
Rutin Ameliorates Acrylamide-Induced Hepatotoxicity and Biochemical Disturbance in Male Albino Rats

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Abstract

The present study investigated the effect of rutin on acrylamide (ACR) toxicity was evaluated in experimental animal rats. The rats were feeding on basal diet and received oral 20 mg/kg b.w of acrylamide with 100, 50 and 25 mg/kg w.t of rutin; others were received the acrylamide concentrations only. The present results showed that 8-hydroxydeoxy guanine (8-OHdG), protein carbonyl (PC), reactive oxygen species (ROS), glutathione peroxidase (GPx), tumor necrosis factor- α (TNF- α), nitric oxide (NO) myeloperoxidase (MPO), caspase-3, bax, P53 and cytochrome-C in liver tissues were increased in rats-treated with acrylamide. This effect of acrylamide was attenuated by rutin. In conclusion, rutin ameliorates hepatotoxicity induced by acrylamide in male albino rats. It can be recommend the use of rutin to mitigate the acrylamide concentration in food products.

Key words: Acrylamide, rutin, oxidative stress

Introduction

Diabetes In 2002, high level of ACR was detected in some fried and grilled starchy foods, which may be produced by the reaction between asparagine and reducing sugars or reactive carbonyls at temperatures above 120 °C (Mottram *et al.*, 2002). The discovery of ACR in cooked starchy foods prompted concerns about the carcinogenicity of those foods. Genetic toxicology studies have found that ACR has mutagenic effects both *in vivo* and *in vitro* experiments, and it can cause gene mutation and chromosomal abnormalities of somatic and germ cells of mammalian, such as the formation of micronuclei, and sister chromatid conversion (Camacho *et al.*, 2012). Moreover, it has been reported that epoxypropionamide, a metabolites of ACR, is the main material of mutagenic activity, which is easy combined with guanine of DNA then forming an adduct. International Agency for Research on Cancer (IARC) have evaluated the carcinogenicity of AA and listed it as a category carcinogen (Forstova *et al.*, 2014). Toxicity evaluation of pollutants is mainly carried out through the biological tests which spend a long time and high cost. What is more, it is difficult to test various amounts of contaminants in water

individually. In comparison, DNA biosensors providing a cheap and fast approach for the study of toxicity of pollutants have attracted much research interest Liu *et al.* (2012).

Rutin (quercetin-3-rutinoside), a natural flavonoid glycoside, being already clinically used for strengthening of capillaries (Wadworth and Faulds, 1992), possesses as other flavonoids iron/copper chelating and antioxidant activity which based on the mentioned known pathological mechanism, appears to be a useful drug.

Flavonoids are a large family of over 4000 ubiquitous secondary plant metabolites, comprising seven subclasses, anthocyanins (e.g. europolinidin), flavones (e.g. apigenin), flavonols (e.g. quercetin), flavanonols (e.g. taxifolin), isoflavones (e.g. genistein), flavanones (e.g. naringenin) and flavonol glycosides (e.g. rutin). Flavonoids are known to possess strong antioxidant properties, which can be due to both their radical-scavenging effects and metal-chelating properties, of which the former may dominate. Owing to the antioxidant properties, they can prevent deoxyribonucleic acid (DNA) and cells from oxidative damage, and thus possess a wide range of pharmacological

activities such as anti-tumor, anti-inflammatory and anti-aging (Undeger *et al.*, 2004).

Rutin (Scheme 1), is one of the most abundant flavonoids in the human diet, also known as vitamin P, has been shown to act as a scavenger of various oxidizing species, i.e. hydroxyl and peroxy radicals. As a result of these biological effects, its pharmacological activities have been widely explored including antibacterial, anti-inflammatory, antitumor, antiallergic, antiviral and antiprotozoal properties (Blasco *et al.*, 2004; Hasumura *et al.*, 2004).

MATERIALS AND METHODS

- MATERIALS

- Chemicals

The chemicals used in the present study as well as their sources are displayed as follows:

rutin was obtained from Sigma Chemical Company, USA. All other chemicals used in the present investigation were of bioanalytical grade.

Experimental animals

Sixty adult male albino rats weighing between 160-180g were used. The rats were obtained from the animal house of the National Organization for Drug Control and Research Egypt. The animals were kept under standard laboratory conditions of light/dark cycle (12/12 h) and temperature (20 - 25°C) for one week. They were provided with a nutritionally adequate basal laboratory diet. The basal diet consists of casein 10%, cotton seed oil 4%, salt mixture 4%, vitamin mixture 1%, carbohydrates (sucrose, starch 1:1) 80.8% and Choline Chloride 0.2% (American Institute of Nutrition., 1980).

Grouping of animals

A total number of 60 adult male albino rats were divided to equally sex groups as follow:

1-Normal control (NC): Rats were feeding basal diet. 2-Rats were feeding basal diet and received orally 20mg/kg acrylamide alone. 3-Rats were feeding basal diet and received orally 20mg/kg acrylamide (ACR) and 100 mg/kg rutin (RU). 4-Rats were feeding basal diet and received orally 20mg/kg acrylamide (ACR) and 50 mg/kg rutin (RU). 5- Rats were feeding basal diet and received orally 20mg/kg acrylamide (ACR) and 25 mg/kg rutin (RU). 6- Rats were feeding basal diet and received orally 100 mg/kg rutin (RU) alone.

Handling of Tissue Samples

Liver was collected and weighed. The liver was homogenized (10%, w/v) separately in ice-cold KCl (1.15%) sodium phosphate buffer (0.01M, pH 7.4) with a Potter-Elvehjem glass homogenizer (EZ stir, Korea). The homogenate was centrifuged at 10,000 ×g for 20 min at 4°C (Sigma 3k30, Germany). The supernatant of the liver was divided into three aliquots and collected into sterilized Eppendorf tubes and stored at -80°C until analysis. The supernatant was used for the estimation the

level of NO (Wang *et al.*, 2002) using Griess reagent, Protein Carbonyl (Levine *et al.*, 1990) Levels of TNF- α , 8-OHdG, cytochrome-c, caspase-3, Bax, and p53 were quantified using ELISA (BioTek, ELx-50, USA) kits according to the manufacturer's instructions and guidelines. Protein content in the supernatant was estimated by the method of Lowry *et al.* (1951).

Histological examination

Part of the liver was placed in well-sealed containers containing 10% formalin saline solution for 10 h at least, and then washed in tap water for 12 h. Serial alcohols (methyl, ethyl and absolute ethyl alcohols) were used for dehydration of the tissue samples. Tissue specimens were cleared in xylene and embedded in paraffin. The paraffin blocks were sectioned at 3 micron thickness by slide microtome. The obtained tissue sections were collected on the glass slides and stained by haematoxylin and eosin stains (Banchroft *et al.*, 1996) for histological examination by the light microscope.

METHODS

Bioassay

Determination of Myeloperoxidase (MPO)

Activity: Myeloperoxidase activity was determined using Bradley *et al.* (1982) method.

Determination of GPx activity: The activity of GPx was determination according to Brigelius-Flohe (1999).

Quantification of reactive oxygen species (ROS)

content: A modified version of a previously described assay for the intracellular conversion of nitro blue tetrazolium (NBT) to formazan by superoxide anion was used to measure the generation of reactive oxygen species (Vrablic *et al.*, 2001).

Determination of Nitric Oxide content: Nitric oxide (NO) level was measured by the determination of total nitrate and nitrite concentrations in the sample using the method of (Wang *et al.*, 2002, using Griess reaction).

Determination of lipid peroxidation content:

Lipid peroxides formation was determined in serum and brain homogenate as thiobarbituric acid reactive substances (TBARS). It was determined according to the method of (Uchiyama and Mihara 1978).

Determination of Protein Carbonyl

Concentration: Protein carbonyl concentration was measured in liver and heart by first forming labelled protein hydrazone derivatives using 2, 4-dinitrophenyl hydrazide (Smith *et al.*, 1991).

Determination of tumor necrosis factor- α (TNF- α). The activity of TNF- α was calculated according to (Bonavida 1991). The Rat TNF- α ELISA (Enzyme-Linked Immunosorbent Assay) kit is an *in vitro* enzyme-linked immunosorbent assay for the quantitative measurement of rat TNF- α in cell lysate and tissue lysate.

Determination of 8-hydroxy-2 deoxyguanosine (8-OHdG). The activity of 8-OHdG was calculated according to (Nadja *et al.* 2001).

Determination of Cytochrome-C Rat ELISA Kit. Determination of Cytochrome-C Rat ELISA Kit was measured According to Abcam's Cytochrome C kit designed for the quantitative measurement of Cytochrome C protein rat tissues.

Determination of Bax (Apoptosis regulator Bax) ELISA Kit. Biosource kit is designed for the quantitative measurement of Bax in tissues according to the manufacturer's instructions and guidelines.

Statistical analysis

The data were subjected to statistical analysis using SPSS program version 20. Independent t-test was adopted to compare between two groups. The values were expressed as mean \pm S.E. and were considered significantly different if *P* values were <0.05 (Bailey, 1995).

Results and Discussion

Effect of acrylamide on oxidative stress parameters:

The data obtained from Table (1) revealed that the effect of 20 mg/kg ACR and group received 20 mg ACR with 25 mg/kg RU on liver, indicated that 8-hydroxydeoxy guanine (8-OHdG), protein carbonyl (PC), lipid peroxidation (LPO), reactive oxygen species (ROS) were increased along with decreased in glutathione peroxidase (GPx) compare to control. On the other hand groups treated with 20 mg ACR plus 100 or 50 mg RU showed non-significant effect compare with control. Our result is similar to the results of Harborne and Williams (2000) showed numerous biological activities of interest, for example, antioxidant capacity, anti-inflammatory action and stimulation of the immune system. Complementary studies of cytotoxicity, genotoxicity and mutagenicity indicated that the majority of flavonoids (such as catechins, genistein, daizein and rutin, among others) show negative results in these tests (Hollman and Katan, 1996; Nijveldt *et al.*, 2001; Birt *et al.*, 2001; Yao *et al.*, 2004) Ghorbel *et al.* (2015) showed that acrylamide (20 mg/kg body weight) for 3 weeks increase malondialdehyde, advanced oxidation protein product, H₂O₂ and decrease in GSH and Non-protein thiol levels in erythrocytes. In addition, activities of catalase, glutathione peroxidase and superoxide dismutase were decreased in all treated rats.

The enzymatic antioxidant defense systems are the natural protectors against lipid peroxidation, Glutathione peroxidase, (GPx) is an important scavenger of H₂O₂ and prevents the generation of hydroxyl radical and protect the cellular constituents from oxidative damage (Scott *et al.*, 1991) Tables (1).

Effect of acrylamide (ACR) on inflammatory marker parameters:

The results in Table (2) showed that inflammation marker revealed that the effect of ACR on liver rats. Groups which received 20 mg/kg ACR alone showed increase in tumor necrosis factor- α (TNF- α), Nitric Oxide (NO) and myeloperoxidase (MPO) compare with other groups. Our results revealed that MPO was increased significantly due to ACR toxicity, which are in agreement with the findings of Alturfan *et al.* (2012). Nakazato *et al.* (2007) demonstrated that MPO derived chlorinated oxidants and hydroxyl radicals play key roles in inducing oxidative stress mediated apoptosis in myeloid leukemia cells, with ROS being responsible for triggering apoptosis. After exposure to ACR, hepatic pro-inflammatory cytokines expression including tumor necrosis factor- α , interleukin-6, interleukin-1 β was increased suggesting leucocytes infiltration in the liver. A random DNA degradation was observed on agarose gel only in the liver of exposed rats to acrylamide treatment (Ghorbel *et al.*, 2015). On the other hand the groups which received 20 mg/kg ACR with 100 or 50 mg/kg RU represent non-significant effects compare with control group. Jyh-Ming-Chow *et al.* (2005) evaluated the cytotoxic potential of rutin in RAW264.7 cells (mouse bone marrow cell line) utilizing the MTT assay. After 12 h of treatment, rutin at concentrations of 25, 50, and 100mM did not have any toxic effect in this cell line. The lack of cytotoxic effect was also observed by Iffert *et al.* (2000) utilizing the MTT assay in three different breast cancer cell lines (MaTu p19, MaTu 0.1 and MaTu 0.25), after 24 h of exposure to 94.8 mM rutin. These authors recommended eating foods with the flavonoid rutin, since it does not have any harmful effect on health, while protecting tissues against free radicals.

Complementary studies in vivo showed that mice treated by gavage with 625, 1250 and 2500 mg/kg rutin for two consecutive days did not cause primary damage in DNA and or the formation of micronuclei in bone marrow cells (Silva *et al.*, 2002) In the evaluation of the protective effect of rutin in MNCTB assay against the DNA damage-inducing agent benzo[a]pyrene only the three highest concentrations, 90, 270 and 810mM produced statistically significant protection, where the percent reduction in damage was 36.%, 51.4% and 54.%, respectively.

Effect of acrylamide (ACR) on apoptosis marker parameters:

The data obtained from Table (3) revealed that the effect of RU by three doses (100, 50 and 25 mg/kg) in caspase-3, Bax, P53 and cytochrome-C on rats treated with ACR after 30 days. The rats which received 20 mg/kg ACR with 100 and 50 mg/kg RU represent non-significant compare with control group. On the other hand, groups which received

20 mg/kg ACR alone indicated increased in Caspase-3, Bax, P53 and Cytochrome-C compare with control group. These results according to **Hyun-Jung *et al.* (2000)**, there two categories of possible mechanisms involved in the protection of hepatic cells (HepG2, HTC) against DNA damage by B[a]P:1) inactivation of the biotransformation enzymes of the cytochrome P450 system (phase I) which leads to the reduced formation of reactive metabolites and DNA adducts; 2) direct interaction with the derivatives of polycyclic aromatic hydrocarbons. However, further studies are needed to determine the mechanism of action for the protective effect of flavonoids.

Acrylamide produces genotoxic effect through its reactive metabolite, glycidamide, generated by catalysis with cytochrome P450 (**Sumner *et al.*, 1999**). The resulting metabolite is an epoxide derivate i.e glycidamide which is more reactive toward DNA and proteins than the parent compound, acrylamide (**Besartinia and Pfeifer, 2007**).

Glycidamide is regarded as a cancer initiating species reacting with DNA to cause mutagenicity and clastogenic (**Adler *et al.*, 2000**). The clastogenic potential is responsible for dominant lethality targeting male gonadal spermatocytes to result in reproductive toxicity in rodent (**Xiao and Tates, 1994; Adler *et al.*, 2000**). Several authors reported that ACR has the ability to induce biochemical changes and hormone disturbances (**Park *et al.*, 2002; Yousef and El- Demerdash, 2006; Parzefall, 2008**).

The experimental data shown in Table (3) no significant effect Caspase-3, Bax, P53 and Cytochrome-C content in concentrated (100, 50 mg/kg) RU with 20 mg/ kg ACR compared with control groups. On the other hand, rats treated with ACR 20 mg/kg alone indicated a highly significant increase in Caspase-3, Bax, P53 and Cytochrome-C levels in rats compare with control.

Conclusion

The present study declares that, rutin has an appreciable ability to prevent hepatotoxicity caused by acrylamide, possibly via its chemical constituents which has a free radical scavenging properties.

Table (1): Ameliorative effects of quercetin against acrylamide-induced oxidative stress in rat liver

Groups	Oxidative stress in liver			
	GPx (IU/g protein)	ROS (μ mol NBT/g tissue)	PC (nmol /mg)	8-OHdG (ng/mg protein)
Control	25.63 \pm 0.59 ^a	19.20 \pm 0.18 ^a	28.0 \pm 0.58 ^a	4.61 \pm 0.21 ^a
20 mg (ACR)	16.83 \pm 0.45 ^c	23.36 \pm 0.59 ^c	39.7 \pm 0.81 ^c	8.51 \pm 0.39 ^c
20mg ACR+ 100 mg RU	25.83 \pm 0.15 ^a	19.00 \pm 0.68 ^a	28.6 \pm 0.32 ^a	4.14 \pm 0.58 ^a
20 mg ACR+ 50 mg RU	25.10 \pm 0.41 ^a	19.13 \pm 0.55 ^a	28.8 \pm 0.21 ^a	4.66 \pm 0.63 ^a
20 mg ACR+ 25 mg RU	20.31 \pm 1.04 ^b	19.89 \pm 0.61 ^b	32.4 \pm 0.80 ^b	4.81 \pm 0.53 ^b
100 mg RU alone	26.17 \pm 0.47 ^a	18.86 \pm 0.38 ^a	27.8 \pm 0.60 ^a	4.18 \pm 0.62 ^a

Values are mean \pm SE of 6 rats. Values with the same letters are not significant at ($P \leq 0.05$). AA= acrylamide, RU= Rutin. Values on the same column not sharing the same superscript letters were significantly different ($P < 0.05$).

Table (2): Ameliorative effects of quercetin against acrylamide-induced Inflammation markers in liver

Groups	Inflammation markers in liver		
	MPO (U/mg protein)	Nitric Oxide (μ mol/mg protein)	TNF- α (pg/mg protein)
Control	10.55 \pm 0.21 ^a	8.90 \pm 0.53 ^a	14.21 \pm 0.46 ^a
20 mg (ACR)	23.38 \pm 0.57 ^c	13.58 \pm 0.90 ^c	37.18 \pm 0.98 ^c
20mg ACR+ 100 mg RU	10.58 \pm 0.39 ^a	8.66 \pm 0.71 ^a	13.94 \pm 0.65 ^a
20 mg ACR+ 50 mg RU	10.96 \pm 0.62 ^a	8.81 \pm 0.68 ^a	14.03 \pm 0.48 ^a
20 mg ACR+ 25 mg RU	12.71 \pm 0.54 ^b	10.37 \pm 0.59 ^b	22.5 \pm 0.21 ^b
100 mg RU alone	10.42 \pm 0.32 ^a	8.59 \pm 0.61 ^a	13.98 \pm 0.37 ^a

Values are mean \pm SE of 6 rats. Values with the same letters are not significant at (P \leq 0.05). AA= acrylamide, RU= Rutin. Values on the same column not sharing the same superscript letters were significantly different (P<0.05).

Table (3): Ameliorative effects of quercetin against acrylamide-induced apoptosis markers in liver

Groups	Apoptosis marker in liver			
	Caspase-3 (ng/mg protein)	Bax (ng/mg protein)	P53 (pg/ml)	Cytochrome-C (ng/ml)
Control	0.93 \pm 0.10 ^a	0.23 \pm 0.06 ^a	74.21 \pm 11.95 ^a	0.46 \pm 0.22 ^a
20 mg (ACR)	2.17 \pm 0.15 ^c	1.38 \pm 0.12 ^c	182.21 \pm 8.39 ^c	0.91 \pm 0.32 ^c
20mg ACR+ 100 mg RU	0.94 \pm 0.18 ^a	0.24 \pm 0.09 ^a	76.00 \pm 21.16 ^a	0.46 \pm 0.51 ^a
20 mg ACR+ 50 mg RU	0.95 \pm 0.08 ^a	0.25 \pm 0.16 ^a	76.26 \pm 16.07 ^a	0.48 \pm 0.24 ^a
20 mg ACR+ 25 mg RU	1.52 \pm 0.17 ^b	0.64 \pm 0.05 ^b	119.60 \pm 16.03 ^b	0.68 \pm 0.31 ^b
100 mg RU alone	0.91 \pm 0.13 ^a	0.21 \pm 0.03 ^a	72.21 \pm 7.84 ^a	0.42 \pm 0.08 ^a

Values are mean \pm SE of 6 rats. Values with the same letters are not significant at (P \leq 0.05). AA= acrylamide, RU= Rutin. Values on the same column not sharing the same superscript letters were significantly different (P<0.05).

Histopathological findings of liver

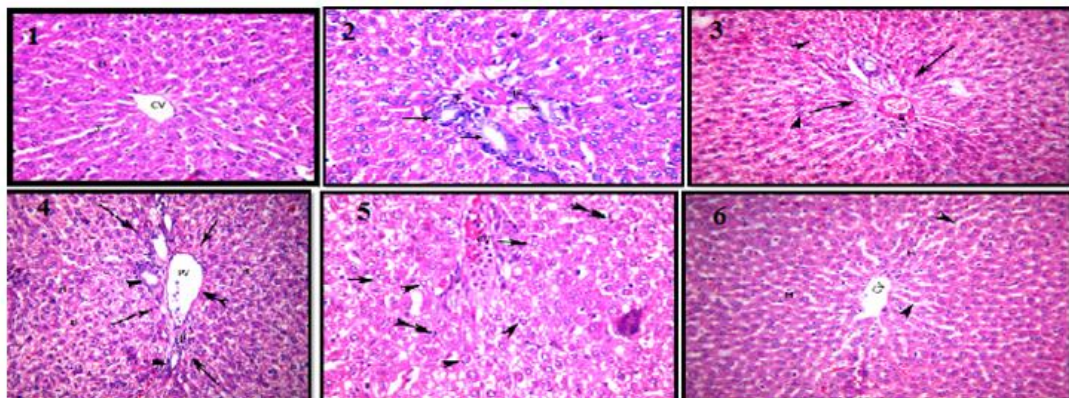


Fig (1): Photomicrograph of liver section taken from control animals showing normal architecture with normal appearance of hepatocytes (H), sinusoid (S) and vascular channels as central vein (CV). Fig (2): Photomicrograph of liver section taken from animals treated 20 mg acrylamide alone showing most portal area were constricted with proliferated bile duct (arrow) and periportal inflammation (IF). Proliferated van kupfer cells also were seen. Fig (3): A photomicrograph of liver section taken from animals treated 20 mg acrylamide with 100 mg Rutin showing constricted portal area (arrow) with mild vacuolated hepatocytes (head of arrow) and mild aggregation of inflammatory cells (IF). Fig (4): A photomicrograph of liver section taken from animals treated 20 mg acrylamide with 50 mg rutin showing expanded portal area (arrow), mild dilated congested portal vein (PV) and proliferated bile duct (head of arrow) together with moderate vacuolated hepatocytes (H). Mild inflammatory cells infiltrations in periportal were also seen (IF). Fig (5): A photomicrograph of liver section taken from animals treated 20 mg acrylamide with 25 mg rutin showing sever vacuolated hepatocytes (arrow) with magnetite chromatin (head of arrow). Proliferated vankuffer cells (2 head of arrow) and dilated congested central vein (CV) were also seen. Fig (6): A photomicrograph of liver section taken from animals' treated 100 mg rutin alone showing normal hepatocytes (H), central vein (CV) and sinusoidal (head of arrow).

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