

Effect of Citric Acid and Saccharomyces Administration on the Growth Performance, some Biochemical and Histopathological Parameters on Broiler Chicks.

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Abstract

The present study was carried out to assess the efficacy of yeast probiotics and citric acid supplementation on the growth performance, some biochemical profiles, and histopathological alterations of broiler chicks. A total of 240 one-day-old Cobb 500 broiler chicks were purchased and randomly assigned into four groups (n=60) for the experimental trial. The four groups are, the control group (C-gr), the citric acid group (CA-gr), the probiotics group (PB-gr), and the combination group (CO-gr). The control group Fed on a standard diet and tap water. The CA group was given a standard diet and acidified water (at pH 4.5) using citric acid monohydrate (0.8 g /liter) in drinking water. Probiotic (Saccharomyces) was given to the PB group at a rate of 2x10⁹ CFU/liter, while the combination group received combination of PB 2x10⁹ CFU/liter and CA in their drinking water 0.8 g /liter. The trial was lasted for 5 weeks. Results revealed that live body weight (LBW) of the combination group was significantly higher than other groups. Comparing CA gr to other groups, the gizzard weight was significantly (P<.05) high. Liver weight revealed significant increasing in all treated groups. Likewise, heart weight showed a significant increase in the CA and PB groups. MDA level showed significant decrease in all treated groups. GSH level and Serum T3 concentration were significantly (p<0.05) increased only in PB and combination groups, while serumT4 concentration was obviously elevated in all treated groups. The CA, PB, and CO groups showed a significant (p<.05) decrease in TC and TG. In all experimental groups, HDL-C was significantly (p<.05) increased, but LDL-C showed a significant reduction especially in the PB group compared to C-gr. CA and PB groups showed long villi than the C-gr and CO-gr at 3rd and 5th weeks. In general, a combination of PB and CA has a promising effect and may boost the growth performance and biochemical profiles of broiler chicks after short-term supplementation.

Keywords: Broilers, Probiotics, Citric acid, Biochemical, Histopathological parameters

1. Introduction

Poultry meat and their products are widely consumed in Egypt. Large-scale consumption of poultry meat is due to their competitive pricing in comparison with other animal meat products. One goal

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of every poultry producer is to maintain the balance between low-cost diets and maximizing output. Poultry production is a company that, like other businesses, aims to make a profit [1]. Thus, the industry works toward obtaining the greatest amount of weight gain for the lowest production cost [2]. The production of poultry still experiences significant losses because of pathogenic bacterial invasion and the harmful effects of these organisms on the birds includes decrease weight

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gain or higher mortality, poor feed efficiency, and weak chicken performance [3]. All of these effects, including the load in the environment and gastrointestinal tract, are tightly correlated with the host bird's qualitative and quantitative microbial load [4].

Antibiotics as antimicrobial feed additives are usually used to eliminate or suppress the hazardous intestinal microorganisms and to hasten growth and feed efficiency [5]. Recently, confidence has diminished about the use of antibiotics due to fears over bacterial resistance and possible transmission of these antibiotic residues into the human food chain [6]. These residues may have a potential health hazard to humans.

The Aim of work is to study the effect of citric acid and saccharomyces cerevisiae on body weight, Edible organs weight, hepatic malondialdehyde (MDA), reduced glutathione (GSH), thyroid hormones (triiodothyronine T3 and thyroxine T4), Lipid profile (TC, TG, HDL-C and LDL-C) and histomorphological and morphometrical changes in different segments of intestine.

2. Materials and methods:

2.1. Birds and management

A total of 240 one-day-old apparent healthy Cobb 500 broiler chicks of both sexes, weighing 50 ± 5 g has been purchased from Ismailia-Miser for Poultry Company, Egypt. Chicks were raised in a well-ventilated, clean environment with a range of temperatures (32–35 °C). Electric heaters were employed to change the temperature, and electric lamps had been used as a source of light. Chicks received light for the entirety of the day (24 h). Vaccinations were supplied to chicks, against Newcastle disease (ND) and Infectious Bursal Disease (Gumboro) at some point of the experimental period as ordinary.

2.2. Experimental Diet

A starter between the ages of 1 and 17 days and a finisher between the ages of 18- and 35-days' meals were offered to the experimental birds meanwhile the trial as provided in table 1. According to [7], both diets have been designed to meet the nutritional needs of broilers.

2.3. Experimental Design:

Experimental chicks were divided into 4 equal groups, each contributing 60 chicks. Control group (C-gr) provided with a normal diet and tap water. While citric acid (CA-gr) was produced using citric acid monohydrate (Sigma, Australia) with a normal diet and drinking water acidified to (pH 4.5) [8]. Probiotic group (PB-gr) received a basal diet and probiotics 2x10⁹ CFU / liter in their drinking water. A combination group (CO-gr) received mixture of probiotics 2x10⁹ CFU / liter and citric acid in their drinking water 0.8 g /liter. The treatment continued for 35 days. Body weight gain of each chick were measured at 28 and 35 days.

Table 1: Compositions of the basal diets								
Ingredients (%)	Starter (1-17	Finisher						
	days)	(18-35 days)						
Yellow corn	61.77	73.00						
Soybean meal	24.50	13.60						
(44 % CP)								
Corn gluten	10.00	10.00						
meal								
Bone meal	2.60	2.0						
Limestone	0.30	0.60						
Vit. & Min	0.30	0.30						
Premix								
NaCl	0.25	0.25						
Lysine	0.18	0.20						
Dl- Methionine	0.10	0.05						
Total	100	100						

2.4. Blood sampling:

Wing vein blood samples were obtained at the age of 28 days and at the end of the experimental period (at 35 days) after an overnight fast, serum samples were drawn into simple tubes and kept at -20°C till lipid and hormonal profile analysis.

2.5. Edible organs weight:

The edible visceral organs (heart, liver, and gizzard) of the slaughtered chickens were removed, cleaned, and weighed at the end of the trial.

2.6. Thyroid Hormones:

Using Taytec Incorporation's (Canada) commercial enzyme-linked immunoassay kits, serum levels of T3 and T4 have been measured in accordance with the manufacturer's protocol.

2.7. Lipid profile:

Using commercial kits (Diamond, Egypt), the levels of high-density lipoprotein cholesterol (HDL-c), triglycerides (TGs) and total cholesterol (TC) were calorimetrically determined. Enzymatic calorimetric kits (QCA Co., Spain) were used to measure serum low-density lipoprotein cholesterol (LDL-c) via accordance with [9].

2.8. Histopathological Examination:

Small intestine samples were obtained shortly after the chicks were slaughtered and preserved in 10% formalin saline for 24 h. Hematoxylin and eosin staining of 5- μ m thick sections was done using standard histological laboratory methods [10]. Using Image J software, a morphometric analysis of intestinal villus height (VH, m) and crypt depth (CD, m) was performed. [11]. The CD was calculated as the depth of the invagination between two adjacent villi whereas the VH was measured from the villus top to the lamina propria [12].

2.9. Statistical analysis:

Data collected from treated groups were statistically analyzed in comparison to control group for the mean and standard error using statistical software program (SPSS for Windows, version 16, USA) according to [13]. Differences between means of different groups were carried out using oneway ANOVA followed by Duncan Multiple Range tests [14]. Differences were considered significant at (P<0.05) and highly significant at (P<0.01).

3. Results

3.1. Effects of dietary Citric acid and/or yeast probiotic supplement on body weight and internal organs of broilers

Table (2) demonstrated that LBW, gizzard weight showed a significant (P<0.05) improvement in citric acid, yeast probiotic and combination group

than control after 28 days, Combination group showed significant (P<0.05) higher body weights than yeast probiotic and citric acid group. Liver and heart weights resulted in non-significant (P>0.05) alternation between all groups at 28 days. At 35 days, citric acid, yeast probiotic and citric acid and yeast probiotic groups exhibited significant (P<0.05) higher body weights, liver weight than control but no significant variations were observed between them. Heart weight exhibited significant (P<0.05) higher values in citric acid and yeast probiotic groups than control and combination groups while the gizzard weight exhibited non-significant (P>0.05) alternation at 35 days among all groups.

3.2. Effects of dietary Citric acid and/or yeast probiotic supplement on malondialdehyde content (MDA) and glutathione content(GSH)

Table (3) demonstrated that the levels of hepatic MDA, as an indicator for lipid peroxidation, was significantly (P<0.05) reduced in citric acid, yeast probiotic and combination groups than control at 28 days and 35 days. There were nonsignificant (P>0.05) differences observed among treated groups at 28 days or 35 days. Nonetheless, at 28 days of experiment, the activity of hepatic GSH revealed a significant promotion (P<0.05) in PB-gr and CO-gr than control and citric acid groups. At 35 days, the activity of GSH in serum showed significant (P<0.05) elevation in yeast probiotic and combination groups than citric acid treated group and control group. No significant (P>0.05) differences observed between citric acid and control groups as well as yeast probiotic and combination groups.

3.3. Effects of dietary Citric acid and/or yeast probiotic supplement on serum thyroid hormones (T3 and T4)

As indicated in Table 4, the level of T3 was significantly (P<0.05) elevated in yeast probiotic and combination groups than control and citric acid groups at 28 days and 35 days. The combination group showed superior T3 level than yeast probiotic group at 28 days and 35 days. The T4 levels were significantly (P<0.05) improved in citric acid,

Table 2. Enects of dictary entrie acid and yeast problems supprementent biology weight and internationgans weight of biolets									
Tested	L	After 28 days				After 35 days			
parameters									
/ days	C-gr	CA-gr	PB-gr	CO-gr	C-gr	CA-gr	PB-gr	CO-gr	
Body	920.00 ^d	1153.33 ^c	1286.66 ¹	° ± 516.00 ^a	1536.25 ^b	2104.50 ^{<i>a</i>}	2098.25 ^{<i>a</i>}	2270.00 ^{<i>a</i>}	
weight (g)	± 30.55	± 17.63	6.66	± 23.43	± 27.65	± 61.16	± 99.69	± 88.53	
Gizzard (g)	$19.00^b \pm$	22.33 ^a	$23.34^{a} \pm$	$26.66^a \pm$	$23.41^a \pm$	29.51 ^{<i>a</i>}	28.90^{a} ±	28.34^a ±	
	2.08	± 2.18	1.66	0.88	0.93	± 0.51	1.00	1.23	
Liver (g)	$21.67^{a}\pm$	20.00 ^{<i>a</i>}	$21.66^a \pm$	$16.67^a \pm$	$42.93^{b} \pm$	50.51 ^a	52.61^{a} ±	49.98^{a} ±	
	1.66	± 2.88	1.66	1.65	2.32	± 1.78	1.24	2.42	
Heart (g)	$5.37^a \pm$	5.26 ^{<i>a</i>}	$5.73^a \pm$	6.11^{a} ±	$7.55^b \pm$	10.43 ^{<i>a</i>}	11.49^{a} ±	$7.96^b \pm$	
	0.26	± 0.18	0.27	0.40	0.37	± 0.48	0.65	0.49	

Table 2: Effects of dietary citric acid and yeast probiotic supplement on body weight and internal organs weight of broilers

* Values are significant at p < 0.05

Table 3: Effects of dietary citric acid and yeastprobiotic supplement on malondialdehyde (MDA) and glutathione content (GSH) ofbroilers

	After 28 days				After 35 days			
Groups	C-gr	CA-gr	PB-gr	CO-gr	C-gr	CA-gr	PB-gr	CO-gr
MDA	$0.82^{a} \pm$	0.54^{b}	$0.66^{b} \pm$	$0.55^b \pm$	$0.80^{a} \pm$	$0.48^b \pm$	0.56^{b}	$0.53^{b} \pm$
(nmol/mg)	0.02	± 0.06	0.03	0.03	0.01	0.05	± 0.03	0.02
GSH (nmol/g)	$6.17^{b} \pm$	7.63^{b}	$8.81^a \pm$	$8.83^a \pm$	$8.87^{b} \pm$	10.70^{b}	12.77 ^a	14.17 ^a
	0.27	± 0.43	0.61	0.71	0.61	± 0.62	± 0.22	± 0.58

* Values are significant at p < 0.05

yeast probiotic and combinations group than control group at 28 days and 35 days.

3.4. Effects of dietary Citric acid and/or yeast probiotic supplement on lipid profile parameters

At 28 days, the yeast probiotic group had significantly lower total cholesterol (TC) and triglycerides (TG) than the control group (P<.05). All experimental groups revealed non-significant changes in High-density lipoprotein cholesterol (HDL-c) and Low-density lipoprotein cholesterol (LDL) compared to the control.

At 35 days, the CA, PB, and combination groups had significantly lower levels of TC and TG than the control group. In comparison to control, HDL-C revealed a significant increasing (p<0.05) in all treated groups. Compared to the control, LDL-C significantly decreased in all experimental groups (CA, PB, and combination groups), with highly significant reduction in PB as compared with CA and combination group (Table 5).

3.5. Effects of dietary Citric acid and/or yeast probiotic supplement on histology of intestine at 3rd and 5th week

Control group showed normal healthy intestinal villi of all segments (duodenum, jejunum and ilium. Citric acid fed group showed normal healthy long intestinal villi with increase in number of goblet cells lining the intestinal villi. The probiotic fed group showed normal histological architecture, including long villi with normal epithelial lining and normal intestinal glands. The combination group had normal histological intestinal villi. Figures (1-6).

4. Discussion

In many countries throughout the world, the use of antibiotics as a growth enhancer in poultry feed has been prohibited.

In order to improve growth and prevent diseases in poultry, it is important to look for al-

	After 28 days				After 35 days			
Groups	C-gr	CA-gr	PB-gr	CO-gr	C-gr	CA-gr	PB-gr	CO-
								gr
T3	3.98 ^c	4.91 ^{<i>a</i>}	4.73 ^{<i>a</i>}	4.45^{b}	4.13 ^c	5.37 ^{<i>a</i>}	5.16 ^{<i>a</i>}	4.84^{b}
(ng/ml)	± 0.09	± 0.11	± 0.18	± 0.15	± 0.09	± 0.01	± 0.02	± 0.08
T4	0.49^{b}	0.59 ^{<i>a</i>}	0.54^{a}	0.60 ^{<i>a</i>}	0.47^{b}	0.58^{a}	0.64^{a}	0.61 ^{<i>a</i>}
(ng/ml)	± 0.01	± 0.03	± 0.03	± 0.03	± 0.01	± 0.02	± 0.03	± 0.03

Table 4: Effects of dietary citricacid and yeast probiotic supplement on triiodothyronine (T3) and thyroxine (T4) levels of broilers

* Values are significant at p < 0.05

Table 5: Effects of dietary citricacid and yeast probiotic supplement on Lipid profile parameters of broilers

After 28 days					After 35 days				
Groups	C-gr	CA-gr	PB-gr	CO-gr	C-gr	CA-gr	PB-gr	CO-gr	
TC	184.00 ^{<i>a</i>}	164.00 ^{<i>a</i>}	$148.30^b \pm 5.5$	50167.00 ^a	140.00 ^{<i>a</i>}	100.00^{b}	85.00^{b}	90.00^{b}	
	± 4.50	± 8.50		± 2.65	± 2.04	± 8.17	± 6.12	± 4.08	
TG	120.00 ^{<i>a</i>}	113.33 ^{<i>a</i>}	$98.30^{b}\pm2.33$	3 113.00 ^a	120.80 ^{<i>a</i>}	73.10^{b}	63.00^{b}	63.20^{b}	
	± 5.60	± 2.18		± 1.0	± 2.04	± 1.22	± 1.23	±1.23	
HDL-	120.30 ^{<i>a</i>}	108.30 ^{<i>a</i>}	100.00^{a}	103.00 ^{<i>a</i>}	40.00 ^c	65.00^{a}	63.00 ^{<i>a</i>}	55.00^{b}	
С	± 6.90	± 4.88	± 7.50	± 1.20	± 1.29	± 2.06	± 1.23	± 2.39	
LDL-	39.70 ^{<i>a</i>}	33.00 ^{<i>a</i>}	28.70 ^{<i>a</i>}	41.30 ^{<i>a</i>}	76.00^{a}	20.00^{b}	9.00 ^c	22.00^{b}	
С	± 4.30	± 3.20	± 2.40	± 1.70	± 1.03	± 0.70	± 1.05	± 0.89	

* Values are significant at p < 0.05

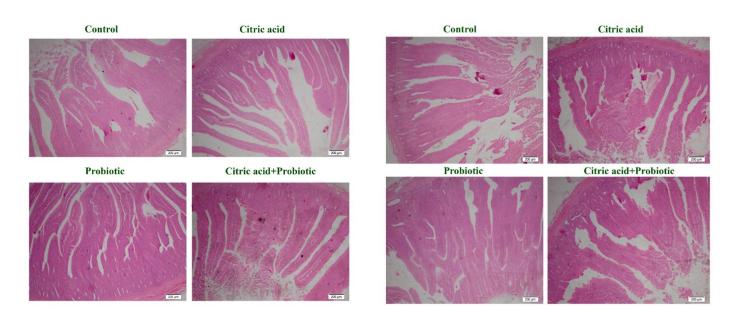


Figure 1: Duodenum at 3rd week showing elongation of villus heights in all treated groups especially probiotic fed group. H&E. Bar=200 μ m.

Figure 2: Duodenum at 5th week showing elongation of villus heights in all treated groups especially probiotic fed group. H&E. Bar=200 μ m.

ternatives. Probiotics are considered as alternative feed additives to antibiotics and microbial food supplements that have a beneficial effect on the host by improving its intestinal microbial balance [15]. To increase production or stop the spread of diseases, organic acids have been introduced to broiler chick's food or drinking water. OAs have been applied for antimicrobial, disinfecting and hygienic purposes in animal feed industry. Supplementation of a combination of probiotic and organic acids shows a significant improvement in the performance of poultry [16]. Thyroid hormones, especially T3, which have a physiological impact on all major metabolic pathways and can, lead to improved body weight. The potential impact of this study of PB and CA as feed additives on the body weight, weight of their edible organs, feed intake, lipid peroxidation (MDA), glutathione ratio (GSH), serum T3 and T4 and Histopathological examination of intestine.

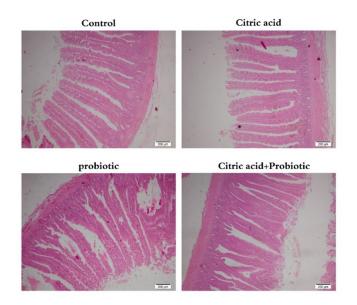


Figure 3: Duodenum at 5th week showing elongation of villus heights in all treated groups especially probiotic fed group. H&E. Bar=200 μ m.

The results of body weight were in accordance with those of [17–21], [23].

Concerning the internal organs of broilers results of the current study revealed that, for the gizzard weight, the results agree with [18], [22], [23– 27], and disagree with [19], [30]. For liver weight, the results agree with [19], [27], and disagree with [31–33]. And for the heart weight, the results were in accordance with those of [19], [34] and disagree with [27].

The improvement in body weight and edible organs weight of birds supplemented with CA may be due to improved utilization of minerals, protein proteolysis and amino acids digestibility [35]. The more powerful effect of CA could be attributed to citrate, the conjugate base of CA as one of a series of compounds involved in the physiological oxidation of acetate from fats, proteins, and carbohydrates. The acetate from these macronutrients is converted into the intracellular energy of ATP, as well as the common by products carbon dioxide, and water. This series of chemical reactions is central to nearly all metabolic reactions and is the source of two-thirds of the food-derived energy in higher organisms [36].

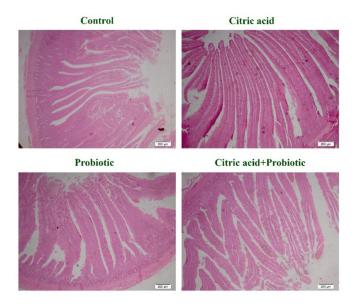


Figure 4: Jejunum at 5th week, showing pronounced elongation of villi in all supplemented groups. H&E. Bar=200 μ m.

The results related to hepatic MDA level were similar to those obtained by [37- 43] and disagree with [44].

The typical shelf life of fresh meat is only a few days due to the quick oxidation of the lipid and protein contents [45]. MDA has been recognized as the primary byproduct of lipid oxidation by reactive oxygen species (ROS), which is typically used to measure the level of lipid peroxidation in poultry flesh [46]. In this study, dietary supplementation with OA might reduce lipid peroxidation, as seen by decreased serum MDA, which could enhance the quality of the meat and increase its shelf life.

The results showing the activity of hepatic GSH are in agreement with [37], [40- 44].

The antioxidant enzyme GSH acts as the first line of defense against oxidative damage, relieving it by removing the excessively produced free radicals. It has been observed that dietary probiotics in diets were valuable to the antioxidant capacity of broilers, which is consistent with our results [45]. The peroxides end up turning out to be nonoxidative, hazardous, or more stable metabolites in the metabolic pathway of the GSH process, which involves GSH oxidation. It has been discovered that including dietary probiotics in diets may increase the body's capacity for antioxidants [47].

Concerning the concentration of the T3 serum, the results of the current study are in agreement with [35], [48-49].

Concerning the concentration of the T4 serum, the results are in agreement with [50- 53] and disagree with [19].

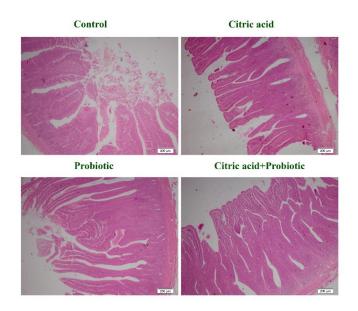


Figure 5: ileum at 3rd week showing elongated villi along with hyperplastic lymphoid follicles. H&E. Bar=200 μ m.

Similar to mammals, birds' thyroid glands generate hormones that effectively regulate the metabolism and growth of body tissues [54]. Previous investigations suggested that the indirect effects of normal thyroid hormone levels on bird

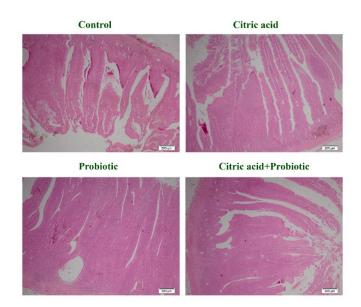


Figure 6: ileum at 5rd week showing pronounced elongated villi along with distinct hyperplastic lymphoid follicles. H&E. Bar=200 μm

growth were caused by the stimulatory effects of growth factors like insulin-like growth factor [55]. According to the results of various research, including thyme in the diet of poultry improved feed conversion and increased broiler weight [56-58]. In a similar way, numerous studies revealed that the use of probiotics increased the growth performance of poultry [59], [60].

Over 90% of the thyroid hormones produced by chicken's thyroid gland are T4 while the majority of T3 hormones are produced by the pituitary gland's conversion of T4 to T3 [61]. Probiotic use, however, significantly raised blood T4 levels. The probiotics utilized in this study appeared to have a greater impact on the creation and secretion of T4 because of their positive effects on the digestive tract enhanced digestibility, nutrient absorption, and nitrogen retention materials [62]. Previous studies showed that the thyroid gland is more successful at producing T4 than T3 when digestion and absorption of nutrients for the synthesis of thyroid hormones are optimal in the diet of poultry. [63], [64].

The findings of lipid profile parameters are in agreement with [34], [65-70].

The reduction in lipid profile parameters may be explained as the observed lower feed consumption during the period of growth and consequently lower fat intake resulted in fat depletion may also contribute in reducing blood lipid content. Moreover, the observed hyperthyroidism associated with water acidification could also explain the observed reduction in serum lipid profile [71] where thyroid hormones regulate lipid synthesis, mobilization and degradation, even though degradation is influenced more than synthesis [72]. T3, T4 favor lipolysis in adipose tissue resulting in a decrease in plasma cholesterol content and they may have an indirect effect on lipogenesis [73]. They favor lipolysis in adipose tissue resulting in a decrease in plasma cholesterol content and they may have an indirect effect on lipogenesis [74]. They also can stimulate liver gluconeogenesis through gene activation [75] and [76].

5. Conclusion

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By incorporating CA and PB to the broilers' water, researchers were able to influence the health of their liver, intestinal mucosa, and thyroid hormones (T3 and T4), were improved and promoted. That illustrates how CA and PB have a stronger impact on balancing body internal homeostasis.

References

- [1] E. U. Ahiwe, A. A. Omede, M. B. Abdallh, P. Iji, Managing dietary energy intake by broiler chickens to reduce production costs and improve product quality. Animal husbandry and nutrition (2018).
- [2] H. Qaqaya, G. Lipimile, ., The effects of anti-competitive business practices on developing countries and their development prospects. United Nations Conference on Trade and Development (2008). United Nations. No. UNCT AD/DITC/CLP/2008/2. (2008).
- [3] M. Gunal, G. Yayli, O. Kaya, N. Karahan, O. Sulak, The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broilers, Int. J. Poult. Sci 5 (2) (2006) 149–155.
- [4] M. Z. Islam, Z. H. Khandaker, S. D. Chowdhury, K. M. S. Islam, Effect of citric acid and acetic acid on the performance of broilers, J. Bangladesh Agril. Univ 6 (2) (2008) 315–335.
- [5] P. Gao, C. Ma, Z. Sun, L. Wang, S. Huang, X. Su, J. Xu, H. Zhang, Feed-additive probiotics accelerate yet antibiotics delay intestinal microbiota maturation in broiler chicken. Microbiome (2017).

- [6] Y. Mehdi, M. P. Létourneau-Montminy, M. L. Gaucher, Y. Chorfi, G. Suresh, . R. T, S. Godbout, Use of antibiotics in broiler production: Global impacts and alternatives, Animal nutrition 4 (2) (2018) 170–178.
- [7] National research center (NRC) (1994).
- [8] Z. Açıkgöz, H. Bayraktar, Ö. Altan, Effects of formic acid administration in the drinking water on performance, intestinal microflora and carcass contamination in male broilers under high ambient temperature. Asian-Aust, J. Anim. Sci 24 (1) (2011) 96–102.
- [9] A. Galler, G. Gelbrich, J. Kratzsch, N. Noack, T. Kapellen, W. Kiess, Elevated serum levels of adiponectin in children, adolescents and young adults with type 1 diabetes and the impact of age, gender, body mass index and metabolic control: a longitudinal study, Eu. J. Endocrinol 157 (4) (2007) 481–489.
- [10] M. F. Gridley, Manual of Histologic and Special Staining Technique, 1960, PP 28-29 and 82-83, Mc-Graw-Hill Book Company, New York, NY.
- [11] W. S. Rasband, J. U. S. Image, National Institutes of Health (1997).
- [12] P. Iji, A. Saki, D. Tivey, Intestinal development and body growth of broiler chicks on diets supplemented with non-starch polysaccharides, Anim. Feed Sci. Technol (89) (2001) 175–188.
- [13] S. J. Coakes, L. Steed, SPSS: Analysis without anguish using SPSS version 14.0 for Windows (2009).
- [14] A. E. Permanasari, D. R. A. Rambli, P. D. D. Dominic, Forecasting method selection using ANOVA and Duncan multiple range tests on time series dataset, International Symposium on Information Technology (2) (2010) 941–945.
- [15] R. U. Khan, N. S, The applications of probiotics in poultry production, Poult. Sci. J 69 (3) (2013) 621–632.
- [16] A. W. Youssef, H. M. A. Hassan, H. M. Ali, M. A. Mohamed, Effect of probiotics, prebiotics and organic acids on layer performance and egg quality, Asian J Poult Sci 7 (2) (2013) 65–74.
- [17] E. R. Kamel, L. S. Mohamed, Effect of dietary supplementation of probiotics, prebiotics, synbiotics, organic acids and enzymes on productive and economic efficiency of broiler chicks, Alex. J. Vet. Sci 50 (1) (2016) 8– 17.
- [18] A. M. Shareef, A. S. A. Dabbagh, Effect of probiotic (Saccharomyces cerevisiae) on performance of broiler chicks. Iraqi, J. vet. Sci 23 (1) (2009) 23–29.
- [19] T. Aluwong, F. Hassan, T. Dzenda, M. Kawu, J. Ayo, Effect of different levels of supplemental yeast on body weight, thyroid hormone metabolism and lipid profile of broiler chickens, J. Vet. Med. Sci 75 (3) (2012) 291–298.
- [20] G. Srinivas, B. Swathi, S. Raju, R. Tungani, Influence of dietary supplementation of organic acids, probiotics and their combinations on growth, carcass traits and serum parameters in broiler chickens, Indian. J. Anim. Nutr 35 (2) (2018) 201–205.
- [21] A. Alcicek, M. Bozkurt, M. Cabuk, The effect of a mixture

of herbal essential oils, an organic acid or a probiotic on broiler performance. South African, J. Anim. Sci 34 (4) (2004) 217–222.

- [22] A. Masud, M. D. S. Ali, M. U. Ahammad, Combined use of dietary probiotic and acidifier for the production of antibiotic free broiler, Res. Agric. Livest. Fish 3 (1) (2016) 127–137.
- [23] A. W. Zhang, B. D. Lee, S. K. Lee, K. W. Lee, G. H. An, K. B. Song, C. H. Lee, Effects of yeast (Saccharomyces cerevisiae) cell components on growth performance, meat quality and ileal mucosa development of broiler chicks, Poult. Sci 84 (2005) 1015–1021.
- [24] E. Marin-Flamand, A. Vazquez-Duran, A. Mendez-Albores, Effect of organic acid blens in drinking water on growth performance, blood constituents and immune response of broiler chickens, J. Poult. Sci 51 (2014) 144– 150.
- [25] I. M. I. Youssef, A. S. Mostafa, M. A. Abdel-Wahab, Effect of dietary inclusion of probiotics and organic acids on performance, intestinal microbiology, serum biochemistry and carcass traits of broiler chickens, J. World Poult. Res 7 (2) (2017) 57–71.
- [26] H. E. E. Malik, R. H. H. Hafzalla, M. M. O. Elhassan, B. M. Dousa, A. M. Ali, K. M. Elamin, Effect of probiotics and acidifiers on carcass yield, internal organs, cuts and meat to bone ratio of broiler chicken, J. Agric. Vet. Sci 9 (2) (2016) 18–23.
- [27] W. A. Awad, K. Ghareeb, S. Abdel-Raheem, J. Bohm, Effects of dietary inclusion of probiotic and synbiotic on growth performance, organ weights and intestinal histomorphology of broiler chickens, Poult. Sci 88 (2009) 49–55.
- [28] T. Aluwong, M. Kawu, M. Raji, T. Dzenda, F. Govwang, V. Sinkalu, J. Ayo, Effect of yeast probiotic on growth, antioxidant enzyme activities and malondialdehyde concentration of broiler chickens, J. Antioxidants 2 (2013) 226–239.
- [29] J. H. Park, I. H. Kim, Supplemental effect of probiotic Bacillus subtilis B2A on productivity, organ weight, intestinal Salmonella microflora, and breast meat quality of growing broiler chicks, Poult. Sci 93 (8) (2014) 2054– 2059.
- [30] K. Celik, K. Ugur, A. H. M. E. T. Uzatici, Effect of supplementing broilers diets with organic acid and whole grain, Asian J. Anim. Vet. Adv 3 (5) (2008) 328–333.
- [31] M. Bozkurt, K. Küçükyılmaz, A. U. Çatlı, M. Çınar, The effect of single or combined dietary supplementation of prebiotics, organic acid and probiotics on performance and slaughter characteristics of broilers, South African J. Anim. Sci 39 (3) (2009) 197–205.
- [32] M. A. Mohamed, E. F. El-Daly, A. El-Azeem, N. A. Youssef, A. W. Hassan, H. M. A, Growth performance and histological changes in ileum and immune related organs of broilers fed organic acids or antibiotic growth promoter, Int. J. Poult. Sci 13 (10) (2014) 602–610.
- [33] M. Denli, F. Okan, K. Celik, Effect of dietary probiotic,

organic acid and antibiotic supplementation to diets on broiler performance and carcass yield, Pakistan J. Nut 2 (2) (2003) 89–91.

- [34] A. A. H. Tollba, S. A. M. Shabaan, M. A. A. Abdel-Mageed, Effects of using aromatic herbal extract and blended with organic acids on productive and physiological performance of poultry 2-the growth during cold winter stress, Egy. Poult. Sci. J 30 (1) (2010) 229–248.
- [35] S. D. Boling, M. W. Douglas, J. L. Snow, C. M. Parsons, D. H. Baker, Citric acid does not improve phosphorus utilization in laying hens fed a corn-soybean meal diet, Poult. Sci 79 (9) (2000) 1335–1337.
- [36] A. El-Hakim, S. A. Cherian, G. Ali, M. N, Use of organic acid, herbs and their combination to improve the utilization of commercial low protein broiler diets. Inter, J. Poult. Sci 8 (1) (2009) 14–20.
- [37] T. Ebeid, I. Al-Homidan, M. Fathi, R. Al-Jamaan, M. Mostafa, O. Abou-Emera, A. El-Razik, M. Alkhalaf, A, Impact of probiotics and/or organic acids supplementation on growth performance, microbiota, antioxidative status, and immune response of broilers, Italian J. Animal. Sci 20 (1) (2021) 2263–2273.
- [38] P. Baghban-Kanani, B. Hosseintabar-Ghasemabad, S. Azimi-Youvalari, A. Seidavi, M. Ragni, V. Laudadio, V. Tufarelli, Effects of using Artemisia annua leaves, probiotic blend, and organic acids on performance, egg quality, blood biochemistry, and antioxidant status of laying hens, J. Poult. Sci 56 (2) (2019) 120–127.
- [39] S. A. Kazemi, H. Ahmadi, K. Torshizi, M. A, Evaluating two multistrain probiotics on growth performance, intestinal morphology, lipid oxidation and ileal microflora in chickens, J. Anim. Phys. Anim. Nut 103 (5) (2019) 1399–1407.
- [40] T. He, S. Long, S. Mahfuz, D. Wu, X. Wang, X. Wei, X. Piao, Effects of probiotics as antibiotics substitutes on growth performance, serum biochemical parameters, intestinal morphology, and barrier function of broilers, Animals 9 (11) (2019) 985–985.
- [41] Y. Wu, B. Wang, Z. Zeng, R. Liu, L. Tang, L. Gong, W. Li, Effects of probiotics Lactobacillus plantarum 16 and Paenibacillus polymyxa 10 on intestinal barrier function, antioxidative capacity, apoptosis, immune response, and biochemical parameters in broilers, Poult. Sci 98 (10) (2019) 5028–5039.
- [42] L. Gong, B. Wang, X. Mei, H. Xu, Y. Qin, W. Li, Y. Zhou, Effects of three probiotic Bacillus on growth performance, digestive enzyme activities, antioxidative capacity, serum immunity, and biochemical parameters in broilers, Anim. Sci. J 89 (11) (2018) 1561–1571.
- [43] K. Bai, C. Feng, L. Jiang, L. Zhang, J. Zhang, L. Zhang, T. Wang, Dietary effects of Bacillus subtilis fmbj on growth performance, small intestinal morphology, and its antioxidant capacity of broilers, Poult. Sci 97 (7) (2018) 2312–2321.
- [44] A. S. Salah, O. A. Ahmed-Farid, M. S. El-Tarabany, Carcass yields, muscle amino acid and fatty acid profiles,

and antioxidant indices of broilers supplemented with synbiotic and/or organic acids, J. Anim. Phys. Anim. Nut 103 (1) (2019) 41–52.

- [45] S. T. Ahmed, M. M. Islam, A. B. M. R. Bostami, H. S. Mun, Y. J. Kim, C. J. Yang, Meat composition, fatty acid profile and oxidative stability of meat from broilers supplemented with pomegranate (Punica granatum L.) byproducts, Food Chemistry 188 (2015) 481–488.
- [46] A. M. Salih, D. M. Smith, J. F. Price, L. E. Dawson, Modified extraction 2-thiobarbituric acid method for measuring lipid oxidation in poultry, Poult. Sci 66 (1987) 1483–1488.
- [47] L. Ragione, R. M. Woodward, M. J. Competitive exclusion by Bacillus subtilis spores of Salmonella enterica serotype Enteritidis and Clostridium perfringens in young chickens, Vet. Microbiology 94 (3) (2003) 245– 256.
- [48] M. H. Tabidi, A. M. Mukhtar, H. I. Mohammed, Effects of probiotic and antibiotic on performance and growth attributes of broiler chicks, Global Journal of Medicinal Plant Research 1 (1) (2013) 136–142.
- [49] D. Chotinsky, R. Mihaylov, Effect of probiotics and avotan on the level of thyroid hormones in the blood plasma of broiler chickens, Bulgarian J. Agric. Sci 19 (4) (2013) 817–821.
- [50] S. S. Elnesr, A. R. H. Abdel-Razik, H. A. Elwan, Impact of humate substances and Bacillus subtilis PB6 on thyroid activity and histomorphometry, iron profile and blood haematology of quail, J. Anim. Phys. Anim. Nut 106 (1) (2022) 110–117.
- [51] A. M. Motlagh, V. Babapour, Z. A. Pirsaraei, N. Sheikhi, Effect of thyme (Zataria Multiflora) extract and probiotic (broilact) feeding on blood thyroid hormones concentration and growth hormone gene expression of liver in broiler chickens, Ind. J. Fund. Applied Life Sci. ISSN 5 (1) (2015) 2231–6345.
- [52] A. E. Khalifa, M. Abdelrahman, K. Gahreeb, Effect of probiotic on growth performance, carcass traits, and clinical health parameters of broilers reared under heat stress in upper Egypt, SVU-Int. J. Vet. Sci 2 (2) (2019) 27– 44.
- [53] H. M. A. Abdelrazek, S. M. M. Abuzead, S. A. Ali, H. M. A. El-Genaidy, S. A. Abdel-Hafez, Effect of citric and acetic acid water acidification on broiler's performance with respect to thyroid hormones levels, Adv. Anim. Vet. Sci 4 (5) (2016) 271–278.
- [54] M. A. Lazar, Thyroid hormone receptors: Multiple forms, multiple possibilities, Endocrine Reviews 14 (2) (1993) 184–193.
- [55] F. A. Mcnabb, D. B. King, Thyroid hormone effects on growth, development, and metabolism, Endoc. Gr. Dev. Met. Vert 10 (1993) 873–885.
- [56] R. Abdulkarimi, A. M. Aghazadeh, M. Daneshyar, Growth performance and some carcass characteristics in broiler chickens supplemented with Thymus extract (Thymus vulgaris) in drinking water, J. American Sci

7 (11) (2011) 400–405.

- [57] G. A. Al-Kassie, Influence of two plant extracts derived from thyme and cinnamon on broiler performance, Pakistan Vet. J 29 (4) (2009) 169–173.
- [58] K. W. Lee, H. Everts, A. C. Beynen, Essential oils in broiler nutrition, Int. J. Poult. Sci 3 (12) (2004) 738–752.
- [59] L. I. Peric, N. I. Milosevic, D. R. Zikic, S. I. Bjedov, D. R. Cvetkovic, S. I. Markov, M. Mohnl, T. Steiner, Effects of probiotic and phytogenic products on performance, gut morphology and cecal microflora of broiler chickens, Archives Animal Breeding 53 (3) (2010) 350–359.
- [60] T. Zhi-Gang, M. Naeem, W. Chao, W. Tian, Z. Yan-Min, Effect of dietary probiotics supplementation with different nutrient density on growth performance, nutrient retention and digestive enzyme activities in broilers, JAPS: J. Anim. and Plant Sci 24 (5) (2014) 1309–1315.
- [61] F. M. Mcnabb, Thyroids. In Sturkie PD. Avian Physiology (1999) 461–469.
- [62] C. Schneitz, T. Kiiskinen, V. Toivonen, M. Nasi, Effect of broilact on the physicochemical conditions and nutrient digestibility in the gastrointestinal tract of broilers, Poult. Sci 77 (3) (1998) 426–432.
- [63] F. A. Mcnabb, J. R. Blackman, J. A. Cherry, The effects of different maternal dietary iodine concentrations on Japanese quail, Domestic Animal Endocrinology 2 (1) (1985) 25–34.
- [64] M. J. Mcnichols, F. A. Mcnabb, Comparative thyroid function in adult Japanese quail and Ring doves: Influence of dietary iodine availability, J. Exp. Zoology 244 (2) (1987) 263–268.
- [65] M. B. Moghadam, M. Rezaei, F. Niknafs, H. Sayyahzadeh, Effect of combined probiotic and organic acid on some blood parameters and immune system of broiler chicks, Proceedings 2nd Mediterranean Summit of World Poult. Sci (2006) 4–7.
- [66] A. M. Kamal, N. M. Ragaa, Effect of dietary supplementation of organic acids on performance and serum biochemistry of broiler chicken, Nat. Sci 12 (2) (2014) 38–45.
- [67] M. I. Haque, N. Ahmed, M. A. Miah, Comparative analysis of body weight and serum biochemistry in broilers supplemented with some selected probiotics and antibiotic growth promoters, J. Adv. Vet. Anim. Res 4 (3) (2017) 288–294.
- [68] H. Wang, X. Ni, X. Qing, D. Zeng, M. Luo, L. Liu, G. Li, K. Pan, B. Jing, Live probiotic lactobacillus johnsonii BS15 promotes growth performance and lowers fat deposition by improving lipid metabolism, intestinal development and gut microflora in broilers, Fron. Microbiol 8 (2017) 1073–1073.
- [69] A. M. El-Baz, N. S. Ibrahim, A. M. Shehata, N. G. Mohamed, E. D. Abdel-Moneim, Impact of multi-strain probiotic, citric acid, garlic powder or their combinations on performance, ileal histomorphometry, microbial enumeration and humoral immunity of broiler chickens, Trop. Anim. health Prod 53 (1) (2021).
- [70] A. Gheorghe, M. Habeanu, G. Ciurescu, N. A. Lefter,

M. Ropota, I. Custura, M. Tudorache, Effects of dietary mixture enriched in polyunsaturated fatty acids and probiotic on performance, biochemical response, breast meat fatty acids and lipid indices in broiler chickens, Agriculture 12 (8) (2022) 1120–1120.

- [71] S. F. El-Afifi, N. M. El-Mednay, M. Attia, Effect of citric acid supplementation in broiler diets on performance and intestinal microflora, Egypt. Poult. Sci 21 (2001) 491–505.
- [72] A. El-Fattah, S. A. El-Sanhoury, M. H. El-Mednay, N. M. Abdel-Azeem, F, Thyroid activity, some blood constituents, organs morphology and performance of broiler chicks fed supplemental organic acids, Int. J. Poult. Sci 7 (3) (2008) 215–222.
- [73] E. Pucci, L. Chiovato, A. Pinchera, Thyroid and lipid metabolism. Inter, J. Obes. Rel. Met Dis 24 (2) (2000) 109–112.
- [74] A. A. Saki, S. M. Eftekhari, P. Zamani, H. Aliarabi, M. Abbasinezhad, Effects of an organic acid mixture and methionine supplements on intestinal morphology, protein and nucleic acids content, microbial population and performance of broiler chickens, Anim. Pro. Sci 51 (11) (2011) 1025–1033.
- [75] B. M. Eshkhatkhah, S. Sadaghian, S. Eshkhatkhah, K. Pourabi, Najafian, Relationship between the blood thyroid hormones and lipid profile in Moghani sheep; influence of age and sex, Comp. Clin. Path 19 (2010) 15– 20.
- [76] P. M. Yen, X. Feng, F. Flamant, Y. Chen, R. L. Walker, R. E. Weiss, O. Chassande, J. Samarut, S. Refetoff, P. S. Meltzer, Effects of ligand and thyroid hormone receptor isoforms on hepatic gene expression profiles of thyroid hormone receptor knockout mice, EMBO Rep 4 (6) (2003) 581– 587.