

**A CONTRIBUTION ON THE EPIDEMIOLOGY OF SOME COMMON  
PARASITES INFESTING CULTURED SEA BASS  
(*DICENTRARCHUS LABRAX*) IN EGYPT**

By

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**ABSTRACT**

This study was carried out to spotlight on the most common parasitic infestation in cultured Sea bass, *Dicentrarchus labrax*. A total number of 120 *Dicentrarchus labrax* were collected from two private marine fish farms in Damietta governorate from September 2016 to August 2017. The prevalence, seasonal variation, as well as the histopathological changes in infested tissues were estimated. The parasitological examination revealed four genera of protozoa including (*Trichodina*, *Epistylis*, *Amyloodinium* and *Microsporidia*), two genera of monogenean (*Diplectanum* and *Pseudohaliotrema* sp.), one genus of crustacean (*Caligus minimus*), unidentified digenetic encysted metacercariae, two genera of adult digenetic trematodes (*Timoniella* and *Bucephalus*) and one genus of Nematode (*Conteracecum*) with total prevalence 84.2%. The seasonal prevalence was different from parasite to another. Histopathological examination revealed different pathological changes in the affected tissues as a reaction to parasitic infestation.

**Keywords:**

*Dicentrarchus labrax*, cultured, parasites, seasonal variation, Histopathology.

**INTRODUCTION**

Fisheries and aquaculture are highly important sources of nutrition, income and livelihoods for millions of people all over the world (FAO, 2016). Aquaculture in Egypt is a promising field with a great capability for expansion and development (Shaalán *et al.*, 2018). Marine aquaculture represents about 14.5 % of the total aquaculture in Egypt. About 80 % of marine aquaculture production in Egypt is located in Damietta Governorate, while the remaining

percent located in the coastal provinces namely, Port Said, Suez, and Alexandria (Wally, 2016). The European Sea bass, *Dicentrarchus labrax*, is the most important commercial fish cultured in Southern Europe, especially in the North Atlantic and the Mediterranean area (FAO, 2014). In Egypt, Sea bass is one of The major marine fish species that are cultured in semi intensive brackish-water ponds, often in polyculture with mullet, mainly in the Diba Triangle Zone located between Damietta and Port Said (Rothuis et al., 2013). Semi-intensive and intensive fish farming systems increase the importance of parasites as disease-causing agents due to their ability to evolve rapidly the most serious limiting factors in aquaculture because of the increased density of cultured fish populations in restricted bodies of water and the ease by which pathogens can be transmitted from one fish to another (Antonelli et al., 2016). Parasites are the most commonly found pathogen with a prevalence rate up to 80% of the total infections in fish on farms (Shaheen et al., 2013). The common parasites infested marine fishes include monogenetic and digenetic trematodes, protozoa namely, *Ichthyophthirius multifiliis*, *Trichodina spp.* and *Chilodonella* as well as the encysted metacercariae (Aly, 2013). The present work was done on cultured Sea bass, (*Dicentrarchus labrax*) from tow private fish farms in Damietta governorate to identify the most common parasites infesting cultured Sea bass, (*Dicentrarchus labrax*), study their prevalence, seasonal dynamics as well as the histopathological changes induced by detected parasites.

## MATERIAL AND METHODS

### **Fish:**

A total number of 120 Sea bass (*Dicentrarchus labrax*) with average body weight of  $150 \pm 30$  g were collected alive from tow private fish farms in Diba triangle in Port Said- Damietta way from September 2016 - August 2017. The collected fish were transported in large tanks filled with water of the same sources and supplied with battery air pumps to Fish Diseases Department Lab., at Animal Health Research Institute, Dokki, and Giza, Egypt. The clinical signs, postmortem changes were recorded and the parasitological examination was applied according to Noga (2010).

### **Clinical and post-mortem examination:**

This examination was applied on spot in the fish farms from which the naturally infested fish were collected. External examination of fish including, the clinical abnormalities and post-mortem changes were recorded according to Noga (2010).

**Parasitological examination and identification:**

The *Dicentrarchus labrax* were examined externally with naked eye and with aid of magnifying glass for detection of parasitism according to Noga (2010). Mucous smears were immediately prepared from the skin and fins with the aid of microscopic slides and covers. Then they were examined under stereomicroscope at X 10, 40 and 100 magnification for investigation of external parasites. Fish were euthanized and gills were carefully removed and placed in separate petri dish containing sterilized marine water to remove any excess gill mucus, Afterwards they were examined for parasitic infection under stereomicroscope. These internal organs were examined by naked eye for the presence of encysted metacercariae or worm and small pieces of each was taken and mixed with few drops of saline, compressed between two glass slides and examined under the microscope to detect the presence of encysted metacercariae. Detected parasites were collected, examined, fixed and stained and identified according to Lucky (1977) ; Paperna and Laurencin (1979); Lom (1995) and Paperna (1996).

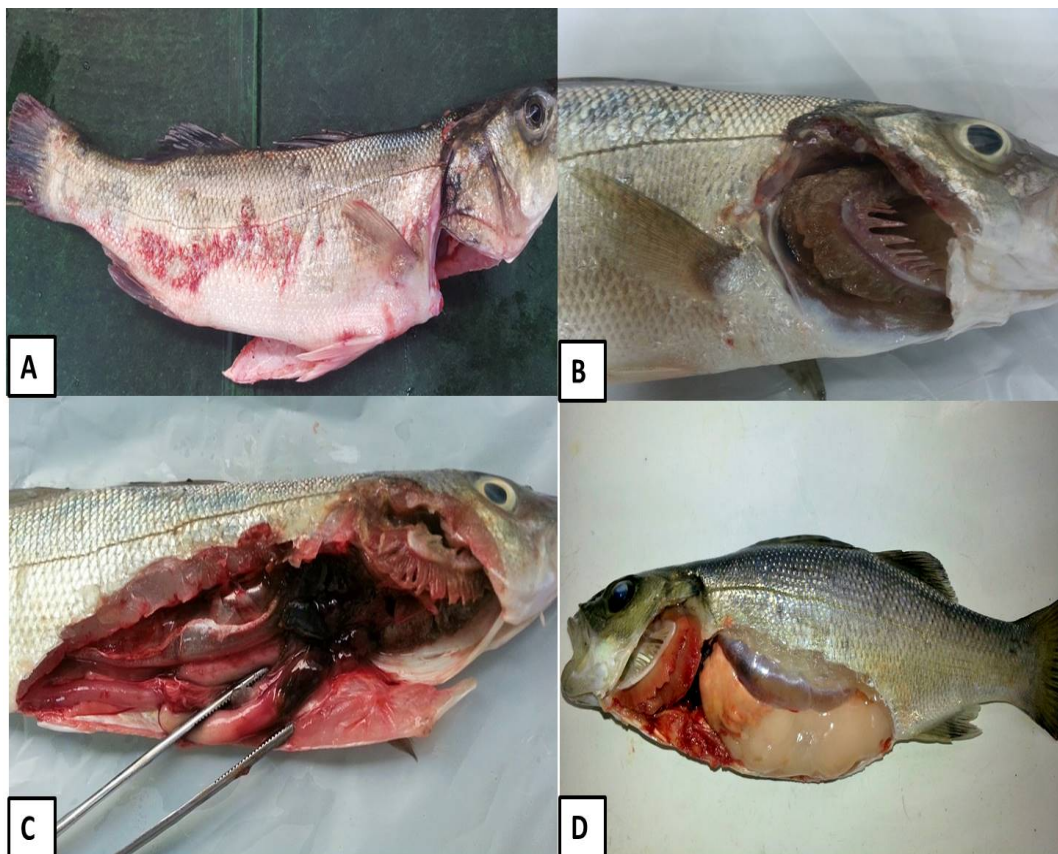
**Histopathological examination:**

Tissue specimens from different organs were fixed in 10% neutral buffered formalin. The fixed samples were washed in tap water overnight then exposed to ascending concentrations of ethanol (70, 80, 90 and 100%), cleared in xylene and embedded in paraffin. Tissue slides of 5µm thick sections were prepared and stained by hematoxylin and eosin (H&E). The histopathological preparation was performed according to Bancroft *et al.* (1996)

**RESULTS**

**Clinical and post-mortem examinations:**

The clinical picture of infested fish in the fish farms was restlessness, gasping of air, rubbing body against hard objects, swam near the surface of the water with increased breathing frequency, stretched gills covers, loss of appetite and increase in mortality rate. Fish infested with Protozoa, monogenea and crustacean showed haemorrhagic skin and fins, areas of detachment of scales and excessive mucous secretion on the body and gills. In case of internal helminthes and digenetic-encysted metacercaria, infested fishes appear emaciated and off food. The post mortem examination of examined fishes showed paleness of gills in some infested fishes. The recorded clinical signs and postmortem changes demonstrated in plate (1).



**plate (1):** A- *Dicentrarchus labrax* infested with mixed infection with *Trichodina heterodontata* and *Caligus minimus* showing haemorrhagic skin with detachment of scales.

B-*Dicentrarchus labrax* infested with a mixed infection monogenea and *Trichodina heterodontata* showing erosion of gill filaments.

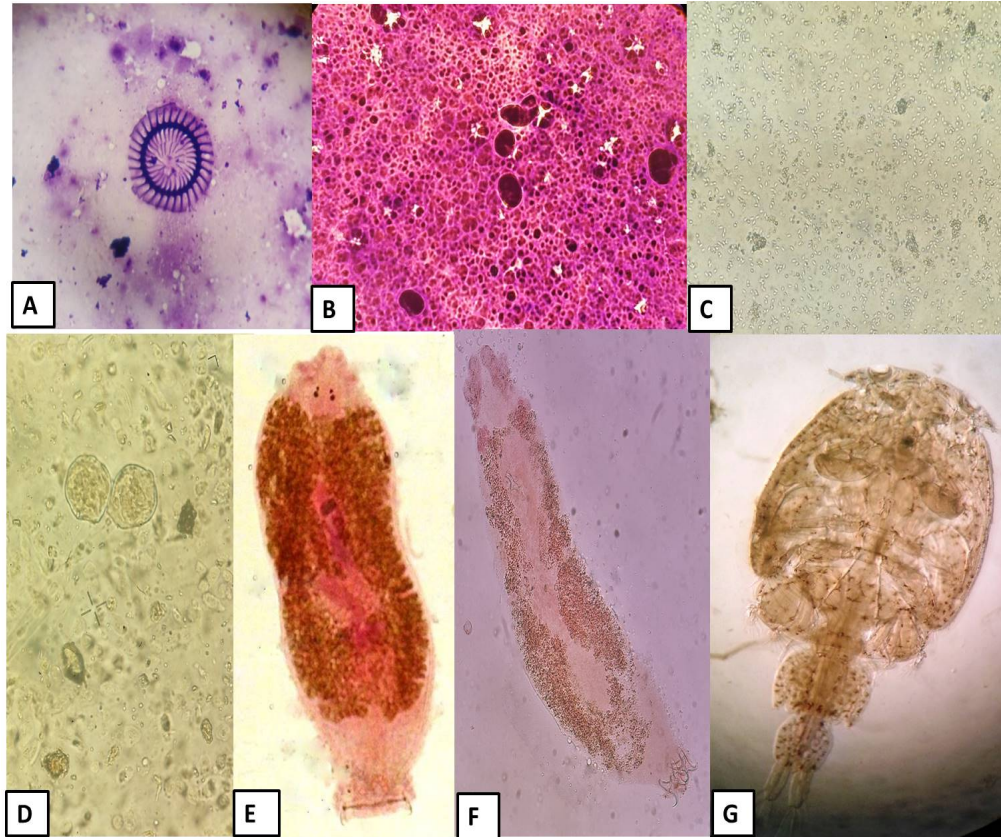
C -*Dicentrarchus labrax* infested with *Timoniella sp.* and *Bucephalus minimus* showing concretion in the gastro-intestinal tract and internal organs.

D-*Dicentrarchus labrax* infested with mixed infestation of *Amyloodinium ocellatum* and unidentified encysted metacercariae in internal organs showing pale and friable liver.

### **Parasitological examination:**

The parasitological examination revealed four genera of protozoa including (*Trichodina*, *Epistylis*, *Amyloodinium* and *Microsporidia*), two genera of monogenean (*Diplectanum* and *Pseudohaliotrema sp.*), one genus crustacean (*Caligus minimus*), unidentified digenetic encysted metacercariae, two genera of adult digenetic trematodes (*Timoniella* and *Bucephalus*) and one genus of Nematode (*Contracecum*) showed in plate (2,3).

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**Plate (2):** A- *Trichodina heterodentata* isolated from gills of *Dicentrarchus labrax* stained with Giemsa stain (X1000).

B- *Amyloodinium ocellatum* detected from *Dicentrarchus labrax* gills stained with Giemsa stain (X400).

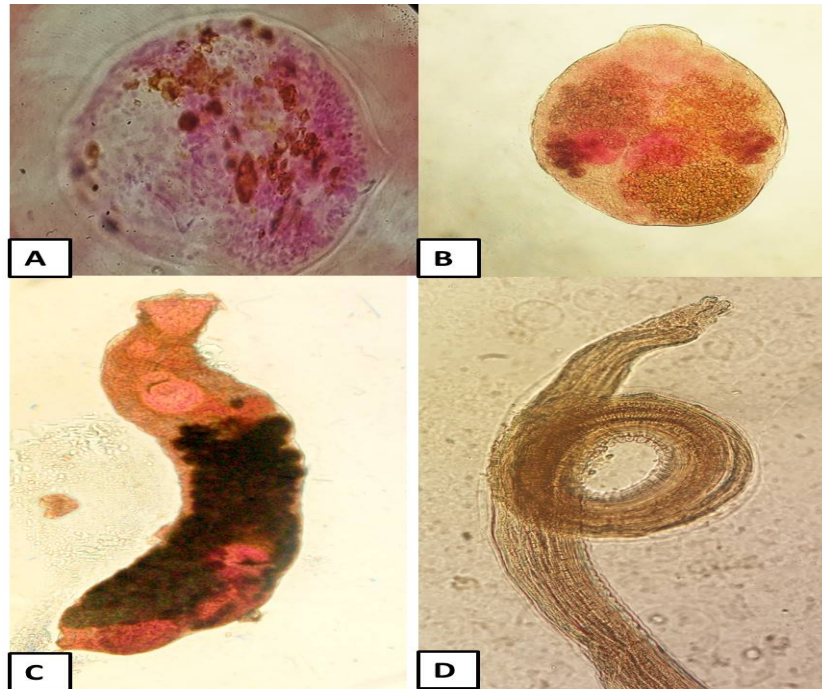
C- *Microsporidia sp.* isolated from gills of *Dicentrarchus labrax* slide stained with Giemsa stain (X400).

D - *Epistylis sp.* wet preparation isolated from *Mugil cephalus* (X400).

E- *Diplectanum sp.* whole parasite isolated from *Dicentrarchus labrax* (X100).

F- *Pseudohaliotrema sp.* whole parasite isolated from gills o of *Dicentrarchus labrax* (X100).

G- Male *Caligus minimus* whole parasite isolated from skin of *Dicentrarchus labrax* (X50).



**Plate (3):** A- Unidentified encysted metacercaria detected from gills of *Dicentrarchus labrax* stained with acid carmine (X400).

B- Adult of *Bucephalus minimus* detected from intestine of *Dicentrarchus labrax* stained with acid carmine (X400).

C- Adult *Timoniella sp.* detected from intestine of *Dicentrarchus labrax* stained with acid carmine (X100).

D- *Contracaecum sp* larvae detected from intestine of *Dicentrarchus labrax* mounted in glycerin alcohol (X100).

### **Prevalence of parasitic infestation:**

In this study, 101 out of examined 120 *D. labrax* (84.2%) were infested with parasites.

The results of prevalence of parasitic infestation were illustrated in (Table 1)

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**Table (1):** prevalence of parasites detected from *Dicentrarchus labrax* from farm I and II.

fish parasites	farm I (n=60)		farm II (n=60)		Total no of examined fish (n=120)	
	No. of infested fish	%	No. of infested fish	%	No. of infested fish	%
<i>Trichodina heterodentata</i>	17	28.3	17	28.3	34	28.3
<i>Epistylis</i>	8	13.3	9	15.0	17	14.2
<i>Amyloodinium ocellatum</i>	3	5.0	21	35.0	24	20.0
<i>Microsporidia</i>	3	5.0	7	11.7	10	8.3
<i>Diplectanum</i>	9	15.0	6	10.0	15	12.5
<i>Pseudohaliotrema</i>	11	18.3	0	0.0	11	9.2
<i>Caligus minimus</i>	10	16.7	18	30.0	18	15.0
<i>unidentified encysted metacercariae</i>	23	38.3	17	28.3	40	33.3
<i>Timoniella sp.</i>	13	21.7	8	13.3	21	17.5
<i>Bucephalus minimus</i>	12	20.0	9	15.0	21	17.5
<i>Conteracecum larvae</i>	3	5.0	2	3.3	5	4.2

\* n= Number of examined fish

**3.3 Seasonal prevalence of parasites:**

Seasonal variation study of all detected parasite were shown in (Table 2).

**Table (2):** Seasonal prevalence of parasites recovered from examined *Dicentrarchus labrax*

parasites \ season		Autumn		winter		spring		summer	
		Farm I (n=15)	Farm II (n=15)	Farm I (n=15)	Farm II (n=15)	Farm I (n=15)	Farm II (n=15)	Farm I (n=15)	Farm II (n=15)
External protozoa	<i>T. heterodentata</i>	53.3	46.7	40	26.7	13.3	20	6.7	20
	<i>Epistylis</i>	33.3	26.7	13.3	20	6.7	6.7	0	6.7
	<i>A. ocellatum</i>	20	53.3	0	26.7	0	40	0	20
	<i>Microsporidia</i>	13.3	20	0	13.3	0	0	6.7	13.3
Monogenean	<i>Diplectanum</i>	26.7	20	13.33	6.7	20	13.3	0	0
	<i>Pseudohaliotrema</i>	33.3	0	13.3	0	26.7	0	0	0
Crustacea	<i>C. minimus</i>	6.7	6.7	0	0	13.3	13.3	46.7	33.3
Helminthes	EMC	40	26.7	33.3	20	26.7	20	53.3	46.7
	<i>Timoniella sp.</i>	26.7	20	40	20	13.3	13.3	6.7	0
	<i>B. minimus</i>	40	33.3	26.7	20	13.3	6.7	0	0
	<i>Conteracecum larvae</i>	20	13.3	0	0	0	0	0	0

**The Histopathological findings:**

**The results of histopathological examination for different organs of naturally infested *D. labrax* revealed the following:**

1- Gills of infested fish with *Trichodina* showing different sections of *Trichodina* between gills filaments while gills with Monogenic infestation showed monogenea attached into groove of fused gill lamellae. The most predominant changes of gills of examined fish were, lifting of respiratory epithelia, proliferation of basal undifferentiated cells, and partial and/or complete fusion of gill lamellae.

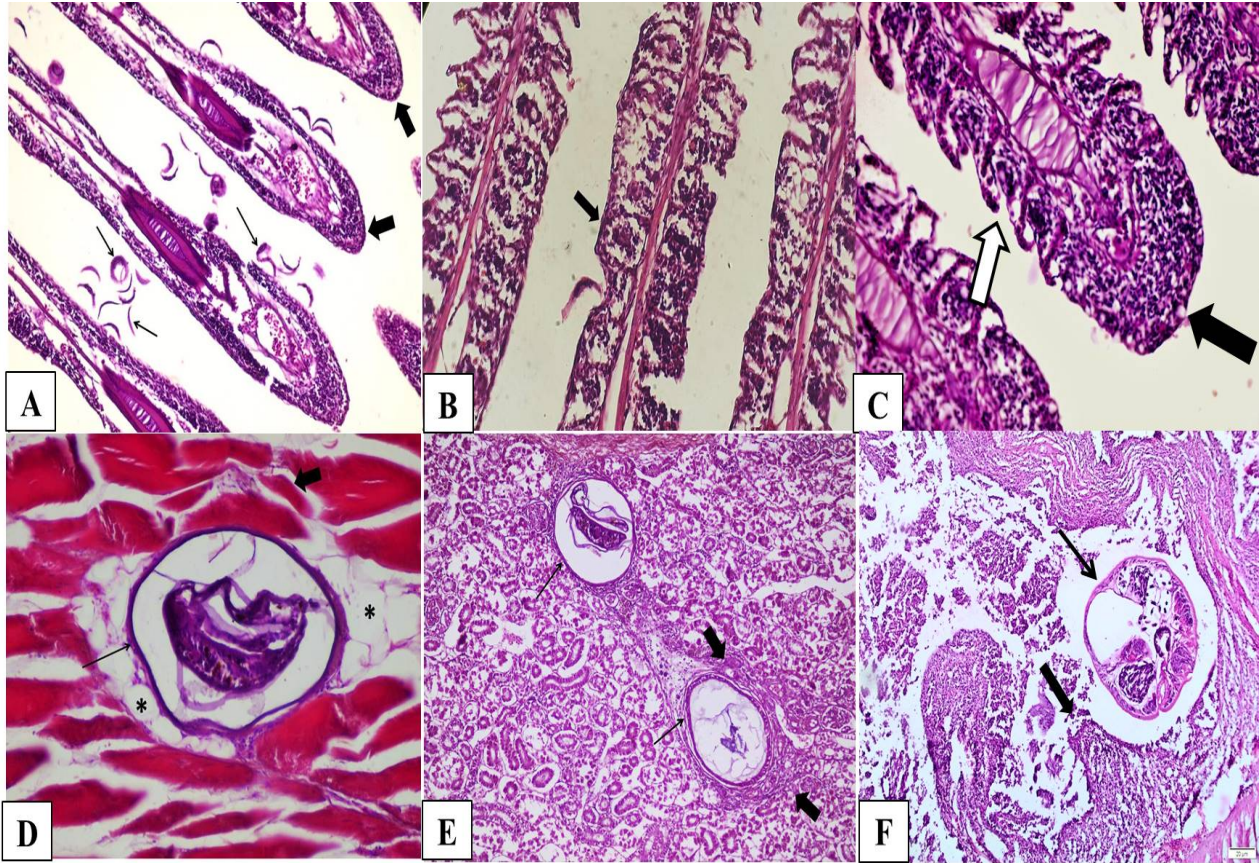
2- Muscle of infested fish with digenetic encysted metacercaria revealed cyst encountered by interstitial edema, some adjacent muscle bundles appeared atrophied with small caliber size.

3- Kidneys showing cystic formation of digenetic-encysted metacercaria within renal tissue, the surrounding tissues showed features of pressure atrophy.

4- Intestine demonstrated variable response to parasites including: massive sub-mucosal inflammatory cells infiltration, proliferation of intestinal villi and some intestinal villi appeared thick and short. Cross sections of parasite were detected in the Intestinal lumen. The results of histopathology showed in Micrograph plate (4).



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**Plate (4):** A- Sagittal section in the gill filaments of *Dicentrarchus labrax* demonstrating heavy parasitic infection with criteria of *Trichodina heterodontata*. Gill lamellae suffer from lifting of respiratory epithelium (thick white arrow) with partial fusion of secondary lamellae (thick black arrow). Gill lamellar tips were exhibiting complete fusion. **H&E X 100.**

B- Sagittal section in the gill filaments' of *Dicentrarchus labrax* revealing attachment of *Diplectanum sp.* to a groove within complete fusion lamellae (thin arrow). **H&E X 100.**

C- Gill lamellae of *Dicentrarchus labrax* exhibiting partial (thick white arrow) and complete fusion of lamellar epithelia (thick black arrow). **H&E.**

D- Longitudinal section of *Dicentrarchus labrax* muscle revealing section of digenetic encysted metacercaria cyst (thin arrow); the myogenic tissue showing interstitial edema (asterisks). Some muscle bundles were atrophied (thick arrow). **H&E X 400.**

E- Renal tissue of *D. labrax* showing cross sections of two digenetic encysted metacercaria cysts (thin arrows); surrounding tissues revealing evidence of pressure atrophy (thick arrows). **H&E X 200.**

F- Intestinal section of *Dicentrarchus labrax* infested with digenetic trematodes showing proliferated of intestinal villi (thick black arrow) and Cross section of digenetic trematodes (thin arrow). **H &E X200.**

## DISCUSSION

Aquaculture is a globally important industry that provides essential food to a growing world population, with a major role in the supply of cheap animal protein. In Egypt there are very rapid developments have been occurred in aquaculture sector because aquaculture is considered as the mainly viable option for reducing the current gap between production and consumption of fish (Soliman and Yacout, 2016). Parasites can seriously affect the marine resources being pathogenic to fish and zoonotic to consumers (Kuhn et al., 2011).

Concerning to the results of the clinical picture recorded in the fish farms, the naturally infected *D. labrax* showed restlessness, gasping of air, rubbing body against hard objects, swimming near the surface of the water with increased breathing frequency, stretched gills covers, loss of appetite, increase in mortality rate, haemorrhagic skin and fins, areas of detachment of scales and excessive mucous secretion in the body and gills.

These manifestation were in accordance with records of (Noga, 2010; Noor El-Deen et al., 2013; Hassanin, 2016 and Seoud et al., 2017). Regarding of postmortem examination showed paleness of gills in some infected fishes friable and pale liver congestion and haemorrhagic patches in intestine this findings supported by finding of (Ahmed, 2017) who observed that *Dicentrachus labrax* and *D. punctatus* infested with *digenetic* trematodes and *encysted metacercariae* showed paleness in liver in some cases and congestion in others also Intestine showed congestion and inflammation.

The prevalence of protozoal infection among the examined fish revealed four genera namely, *Trichodina heterodontata*, *Epistylis*, *Amyloodinium ocellatum* and *Microsporidia* were 28.3%, 14.2%, 20.0% and 8.3% respectively, these results was supported by (Pereira et al., 2011) reported that *Amyloodinium ocellatum* one of the most important ectoparasitic infection in marine aquaculture and Ihwan et al. (2016) recorded that, that trichodinids are highly significant in aquaculture because their responsibility in decreasing of growth rate and immune response for vaccination and chronic mortalities.

Concerning to the results of the seasonal variation for protozoal infection in naturally infected examined fishes the infection rate showed the high peak during autumn in *Trichodina heterodontata*, *Epistylis*, *Amyloodinium ocellatum* and *Microsporidia* infection they was (53.3%, 33.3%, 20% and 13.3%) in farm I and (46.7%, 26.7%, 53.3% and 20% in farm II respectively. on the other hand there infection rate were decrease during summer (6.7%, 0%,

0% and 6.7%) in farm I and (20%, 6.7% ,20% and 13.3%) in farm II respectively , the difference between farm I and II in prevalence rate were very low these may be due to the two farms were located in the same area and use the same water source , also from these results can indicated that climatic changes have a strong effect in infection rate of protozoal diseases . These results in contrast with the record of **Noga (2010)** reported that all protozoan ectoparasites have a direct life cycle, which is faster at higher temperatures. This defrance may be attributet to difference in weather of countries and temprature in different seasons. The results of Monogenean infestation in this study reviled two genera of monogenean namely (*Diplectanum* and *Pseudohaliotrema*) with total prevalence rate 12.5% and 9.2% respectively. These results were supported the finding of **Ahmed (2017)** found that, the prevalence of *D. bocqueti* detected from *D. labrax* were 20% . On the other hand they were lower than the observation of **El-khatib et al. (2014)** found prevalence of *Diplectanum sp* . isolated from *D. labrax* collected from El - Fayoum governorate and were infected also with bacteria was 100 % the difference between the results may be due to difference in water quality between El Fayoum and Damietta . The result of prevalence of *Pseudohaliotrema* were lower than the finding of **(Abdel-Mawla and Abo-Esa, 2011)** found that, the prevalence rate of *Pseudohaliotrema sp.* detected from *Siganus revulatus* from Suez canal was 31.7 this may be due to difference in fish spices and locality of collected samples. The seasonal variation of *Diplectanum sp.* showed the highest peak of infestation during autumn (26.7% and 20 %) followed by spring (20.0% and 13.3%) respectively in farm I and farm II and not detected in both farms in summer. these results show some agreement with the observation of **Ahmed (2017)** concluded that, the prevalence of *D. bocqueti* affecting *D. labrax* and *D. punctatus* was high in winter followed by spring then in summer and not detected in the autumn the difference between the results may be due to difference in the locality of collected samples. In case of *Pseudohaliotrema sp.* infestation examined *D. labraxs* were detected only in farm I with high prevalence rate during autumn (33.3 % ) followed by spring (36.7 %). these results was in contrast with **Abdel-Mawla and Abo-Esa (2011)** recorded that, the highest prevalence of *Pseudohaliotrema sp.* isolated from *Siganus revulatus* was in spring followed by summer while the lowest prevalence was in winter difference between result may be due to difference in fish species. Reading of infestation rate of crustacean ectoparasites infestation in naturally infested *D. labrax* revealed that only one genus, namely *Caligus minimus* with total prevalence rate of 15.0%. These finding was

higher than the results of Wang *et al.* (2001) and El-Raziky (2009) detected that, the infestation rate with crustacean ectoparasites from *Mugil cephalus* were 5.3% and 7% respectively, this difference may be due to difference in fish species. The Seasonal variation of *C. minimus* among examined *D. labrax* (farm I and II) showed that infestation rate increases in summer (46.7% and 33.3%), respectively and not detected in winter. These results in accordance with (Eissa *et al.*, 2012) and Hassanin (2016) reported that, the highest peak of *Caligus Spp.* infestation isolated from *D. labrax* and *M. cephalus* was in summer followed by autumn and decrease in winter. Increase infestation during summer may be due to increase in salinity of water in this locality during summer, this theory supported by (Karlsbakk *et al.*, 2001) reported that sea lice can cause disease outbreaks on non-salmonids fish when they are cultured in full salinity seawater. In case of infestation with unidentified encysted metacercaria (EMC) detected from naturally infested *D. labrax* the prevalence rate of infested with was 33.3 %. These results were higher than Ahmed (2017) recorded that, the infestation rate of *D. labrax* collected from private farm in Ismailia provinces was 19% , these fluctuation of prevalence may be due to the amount of intermediate host located in fish farm . infestation with EMC were detected all over the year with maximum prevalence during summer (53.3% and 46.7%) while the minimum prevalence were detected during spring (26.7% and 20.0%) respectively in farm I and farm II. These finding supported the finding of Elsheikha and Elshazly (2008) and Ghobashy *et al.* (2010) found that the highest rate was in summer , while they were in contrast with Kotb *et al.* (2014) and Ahmed (2017) proved that the seasonal prevalence of EMC was higher during winter followed by spring and decrease during summer. These differences could be explained by the season of migration of the migratory birds that act as definitive hosts for many of the metacercariae and presence of snails witch complete the life cycle of digenean. Concerned to the result of the adult digenetic trematodes detected from gastrointestinal tract of examined fish two genera (*Timoniella sp.* and *Bucephalus minimus*) were isolated with total prevalence rate 17.5% and 17.5% respectively. These results were nearly similar to observation of Ahmed (2017) noted that, the prevalence of *Timoniella sp.* affecting *D. labrax* was 25%, and also supported by Pina *et al.* (2009) recorded that, the adult stage of *B. minimus* develops in the Sea bass *Dicentrarchus labrax* as the final host the seasonal prevalence of *Timoniella* infestation showed the highest prevalence during winter (40.0% and 20.0%) in farm I and farm II

respectively while the lowest prevalence rate showed during summer (6.7% and 0.0%) in farm I and farm II respectively. These results partially disagree with **Ahmed (2017)** noted that seasonal prevalence of *Timoniella sp.* Affecting *D. labrax* and *D. punctatus* the highest peak was in spring followed by autumn and the lowest rate was in summer. On the other hand the infestation with *Bucephalus minimus* showed highest prevalence during autumn (40.0 % and 33.3%) followed by winter (26.7% and 20.0%) in farm I and farm II respectively and not detected during summer. these results was in contrast with the finding of **Pina et al. (2009)** recorded that prevalence of adult *B. minimus* isolated from Sea bass during spring and summer was 25%. This difference may be probably due to deference in geographical area of collected samples. In this study only one genus of nematode was detected namely *conteracecum* larvae from gastrointestinal tract of *D. labrax* with prevalence rate 4.2% , in the recent study of wild European sea bass, which reported prevalence ranging between 65 and 89.36 % in relation with body weight, the fish were caught from the northeastern Atlantic Ocean (**Bernardi, 2009**). This may explain by the different in origin of collected fish samples. Infestation with nematode namely *Conteracecum* larvae detected only during autumn with prevalence rate (20.0% and 13.3%) in farm I and farm II respectively. On the other hand another study reported that, the high prevalence of *Conteracecum sp.* isolated from *Lethrinus lentijan* and *Lutjanus spp.* was in summer (36.7% and 30.0%) respectively (**Abo-Esa and Abdel-Mawla 2012**). This difference may be due to difference of locality of collected samples and fish species. Fish gills are highly sensitive organ that provide the largest surface area get in contact with external environment, so easily damaged (**Karlsson, 1983**).Gills are incorporated in many vital functions; respiration, excretion and osmotic regulation, so histopathological changes in its structure will affect vitally on the fish health and behavior (**Cerqueira and Fernandes, 2002**) that approve the histopathological observations detected in these study, which represented in the form of lifting of respiratory epithelia, proliferation of basal undifferentiated cells, and partial and/or complete fusion of gill lamellae appearance of parasite or cyst of parasites in between gill filaments . Gill filaments epithelial hyperplasia and /or fusion and even aneurysm; that detected as a common feature in the present study, represent an indicator for infectious conditions (**Thiyagarajah et al., 1996**). This change give a false indication of increase surface area (in case of epithelial hyperplasia) (**Molnár et al., 2016**), as it lead to disturbance in blood fellow (**Stentiford et al., 2003**) and less efficient respiration, but provide good condition for parasite transmission (**Molnár et al.,**

2016). In the present study, encysted metacercaria could demonstrate in different body organs including; muscle, kidneys, spleen and liver. as The fish defense mechanisms of fish against infestation nodular formation and melanin pigmentation were detected and end result were encapsulation of parasite, so the functions of the invested organ (s) not affected in most cases (Molnár *et al.*, 2016). The present study demonstrated variable response to parasitic infestation including: massive sub-mucosal inflammatory cells infiltration, proliferation of intestinal villi and some intestinal villi appeared thick and short. Cross sections of parasite were detected in the Intestinal lumen. these results were supported by the findings of (Ashour *et al.*, 2014; Dezfuli *et al.*, 2015) recorded that a massive hyperplastic granulocyte response involving mast cells (MCs) and neutrophils and formed in close approximately to the helminths, meanwhile the associated desquamation of intestinal villi represent a common feature for intestinal parasitic infestation.

## REFERENCES

- Abdel-Mawla, H.I. and Abo-Esa, J.F.K. (2011): The most common parasitic diseases in *siganus revulatus* in sues canal area. J. Egypt. Vet. Med. Assae 71, 257-270.
- Abo-Esa, J.F.K. and Abdel-Mawla , H.I. (2012): Studies on prevailing parasitic infection in some marine fishes from Red Sea,Suez Governorate. Egyptian Journal for Aquaculture. 2, 1-16.
- Ahmed, M.M. (2017): Studies on prevailing parasitic trematodiasis affecting some cultured marine fishes. Aquaculture Diseases control Dept., Fish farming and technology institute , Suez canal university.
- Aly, S.M. (2013): A review of fish diseases in the Egyptian aquaculture sector: Working report.
- Antonelli, L., Foata, J., Quilichini, Y. and Marchand, B. (2016): Influence of season and site location on European cultured sea bass parasites in Corsican fish farms using indicator species analysis (IndVal). Parasitology research 115, 561-568.
- Ashour, D.S., Othman, A.A. and Radi, D.A. (2014): Insights into regulatory molecules of intestinal epithelial cell turnover during experimental infection by *Heterophyes heterophyes*. Experimental parasitology 143, 48-54.
- Bancroft , D., Stevens, A. and Turner, R.(1996): Theory and Practice of Histological Techniques, 2nd Edition. Churchill Livingstone, Edinburgh, London, Melbourne.
- Bernardi, C. (2009): Preliminary study on prevalence of larvae of Anisakidae family in European sea bass (*Dicentrarchus labrax*). Food control 20, 433 - 434.

- Cerqueira, C.C. and Fernandes, M.N. (2002):** Gill tissue recovery after copper exposure and blood parameter responses in the tropical fish *Prochilodus scrofa*. *Ecotoxicology and environmental safety* 52, 83-91.
- Dezfuli, B.S., Bo, T., Lorenzoni, M., Shinn, A. and Giari, L. (2015):** Fine structure and cellular responses at the host–parasite interface in a range of fish - helminth systems. *Veterinary parasitology* 208, 272-279.
- Eissa, I.A.M., El-Lamie, M. and Zakai, M. (2012):** Studies on Crustacean Diseases of Seabass, *Morone Labrax*, in Suez Canal, Ismailia Governorate. *Life Science Journal* 9.
- El-khatib, N.R., Abd El-Ghany, N.A. and Salama, S.S.A. (2014):** Trials to control sea bass mortalities due to concomitantly microbial and parasitic infection. In: 4th Conference of Central Laboratory for Aquaculture Research pp. 283-302.
- El-Raziky, E. (2009):** Studies on the prevailing external parasitic diseases among *Mugil cephalus*. M.V.Sc. Thesis. Fac. Vet. Med. Suez Canal Univ.
- Elsheikha, H.M. and Elshazly, A.M. (2008):** Host-dependent variations in the seasonal prevalence and intensity of heterophyid encysted metacercariae (Digenea: Heterophyidea) in brackish water fish in Egypt. *Veterinary parasitology* 153, 65-72.
- FAO (2014):** The state of World Fisheries and Aquaculture. Food and Agriculture Organization of the United Nations Publication .
- FAO, I. (2016):** WFP (2015), The State of Food Insecurity in the World 2015. Meeting the 2015 international hunger targets: taking stock of uneven progress. Food and Agriculture Organization Publications, Rome.
- Ghobashy, M., MFM, S. and Hassan, E. (2010):** Responses of the Mullet, *Liza auratus* and the Cichlid, *Oreochromis tiloticus* from Lake Manzala (Egypt) to Heterophyid Infection. *International Journal of Zoological Research* 6, 13-23.
- Hassanin, D.A.(2016):** Studies on Prevailing Problems Affecting Cultured Marine Fishes at Port-Said Governorate. M.sc. Fish Diseases and Management Dept. Fac. Vete. Med. Suez Canal Univ.
- Ihwan, M., Syahnon M., Fakhrulddin IM. and MA., M.H.A.A.(2016):** New Report on Trichodiniasis (Protozoa: Ciliophora: Peritrichida) in Jade Perch; *Scortum barcoo* from Peninsular Malaysia. *Journal of Fisheries and Aquatic Science* 11, 437-443.
- Karlsbakk, E., Otterlei, E., Hoie, H. and Nylund, A. (2001):** Parasites of cultured cod (*Gadus morhua*) postlarvae fed natural zooplankton. *Bulletin of the European Association of Fish Pathologists* 21, 63-70.
- Karlsson, L. (1983):** Gill morphology in the zebrafish, *Brachydanio rerio* (Hamilton-Buchanan). *Journal of fish biology* 23, 511-524.

- Kotb, H.L., Mahdy, O.A. and Shaheed, I.B. (2014):** Parasitological and histopathological study of digenetic Trematodes in mullets from Lake Qarun, Egypt. *Global Veterinaria* 13, 202-208.
- Kuhn, T., García-Márquez, J. and Klimpel, S. (2011):** Adaptive radiation within marine anisakid nematodes: a zoogeographical modeling of cosmopolitan, zoonotic parasites. *PLoS One* 6, e28642.
- Lucky, Z. (1977):** Methods for the diagnosis of fish diseases, Amerind. publishing Co. PV T. Ltd., New Delhi, Bombay, India.
- Molnár, K., Gibson, D.I., Majoros, G., Székely, C., Sándor, D. and Cech, G. (2016):** Malformations of the gill filaments of the ruffe *Gymnocephalus cernuus* (L.) (Pisces) caused by *echinostomatid metacercariae*. *Journal of fish diseases* 39, 1357-1367.
- Noga, E. (2010):** Fish disease: Diagnosis and treatment.. Watsworth Publishing, second edition, Co., USA.
- Noor El-Deen, A., Mahmoud, A.E. and Hassan, A.H. (2013):** Field studies of caligus parasitic infections among cultured seabass (*Dicentrarchus labrax*) and mullet (*Mugil cephalus*) in marine fish farms with emphasis on treatment trials. *Global Veterinaria* 11(5), 511-520.
- Paperna, I. and Laurencin, F.B. (1979):** Parasitic infections of sea bass, *Dicentrarchus labrax*, and gilt head sea bream, *Sparus aurata*, in mariculture facilities in France. *Aquaculture* 16, 173-175.
- Pereira, J., Abrantes, I., Martins, I., Barata, J., Frias, P. and Pereira, I. (2011):** Ecological and morphological features of *Amyloodinium ocellatum* occurrences in cultivated gilthead seabream *Sparus aurata* L.; A case study. *Aquaculture* 310 289-297.
- Pina, S., Barandela, T., Santos, M.J., Russell-Pinto, F. and Rodrigues, P. (2009):** Identification and description of *Bucephalus minimus* (Digenea: *Bucephalidae*) life cycle in Portugal: morphological, histopathological, and molecular data. *Journal of Parasitology* 95 : (2), 353-359.
- Rothuis, A., van Duijn, A.P., Roem, A., Ouwehand, A., van der Pijl, W. and Rurangwa, E. (2013):** Aquaculture business opportunities in Egypt (Wageningen UR).
- Seoud, S.S.M., Zaki, V.H., Ahmed, G.E. and Abd El-Khalek, N.K. (2017):** Studies on Amyloodinium Infestation in European Seabass (*Dicentrarchus labrax*.) Fishes with Special Reference for Treatment. *International Journal of Marine Science* 7.
- Shalan, M., El-Mahdy, M., Saleh, M. and El-Matbouli, M. (2018):** Aquaculture in Egypt: Insights on the Current Trends and Future Perspectives for Sustainable Development. *Reviews in Fisheries Science and Aquaculture* 26, 99-110.



- Shaheen, A., Seisay, M. and Nouala, S. (2013):** An industry assessment of tilapia farming in Egypt. African Union, International Bureau for Animal Resources (AU-IBAR).
- Soliman, N.F. and Yacout, D.M. (2016):** Aquaculture in Egypt: status, constraints and potentials. Aquaculture International 24, 1201-1227.
- Stentiford, G., Longshaw, M., Lyons, B., Jones, G., Green, M. and Feist, S. (2003):** Histopathological biomarkers in estuarine fish species for the assessment of biological effects of contaminants. Marine Environmental Research 55, 137-159.
- Thiyagarajah, A., Hartley, W.R., Major, S.E. and Broxson, M.W. (1996):** Gill histopathology of two species of buffalo fish from a contaminated swamp. Marine Environmental Research 42, 261-266.
- Wally, A. (2016):** The State and Development of Aquaculture in Egypt. (Global Agricultural Information Network. USDA Foreign Agriculture Service).
- Wang, J.-H., Lee, J.-L., Chang, H.-Y., Wang, C.-T., Wu, C.-H. and Tasi, K.-H. (2001):** A survey of the diseases in cultured brood grey mullet (*Mugil cephalus* Linnaeus) in the Yun-Chia-Nan area of Taiwan. JOURNAL-CHINESE SOCIETY OF VETERINARY SCIENCE 27, 89-93.