{\circ}$ C for 2-3 min. All obtained flake samples are then stored in plastic bags at room temperature for further analysis.

2. 2.4. Chemical analysis

Proximate chemical composition

Moisture, crude protein, ether extract, crude fiber, and ash were determined in cornquinoa flakes samples according to AOAC (24). Carbohydrates were calculated by difference as follows: Carbohydrates (%) = 100 - (% protein + % fat + % crude fiber + % ash).

Determination of amino acids profile

Amino acids of corn-quinoa flake samples were determined according to the method described in AOAC (**25**) using the automatic amino acid analyzer Model (Beckman 7300/G 300). Amino acid score (AAS) was calculated according to FAO/WHO (**26**). As follows: AAS% = $\frac{\text{mg of amino acid in 1g tested protein}}{\text{mg of amino acid in reference protein}} x 100$

Determination of minerals content

Calcium (Ca), potassium (K), and magnesium (Mg) were determined according to the method of Jackson (27). Phosphorus (P) was determined according to the procedure described by Tan (28). Iron (Fe) and zinc (Zn) were determined using Atomic Absorption GBC 909 AA, as described in AOAC (25).

Determination of total phenolic content

The total amount of phenolic compounds in corn-quinoa flakes samples was determined in the extract flour samples by the Folin-Ciocalteu method. Gallic acid was used as a standard, expressed in mg of gallic acid equivalents (GAE) per 100 gram dry weight according to (**29**).

Determination of total flavonoids content

The total amount of flavonoid compounds in corn-quinoa flakes samples was analyzed according to the method described (**30**). The total flavonoids content was expressed as mg rutin equivalents/100 g (dry weight basis).

Determination of antioxidant activity

The antioxidant activity of cornquinoa flake samples was determined according to 2-2-diphenyl-1-picrylhydrazyl (DPPH). The DPPH method is based on the destruction of DPPH by the antioxidant

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substances present in the sample according to the following equation by Beta et al. (31):

% Inhibition= [(Abs control – Abs samples) / Abs control] x 100.

2. 2.5. Determination of color quality of corn-quinoa flakes

The color of corn-quinoa flake samples was determined by measuring L (whiteness/ darkness), a (redness/greenness), and b (yellowness/blueness) parameters using Hunter lab colorimeter according to the method of Akesowan (**32**).

2. 2.6. Sensory evaluation

Sensory evaluation of corn-quinoa flakes samples was determined using twenty members from Flavor and Aroma Chemistry Department, Food Industries and Nutrition Division, National Research Centre. Each panellist was asked to assign scores 0–10 for appearance, color, taste, odour, texture, and overall acceptability. A sensory score of 5 or above was deemed acceptable, and a score below five was considered unacceptable (33).

2. 2.7. Statistical analysis

The results obtained were statistically analyzed using one-way analysis of variance (ANOVA) followed by Duncan's test according to the procedure of (**34**) using SPSS version 20.0 software computer program.

4. Results and Discussion

4. 1. Chemical composition of Corn-Quinoa flakes

The chemical composition of prepared corn-quinoa flakes listed in Table (2). As shown in the obtained results (Table 2), it could be noticed that substitution of corn-quinoa flakes produced from YCF substituted by different levels (20, 40, 60, 80, and 100%) of QF caused a significant increase (P < 0.05) in crude protein (10.55, 12.07. 13.42, 14.85 and 16.35%, respectively), ether extract (2.87, 3.71, 4.34, 5.02 and 5.90%, respectively), crude fiber 1.75, 1.95, 2.15 and 2.36%, (1.48.respectively) and ash (1.54, 2.03, 2.43, 2.85 and 3.27%, respectively) when compared with control sample prepared from 100% (9.22, 2.29, 1.26 YCF and 1.14%, respectively). On the other hand, the carbohydrates content decreased significantly $(P \le 0.05)$ as the substitution levels increased from 20% to 100% in produced flakes, which reached (72.12%) in flakes containing 100% QF compared with (86.09%) control samples containing 100% YCF.

These results are following those obtained by Rosell et al. (22), Khalil et al. (35), and El-Sohaimy et al. (36). They reported gradual increases in protein, fat, fiber, and ash content, parallel to the rise in the substitution level of quinoa flour in bakery products such as pan bread and cookies. In addition, Quinoa seeds are a complete food with high nutritional value (37) due mainly to their high content of good-quality protein (9).

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Samples	Moisture	Crude Protein	Ether Extract	Crude Fiber	Ash	Total carbohydrates'
Control	5.96	9.22	2.29	1.26	1.14	86.09
(100%YCF)	$\pm 0.04^{f}$	±0.33 ^f	±0.13 ^f	$\pm 0.06^{f}$	$\pm 0.02^{f}$	$\pm 0.15^{\mathrm{a}}$
80% YCF	6.08	10.55	2.87	1.48	1.54	83.56
+20% QF	±0.02 ^e	±0.24 ^e	±0.08 ^e	±0.04 ^e	±0.06 ^e	±0.24 ^b
60% YCF	6.13	12.07	3.71	1.75	2.03	80.44
+40% QF	±0.06 ^d	±0.16 ^d	±0.16 ^d	$\pm 0.04^{d}$	±0.06 ^d	±0.10 ^c
40% YCF	6.24	13.42	4.34	1.95	2.43	77.86
+60% QF	±0.02 ^c	±0.24 ^c	±0.05 ^c	±0.03 ^c	±0.04 ^c	±0.28 ^d
20% YCF	6.39	14.85	5.02	2.15	2.85	75.13
+80% QF	±0.05 ^b	±0.12 ^b	±0.09 ^b	±0.02 ^b	±0.02 ^b	±0.50 ^e
100% QF	6.57	16.35	5.90	2.36	3.27	72.12
	$\pm 0.08^{a}$	$\pm 0.09^{a}$	±0.06 ^a	±0.04 ^a	±0.03 ^a	$\pm 0.33^{f}$

(2): Chemical composition (g/100g) of flakes produced from yellow corn and quinoa flour(on a dry weight basis)

Means \pm standard deviations; the means within the same column having different superscripts are significantly varied (P < 0.05) .; YCF: Yellow Corn Flour; QF: Quinoa Flour; • Total Carbohydrates were calculated by difference.

4. 2. Essential amino acids content of Corn-Quinoa flakes

The nutritional quality protein of Corn-Quinoa flakes samples was determined according to the reference protein pattern of FAO/WHO (26) and listed in Table (3). From the obtained results in Table (3), it could be shown that the essential amino acids of produced corn-quinoa flakes were significantly increased (P < 0.05) in essential amino acids (Histidine, Lysine, Isoleucine, Threonine, and Methionine+Cystein by increasing levels of added QF from (20% to 100%) in corn-quinoa flakes samples, it was presented (2.01 to 3.28, 3.13 to 5.83, 4.22 to 5.15, 3.49 to 4.86 and 3.34 to 6.16 g/100g protein, respectively) when compared with the control sample (1.69, 2.41, 3.99, 3.21 and 2.62 g/100g protein, respectively).

Also, As given in Table (3), it could be observed that the (%) amino acids score of Corn-Quinoa flakes samples recorded a (P<0.05) significant increase by replacement levels of QF (20, 40, 60, 80, and 100%), it was presented (99.62, 105.47, 111.18, 116.89 and 122.88 %, respectively) as compared with corn-quinoa flakes control sample (93.82%). This increase in essential amino acids of Corn-Quinoa flakes samples may be due to the highest content of QF from essential Amino acids: Histidine, Lysine. Valine. Leucine Isoleucine, Threonine, Methionine + Cysteine, and Phenylalanine + Tyrosine (3.16, 5.77, 4.90, 7.05, 5.03, 4.14, 5.13 and 7.31 g/100g of QF) as shown in Table (3). This result was accordance with Repo-Carrasco et al. (9), El-Sohaimy et al. (36), and Demir and Bilgicll(38).

Table (3): Essential amino acids content (g/100g protein) of QF and flakes produced from corn and quinoa flour as compared by the provisional amino acid pattern of FAO/WHO (1973).

	QF							
Amino acids g/100g protein	g/100g protein	Control ▲	20%	40%	60%	80%	100%	FAO/WHO g/100g protein
Histidine	3.16±	1.69*±	2.01±	2.35±	2.67±	2.95±	3.28±	1.90
	0.08	0.05 ^f	0.08 ^e	0.09 ^d	0.07 ^c	0.10 ^b	0.08 ^a	
Lysine	5.77±	2.41*±	3.13*±	3.79 *±	4.43 *±	5.15*±	5.83±	5.5
	0.10	0.08 ^f	0.06 ^e	0.08 ^d	0.10 ^c	0.06 ^b	0.12 ^a	
Valine	4.70 ±	4.70±	4.72±	4.73 ±	4.75 ±	4.77 ±	$4.80\pm$	3.88
	0.09	0.10 ^a	0.12 ^a	0.10 ^a	0.12 ^a	0.09 ^a	0.06 ^a	
Leucine	7.75±	8.55±	8.43±	8.32±	8.19±	8.06±	7.96±	7.00
	0.07	0.12 ^a	0.06 ^a	0.08 ^{ab}	0.10 ^{ab}	0.08 ^b	0.10 ^b	
Isoleucine	5.03±	3.99*±	4.22±	4.46±	4.70±	4.91±	5.15±	4.00
	0.08	0.04 ^f	0.06 ^e	0.12 ^d	0.08 ^c	0.10 ^b	0.12 ^a	
Threonine	4.54±	3.21*±	3.49*±	3.80*±	4.11±	4.47 ±	4.86±	4.00
	0.09	0.06 ^f	0.12 e	0.08 ^d	0.10 °	0.09 ^b	0.08 ^a	
Methionine	5.93±	2.62*±	3.34*±	$4.07\pm$	4.81±	5.50±	6.16±	0.70
+Cysteine	0.08	0.05 ^f	0.08 ^e	0.12 ^d	0.08 ^c	0.09 ^b	0.10 ^a	3.50
Phenylalanine	6.31±	7.15±	7.10±	7.06±	7.01±	6.95±	6.91±	6.80
+ Tyrosine	0.09	0.10 ^a	0.12 ^a	0.10 ^a	0.08 ^a	0.10 ^a	0.12 ^a	
Total Essential	<i>43.19</i> ±	34.32*±	36.44*	38.58±	40.67±	42.76±	44.95±	36.58
Amino acids	0.12	0.15 ^f	±	0.12 ^d	0.10 °	0.16 ^b	0.18 ^a	
			0.17 ^e					
Amino acid Score (%)	118.07	93.82	99.62	105.47	111.18	116.89	122.88	100%

Means \pm standard deviations; the means within the same row having different superscripts are significantly varied (P < 0.05).;

Control sample prepared from 100% yellow corn flour; QF: Quinoa Flour. *: Limiting amino acids. - Tryptophan was not determined

4.3. Minerals content of Corn-Quinoa flakes

The mineral content of corn-quinoa flakes samples was determined, and results were presented in Table (4). Obtained data in Table (4) indicated that the contents of Ca, P, Mg, K, Fe, and Zn were increased significantly (P < 0.05) with increasing the substitution levels of quinoa flour (QF) from 20% to 100% in all the tested samples when compared with control. In this concern, flakes containing 100% QF exhibited higher content of Ca (118.74 mg/100g), P (405.42 mg/100g), Mg (199.08 mg/100g), K (795.58 mg/100g), Fe (8.44 mg/100g) and Zn (4.62 mg/100g) than the control flakes made from 100% YCF. This increase in the mineral content of produced flakes may be attributed to the high concentration of these minerals in quinoa seeds flour (39 and 40).

These results agree with Khalil et al. (35) found that incorporation of quinoa flour with corn flour at levels 25, 50, 75 and 100% caused an increase in K, Ca, P, Mg, Fe, Mg, Na and Na and Zn content of produced free gluten pan bread. Also, (41) indicated that the mineral content of corn flakes was increased with increasing additional levels of peanut flour up to 30%.

Samples	Minerals content (mg/100g)							
	Ca	Р	Mg	K	Fe	Zn		
Control (100% YCF)	48.74±0.69 ^f	231.11±2.71 ^f	125.55 ± 1.17^{f}	320.94±2.75 ^f	3.09±0.02 ^f	1.45 ± 0.02^{f}		
80% YCF +20% QF	63.00±0.84 ^e	266.46±1.53 ^e	139.61±0.66 ^e	411.01±3.92 ^e	4.15±0.03 °	2.02±0.07°		
60% YCF +40% QF	77.19 ± 0.82^{d}	300.55±2.68 ^d	155.60±2.28 ^d	520.41±1.15 ^d	5.27±0.01 ^d	2.79±0.03 ^d		
40% YCF +60% QF	92.19±1.88°	335.80±3.71°	168.67±1.90°	609.76±2.10°	6.31±0.02 °	3.41±0.03°		
20% YCF +80% QF	103.71±2.20 ^b	372.34±2.91 ^b	183.55±2.44 ^b	701.91±3.40 ^b	7.37±0.04 ^b	4.05±0.04 ^b		
100% QF	118.74±1.97 ^a	405.42±2.95 ^a	199.08±2.61 ^a	795.58±3.28 ^a	8.44±0.01 ^a	4.62±0.02 ^a		

Table (4): Minerals content (mg/100g sample) of corn-quinoa flakes samples produced from yellow corn and quinoa flours (on a dry weight basis).

Means \pm standard deviations; the means within the same column having different superscripts are significantly varied (P < 0.05); YCF: Yellow Corn Flour; QF: Quinoa Flour.

4.4. Phenolics, flavonoids content, and the antioxidant activity of developed cornquinoa flakes

The means value of total phenolics (mg GAE/100 g), total flavonoids (mg rutin /100g), and antioxidant activity by DPPH (on a dry weight basis) of corn flakes produced from different levels of yellow corn flour (YCF) and quinoa flour (QF) were determined and are represented in Table (5). From the obtained data, as illustrated in Table (5), it could be seen that the total content of phenolic and flavonoid compounds and antioxidant activity of prepared flakes were increased significantly (p < 0.05) by increasing of QF ratio in the flakes formulation. Total phenolics and total flavonoids content were increased from (128.15 and 7.29 mg/100g) in flakes enriched with 20% QF to (162.10 and 10.82 mg/kg) in flakes containing 100% QF as compared with control samples prepared from 100% YCF, which recorded 119.45 and 6.35 mg/100g); respectively. In this concern, the antioxidant activity of produced flakes increased significantly (p<0.05) from 20.89% in flakes samples containing 20% QF to reach 36.52 % in flakes made from 100% QF compared with control samples which recorded 16.95%. This increase in the content of total phenolics, total flavonoids, and antioxidant activity values of the corn flakes enriched with quinoa flour is most

likely attributable to the fact that quinoa seeds flour is very rich in phenolics compounds and antioxidant capacity (42 and 43). Several studies have noted significant increases in antioxidant activity and total phenolic content following the addition of quinoa flour in bakery products (35, 47, and 38).

Table (5): Total phenolics, total flavonoids content, and antioxidant activity of cornquinoa flakes samples produced from corn and quinoa flour

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Samples	Total Phenolics (mg GAE/100g)	Total Flavonoids (mg rutin/100g)	Antioxidant Activity (%)					
Control	119.45 ±	6.35 ±	16.95 ±					
(100% YCF)	1.12 ^f	0.25 ^f	0.23 ^f					
80% YCF	128.15 ±	7.29 ±	$20.89 \pm$					
+20% QF	0.98 ^e	0.20 ^e	0.92 ^e					
60% YCF	137.10 ±	8.16 ±	$24.70 \pm$					
+40% QF	0.87 ^d	0.57 ^d	0.59 ^d					
40% YCF	145.92 ±	9.06 ±	28.65 ±					
+60% QF	1.02 ^c	0.35°	0.65°					
20% YCF	153.89 ±	9.95 ±	32.62 ±					
+80% QF	0.55 ^b	0.08 ^b	0.20 ^b					
100% QF	$162.10 \pm$	$10.82 \pm$	$36.52 \pm$					
	0.68 ^a	0.55ª	0.45 ^a					

 $\begin{array}{ll} \mbox{Means}\pm\mbox{standard}\ \mbox{deviations};\ \mbox{the means within the same column}\\ \mbox{having different superscripts}\ \mbox{are significantly varied}\ (P{<}0.05)\ .;\\ \mbox{YCF: Yellow corn Flour;} & \mbox{QF: Quinoa Flour.} \end{array}$

4. 5. Color quality of Corn-Quinoa flakes

Color values of control flakes prepared from 100% YCF and flakes samples containing quinoa flour (QF) at substitution levels of 20, 40, 60, 80, and 100% were determined, and results are summarized in Table (6). *L**(light/dark) and b^* (blue/yellow) values of produced flakes decreased gradually with the substitution amount from 20 to 100% quinoa flour compared with control corn flakes. While, a^* (green/red) values increased significantly with increasing the replacement levels of quinoa flour compared with control. These changes in the L*, a* and b* values of produced corn-quinoa flakes may be attributed to Millard reactions during baking (44). Similar results have been reported by Malik et al. (41), Priyanka et al. (45), and Dewidar et al. (46).

 Table (6): Color quality of RTE-flakes produced

 from yellow corn and quinoa flour

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Samples	Parameter						
Samples	L^*	a^*	b^*				
Control	67.46±	16.59±	24.70±				
(100%							
YCF)	1.23 ^a	0.53 ^e	0.86 ^a				
80% YCF	$67.02 \pm$	16.84±	24.96±				
+20% QF	1.10 ^a	0.67 ^e	0.65 ^a				
60% YCF	65.18±	18.35±	23.16±				
+40% QF	0.26 ^b	1.14 ^d	0.27 ^b				
40% YCF	64.06±	21.46±	21.55±				
+60% QF	0.12 ^b	1.08 ^c	1.10 ^c				
20% YCF	62.92±	23.20±	19.87±				
+80% QF	0.08b ^c	0.83 ^b	0.29 ^d				
100% QF	58.77 ±	24.89±	18.09±				
	0.51 ^d	0.50 ^a	0.50 ^e				

Means \pm standard deviations; the means within the same column having different superscripts are significantly varied (*P*<0.05) YCF: Yellow corn Flour; QF: Quinoa Flour. L^{*} degree of lightness; a^{*} degree of redness; b^{*} degree of yellowness; ΔE is a single value that considers the differences between the *L*, *a*, and *b* of the sample and standard.

4. 6. Organoleptic characteristics of Corn-Quinoa flakes

The sensory characteristics of the flakes prepared from yellow corn (YCF) and quinoa flour (QF) were evaluated, and the results are presented in Table (7). The organoleptic attributes (appearance, color, odor. texture. taste. and overall corn-quinoa flakes acceptability) of containing different levels of QF were significantly affected. The overall acceptability of flakes was determined by taking average of all the value pertaining to appearance, color, taste, texture and odor. It was found that corn flakes containing 20 and 40% of QF were found to secure maximum score, while the minimum score was found in samples containing 100% QF compared with the control sample. Corn-quinoa flakes samples containing 20 and 40 % QF showed no significant differences in all their sensory properties and were as acceptable as compared to control flakes prepared from 100% YCF, while further increase in substituted levels results in drastic reduction in most sensory properties. These results are approximately similar to those reported by Khalil et al. (35), Priyanka et al. (45), and El-Sebeay et al. (47).

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Samples	Appearance	Color	Taste	Odor	Texture	Overall acceptability
Control (100% YCF)	9.55±0.12ª	9.40±0.03ª	8.90±0.06ª	9.00±0.02ª	8.50±0.91ª	8.10±0.02ª
80% YCF +20% QF	9.50±0.04ª	9.38±0.09ª	8.87±0.10 ^a	9.10±0.08ª	8.55±0.07 ^a	8.00±0.16 ^a
60% YCF +40% QF	9.53±0.08ª	9.39±0.12ª	8.20±0.02 ^b	9.00±0.02ª	8.10±0.02 ^b	8.00±0.05ª
40% YCF +60% QF	8.50±0.13 ^b	8.76±0.08 ^b	7.85±0.08°	9.05±0.05 ^b	7.80±0.45°	7.60±0.02 ^b
20% YCF +80% QF	8.24±0.02°	8.19±0.02°	7.46±0.03 ^d	8.70±0.09°	7.25±0.13 ^d	7.55±0.08 ^b
100% QF	7.95±0.05 ^d	8.00±0.03d	7.10±0.05 ^e	8.15±0.04 ^d	7.00±0.09e	7.00±0.12 ^c

 Table (7): Organoleptic properties of corn-quinoa flakes produced from yellow corn and quinoa flour

Means \pm standard deviations; the means within the same column having different superscripts are significantly varied (P < 0.05) .; YCF: yellow corn flour; QF: Quinoa Flour.

5. Conclusion

In this research, incorporating quinoa flour into corn flakes significantly increased crud protein, ether extract, crud fiber, and ash content. Also, significant improvements were observed in minerals, amino acids contents and some phytochemicals, including phenolics, flavonoids, and antioxidant activities of produced corn-quinoa flakes. Color values showed no significant changes with replacing yellow corn flour with quinoa flour up to 40%. Corn flakes enriched with quinoa flour up to 40% were found to be the most accepted samples from the sensory point of view compared to the control samples. Overall, it could be recommended that the feasibility utilization of quinoa seeds flour in bakery products and other food products is an excellent source of essential amino acids and other nutrients.

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Conflict of Interests

The authors declare no conflict of interest, financial or otherwise

6. References:

1-Dendy, D. A. V. and Dobraszczyk, B. J. (Eds.). (2001). Cerealesy productos derivados. Químicay tecnologi a. Zaragoza: Acribia.

2- Edible Flakes Market Size, Share & Trends Analysis Report By Product (Corn, Oat, Wheat), By Distribution Channel (Offline, Online), By Region (North America, Europe, APAC, CSA, MEA), And Segment Forecasts, 2020 – 2027. (2019). Report ID: GVR-4-68038-213-6. https://www.grandviewresearch.com/industr y-analysis/edible-flakes-market.

3-Miller, H. E.; Rigelhof, F.; Marquart, L.; Prakash, A. and Kanter, M. (2000). Antioxidant content of whole grain breakfast cereals, fruits and vegetables. J AmColl Nutr;19:312S–9S.

4-Ryan, L.; Thondre, P. and Henry, C. (2011). Oat-based breakfast cereals are a rich source of polyphenols and high in antioxidant potential. J Food Compos Anal; 24: 929–34.

5-Rouf Shah, T.; Prasad, K. and Kumar, P. (2016). Maize a potential source of human nutrition and health: A review. Cogent Food & Agriculture, 2: 1166995. <u>http: //dx.doi.org/</u> 10.1080/ 23311932.2016. 1166995.

6-FAO (2001). FAO-STAT Agriculture Data. h t t p : // w w w . F a o . o r g / p a g e /collections?subset=agriculture (20 Nov. 2001).

7-Azevedo, R. A.; Arruda, P.; Turner, W. L.; Lea, P. J. (1997). The biosynthesis and metabolism of the aspartate derived amino acids in higher plants. Phytochemistry, v.46, p.395-419.

8-Bhargava, A.; Shukla, S. and Ohri, D. (2006). Chenopodium quinoa An Indian perspective. Industrial Crops and Products. 23:73–87.

9-Repo-Carrasco, V. M. and Serna, L. A. (2011). Quinoa (Chenopodium quinoa, Willd.) as a source of dietary fiber and other functional components. Cienc. Tecnol. Aliment., Campinas, 31(1): 225-230.

10-González Martín, M. I.; Wells Moncada, G.; Fischer, S. and Escuredo, O. (2014). Chemical characteristics and mineral composition of quinoa by nearinfrared spectroscopy. J Sci Food Agric 94: 876-881.

11-Nowak, V.; Du, J. and Charrondière, U. R. (2016). Assessment of the nutritional composition of quinoa (Chenopodium quinoa Willd.). Food Chemistry 193:47–54.

12-Tang, Y.; Li, X.; Zhang, B.; Chen, P. X. and Liu, R. (2015) Characterisation of

phenolics, betanins and antioxidant activities in seeds of three Chenopodium quinoa Willd. genotypes. Food Chem 166: 380-388.

13-Bertazzo, A.; Comai, S. and Brunato, I. (2011). The content of protein and nonprotein (free and protein-bound) tryptophan in Theobroma cacao beans. Food Chem. 124: 93-96.

14-Ruales, J., De-Grijalva,Y.; Lopez-Jaramillo, P. and Nair, B. M. (2002). The nutritional quality of infant food from quinoa and its effect on the plasma level of insulin-like growth factor-1 (IGF-1) in undernourished children. International Journal of Food Science & Nutrition: 53:143-54.

15-Farinazzi-Machado, M. F. V.: Barbalho, S. M.; Oshiiwa, M.; Goulart, R, and Junior, P. O. (2002). Use of cereal bars with quinoa (Chenopodium quinoa Willd) to reduce risk factors related to cardiovascular diseases. Cienc Technol Aliment Campinas, 32(3): 239-44.

16-Pasko, P.; Barton, H.; Zagrodzki, P.; Izewska, A.; Krosniak, M.; Gawlik, M. and Gawlik, M. (2010). Effect of diet supplemented with quinoa seeds on oxidative status in plasma and selected tissues of high fructose-fed rats. Plant Foods Hum. Nutr. 65 (2), 146-151.

17-Foucaultm A.; Even, P.; Lafont, R.; Dioh, W.; Veillet, S.; Tome, D.; Huneau, J.; Hermier, D. and Quignard-Boulange, A. (2014). Quinoa extract enriched in 20hydroxyecdysone affects energy homeostasis and intestinal fat absorption in mice fed a high-fat diet. Physiol Behav. 10;128:226-31. doi: 10.1016/ j.physbeh. 2014.02.002. Epub 2014 Feb 15.

18-Darwish, A. M. G.; Al-Jumayi, H. A. O. and Elhendy, H. A. (2020). Effect of germination on the nutritional pofile of Quino (Chanopodium quinoa Willd) seeds and its anti-anemic potential in spragus-Dawley male albino rats. Cereal Chemistry. 3586, 1575.

19-Sezgin, A. C. and Sanlier, N. (2019). A new generation plant for the conventional cuisine quinoa (Chanopodium quinoa Trends in Food Science and Willd). Technology. vol 86 pag 51-58.

20-Srujana, M. N. S.; Kumari, B. A.; Suneetha, W. J. and Prathyusha, P. (2019). Processing technologies and health benefits of quinoa. The Pharma. Innovation Journal, 8(5): 155-160.

21-Zevallos, V. F.; Herencia, L. I.; Chang, F.; Donnelly, S.; Ellis, H. J. and Ciclitira, P. J. (2014). Gastrointestinal effects of eating quinoa (Chenopodium quinoa Willd.)in celiac patients. The American Journal of Gastroenterology. 109(2):270-278. Doi:10.1038/ajg.2013.431.

22-Rosell, C. M.; Cortez, G. and Repo-Carrasco, R. (2009). Bread making use of Andean Crops Quinoa, Kaniwa, Kiwicha, and Tarwi. Cereal Chemistry, 86(4): 386-392.

23-Leusner, S. J. (1999). Flaked RTE cereals and method of preparation. United State Patent, 5919503.

24-A.O.A.C., (2011). Official Methods of Analysis of AOAC International. 18th ed. Published by the AOAC International (revised edition), Gaithersburg, Maryland, U.S.A.

25-AOAC (2008). Official Methods of Analysis 16th ed. Association of Official Agriculture, Washington DC. Analytical Chemists International Arligton, Virginia, U.S.A.

26-FAO/WHO. (1973). Energy and Protein Requirements. Report of a Joint FAO/WHO Ad Hoc Expert Committee. WHO Tech. Report Series 522, FAO Nutrition Meetings Report Series, 52, Rome, Italy.

27-Jackson, M. L. (1973) Soil Chemical Analysis. Prentice Hall of Ind Private Limited, New Delhi.

28-Tan, K. H. (1996). Soil Sampling, Preparation and Analysis. Marcel Dekker,

Egypt. J. Chem. 66, No. SI 13. (2023)

Inc.New York, NY.

29-Gamez-Meza, N.; Noriega-Rodriguez, J. A.; Medina-Juarez, L. A.; Ortega Garcia, J.; Cazarez-Casanova, R. and Angulo-Guerrero, O. (1999). Antioxidant activity in soybeen oil of extracts from Yhompson grape bagasse. Journal of the American Oil Chemisis Society, 76: 1445-1447.

30-Bahorun, T., A. L. uximon-Ramma, A. Crozier and O. I. Aruoma, (2004).Total phenol, flavonoid, proanthocyanidin and vitamin C levels and antioxidant activities of Mauritian vegetables. J. Sci. FoodAgric., 84: 1553–1561.

31-Beta, T.; Nam, S.; Dexter, J. E. and Sapirstein, H. D. (2005). Phenolic content and antioxidant activity of pearled wheat and roller-milled fractions. Cereal Chemistry, 82: 390-393.

32-Akesowan, A. (2010). Effect of konjac incorporated with soy protein isolate on quality characteristics of reduced-fat chiffon cakes. African Journal of Biotechnology, 9(28): 4386- 4391.

33-Zabik, M. E. and Hoojjat, P. (1984). Sugar-shap cookies 95: 9-18. prepared with wheat-navy bean sesame seed flour 30. Berski, W., D. Litwinek, H. Gambus and A. Nowotna, blends. Cereal Chem., 61: 41-44.

34-Armitage, P., (1971). Statistical Methods in Medical Research. Blackwell Scientific Publications, London.

35-Khalil, M. M.; Ghoneim, G. A.; Youssif, M. R. and Tamimy, M. (2018). Effect of adding corn flour to quinoa flour to produce gluten free pn bread. Journal of Food and Dairy Science, 3, 161-167.

36-El-Sohaimy, S. A.; Shehata, M. G. and Toshev, A. D. (2020). Development and characterization of functional pan bread supplemented with quinoa flour. Journal of Food Processing and Preservation. 45(1), 1-10.

37-Lilian, E. and James, A. (2009). Quinoa (Chenopodium quinoa Willd.): Composition, Chemistry, Nutritional, and Functional Properties. Food and Nutrition Research, 58: 1 -31.

38-Demir, B. and Bilgicll, N. (2020). Utilization of Quinoa flour (*Chenopodium Quinoa Willd*) in gluten-free pasta formulation: Effect on nutritional and sensory properties. Food Science and Technology International. 27 (3): 242-250.

39-Hager, A. S.; Wolter, A.; Jacob, F.; Zannini, E. and Arendt, E. K. (2012). Nutritional properties and ultra-structure of commercial gluten free flours from different botanical sources compared to wheat flours. Journal of Cereal Science, 56(2): 239-247.

40-Nascimento, A. C.; Mota, C.; Coelho, I.; Gueifão, S.; Santos, M.; Matos, A. S., Gimenez, A.; Lobo, M.; Samman, N. and Castanheira, I. (2014). Characterisation of nutrient profile of quinoa (Chenopodium quinoa), amaranth (Amaranthus caudatus), and purple corn (Zea mays L.) consumed in Argentina: the North of proximates, minerals and trace elements. Food Chemistry, 148: 420-426.

41-Malik, A. A.; Bhat, A.; Kour, H.; Ahmad, N. and Gupta, N. (2017) Processing and Assessment of Quality Characteristics of Corn-peanut Flakes. Int. J. Food. Ferment. Technol. 7(2): 287-294.

42-Vilcacundo, R.; Miralles, B.; Carrillo, W. and Hernandez-Ledesma, B. (2018). In vitro chemopreventive properites of peptides released from quinoa protein under simulated gastrointestinal digestion. Food Research International.105:403-411.

43-Abdelaleem, M. A. and Elbassiony, K. R. A. (2020). Evaluation of phytochemicals and antioxidant activity of Gamma irradiated quinoa (Chenopodium quinoa). Brazilian Journal of Biology. 17: 1-8.

44-Nizamlioglu, N. M. and Nas, S. (2010). The phenolic compounds in Vegetables and fruits; structures and their importance. Electronic Journal of Food Technology. 5(1), 20-35.

45-Priyanka, M.; Jessie-Suneetha W, Maheswari, K.; Suneetha, K. B. and B Anila Kumari (2017). Development and evaluation of quinoa based snack items. Journal of Pharmacognosy and Phytochemistry 2017; 6(4): 831-833.

46-Dewidar, O. M. and El-ghandour, H. M. A. (2020). Development of extruded snacks and corn flakes using yellow corn and by product broken beans. Midell East Journal of Applied Sciences. 10(2), 390-406.

47-El-Sebeay, A.S. & Lamia M. Hafez (2018). Chemical and Biological Evaluation of Some Products from Quinoa-Based Blends for Celiac Disease. Alex. J. Food Sci. & Technol., 15(1), 23-3.