

EFFECT OF COMMERCIAL ENZYMES AND/OR MANNAN OLIGOSACCHARIDE SUPPLEMENTATION ON PRODUCTIVE PERFORMANCE, NUTRITIVE VALUES AND METABOLIC INDICES OF BROILER CHICKS

Maha A. Abd El Latif

Animal and Poultry Production Department, Faculty of Agriculture, Minia University, El Minia, Egypt.

Running title: *Effect of enzymes and/or MOS on performance*

Submitted: 25/1/2023 ; **Accepted:** 11/4/2023 ; **Published online:**8/8/2023

SUMMARY

This study was conducted to evaluate the effect of using feed additives in Sasso broiler chicks as growth promoters on productive performance, digestibility coefficient of nutrients, blood indicators, carcass and gastrointestinal measurements and economic return. A total number of 120, one - day old, unsexed Sasso chicks were assigned randomly to four groups. Each group included three replicates. The first group was kept as control and fed the basal diet. The 2nd, 3rd and 4th groups were fed the basal diet (+ 1.00 g Frazyme), (2.50 g mannan oligosaccharides) and (1.00 g Frazyme + 2.50 g MOS) kg diet respectively during whole experimental period (1-58) days of age. The obtained results showed that the better ($P < 0.05$) values of LW, WG and FCR were recorded to group four followed by group three at the end of experimental period. There were no significant differences among all experimental treatments in feed intake during the whole experimental period. Significant ($P < 0.05$) improvement has been recorded in digestibility of CP and CF for group four and two respectively. Group four showed the highest values ($P < 0.05$) of breast yield and spleen weight compared to other groups. There were no significant differences of gastrointestinal lengths among all experimental groups. Regarding blood parameters, broilers in group two had significantly ($P < 0.05$) Hct, Lipase and Amylase. While, group four broiler's were recorded the highest ($P < 0.05$) TP, Alb., Hb and TAC compared to others. The chemical analysis of meat revealed that greatest ($P < 0.05$) CP of breast meat for group four. However, Thigh meat analysis for birds in control and group four achieved high significant values of fat and ash contents. The lowest mortality and highest PI and economic efficiency were related to groups four and three respectively. On the basis of these results, it can be concluded that adding enzymes combination, MOS or their mixture (Enz+MOS) as feed additives were suitable and had good role in improving productive and economic efficiency of Sasso broiler chicks.

Keywords: *commercial enzymes, mannan oligosaccharides, productive, broiler chicks*

INTRODUCTION

In past, antibiotic growth promoters have been widely used in all types of livestock production systems particularly poultry (Alhidary *et al.*, 2017 and Chand *et al.*, 2018) at sub therapeutic level they are used as feed additives to improve growth and nutrient availability by regulating microbial populations (Abudabos *et al.*, 2017). On the other hand, it has negative effects as it increases microbial resistance to antibiotics and results the residues in chicken meat products which might be harmful to consumers (Diarra *et al.*, 2007 and Koc *et al.*, 2010). Recently, feed additives are being tested to relieve the problems connected with the elimination of antibiotics from food (Attia *et al.*, 2014a). The alternative feed additives include products such as enzymes, prebiotics, probiotics, appetizers, yeast, growth promoters or combinations of all those products, which are classified as non-nutrient feed additives (NNFA). They are added to the feed to

improve or to accelerate the rate of nutrient utilization (Altafur *et al.*, 2007 and Midilli, *et al.*, 2008).

Several studies on the supplementation of exogenous enzymes to broiler diets have been performed. Addition of exogenous enzymes in broilers diets improved broilers performance, daily gain, feed conversion (Wang *et al.*, 2005), they were found to enhance the food digestibility, minimize the anti-nutritional effects. Reduction of the adverse impacts of NSPs, correlated with the degradation of NSPs for better energy utilization and availability in broiler's intestinal tract (Abdollahi *et al.*, 2016). They promoted the productivity indices (Hooge *et al.*, 2010) due to its vital role in the diets. Olukosi *et al.* (2015) found that a combination of enzymes cocktail produced a greater effect in improving energy and protein values, as well as increasing the solubilization of diets given to broilers. Also, Stefanello *et al.* (2015) demonstrated that the growth Performance was improved when broilers were fed diets supplemented with addition of enzymes.

The use of prebiotics instead of antibiotics is becoming increasingly popular in feeding of birds to improve the useful microbial population in the gut (Kermanshahi, 2006). Mannan oligosaccharides are commonly used in the same manner as prebiotics with the due to their ability to selectively enriching the beneficial bacterial populations (Patterson and Burkholder, 2003).

Mannanoligosaccharides (MOS) or Y-MOS is a natural extract from natural yeast compound; it is a derivative of the cell wall of yeast *Saccharomyces cerevisiae* (Chand *et al.*, 2016). Many studies evaluated the effects of MOS on growth performance, feed efficiency, blood chemistry, and intestinal environment. Chand *et al.* (2018) reported that addition MOS to basal diet of broiler improved; the number of goblet cells in the gut, pH alteration of intestine, enzyme production, antagonist for intestinal adhesion receptors, and opposition to feed ingredients. Improved structure of intestine (Iji *et al.*, 2001), and reducing toxin level, and stimulation of the immune system. The main effect of Y-MOS yeast supplementation to basal diet was to increase weight gain and overall growth measurements when compared to control group, Those positive attributes have been reported by Mohamed and Mukhtar (2016) and Habib *et al.*, (2017). Feeding of prebiotic has been useful to improve carcass quality of broiler chickens (Tavaniello, 2018). The productive efficiency can be achieved when obtaining maximum production with minimum cost and using the least amount of resources to produce a given output level (Emara, 2009; Romero *et al.*, 2010 and Mohamed, 2016).

Therefore, the aim of this study was to evaluate the effect of commercial enzymes, mannan oligosaccharides and its mixture on growth parameters, hemato-biochemical parameters, digestibility of nutrients, carcass characteristics, gastrointestinal tract characteristics and economic analysis when fed to Sasso broiler chicken.

MATERIALS AND METHODS

Experimental chicks, Management and Housing:

The current study was carried out at the Animal and Poultry Research Farm, Animal and Poultry Production Department, Faculty of Agriculture, Minia University. All experimental procedures were carried out in accordance with the local Experimental Animal Care Committee and authorized by the Institutional Committee of the Department of Animal

Production, Faculty of Agriculture, Minia University, Egypt. A total number of 120, One day- unsexed Sasso broiler chicks were purchased from Cairo Company for Poultry. The chicks were weighted and randomly allocated to four experimental groups (30 chicks/group). Each group consists of three replicates (10 chicks/replicate). The chicks were housed in an open house, two-tiers floor batteries (100× 60 × 40 cm/cage), for length, width and height, respectfully. Gas heater were placed in the house to control the interior temperature. Brooding temperature started at 33- 34°C during the first 3 days, then 31°C till the end of the first week, followed by reduction of 2°C/week until the temperature reached 28°C at the end of experiment according to Marwa, (2013) and Mohamed, (2016). Feed and water were offered continuously. The lighting was 24 hr of light during first three days, then 23h until the 7th day followed by 20 h light from the 8th day until 15th days of age. Then natural day light only until the end of the experimental period, which was provided according to farmer's guide of Sasso colored broiler chicks.

Experimental Diets:

Chicks were fed on balanced diet as shown in Table 1.

The test diets for the starter and grower phases (1-28 and 29-58 days of age) were formulated to be iso-caloric and iso-nitrogenous, and containing adequate levels of all other nutrients to meet the requirements of starter and grower of Sasso broiler. The first group was fed the basal diet without supplementation (control); while the 2nd, 3rd, and 4th groups were fed basal diet supplemented with 1.00, 2.50 and 1.00 + 2.50 g/kg diet of commercial enzymes mixture (Frazyme), mannan oligosaccharides (MOS) and mixture of frazyme and MOS respectively. Frazyme was purchased from ATCO Pharma Company for Animal Health – Cairo, Egypt. While, MOS was purchased from Khayrat El-Nile Company for feed additives, Cairo, Egypt.

Frazyme components were Alpha-amylase 2100 IU, Xylanase (*Trichoderma Reesi*)16000U, 1.3(4) Beta-glucanase 2400 U, Pictinase 210U, Mannanase 3000U, Proease 600 U and Carrier: Ceplolite up to 1g.

MOS components were yeast cell wall (*Saccharmyces Cerevisiae*) 100%, Beta glucanase 25% and Mannanoligosaccharides 18.5%.

Table 1. The composition and chemical analysis of starter and grower diets

Ingredients %	Starter diet	Grower diet
Yellow corn	50.00	55.00
Soybean meal, 44%	32.00	27.00
Corn gluten, 62%	5.00	4.00
Wheat bran	5.50	6.00
Vegetable oil	4.00	4.90
Limestone	2.00	1.6
Di-calcium phosphate	1.00	1.00
Nacl	0.25	0.25
Vit. & min. *	0.25	0.25
Total	100.00	100.00
Price\ kg	5.50 (EGP)	5.00.(EGP)
Calculated analysis (NRC,1994):		
CP%	22.07	19.67
ME, k cal\kg	2996.00	3100.21
CF%	3.54	3.62
Ca%	1.03	0.95
Av. Ph%	0.69	0.65
Lys. %	1.10	0.95
Meth+Cys.	0.52	0.50
Chemical analysis:		
DM%	93.02	92.78
Ash%	7.20	6.76
CP%	22.00	19.80
CF%	3.35	3.72
EE%	4.69	6.21
NFE%	62.76	63.51

According to NRC,1994 and Sasso farmer's guide* Each 3 kg contains contain: Vitamin A = 12,000,000 IU, D3 = 2,000,000 IU, E = 10,000 mg, K3= 2000mg, B1 = 1000 mg, B2 =5000 mg, B6 =1500 mg, B12= 10mg, Biotin= 50 mg, pantothenic acid= 10000 mg, Nicotinic acid = 30000 mg, Folic acid =1000 mg, Zinc = 50,000 mg, Manganese = 60,000 mg, Iron = 30,000 mg, Copper = 10,000 mg, Iodine =1,000 mg, Selenium = 100 mg, Cobalt = 100 mg, Cobalt = 1000 mg, and Calcium carbonate up to 3 Kg.

Growth measurements:

Live body weight and feed intake were recorded every two weeks, from 1 to 14, from 15 to 28, from 29 to 42 and from 43 to 58 days of age. Body weight gain and feed conversion ratio were calculated for the same periods during starter and grower phases. Mortality rate was estimated at the end of the experimental period by the number of dead birds during the whole period m, divided by the initial number of bird in each group, and multiplied by 100. Performance index (PI) and economic efficiency were calculated according to North (1981) and Waheed and ElTaieb (2005) respectively.

Slaughter test:

Slaughter test was done with randomly selected three birds from every group at the end of experiment (58 days of age) to determine carcass characteristics as dressing, breast, thigh percentages and liver, gizzard, heart, abdominal fat and spleen weights. Gastrointestinal tract was removed after evacuation the carcass and lengths of duodenum, jejunum, ileum, cecum and colon were measured in centimeter.

Chemical analysis of meat, diets and excreta:

Samples of breast and thigh meat from slaughtered broilers and the experimental diets and excreta were dried in Muffle oven at 60° C over night, and then were crushed into fine powder and saved in glass jars to be chemically analysis according to (AOAC, 2010) to determine content of dry matter, crude protein, crude fat and ash.

Digestibility trial:

At the end of the experimental period (8 weeks of age) digestion trial was conducted for 3-days to estimate the digestion coefficient of nutrients; dry matter, organic matter, crude protein, crude fiber, and nitrogen free extract. The feed consumption was recorded, and the manure, which fell on polyethylene sheets, was collected quantitatively for each replicate (6 birds/group) every 24 hour. Fecal nitrogen was determined according to Jakobsen *et al.* (1960) as follow:

- 1- About 2 gm of dried excreta in a 300 ml glass beaker plus 70 ml of distilled water.
- 2- 20 ml of sodium borate and 6 ml of potassium permanganate were added.
- 3- The beaker was placed in a water stirred bath at a temperature of 50°C for an hour.
- 4- The beaker was raised and left to slow down for at least one hour, at room temperature.
- 5- Addition about 30 ml tri-choloro acetic acid (10%) to the beaker and stirred with glass straight stick.
- 6- The beaker was left again for 30 minutes at room temperature, then filtered in ashless filter paper (15 cm).
- 7- The residual washed four times with 25-30 ml tri-choloro acetic acid (2%).
- 8- The filter paper with the sample was dried in an oven at 90°C for three hours.
- 9- Finally, determination the nitrogen content of faeces with kjeldahle method.
- 10- Calculation digestion coefficient = digested feed/feed intake×100.

Blood measurements:

Blood samples were taken during slaughter into two tubes/bird to obtain plasma and serum, from coagulated blood. Plasma or serum were separated by centrifugation of the blood at 3000 rpm for 20 minutes and stored at -20°C for later analysis. Heparinized tube was used to estimate the total count of red blood cells (RBC) white blood cells (WBC), packed cell volume (PCV), and hemoglobin (Hb%). Un heparinized tube was used to determine serum biochemical parameters as total protein (TP), albumin (Alb.), globulin (Glob.) was calculated, total lipids (TL), glucose (Glu), uric acid (UR), alkaline phosphatase (ALP), tri-iodothyronine (T3), thyroxin (T4), total antioxidants capacity (TAC), and some digestive enzymes like as lipase, amylase and protease. The colorimetric methods were used to assess blood biochemistry and radioimmunoassay technique to concentration of total T3 and T4 using commercial test kits that were obtained from Biomed Diagnostic Company, Giza, Egypt.

Statistical analysis:

The obtained results were subjected to statistical analysis using one-way analysis of variance (ANOVA) using the SAS, 2002. The significance of differences among means was determined by using

Duncan's new multiple range test (Duncan, 1955.) and the following statistical model was used,

$$Y_{ik} = \mu + T_i + e_{ik}$$

Where:

Y_{ik} = Experiment observations.

μ = the overall mean.

T_i = the effect of dietary treatment.

e_{ik} = the experimental error.

RESULTS

Productive performance:

Growth performance:

The effect of different additives on productive performance indicators such as live weight (LW), weight gain (WG), feed intake (FI) and feed conversion ratio (FCR) at different ages for Sasso broilerchicks is shown in Table 2. Sasso broiler chicks fed basal diet supplemented with feed additives had significantly ($P < 0.05$) recorded linear increase in LW and WG at (1-14), (15-28), (29-42), (43-58) days of age and total experimental period (1-58 days of age) compared to broiler chicks fed control basal diet without supplements. The greater LW and WG was noted in chickens of group four that were supplemented with mixture of (1.00g Frazyme + 2.50g MOS/kg) followed by broiler chicks in group three that were fed (1.00g MOS/kg) compared to control chicken group. Moreover, Chicks fed the basal diet or basal diet with feed additives had no significant differences in feed intake (FI). The lowest value of FI has been recorded for broilers that were fed diet with 2.5 g MOS (4330.54g/bird) followed by broilers fed diet with enzyme + MOS (4362.76 g/bird) compared to broiler fed the control diet (4455.47 g/bird) or broilers that were fed control diet plus commercial enzymes addition (4433.90 g/bird) during whole experimental period (1-58 days of age). The enhancement in FCR values was significant ($P < 0.05$) in line of treated groups during (1-14), (43-58) and whole period (1-58) days of age. The best value of FCR was calculated for broiler chicks fed diet with MOS (group three) at starter phase, while, during grower phase the significant improve was shared for birds in group four (Frazyme + MOS) then birds in group three (MOS) compared to control or enzyme groups during periods (43-58) and (1-58 days of age).

Table 2. Effect of different additives on productive performance of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz.+ MOS	SEM	P-value
LW at 01	40.83	41.30	40.17	40.00	0.86	0.699
At 14	200.23 ^b	213.51 ^{ab}	220.98 ^a	224.84 ^a	4.15	0.013
At 28	618.16 ^b	634.84 ^{ab}	641.08 ^{ab}	670.23 ^a	13.69	0.132
At 42	1328.34 ^b	1351.97 ^{ab}	1354.90 ^{ab}	1395.03 ^a	17.56	0.135
At 58	2087.75 ^b	2109.61 ^b	2204.45 ^b	2392.20 ^a	39.73	0.002
WG (1-14)	158.39 ^b	171.51 ^a	180.53 ^a	183.73 ^a	3.90	0.007
(15-28)	418.10	421.15	422.19	446.10	14.67	0.539
(29-42)	698.68	715.31	713.05	723.14	19.85	0.847
(43-58)	719.41 ^c	757.64 ^{bc}	849.53 ^b	995.83 ^a	33.33	0.001
(1-58)	2039.24 ^b	2067.94 ^b	2164.00 ^b	2350.13 ^a	38.97	0.001
FI (1-14)	313.27	308.22	308.24	320.35	4.65	0.282
(15-28)	844.83	838.82	803.00	817.20	26.54	0.672
(29-42)	1440.27	1442.23	1408.10	1404.96	34.06	0.805
(43-58)	1857.10	1844.20	1811.86	1818.26	24.25	0.540
(1-58)	4455.47	4433.90	4330.54	4362.76	51.49	0.338
FC (1-14)	1.98 ^a	1.79 ^b	1.67 ^b	1.74 ^b	0.05	0.014
(15-28)	1.97	1.98	1.90	1.84	0.07	0.484
(29-42)	2.06	2.03	1.98	1.94	0.06	0.630
(43-58)	2.60 ^a	2.45 ^{ab}	2.13 ^{bc}	1.82 ^c	0.11	0.006
(1-58)	2.19 ^a	2.14 ^{ab}	2.01 ^{bc}	1.86 ^c	0.04	0.005

a.,b and c means in the same rows for each treatment having different letter(s) are significantly different ($p < 0.05$), LW= live weight, WG= weight gain, FI= Feed Intake, FC= Feed conversion, Con.= control, Enz.= enzymes mixture and MOS=mannan oligosaccharides

Carcass and intestinal parameters:

Effects of different additives on carcass parameters of Sasso broiler chicks are shown in Table 3. Addition of commercial enzymes, MOS and their mixture had insignificant effects on dressed weight (DW) or dressing percentage (Dress%), thigh weight (ThW), thigh yield%, liver weight (Liv.W), gizzard weight (Giz.W), heart weight (Hrt. W),

abdominal fat weight (Ab, Fat) compared to unsupplemented group. Sasso broiler chicks fed control diet supplemented with mixture of (Enz.+ MOS) had significant ($P < 0.05$) values in breast weight (BW), breast yield % and spleen weight followed by chicks in group three that fed MOS compared to the others.

Table 3. Effect of different additives on carcass organs weights of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz. + MOS	SEM	P-value
LBW, g	2070.00	2179.33	2212.67	2339.00	91.15	0.292
DW, g	1415.47	1496.87	1549.03	1765.80	102.07	0.171
Dress. %	68.41	68.57	70.25	75.15	2.98	0.398
Breast W., g	360.07 ^b	409.59 ^b	461.92 ^{ab}	541.27 ^a	38.10	0.048
Breast %	25.38 ^b	27.17 ^b	29.86 ^a	30.60 ^a	0.742	0.003
Thigh W., g	234.71	258.09	256.21	276.020	14.31	0.313
Thigh %	16.41	17.23	16.47	16.05	0.77	0.750
Liv. g	47.28	47.69	49.45	53.90	6.48	0.881
Giz., g	35.53	37.97	34.48	34.15	4.44	0.925
Hrt, g	13.26	11.31	15.98	10.71	2.29	0.414
Spleen, g	4.75 ^b	4.11 ^{ab}	4.71 ^{ab}	5.20 ^a	0.26	0.098
Ab. Fat, g	28.45	25.91	30.33	21.76	4.28	0.552

a,b and c means in the same rows for each treatment having different letter(s) are significantly different ($p < 0.05$), LBW= live body weight, DW= dressed weight, Dress= dressing, Liv= liver, Giz =gizzard, Hrt=heart, Ab.fat= abdominal fat, Con.= control, Enz.= enzymes mixture and MOS=mannan oligosaccharides

The results in Table 4 showed that feed additives of Enz., MOS and Enz. +MOS as growth promoters

for Sasso broiler had not significant differences on gastrointestinal tract lengths (cm) or its percentages

i.e. duodenum, jejunum, ileum, colon or cecum in comparison with the control group.

Table 4 Effect of different additives on intestinal parts lengths of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz. + MOS	SEM	P-value
Doud., cm	27.83	32.86	29.00	28.66	1.73	0.250
Doud. %	12.85	13.94	12.68	12.50	0.97	0.733
Jej., cm	83.86	86.43	83.06	89.66	5.63	0.839
Jej., %	38.73	36.42	36.23	38.61	1.83	0.666
Ileum, cm	88.33	90.40	90.40	86.73	5.64	0.958
Ileum %	40.86	38.26	38.96	37.51	1.86	0.634
Colon, cm	7.23	7.56	8.16	8.33	0.81	0.757
Cecum, cm	16.33	19.33	19.83	18.00	1.36	0.333

Doud. = duodenum, jej. = jejunum, Con.= control, Enz. = enzymes mixture and MOS=mannan oligo-saccharides

Mortality, production index and economical return:

Results in Table 5, enclosed values of mortality rate, production index, total feed costs, total revenue, net revenue, economic efficiency and relative economic efficiency. Sasso broiler chicks fed diet

supplemented with Enz.+MOS (group four) and MOS (group three) achieved the lowest mortality %, the highest production index, economic efficiency and relative economic efficiency respectively rather than control (group one) or Enz. (group two).

Table 5. Effect of different additives on mortality, production index, economic efficiency and relative economic efficiency of Sasso broiler chicks

Parameters	Treatments			
	Con.	Enz.	MOS	MOS+Enz.
Mortality%	10.00	6.66	3.33	3.33
Production index*%	95.29	98.55	109.65	128.60
FI, kg(starter)	1.158	1.147	1.111	1.137
Price of starter, EGP	6.369	6.320	6.188	6.244
FI, kg(grower)	3.297	3.286	3.219	3.22
Price of grower, EGP	16.49	16.46	16.29	16.37
Total feed price, EGP [#]	22.86	22.78	22.48	22.72
Total revenue**	54.26	54.83	57.30	62.19
Net revenue	31.40	32.05	34.82	39.47
EE	1.37	1.41	1.55	1.74
REE	100.00	102.92	113.14	127.00

*Production index value was calculated throughout the experimental period according to (North, 1981) as follow $PI = (\text{body weight, kg}/\text{FCR}) \times 100$, ** total revenue= LW(kg)× price (26.00 EGP kg LW), one kg frazyme= 100.00 EGP (using one g/kg diet), one kg MOS = 65.00 EGP (using 2.5 g/kg), # prices of feed ingredients, feed additives and live broilers during experiment time

Digestibility of nutrients:

The apparent digestibility of nutrients of Sasso broiler chickens fed diet supplemented with enzyme, MOS or their mixture during the whole period of study (1-58 days of age) are shown in Table 6. The obtained data cleared that addition of growth promoter (Frazyme, MOS and Frazyme + MOS) had insignificant effect on digestibility of nutrients such as dry matter (DM), organic matter (OM), ether

extract (EE) or nitrogen free extract (NFE) except crude protein (CP) and crude fiber (CF) which were significant. Sasso broiler chicks fed basal diet supplemented with additives had significantly ($P < 0.05$) better digestibility values of CP and CF than birds in control group. The highest values of digestibility coefficients of CP and CF were pertained to group four and group two respectively in comparison of the other groups.

Table 6. Effect of different additives on nutrients digestibility's of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz. + MOS	SEM	P-value
DM%	69.42	72.38	72.63	75.94	2.63	0.433
OM%	72.65	71.75	70.83	70.50	1.69	0.804
CP%	73.87 ^b	78.15 ^{ab}	77.39 ^{ab}	80.67 ^a	1.68	0.018
CF%	24.25 ^b	37.21 ^a	36.99 ^a	32.98 ^a	2.05	0.060
EE%	74.10	78.73	73.41	78.46	1.63	0.098
NFE%	72.31	74.08	73.81	74.99	1.68	0.732

a,b and c means in the same rows for each treatment having different letter(s) are significantly different ($p < 0.05$), DM= dry matter, OM= organic matter, CP= crude protein, CF= crude fiber, EE=ether extract, NFE=nitrogen free extract, Con.= control, Enz.= enzymes mixture and MOS=mannan oligosaccharides

Chemical analysis of meat:

The chemical composition of breast and thigh meats as affected by feed additives is presented in Table 7. Dry matter, fat, and ash of breast meat not affected significantly with supplements. Breast Crude protein had high significant ($P<0.05$) differences among treated groups and control group. The greatest ($P<0.05$) crude protein of breast meat was related to

Enz. + MOS group (79.73%) followed by (76.76%) for MOS group compared to Enz. Group or control group. Dry matter, crude protein and fat of thigh meat content had no significant variances with insignificant increase for fat thigh content compared to the supplemented groups. Ash percentage of thigh meat was increased significantly with adding the mixture (group four).

Table 7. Effect of different additives on chemical composition of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz. + MOS	SEM	P-value
Breast meat						
Most. %	73.40	73.60	73.16	72.800	0.84	0.916
DM%	26.60	26.40	26.50	27.20	0.85	0.908
CP%	73.40 ^c	73.33 ^c	76.76 ^b	79.73 ^a	0.74	0.0008
Fat%	4.60	4.03	4.13	3.81	0.35	0.591
Ash%	3.86	4.16	4.56	5.00	0.42	0.326
Thigh meat						
Most. %	75.50	74.83	75.55	75.06	0.273	0.259
DM%	24.50	25.16	24.44	24.93	0.273	0.259
CP%	64.66	64.78	65.76	67.80	1.13	0.271
Fat%	6.26 ^a	5.17 ^{ab}	4.66 ^b	5.53 ^{ab}	0.398	0.104
Ash%	4.43 ^b	5.10 ^{ab}	5.93 ^a	6.02 ^a	0.288	0.013

a, b and c means in the same rows for each treatment having different letter(s) are significantly different ($p<0.05$), Moist. = moisture DM= dry matter, CP= crude protein, Con.= control, Enz. = enzymes mixture and MOS=mannan oligosaccharides

Blood parameters:

The hematological and biochemical blood constituents, digestive enzymes and total antioxidants of Sasso broilers are shown in Table 8. Effect of Enz., MOS or Enz.+ MOS was insignificant on values of total counts of white blood cells and red blood cells, albumin, globulin, albumin: globulin ratio, total lipids, glucose, uric acid, alkaline phosphatase, T3, T4, and protease. Sasso broilers fed

basal diet with Enz. Had high significant values of hematocrit (Hct%), lipase and amylase. While, birds fed Enz. + MOS diet significantly ($P<0.05$) recorded the highest value for hemoglobin (Hb%), total protein (TP), and total antioxidants capacity (TAC), those broilers in the same group had insignificant increase of albumin (Alb.) and protease enzyme values. Broiler chicks fed diet with MOS recorded insignificantly increase of red blood cells count.

Table 8. Effect of different additives on hematological and biochemical blood parameters and digestive enzymes of Sasso broiler chicks

Parameter	Treatments				Statistics	
	Con.	Enz.	MOS	Enz.+ MOS	SEM	P-value
WBC $\times 10^3$	24.16	26.73	25.73	26.17	0.75	0.1710
RBC $\times 10^6$	4.03	4.53	5.73	4.50	0.74	0.189
Hct%	35.70 ^b	42.80 ^a	37.50 ^b	37.93 ^b	1.20	0.016
Hb%	11.90 ^b	12.63 ^b	12.50 ^b	14.27 ^a	0.40	0.017
TP(g/dl)	4.17 ^c	4.57 ^{bc}	5.13 ^{ab}	5.63 ^a	0.28	0.024
Alb. (g/dl)	1.34 ^b	1.50 ^{ab}	1.48 ^{ab}	1.99 ^a	0.17	0.122
Glob. (g/dl)	2.72	3.03	3.44	2.74	0.47	0.517
Alb\Glob	0.50	0.52	0.43	0.70	0.11	0.365
TL(mg/dl)	563.87	570.48	580.41	540.93	44.46	0.741
Glu(mg/dl)	209.70	269.30	240.00	253.40	20.50	0.281
UR(mg/dl)	3.86	4.23	4.30	4.87	0.60	0.711
ALP (U/L)	71.66	83.33	68.04	76.70	15.96	0.9156
T3 (ng/ml)	2.58	3.55	2.970	3.33	0.44	0.4799
T4(ng/ml)	13.93	14.65	16.00	14.66	1.11	0.6315
Lipase (U/L)	30.00 ^b	48.77 ^a	36.00 ^{ab}	39.53 ^{ab}	4.45	0.0391
Amylase(U/L)	52.33 ^b	89.00 ^a	53.34 ^b	53.00 ^b	5.92	0.005
Protease(U/L)	51.00	67.00	61.33	70.45	8.62	0.455
TAC (Mmol/L)	0.96 ^b	1.10 ^b	1.21 ^b	1.72 ^a	0.09	0.002

a, b and c means in the same rows for each treatment having different letter(s) are significantly different ($p<0.05$), WBC= white blood cells, RBC= red blood cells, Hct=hematocrit, Hb= hemoglobin, TP= total protein, Alb =albumin, Glob= globulin, Alb\Glob=albumin: globulin ratio, TL= total lipids, Glu= glucose, UR=uric acid, ALP= alkaline phosphatase, TAC= total anti-oxidant capacity, Con.= control, Enz.= enzymes mixture and MOS=mannan oligo-saccharides.

DISCUSSION

Productive performance :

Growth measurements:

In the present study, the performance traits were improved in the treatment groups compared to the control. Higher live body weight and weight gain in broiler of group 3 and group 4 may be due to MOS feeding which increases the beneficial bacteria such as *Bifidobacteria* and *Lactobacilli* in the gut of broiler (Baurhoo *et al.*, 2007), Improvement in microbial population in the gut causes improvement in nutrient absorption and decrease in nutrients loss (Sultan *et al.*, 2015) , in addition to provide the energy for intestinal epithelial cells via short chain fatty acids production which relate to the health of gut lining (Ferket *et al.*, 2005), or may be due to feeding (Enz. + MOS) caused alteration the chicken intestine PH in addition to increase digestive enzymes like protease, amylase, and lipase which enhance nutrient digestion, absorption and utilization of essential minerals and amino acids in the gastrointestinal tract by providing a larger surface area for efficient nutrients absorption (Xu *et al.*, 2003; Yang *et al.*, 2009; Santoso *et al.*, 2001; Karimi *et al.*, 2010 and Ghazalah *et al.*, 2011). The present study concluded that significant enhancement of FCR values for all treated groups (Enz., MOS or Enz. + MOS) compared to control group, this result is in agreement with the findings of Dizaji *et al.* (2013), Fernandes *et al.* (2014) and Akoy (2015). Also, Luo *et al.* (2009), Plessis, (2014) and Alqhtani *et al.* (2022) indicated that FCR improved by enzymatic supplementations alone or with other feed additives supplementation compared to control group (at 21-day, 35 day and at 42 day of age). The high enhancement in FCR for feed additives groups (MOS or MOS+ Enz.) may be attributed to stimulated growth of the beneficial microflora in the gastrointestinal tract (GIT) induced by dietary supplementation of MOS or MOS+ Enzyme (Cinar *et al.*, 2009), or could be attributed to the growth promoting ability via the prevention and treatment of subclinical infections (Ao *et al.*, 2011a, 2011b and Toghyani *et al.*, 2011). The current results in disagreement with Salehimanesh *et al.* (2016) who suggested that, the dietary supplementation of MOS and B-glucan did not affect body weight and gain of broiler and (Mahmoud *et al.*, 2020) who indicated that the addition of MOS with B-glucan had no significant effect on duck body weight and gain. Also, Kamel and Mohamed (2016) and Garipoglu *et al.* (2006) showed that the lowest value of final body weight and weight gain at six weeks of age found for checks fed basal diet with enzyme supplementation compared to control or other feed additives. Likewise, Rabie and Abo El-Maaty (2015) clarified that final live BW and BW gain of growing Japanese quails fed diet supplemented with Bio-Feed® Pro

enzyme were significantly depressed. The obtained results showed that no significant differences in feed intake among all groups. In contrast, Abd-El Hamed *et al.*, (2017) found that the highest total feed intake value was found for group treated with enzyme, while the lowest value was found for group treated with MOS. Also, Tufail *et al.* (2019) found significantly higher feed intake was recorded for group MOS-100g/kg feed during week 4th and 5th compared to control or other level of MOS.

Carcass and intestinal parameters:

In connection with carcass characteristics and gastrointestinal tract lengths, there were insignificant effects of different feed additives on dressed, liver, gizzard, heart and abdominal fat or lengths of intestinal tract (duodenum, jejunum, ileum, colon and cecum) in Tables 3 and 4, respectively. The present results in conformity with Algedawy *et al.* (2011) who concluded that weights of the gizzard, heart and liver showed no significant ($p < 0.05$) differences between the feed additive (Natuzyme or Biogen) supplemented groups and the control group. However, as current study, relative weight of the spleen was significantly greater ($p < 0.05$) in Biogen® supplemented birds than in Natuzyme® supplemented birds and the control group. Enhancement in spleen group 4 (Enz+ MOS) may be indicating higher immune competence for these feed additives for broilers. Also, Abdel-Fattah and Fararh, (2009) demonstrated a slight improvement in dressing percentage in birds fed diets supplemented with MOS compared to control birds in the line of current study. Similarly, there was no significant difference in the relative weights of carcass organs for broiler chicks fed on *Lactobacillus spp* (Awad *et al.*, 2009 and Zamanzad-Ghavidel *et al.*, 2011), or those fed on *Saccharomyces cerevisiae* (Celik *et al.*, 2007), compared to the control group. Likely, Abdel-Hafeez *et al.* (2017) and Rehman *et al.* (2020) reported that, prebiotic appeared to have insignificant influence on all carcass parameters of broilers. Sojoudi *et al.* (2012) demonstrated that broiler fed prebiotic at different levels showed no significant difference between treatments in inner organs weights except spleen weight, it has been significantly increased. However, the relative weight of liver and gizzard have been significantly increased by feeding prebiotic (Abdel-Hafeez *et al.* 2017). Biswas *et al.* (2021) included that better ($p < 0.05$) thigh and breast weights (% of live weight) were recorded in (0.2% MOS) group followed by (0.1% MOS) group as compared to control. Similar result has been recorded by Toghyani *et al.* (2011).

In this study, the improvement in breast weight or percentage for group 4 may be attributed to that prebiotic has a positive effect on muscle weight including improvement of calcium, phosphorus and magnesium absorption (Cummings and Macfarlane,

2002). The present findings of intestinal parts size are in contradiction with the findings of Asif, *et al.* (2022) who showed that MOS had significantly ($P<0.05$) affected the length of intestinal sections; duodenum, jejunum and ileum. Similar results were found by Padihari *et al.* (2014) and Castillo *et al.* (2008) they observed that addition of MOS to basal diet at a level of 0.5g/kg significantly increased the duodenum length and no effect on jejunum length as compared control group (Dimitroglou *et al.*, 2010; Padihari *et al.*, 2014 and Chand *et al.*, 2019).

Mortality, production index and economical return:

The current findings about mortality, production index, economic efficiency and relative economic (Table 5) concluded that the best economical measurements were of the share of group four followed by group three then group two compared to un-supplemented control group. This improvement could be due to improving BW and FCR. These results are similar to those reported by Jahan *et al.* (2006), Attia *et al.* (2014b), Mostafa *et al.* (2015) and El-Kelawy *et al.* (2017) who indicated that production index and economic efficiency increased with supplementing multienzyme and or other additives (as Bio-Mos) to the diets compared with control diets. Unlike, Kamel and Mohamed (2016) found that the lowest value of total revenue was found for the enzyme group compared to other groups.

Digestibility of nutrients:

Effect of different feed additives on digestibility coefficients of DM, OM, CP, CF, EE and NFE are presented in Table 7. The improvement in CP digestibility for group 4 in the current study may be mediated to improvement in blood serum TP and protease activity values (Table 8). The digestibility of basal diet is limited to about 70- 80% for broilers, which is mainly owing to the existence of insoluble non-starch polysaccharides (NSPs) in cell walls of corn and SBM (Ward, 2021), causing inhibition of nutrient digestion specialty crude fiber, these substances affect the growth performance of broiler chickens and are indigestible in poultry due to the absence of adequate endogenous activity of enzymes (Shang *et al.*, 2018). Therefore, from the current results, it has been concluded that supplementing exogenous enzymes targeting these indigestible compounds to improving the digestive enzymes to the cell-wall-encapsulated nutrients and better coefficient digestibility of starch (as CF%) in group 2. El-Kelawy *et al.* (2017) showed that *chicks* treated with enzymes addition followed by chicks treated with (enzyme + probiotic) had significantly greater digestibility of crude fiber than untreated chicks (control). Many previous studies investigated that

adding different types of commercial enzyme preparations with or without (prebiotic or probiotic) to broiler diets to achieve a positive impact on the digestibility of CP and crude fiber compared to those fed the control diets (Sherif, 2009a; Sherif, 2009b; Wang *et al.*, 2005 and Amerah *et al.*, 2017). While, exogenous enzyme supplementation had no significant effect on values of digestibility coefficient of EE, CF, CP, and NFE (Shalash *et al.*, 2009 and Sharifi *et al.* (2012)).

Chemical analysis of meat:

Concerning to chemical composition of meat, the previous data showed significant increase and decrease of protein and fat of breast and thigh meats respectively for additive groups compared to control group. Semi-similarly, Ibrahim *et al.* (2021) reported that a significant reduction ($P<0.05$) in fat content of broiler breast and thigh meat of the prebiotics and other feed additives treated groups compared to untreated group (Ooi and Liong, 2010; Weitkunat *et al.*, 2015; Ilham *et al.*, 2019; Okrathok and Khempaka, 2020; and Biswas *et al.*, 2021). While, there was a significant increase ($P<0.05$) in protein content of broiler breast meat of treated groups which has been reported by (Ibrahim *et al.*, 2021). The reduction in breast or thigh meat fat content may be related to the absorption process of bile acids and cholesterol from digestive tracts depending on level of soluble fiber in the ration of MOS (Ilham *et al.*, 2019).

Blood parameters:

Results obtained (Table 8) revealed that significant differences in Hct, TP, Alb., Hb, lipase, amylase and TAC values between supplemental groups and control. In the line of our study, Ismail *et al.* (2011) revealed that Hubbard broiler chicks fed either feed additives or Natuzyme® supplemented diets achieved significantly higher total protein and albumin levels which may be attributed to the favorable environment in the intestinal tract created by the feeding of these additives, in addition it might have helped to digest and absorb more nitrogen (Mohan *et al.*, 1995 and Panda *et al.*, 2006). Hemoglobin values increased in treated groups (four and three) than control group, this result may be due to addition of these additives might have stimulated iron absorption and activity of the hematopoietic organs and causes erythropoiesis, also high environmental temperature may have influenced the hematological parameters (Hasan *et al.*, 2015).

Value of RBCs ranged from (4.03 x 10⁶ / μ l) to (5.73 x10⁶ / μ l) for all groups. There were no significant differences among treatments with simple increase to MOS group. This result semi-agreed with Sosan *et al.* (2010) they concluded that there was a significant increase in erythrocyte count due to

prebiotic supplementation, while disagreed with Abeer and Soltan (2015) who found that value of RBCs decreased by prebiotic supplementation. Also, Chuka (2014) concluded that value of RBCs was lower in enzyme group compared with the control, while disagree with Rahman *et al.* (2013) they reported that value of RBCs increased by enzyme supplementation. The benefit effects of enzymes supplementation preparation that have additional benefit effects with MOS supplementation, these benefits when applied to diet, in addition to the protease component of the enzyme's combination may have a positive effect on trypsin inhibitor (Cowieson and Ravindran, 2008) and causes a greater activity of digestive protease in blood serum. The significant increase in TAC values for supplemented groups in comparison to control may be related to that supplementation. Improved quality and number of useful gut microfloramay have caused release of some bioactive substances that could potentially prevent oxidative damage with an increase in the activity of antioxidant response system (Tan *et al.*, 2010). On the other hand, Sohail *et al.* (2011) demonstrated that Supplementation of the MOS with or without probiotic to heat stressed broiler chicks caused a decrease in ($P < 0.05$) total antioxidant activity and concentrations compared to control.

CONCLUSION

The results of the this study supported the notion that the mixture of (frazyme + mannanoligosahharides) could have more potential value than the commercial enzyme (Frazyme®) or mannanoligosaccharide (Y- MOS) alone as an alternative dietary additive to improve productive performance of broilers. Therefore, it had positive effect on total return, net return, survival, and relative economic efficiency.

REFERENCES

- Abd-El Hamed A.M., E.R. Kamel, M.E. Abo-Salem and S.T. Atallah, 2017. Comparative study on the effect of organic acids, prebiotics and enzymes supplementation on broiler chicks 'economic and productive efficiency. Benha Journal of Applied Sciences (BJAS) Vol. (2), I. (2), 1-8.
- Abdel-Fattah, F.A. and K.M. Fararh, 2009. Effect of dietary supplementation of probiotic, prebiotic and synbiotic on performance, carcass characteristics, blood picture and some biochemical parameters in broiler chickens. Benha Veterinary Medicine Journal, 20, 9-23.
- Abdel-Hafeez, H.M., E.S. Saleh, S.S. Tawfeek, I.M. Youssef, and A.S. Abdel-Daim, 2017. Effects of probiotic, prebiotic, and synbiotic with and without feed restriction on performance, hematological indices and carcass characteristics of broiler chickens. Asian-Australasian journal of Animal Sciences, 30 (5), 672.
- Abdollahi, M.R., V. Ravindran, and A.M. Amerah, 2016. Influence of partial replacement of ground wheat with whole wheat and exogenous enzyme supplementation on growth performance, nutrient digestibility and energy utilization in young broilers. Journal of Animal Physiology and Animal Nutrition, 100, 929–937.
- Abeer M.E., and M. A. Soltan, 2015. Effect of dietary probiotic and/or prebiotic supplementation on growth performance, carcass traits and some serum biochemical alterations in broiler chicken. Animal Science Advanced Journal. 5, 1480-1492.
- Abudabos A.M., A.H. Alyemni, Y.M. Dafalla, and R.U. Khan, 2017. Effect of organic acid blend and *Bacillus subtilis* alone or in combination on growth traits, blood biochemical and antioxidant status in broiler exposed to *Salmonella typhimurium* challenge during the starter phase. Journal of Applied Animal Researches, 45: 538-542.
- Akoy R.A.M., 2015. The effects of probiotics, prebiotics and synbiotics on gut flora, immune function and blood characteristics of broilers. A thesis submitted to the University of Plymouth in partial fulfilment for the degree of Plymouth in partial fulfillment for the degree of doctor of philosophy. School of Biological Sciences Faculty of Science and Engineering.
- Algedawy, S. A., K. S. Magdy, I. M. Elashmawy and Y. S. El-Sayed, 2011, Effect of commercial probiotic and exogenous enzymes on the growth performance, immune responses, and hemato-biochemical parameters of broiler chickens. Foreign Agricultural Relations (FAR), pp. 430 – 445.
- Alhidary I.A., Z. Rehman, R.U. Khan and M. Tahir, 2017. Anti-aflatoxin activities of milk thistle (*Silybum marianum*) in broiler. World's Poultry Science Journal, 73: 559-566.
- Alqhtani A. H, A. R. Al Sulaiman, A. S. Alharthi and A. M. Abudabos, 2022. Effect of exogenous enzymes cocktail on performance, carcass traits, biochemical metabolites, intestinal morphology, and nutrient digestibility of broilers fed normal and low-energy corn–soybean diets. Animals Journal, 12, 1094.
- Altaf-ur-Rahman, M. Ali, S. Sultan and N. Ahmed, 2007. Effect of dietary probiotic supplementation on performance of broilers. Sarhad Agriculture Journal, Pakistan, 23(2):481-484.
- Amerah, A. M., L. F. Romero, A. Awati and V. Ravindran, 2017. Effect of exogenous xylanase, amylase, and protease as single or combined activities on nutrient digestibility and growth

- performance of broilers fed corn/soy diets. *Poultry Science*, 96:807–816.
- Ao, X., J. S. Yoo, T. X. Zhou, J. P. Wang, Q. W. Meng, L. Yan, J. H. Cho and I. H. Kim, 2011a. Effects of fermented garlic powder supplementation on growth performance, blood profiles and breast meat quality in broilers. *Livestock Science*. (10),10- 16.
- Ao, X., T. X. Zhou, H. J. Kim, S. M. Hong and I. H. Kim, 2011b. Influence of fermented red ginseng extract on broilers and laying hens. *Asian-Australians Animal Science Journal*, (24): 993-1000.
- AOAC, 2010. Association of Official Analytical Chemists, Official methods of analysis of the AOAC international (18th ed.).
- Asif, M., Z. Hayat, A. Rahman, M. F. Qamar, S. Nawa, M. Ijaz, I.H. Badar, A.R. Asif and M. K. Yar ,2022. Effects of mannan-oligosaccharide supplementation on gut health, immunity, and production performance of broilers. *Brazilian Journal of Biology*. vol. 84.
- Attia, Y. A., A. E. Abd Al-Hamid, M. S. Ibrahim, M. A. Al-Harhi, F. Bovera and A. Sh. Elnaggar, 2014a. Productive performance, biochemical and hematological traits of broiler chickens supplemented with propolis, bee pollen, and mannan oligosaccharides continuously or intermittently. *Livestock Science*, (03),005i.
- Attia, Y. A., W. S. El-Tahawy, A. E. Abd El-Hamid, A. Nizza, F. Bovera, M. A. Al-Harhi and M. I. El-Kelaway, 2014 b. Effect of feed form, pellet diameter and enzymes supplementation on growth performance and nutrient digestibility of broiler during days 21–37 of age. *Archives Animal Breeding*. 57:1–11.
- Awad, W. A., K. Ghareeb, S. Abdel-Raheem and J. Bohm, 2009. Effects of dietary inclusion of probiotic and synbiotic on growth performance, organ weights, and intestinal histomorphology of broiler chickens. *Poultry Science*. vol. (88): 49-56.
- Baurhoo, B., L. Phillip and C. A. Ruiz-Feria, 2007. Effects of purified lignin and mannan oligosaccharides on intestinal integrity and microbial populations in the ceca and litter of broiler chickens. *Poultry Science*. 86: 1070-1078
- Biswas, A., N. Mohan, k. Dev, M. N. Akbar and A.K. Tiwari, 2021. Effect of dietary mannan oligosaccharides and fructo-oligosaccharides on physico-chemical indices, antioxidant and oxidative stability of broiler chicken meat. *Avian Nutrition and Feed Technology Division, Scientific Reports*, 11:20567.
- Castillo, M., S. Martin-Orue, J. Taylor-Pickard, J. Perez and J. Gasa, 2008. Use of mannanoligosaccharides and zinc chelate as growth promoters and diarrhea preventative in weaning pigs: effects on microbiota and gut function. *Journal of Animal Science*, vol. 86, no. 1, pp. 94-101.
- Celik, K., M. Mutluay and A. Uzatici, 2007. Effects of probiotic and organic acid on performance and organ weights in broiler chicks. *Archiva Zootechnica*. vol. (10): 51- 56.
- Chand, N., H. Faheem, R.U. Khan, M.S. Qureshi, I.A. Alhidary and A.M. Abudabos, 2016. Anticoccidial effect of mannanoligosaccharide against experimentally induced coccidiosis in broiler. *Environmental Science and Pollution Research*, 23: 14414-14421.
- Chand, N., S. Shamsullah, R. Rafiullah, R.U. Khan, M. Mobashar, S. Naz, I. Rowghani and M.A. Khan, 2018. Mannanoligosaccharide (MOS) in broiler ration during the starter phase: 1. Growth performance and intestinal histomorphology. *Pakistan Journal of Zoology*, (In press).
- Chand, N., S. Shamsullah, R. Rafiullah, R.U. Khan, M. Mobashar, S. Naz, I. Rowghani and M.A. Khan, 2019. Mannanoligosaccharide (MOS) in Broiler Ration during the Starter Phase: 1. Growth performance and intestinal histomorphology. *Pakistan Journal of Zoology*, vol. 51, no. 1, pp. 173-176.
- Chuka, E., 2014. Comparative study of the effects of probiotic and commercial enzyme on growth rate, haematology and serum biochemistry of broiler chicken. *Journal of Food Process Technology*, 5:9.
- Çınar, M., A.U. Çatlı, K. Küçükyılmaz and M. Bozkurt, 2009. The effect of single or combined dietary supplementation of prebiotics, organic acid and probiotics on performance and slaughter characteristics of broilers. *South African Journal of Animal Science*, 39(3).
- Cowieson, A. J., and V. Ravindran, 2008. Effects of exogenous enzymes in maize-based diets varying in nutrient density for young broilers; growth performance and digestibility of energy, minerals and amino acids. *British Poultry Science*. 49:34-44.
- Cummings, J. H., and G. T. Macfarlane, 2002. Gastrointestinal effects of prebiotics. *British Journal of Nutrition*. 87S, 145–151.
- Diarra, M. S., G. S. Fred, D. Fatoumata, P. Jane, M. Luke, B. Roland, B. Claudie, D. Pascal, B. Susan, J. S. Brent and T. Edward, 2007. Impact of feed supplementation with anti-microbial agents on growth performance of broiler chickens, clostridium perfringens and enterococcus counts, and antibiotic resistance phenotypes and distribution of antimicrobial resistance determinants in *Escherichia coli* isolates.

- American Society for Microbiology, 73:6566-6576.
- Dimitroglou, A., D.L. Merrifield, P. Spring, J. Sweetman, R. Moate, and S. J. Davies, 2010. Effects of mannan oligosaccharide (MOS) supplementation on growth performance, feed utilisation, intestinal histology and gut microbiota of gilthead sea bream (*Sparus aurata*). *Aquaculture (Amsterdam, Netherlands)*, vol. 300, no. 1-4, pp. 182-188.
- Dizaji, B.R., A. Zakeri, A. Golbazfarsad, S. Faramarzy, O. Ranjbari, 2013. Influences of different growth promoters on intestinal morphology of broiler chickens. *European Journal of Experimental Biology*. 3(2):32-37.
- Duncan, D. B., 1955. The multiple ranges and multiple F Test. *Biometrics*.11: 1-42.
- El-Kelawy, M. I., A. S. El-Shafey and M. A. Reham, 2017. Impact of dietary supplementation with multi-enzyme and/or probiotics on productive performance and nutrients digestibility of broiler chickens. *Egyptian Journal of Nutrition and Feeds*, 20(3): 535-543.
- Emara, F. A. H., 2009. Effect of some feed additives on economic and productive efficiency of broilers. Master degree, Faculty of Veterinary Medicine, Zagazig University Egypt.
- Ferket, P.R., A.A. Santos and E.O. Oviedo-rondón, 2005. Dietary factors that affect gut health and pathogen colonization. In *Proceedings of the 32nd Annual Carolina Poultry Nutrition Conference*, Raleigh, NC, USA.
- Fernandes, B.C.S., M.R.F.B Martins, A.A. Mendes, E. L. Milbradt, C. Sanfelice, B. B. Martins, E. F. Aguiar, C. Bresne, 2014. Intestinal integrity and performance of broiler chickens fed a probiotic, a prebiotic, or an organic acid. *Brazilian journal of Poultry Science*. (4). 417-424
- Garipoglu, A.V., B.Z. Saricicek, U. Kilic, 2006. Effects of the commercial enzyme supplementation to the rations on broiler performance. *Asian Journal of Animal and Veterinary Advances*. 1:42-48.
- Ghazalah, A.A., A.M. Atta, Kout Elkloub, M. E. L. Moustafa, F. H. Riry, 2011. Effect of dietary supplementation of organic acids on performance, nutrients digestibility and health of broiler chicks. *International Journal of Poultry Science*. 10 (3): 176-184.
- Habib, A. B., A. M. Eltrefi, A. A. Mohamed, E. S. Abu shulukh, A. A. Abubaker, H. H. Abdelwahid, A. I. Ghada, M.A. Nafisa and E. O. Basheer, 2017. Impact of different dietary supplementation with (Y-MOS) yeast on performance and carcass characteristics of broiler chickens. *International Journal of Multidisciplinary and Current Research*, Vol; 5, 1337- 1341
- Hasan, S., M. M. Hossain and M. E. R. Bhuiyan, 2015. growth performance and hemato-biochemical parameters in broiler during high environmental temperature by the application of synbiotic. *international journal of Innovation and Scientific Rese* 13, 157-162.
- Hooge, D. M. P., K. M. McBride and P. J. Rigolin, 2010. Metaanalysis of broiler chicken trials using diets with or without Allzyme SSF enzyme complex. *International Journal Poultry Science*, Vol.9, pp.819-823.
- Ibrahim, N. S. K., A. N. S. Ahmed and A. E. S. Ghada, 2021. Impact of dietary supplementation of prebiotic on the growth performance and immunity in broilers fed low protein diets. *Assiut Veterinary Medicine Journal*, Vol. 67, 103-119
- Iji, P.A., A.A. Saki and D. R. Tivey, 2001. Intestinal structure and function of broiler chickens on diets supplemented with a mannan oligosaccharide. *Science of Food and Agriculture*, 81: 1186-1192.
- Ilham, I., N. Nurliana, M. Delima, S. Sugito and S. Samadi, 2019. Administration of various feed additives on cholesterol content of meat and fat abdomen of local chicken (*Gallus domesticus*). *Animal Production journal*, 20(1), 45-52.
- Ismail, F. S. A., H. M. A. Abo El-Maaty and A. A. M. El-Sayed, 2011. Performance of broiler chicks affected by dietary supplementation with commercial enzymes or probiotics. *Journal of Animal and Poultry Production*, Mansoura University, vol. (2): 23 -32.
- Jahan, M. S., M. Asaduzzaman and A. K. Sarkar, 2006. Performance of broiler fed on mash, pellet and crumble. *International Journal of Poultry Science*, 5(3): 265–270.
- Jakobsen, P. E., K. Gertov and S.H. Nilsen, 1960. Frdjlighed frogmed fierbrae. "Digestibility traits with poultry". Bereting fra for sogslabortoret, Kabenhaven, 56: 1-34.
- Kamel, R. E., and S. L. Mohamed, 2016. Effect of dietary supplementation of probiotics, prebiotics, synbiotics, organic acids and enzymes on productive and economic efficiency of broiler chicks. *Alexandria Journal of Veterinary Sciences*, 50 (1): 8-17
- Karimi, T. M. A., A. R. Moghaddam, S. Rahimi and N. Mojangani, 2010. Assessing the effect of administering probiotics in water or as a feed supplement on broiler performance and immune response. *British Poultry Science* vol. (51): 178-184.
- Kermanshahi, H. A. R.H., 2006. Influence of supplemental dried whey on broiler performance and cecal flora. *International Journal of Poultry Science*, Vol.5, pp.538-543.
- Koc, F., H. Samli, A. Okur, M. Ozduven, H. Akyurek and N. Enkoylu, 2010. Effects of saccharomyces cerevisiae and/or manna

- oligosaccharide on performance, blood parameters and intestinal microbiota of broiler chicks. *Bulgarian Journal of Agricultural Science* 16: 643- 650.
- Luo, F., F. Yang, X. Yang, J. Yao, B. Shi and Z. Zhou, 2009. Effects of xylanase on performance, blood parameters, intestinal morphology, microflora and digestive enzyme activities of broilers fed wheat-based diets. *Asian-Australian Journal of Animal Science*, Vol.22, pp.1288-1295.
- Mahmoud, U.T., R. Hassan, O. Amen, G. Abdel-Raheem and D. Al-Rammamneh, 2020. Effect of prebiotic on the behavioural patterns, performance, carcass characteristics, antibody titer and some blood parameters of mule ducks. *SVU-International Journal of Veterinary Sciences*, 3(1), 27-38.
- Marwa, I. A., 2013. Use of biotic preparations as an alternative to antibiotics in broiler chickens under semifield and field conditions. Master Degree in Veterinary Medicine Science (Poultry Diseases) Benha University.
- Midilli, M. A. M., N. H. Kocabagli, O. H. Muglali, N. Turan, H. Yilmaz and S. Cakir, 2008. Effect of dietary probiotic and prebiotic supplementation on growth, performance and serum IgG concentration of broilers. *South African Journal of Animal Science*, Vol.38, pp.21-27.
- Mohamed, L.S., 2016. Effect of some dietary supplementation on economic and productive efficiency of broiler chickens. Master Degree in Faculty of Veterinary Medicine. Benha University.
- Mohamed, S. A., and A. M. Mukhtar, 2016. Effect of commercial (Y-MOS) yeast on performance and carcass characteristics of broiler chicks. *World Journal of Pharmacy and Pharmaceutical Sciences*. 6, 168-179.
- Mohan, B., R. Kadirvel, M. Bhaskaran and A. Natarajan, 1995. Effect of probiotic supplementation on serum/yolk cholesterol and on egg shell thickness in layers. *British Poultry Science* vol. (36): 799-803.
- Mostafa, M. M. E., H. A. Thabet and M. A. M. Abdelaziz, 2015. Effect of Bio-Mos utilization in broiler chick diets on performance, microbial and histological alteration of small intestine and economic efficiency. *Asianan Journal of Animal and Veterinary Advanced*, 10: 323-334.
- North, M.O. ,1981. Commercial chicken. Production Annual. 2nd Edition, AV., publishing company I.N.C., West post Connecticut, USA.
- NRC, 1994. Nutrient requirements of poultry, 9th Edition, National Academy of Sciences-National Research Council, Washington, D.C.
- Okmathok, S., and S. Khempaka, 2020. Modified-dietary fiber from cassava pulp reduces abdominal fat and meat cholesterol contents without affecting growth performance of broiler chickens. *Journal of Applied Poultry Research*, 29(1), 229-239.
- Olukosi, O. A., L. A. Beeson, K. Englyst and L. F. Romero, 2015. Effects of exogenous proteases without or with carbohydrases on nutrient digestibility and disappearance of non-starch polysaccharides in broiler chickens. *Poultry Science*, 94, 2662–2669.
- Ooi, L.G., and M.T. Liong, 2010, Cholesterol-lowering effects of probiotics and prebiotics: a review of in vivo and in vitro findings. *International journal of molecular sciences*, 11(6), 2499-2522.
- Padihari, V. P., S. P. Tiwari, T. Sahu, M. K. Gendley and S. K. Naik, 2014. Effects of mannan oligosaccharide and *Saccharomyces cerevisiae* on gut morphology of broiler chickens. *Journal of World's Poultry Research*, vol. 4, no. 3, pp. 56-59.
- Panda, A. K., V. R. Savaram, V.L.N. Mantena and S. R. Sharma, 2006. Dietary supplementation of *Lactobacillus sporogenes* on performance and serum biochemicolipid profile of broiler chickens. *Journal of Poultry Science* vol. (43): 235-240.
- Patterson, J. A., and K. M. Burkholder, 2003. Application of prebiotics and probiotics in poultry production. *Poultry Science*, 82:627-631.
- Plessis, Du R. E., 2014. Carbohydrase and protease supplementation increased performance of broilers fed maize-soybean-based diets with restricted metabolizable energy content. *South African Journal of Animal Science*, Vol.44.
- Rabie, M. H., and H. M. A. Abo El-Maaty, 2015. Growth performance of japanese quail as affected by dietary protein level and enzyme supplementation. *Asian Journal of Animal Veterinary Advanced*, 10: 74-85.
- Rahman, M. S., A. Mustari, M. Salauddin and M. M.Rahman, 2013. Effects of probiotics and enzymes on growth performance and haematobiochemical parameters in broilers. *Bangladesh Agriculture University*. 11, 111-118.
- Rehman, A., M. Arif, N. Sajjad, M.Q. Al-Ghadi, M. Alagawany, M.E. Abd El-Hack and A. A. Swelum, 2020. Dietary effect of probiotics and prebiotics on broiler performance, carcass, and immunity. *Poultry Science*, 99(12), 6946-6953.
- Romero, L. F., M. J. Zuidhof, S. R. Jeffrey, A. Naeima, R. Renema and F. E. Robinson, 2010. A data envelope analysis to asses factors affecting technical and economic efficiency of individual broiler breeder hens. *Poultry science*, 89:1769-77.
- Salehimanesh, A., M. Mohammadi and M. A. Roostaei, 2016. Effect of dietary probiotic,

- prebiotic and synbiotic supplementation on performance, immune responses, intestinal morphology and bacterial populations in broilers. *Journal of Animal physiology and Animal Nutrition*, 100(4), 694-700.
- Santoso, U., K. Tanaka, S. Ohtani and M. Sakaida, 2001. Effect of fermented product from *Bacillus subtilis* on feed efficiency, lipid accumulation and ammonia production in broiler chicks. *Asian-Australians Journal of Animal Science*, vol. (14): 333-337
- Shalash, S.M., M.A. Sayed, E. E. Hoda, R. A. Nehad and M. S. Manal, 2009. Nutritive value of distillers dried grains with soluble and broiler performance at starter period. *International journal of Poultry Science*, 8: 783-787.
- Shang, Q. H., X. K. Ma, M. Li, L. H. Zhang, J. X. Hu and X.S. Piao, 2018. Effects of galactosidase supplementation on nutrient digestibility, growth performance, intestinal morphology and digestive enzyme activities in weaned piglets. *Animal and Feed Science Technology*, 236, 48–56.
- Sharifi, S.D., F. Shariatmadari and A. Yaghobfar, 2012. Effects of inclusion of hull-less barley and enzyme supplementation of broiler diets on growth performance, nutrient digestion and dietary metabolisable energy content. *Journal Central European Agriculture*, 13: 193-207.
- Sherif, Kh. El., 2009a. Performance of broiler chicks fed plant protein diets supplemented with commercial enzymes. *Agriculture Science, Mansoura University*, 34 (4): 2819-2834.
- Sherif, Kh. El., 2009b. Effect of using probiotics and enzymes with plant protein diets in broiler performance. *Agriculture Science, Mansoura University*, 34 (5): 4493-4505.
- Sohail, M. U., Z. U. Rahman, A. Ijaz, M. S. Yousaf, K. Ashraf, T. Yaqub, H. Zaneb, H. Anwar and H. Rehman, 2011. Single or combined effects of mannan-oligosaccharides and probiotic supplements on the total oxidants, total antioxidants, enzymatic antioxidants, liver enzymes, and serum trace minerals in cyclic heat-stressed broilers. *Poultry Science* 90 :2573–2577.
- Sojoudi, M. R., M. Dadashbeiki and M. Bouyeh, 2012. Effect of different levels of prebiotics TechnoMos on carcass characteristics of broiler chickens. *Basic Applied Science Researches*, 2(7), 6778-6794.
- Sosan, R. A. I., H. M. Safaa, R. M. Fatma, S. S. Salwa and A. E. M. Hanan, 2010. Influence of probiotic, prebiotic and/or yeast supplementation in broiler diets on the productivity, immune response and slaughter traits. *Journal of Animal and Poultry Production*. 1: 45-60.
- Statistical analyses system, (SAS), 2002. SAS Institute Inc., SAS/STAT software, version 9. SAS Institute, Inc., Cary, NC, USA.
- Stefanello, C., S. L.Vieira, G. O. Santiago, L. Kindlein, J. O. B. Sorbara and A. J. Cowieson, 2015. Starch digestibility, energy utilization, and growth performance of broilers fed corn-soybean basal diets supplemented with enzymes. *Poultry Science*, 94, 2472–2479
- Sultan, A., U. Imran, S. Khan, R. Ullah, H. Khan, N. A. Khan and R. U. Khan, 2015. Effect of yeast derived carbohydrate fraction on growth performance, apparent metabolizable energy, mineral retention and gut histomorphology of broilers during starter phase. *Pakistan Veterinary Journal*, 35: 409- 413.
- Tan, G. Y., L. Y. Yang, Q. Fu, J.H. Feng and M. H. Zhang, 2010. Effects of different acute high ambient temperatures on function of hepatic mitochondrial respiration, antioxidative enzymes, and oxidative injury in broiler chickens. *Poultry Science*, 89:115–122.
- Tavaniello, S., 2018. Prebiotics offered to broiler chicken exert positive effect on meat quality traits irrespective of delivery route. *Poultry Science*. 97, 2979–2987.
- Toghyani, M., M. Toghyani, and S. A. Tabeidian, 2011. Effect of probiotic and prebiotic as antibiotic growth promoter substitutions on productive and carcass traits of broiler chicks. *International Conference on Food Engineering and Biotechnology*. 9, 82–96.
- Tufail, M., N. Chand, R. A. Shakoor, U. R. Khan, M. Mobashar and N. Shabana, 2019. Mannanoligosaccharide (MOS) in broiler diet during the finisher phase: 2. growth traits and intestinal histomorphology. *Pakistan Zoology*, (51);2 pp; 597-602.
- Waheed, A., and I. Eltaieb, 2005. Inclusion of some wastes in rabbit diets. Department of Animal Production, Faculty of Agriculture, Alazhar University. pp 14-26.
- Wang, Z.R., S.Y. Oiao, W.O. Lu and D.F. Li, 2005. Effects of enzyme supplementation on performance nutrient digestibility gastrointestinal morphology and volatile fatty acid profiles in the hindgut of broilers fed wheat-based diets. *Poultry Science*, 84: 875-881.
- Ward, N.E., 2021. Debranching enzymes in corn/soybean meal-based poultry feeds: A review, *Poultry Science*, 100, 765–775.
- Weitkunat, K., S. Schumann, K. J. Petzke, M. Blaut, G. Loh and S. Klaus, 2015. Effects of dietary inulin on bacterial growth, short-chain fatty acid production and hepatic lipid metabolism in gnotobiotic mice. *The Journal of Nutritional Biochemistry*, 26(9), 929-937.
- Xu, Z. R., C. H. Hu, M. S. Xia, X. A. Zhan and M.Q. Wang, 2003. Effects of dietary fructo oligosaccharide on digestive enzyme activities,

- intestinal microflora and morphology of male broilers. Poultry Science 82:1030-1036.
- Yang, Y., P. A. Iji and M. Choct, 2009. Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. World's Poultry Science Journal, 65(1):97-114.
- Zamanzad-Ghavidel, S., K. Z. Adl, N. M. Sis, S. Aharizad, A. Mirzaei-aghsaghali, M. Mohammadian and S. A. Siadati, 2011. Effects of lactobacillus-based probiotic on growth performance, mortality rate and carcass yield in broiler chickens. Animal Biology Researches. vol. (2):325-331.

تأثير إضافة الإنزيمات التجارية مع أو بدون سكريات المنان أوليجو على الدلالات الإنتاجية، الغذائية والتمثيلية لبدارى التسمين

مها أحمد عبد اللطيف عمر

قسم الإنتاج الحيواني والداجنى، كلية الزراعة، جامعة المنيا، المنيا، مصر

أجريت هذه التجربة لاستخدام بعض الإضافات الغذائية كمنشطات للنمو لكتاكيت الساسو وتقييم تأثيرها على الأداء الإنتاجى، معاملات هضم العناصر الغذائية، استجابات الدم وكذلك بعض مقاييس الذبيحة والقناة الهضمية والعائد الإقتصادى، تم استخدام ١٢٠ ككتوت ساسو غير مجنس عمر يوم بمتوسط وزن (0.65 ± 40.65) جم تم توزيعها عشوائياً وتقسيمهم لأربع مجموعات (كل مجموعة ٣٠ ككتوت / ١٠ ككتاكيت فى كل مكررة × ٣ مكررات). المجموعة الأولى تم اعتبارها مجموعة (المقارنة الكنترول). المجموعات الثانية والثالثة والرابعة تم اعطائها عليقة الكنترول المضاف إليها: ١ جم من خليط الإنزيمات التجارية (فرازيم / ١ كجم عليقة، ٢,٥ جم من سكر المنان) موس/كجم عليقة ١,٠ جم من فرازيم مع ٢,٥ جم من الموس/كجم عليقة على التوالى. تمت التربية لمدة ٥٨ يوم وتم أخذ القياسات الخاصة بالتجربة. أوضحت النتائج المتحصل عليها:

١. من حيث الأداء الإنتاجى: أعطت المجموعة الرابعة (نزيمات+ موس) أفضل قيم معنوية من وزن الجسم الحى، الزيادة فى الوزن، ومعامل التحويل الغذائى بينما لم توجد أى اختلافات معنوية فى معدل استهلاك العليقة مقارنة بباقى المجموعات.
٢. سجلت معاملات هضم البروتين الخام والالياف الخام أعلى قيم معنوية لكل من المجموعة الرابعة والثانية على التوالى.
٣. من حيث مواصفات الذبيحة: تبين أن أكبر محصول لأوزان الصدر والطحال كان من نصيب المجموعة الرابعة، بينما لم تظهر اختلافات معنوية لأوزان كل من الفخذ، الكبد، القونصة، القلب، دهون البطن أو نسبة التصافى بين المجموعات الأربعة.
٤. قياسات أطوال أجزاء القناة الهضمية: لم تظهر الإضافات الغذائية المختلفة للعليقة الكنترول أى فروق معنوية بين جميع المجموعات.
٥. قياسات الدم: سجلت المجموعة الثانية (مجموعة الإنزيمات) زيادة معنوية فى كل من قيم الهيماتوكريت، وإنزيمات الليباز والاميليز الهاضمة. بينما سجلت المجموعة الرابعة (الخليط) زيادة معنوية فى قيم البروتين الكلى، الألبومين، الهيموجلوبين وكذلك نشاط مضادات الأكسدة الكلية.
٦. التركيب الكيمياءى للحم: أعلى محتوى معنوى للبروتين فى لحم الصدر تم تسجيله للمجموعة الرابعة. بينما أعلى محتوى للدهن فى لحم الفخذ سجل لمجموعة الكنترول، وسجلت قيمة الرماد فى لحم الفخذ أيضاً زيادة معنوية لطبوع المجموعة الرابعة.
٧. أفضل قيم لمقاييس الكفاءة الإقتصادية تم تسجيلها للمجموعة الرابعة، يليها المجموعة الثالثة ثم المجموعة الثانية وذلك مقارنة بمجموعة الكنترول.

من خلال النتائج السابقة يمكن القول أن: هذه الدراسة توصى باستخدام كل من الإضافات السابقة كمنشطات للنمو دون أى تأثير سلبى على بدارى التسمين حيث أثبتت الدراسة أن استخدام (خليط الإنزيمات التجارية مع الموس) أفضل تأثيراً على الأداء الإنتاجى، والكفاءة الإقتصادية من استخدام كل مادة على حدة لبدارى تسمين الدجاج الساسو.