# **Journal of Current Veterinary Research**



ISSN: 2636-4026

Journal homepage: http://www.jcvr.journals.ekb.eg

**Pharmacology** 

#### Effect of Sodium Butyrate and *Origanium Vulgare* on Growth Performance, Biochemical Profile, Immune Status and Carcass Traits of Broiler Chickens

Hend Kamal Maarek<sup>1</sup>, Taha A. Attia<sup>1</sup>, Mostafa A. Shalaby<sup>2</sup> and Saber A. Elhanbally<sup>1\*</sup>

(1) Pharmacology Department, Faculty of Veterinary Medicine, Sadat City University, Egypt.
(2) Pharmacology Department, Faculty of Veterinary Medicine, Cairo University, Egypt.

\*Corresponding author: saber.ahmed@vet.usc.edu.eg Received: 2/8/2023 Accepted: 13/8/2023.

#### ABSTRACT

The principal goal of this research was to examine how the addition of sodium butyrate (SB) and Origanium vulgare (OV) to basal diet affects various aspects of broiler chicken production, including growth rate, biochemical markers, immune system function, and carcass traits. Five hundred one-day-old chicks of Cobb breed (males and females) were distributed at random way into five dietary treatment groups on floor pens, each group have 4 replicates (25 birds/replicate) and reared at a private farm. Group (G1) received basal diet without any addition (negative control). Birds of groups 2, 3, 4 and 5 received basal diet enriched with 500 g/ton sodium butyrate (SB), 500 g/ton Origanium vulgare (OV), 250 g/ton SB+OV and 500 g/ton SB+OV, respectively. At day 35 of rearing, the growth performance, biochemical profile, immunity parameters and carcass traits were evaluated. Our study demonstrated that supplementing basal diet with SB and OV, alone and in combination, resulted in improvements in body weight gain and feed conversion ratio and decreased levels of alanine aminotransferase (ALT), aspartate aminotransferase (AST), total cholesterol (TC), triglycerides (TG) and malondialdehyde (MDA) but increased total protein (TP), albumin, globulin, albumin/globulin ratio, superoxide dismutase (SOD) and catalase (CAT). It also increased levels of IgG and IgM and enhanced immunity as it elevated phagocytic activity, phagocytic index, lysozyme activity, and hemagglutination inhibition (HI) antibody titer against NDV. It improved carcass traits of broilers as it increased live weight and carcass weight and dressing percent (72.5%). It could be concluded that supplementation of basal diet with SB and OV, alone and in combination, improved growth performance, biochemical parameters, immune status, and carcass traits of broilers. Therefore, the utilization of sodium butyrate and Origanium vulgare as feed additives can be used as an effective and beneficial approach in broiler chicken production. Both have hepatoprotective, antioxidant, and immunostimulant effects. The mechanisms of action underlying these effects require further study in broiler chickens.

**Keywords:** Biochemical parameters, Carcass traits, Growth performance, Immune status, *Origanium vulgare* and Sodium butyrate.

#### **INTRODUCTION**

Egypt is suffering from severe shortage of poultry meat, which is a popular food for

most Egyptians. It is estimated that Egypt's chicken meat production will be approximately 1.59 million tons in the year 2023 (Abdelli et al., 2021). The cost of feed ingredients as cereal grains (maize, sorghum, and barley), tends to increase because of fluctuations in prices and the difficulties of importation. Efforts to enhance the health benefits of chickens remain an ongoing concern in the poultry production sector. Antibiotics have been used as a traditional tool for promoting health and growth performance in poultry. Nevertheless, the risk of bacterial resistance development and the incidence of drug residue have increased over time, leading to concerns about their indiscriminate use. As a result, prophylactic and therapeutic uses of antibiotics in poultry nutrition have been restricted (Ricke *et al.*, 2020). Feed additives are utilized in poultry nutrition to serve a variety of purposes ranging from enhancing the quality and safety of feed, as well as enhancing the quality of animalderived food byproducts. The most widelyused feed additives for poultry diets comprise antimicrobial (Alagawany et al., 2021), acidifiers (Ricke et al., 2020), antioxidants (Hashemi and Davoodi, 2011), antimycotoxin (Olivera et al., 2015 and Adhikari, 2018), prebiotics, probiotics, and phytogenic additives (Abd El-Hack et al., 2017; AL-Khalaifah, 2018; Debnath et al., 2019 and Fathi et al., 2023).

As a safer alternative to antibiotics. SB has been receiving an increasing interest within the field of poultry industry. Sodium butyrate is a crucial substitute for antibiotics in the poultry industry due to its stable and non-odorous properties. Furthermore, it has been found to possess antimicrobial, antiinflammatory, and antioxidant characteristics, making it a beneficial feed additive for maintaining optimal animal health (Song et al., 2017). By reducing oxidative stress triggered by heat stress, SB has a potential to enhance meat quality in poultry (Lan et al., 2020). The previous study specified that incorporating SB into poultry feed can be used as a beneficial approach to enhance liver function, broiler growth performance, and meat quality, particularly in hot climatic conditions.

Origanium vulgare Lin., (Oregano), is an aromatic herb with many bioactive chemical compounds (Ri et al., 2017). Oregano possesses antibacterial, antioxidant, antiviral. immunomodulatory. and antiparasitic properties (Alagawany et al., 2018). Incorporating oregano extracts into chicken diets has been shown to provide several benefits, including increased body weight and feed conversion ratio, improved digestion, decreased disease incidence. improved productive performance, and lowered economical loss (Alagawany et al., 2018). Nevertheless, more researches on oregano as a phytogenic feed additive in poultry is essential to determine its influence on lipid metabolism, meat quality, carcass yield, and blood parameters. The goal of the current research was to assess the impact of both sodium butyrate and Origanium growth performance, vulgare on biochemical profile, immune status and carcass traits in broiler chickens.

# MATERIAL AND METHODS *Ethical approval:*

The current study was accomplished with approval from the Institutional Animal Care and Use Committee (IACUC), Faculty of Veterinary Medicine, Cairo, University, Egypt.

# Feed additives:

Sodium butyrate (SB) trade name is CM3000®. It is a commercial 30% spherical granules coated sodium butyrate. It is released at a slow and steady manner throughout the small and large intestines of poultry. The manufacturer of CM3000® is Hangzhou King Techina Feed Co., Ltd, China. It was introduced to the broiler basal diet at a 500 g/ton feed concentration (Sikandar *et al.*, 2017). *Origanium vulgare* L., (OV), the trade name Oregano is Ropadiar<sup>®</sup>, which is a natural and popular herb used in traditional medicine and as a culinary spice, which is beneficial for poultry for boosting immunity. *Origanium vulgare* L is manufactured by Ropapharm International Co., Ltd, the Netherlands. It was introduced to the broiler basal diet at a 500 g/ton feed concentration (Zhang *et al.*, 2021).

#### Experimental chicks:

Five hundred 1-day-old chicks (Cobb breed) of males and females were provided from a local hatchery. After weighting, the chicks were assigned randomly into five groups each of 4 replicates (25 birds/replicate). The experiment was conducted on floor pens at a at private farm Giza, Egypt. The administered vaccination program to all experimental groups of birds included protection against Infectious Bronchitis (IB), Newcastle Disease (ND), and Gumboro (IBD) diseases.

#### Diets and feeding program:

To meet the nutrient requirements of Cobb broilers, diets comprising corn-soybean meal and basal components were prepared (Cobb manual catalogue 2018). The experimental period was 35 days involved providing access to feed in the form of mash-type diets for three stages - starter, grower, and finisher. Water was provided ad labium. The control group, designated as G1, received a broiler basal diet without any additions. Broiler chicks of the group 2 (G2) were consumed the basal broiler diet with addition of 500 g/ton feed of SB. In the group 3 (G3), the broiler chicks were fed on the basal diet to which 500 g/ton feed of OV were added. Group four (G4) was received the basal diets to which half of the concentrations (250 g/ton feed) of both additives were added. Broiler chicks in the group five (G5) received the basal diet that was enriched with 500 g/ton feed of combination of SB and OV feed additives.

# Growth performance:

On day 35 of dietary period, the chicks were weighed, and their daily feed intake (FI) was reported through the experiment period. Body weight gain (BWG) was computed. The feed conversion ratio (FCR) was calculated as feed intake (g)/ body weight gain (g) using the procedure outlined by Kidane *et al.* (2017).

#### **Biochemical parameters:**

At the end of the experimental period, the blood was withdrawn from the brachial wing vein and serum samples were used to determine the activities aspartate of aminotransferase (AST) and alanine aminotransferase (ALT) using the procedure outlined by Bergmeyer et al., (1978). Measurement of serum total cholesterol (TC) level was carried out using the calorimetric method described by Allain et al.,(1974), while triglyceride (TG) level were determined following Wahlefeld (1974). Using a spectrophotometer, the activities of serum superoxide dismutase (SOD) and catalase (CAT) enzymes were estimated following the technique of Nishikimi et al., (1972) and Aebi (1984), respectively. Malondialdehyde (MDA) level were measured following the procedure given by Ohkawa et al. (1979), while total protein was quantified using commercially available diagnostic kits and the biuret method as per the protocol outlined by Zheng et al., (2017). In accordance with the protocol described by Tothova et al., (2019), an automated electrophoresis system was serum protein employed separate to fractions by zone electrophoresis on an agarose gel. The Enzyme-linked immunosorbent assay (ELISA) technique, as outlined by Engvall and Perlmann (1971) was used to determine the levels of serum IgG and IgM.

#### Evaluation of immune status:

The phagocytosis test was accomplished following Bos and de Souza (2000).

Phagocytic activity (PA) is the proportion of phagocytic cells that have engulfed Candida albicans yeast cells, expressed as a percentage. On the other hand, the phagocytic index (PI) is determined by taking the ratio of the number of yeast cells that have been phagocytized to the number of macrophage phagocytic cells. Serum samples were obtained at 1st day after administration and at 1st and 2nd post administration of Newcastle and Gumboro vaccinations and at the end of the experiment. The lytic activity of lysozyme against the cell wall of Micrococcus lysodeikticus was used as a substrate in the lysozyme assay (LA) method. To conduct the assay, an agarose gel plate lyses method was employed, following the protocol outlined by Peeters and Vantrappen (1977). The lysozyme concentration was determined by generating a logarithmic curve with a standard lysozyme solution. Nitric oxide(NO) assay was accomplished in accordance with Sun et al., (2003) using Griess reaction assay after removing protein via mixture of ZnSO<sub>4</sub> and NaOH. The absorbance at 540 nm exhibits a linear correlation with the concentration of NO present in the sample.

# Carcass characteristics:

At the end of the experiment, twenty birds were randomly chosen from each group (with 5 birds per replicate) and prepared for slaughter. The birds were fasted for 12 hours prior to slaughter and were then slaughtered by bleeding the jugular vein. Once slaughtered, the birds were defeathered and eviscerated. Heart, liver, spleen, thymus, bursa, gizzard, and abdominal fat were removed and weighed to the nearest 0.01 g on a digital scale. After the shanks, head, and offal were removed, the remaining carcass was weighed to obtain the weight of the ready-to-cook carcass. Using this weight, the carcass dressing yield percentage (dressing %) was then calculated according to Rosa *et al* (2007) as follows:

Dressing % = Carcass weight/ Live weight X100

# Statistical analysis:

To analyze the data, IBM SPSS® version 19 software was utilized on a personal computer (2010). The mean  $\pm$  SD were compared with a one-way ANOVA test, with a significance level of P < 0.05, and the Post Hoc Duncan test was applied. following Snedecor and Cochran (1986). Antibody titres against Newcastle (NDV) using Hemagglutination inhibition (HI) test according on days 21 and 35, six broilers from each treatment (one broiler per pen) were randomly selected for hemagglutination inhibition (HI) antibody titer. Blood samples (2 ml per broilers) were drawn from the brachial vein into nonheparinized vacuum tubes (Becton Dickinson Vacutainer Systems, Franklin Lakes) and allowed to clot at 4°C for 2 hr. The serum was separated by centrifugation at 3,000g for 15 min, and stored at -20°C for HI anti-body assay. Briefly, after the serum was inactivated at 56°C for 30 min. twofold serial dilution were made in a 96well V-shaped bottom microtitre plate containing 50 µl of CMF-PBS in each well then 50 µl of NDV antigen (4 HA units) was added into all the wells except the last row as the controls. Serum dilutions ranged from 1:21 to 1:212. The plate was incubated at 37°C for 10 min, then 50 µl of 1% rooster erythrocytes suspension was added to each well and incubated for 30 min. A positive serum, a negative serum, erythrocytes and antigens were also included as controls. The last wells which caused complete inhibition was considered as the endpoint. The geometric mean titer was expressed as reciprocal log 2 values of the last dilution that dislead HI.

# RESULTS

Effect on growth performance:

Table 1 shows that supplementation of basal diet with SB and OV and in combination,

increased BWG and improved feed conversion ratio on day 35 of age of broiler chickens.

Parameters	G1	G2	G3	G4	G5
Initial body weight	1395.13	1411.11	1408.11	1419.31	1418.91
at day 28 (g/bird)	$0.12^{a}$	±0.12 <sup>c</sup>	±0.12 <sup>c</sup>	±0.18 <sup>b</sup>	±0.15 <sup>b</sup>
Final Body weight at day 35 (g/bird)	$1999.38 \pm 0.68^{a}$	2079.58 ±0.53 <sup>c</sup>	$2063.37 \pm 0.55^{\circ}$	$2186.85 \pm 0.61^{b}$	$2111.17 \pm 0.62^{b}$
Weight gain	604.25	668.47	665.26	767.54	775.26
(g/bird)	±0.15 <sup>a</sup>	±0.18 <sup>c</sup>	±0.15°	±0.22 <sup>c</sup>	±0.21 <sup>b</sup>
Feed intake (FI)	986.80	1049.50	1035.31	1189.69	1195.77
(g/bird)	±0.45 <sup>a</sup>	$\pm 0.60^{d}$	$\pm 0.60^{d}$	±0.55°	$\pm 0.58^{b}$
Feed conversion ratio (FCR)	1.63	1.57	1.57	1.55	1.54

 Table (1): Effect of Sodium butyrate and/ or Origanium vulgare on growth performance:

Means  $\pm$ SD in each row with different superscript letters are significantly different at P<0.05 G1= Basal diet (BD)

G2= BD supplemented with 500 g/ton of Sodium butyrate (SB)

G3= BD supplemented with 500 g/ton of Origanium vulgare (OV)

G4= BD supplemented with 250 g/ton of SB plus OV

G5= BD supplemented with 250 g/ton of SB plus OV

FCR (feed conversion ratio) =Feed intake (g/bird) / weight gain (g/bird)

# Effect on biochemical profile:

Supplementation of basal diet with SB and OV alone and in combination significantly decreased AST, ALT, TC and TG in broiler chickens (Table 2).

**Table (2):** Impact of sodium butyrate and/or *Origanium vulgare* on serum AST and ALT activities, TC and TG levels:

Parameters	AST	ALT	TC	TG
Groups	(U/L)	(U/L)	(mg/dL)	(mg/dL)

G1	$211.0 \pm 3.31^{a}$	$3.43\pm0.03^a$	$128.6 \pm 3.02^{a}$	$59.4 \pm 1.01^{a}$
G2	$200.0 \pm 2.37^{b}$	$2.63 \pm 0.06^{b}$	$117.8 \pm 3.00^{b}$	$57.0 \pm 1.16^{b}$
G3	$189.0 \pm 3.17^{\circ}$	$2.01\pm0.02^{\rm c}$	$112.0 \pm 3.00^{\circ}$	$53.0\pm1.15^{\rm c}$
G4	$188.0 \pm 2.11^{\circ}$	$2.00\pm0.02^{\rm c}$	$105.0 \pm 2.10^{d}$	$52.0 \pm 1.14^{\circ}$
G5	$174.0 \pm 1.17^{d}$	$1.80\pm0.01^{d}$	$100.0 \pm 2.10^{e}$	$48.5 \pm 1.10^{d}$

Means  $\pm$  SD in each column with different superscript letters are significant at P  $\leq 0.05$ . G1 = Basal diet (BD)

G1 = Basal ulet (BD)G2 = DD = a table in intersection of the model of the mode

G2= BD containing 500 g/ton of Sodium butyrate (SB)

G3= BD containing 500 g/ton of Origanium vulgare (OV)

G4= BD containing 250 g/ton of SB plus OV

G5= BD containing 500 g/ton of SB plus OV

Our results indicated that addition of sodium butyrate and *Origanium vulgar* alone or in a mixture, to the broilers' basal diet caused significant improvements in serum levels of SOD and CAT antioxidant enzymes (Table 3).

**Table (3)** Impact of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination on serum superoxide dismutase (SOD) and catalase (CAT) antioxidant enzymes levels on day 35 of age of broiler chickens:

Parameters	SOD	САТ
Groups	(U/mL)	(U/mL)
G1	$780 \pm 2.6^{e}$	$4.4 \pm 0.01^{\text{e}}$
G2	$792 \pm 3.1^d$	$4.7 \pm 0.01^{d}$
G3	$795 \pm 1.7^{\circ}$	$5.6 \pm 0.02^{\circ}$
G4	$810 \pm 2.2^{b}$	$5.9\pm0.01^{\text{b}}$
G5	$830 \pm 3.1^{a}$	$6.1 \pm 0.02^{a}$

Mean  $\pm$  SD in each column with different superscript letters are significant at P  $\leq 0.05$ .

G1 = Basal diet (BD)

G2= BD containing 500 g/ton of Sodium butyrate (SB)

G3= BD containing 500 g/ton of Origanium vulgare (OV)

G4= BD containing 250 g/ton of SB plus OV

G5= BD containing 500 g/ton of SB plus OV

Supplementation of basal diet with sodium butyrate (500 g/ton feed) and *Origanium vulgare* (500 g/ton feed) alone or in combination significantly decrease serum malondialdehyde (MDA) on day 35 of broiler age as illustrated in Fig. (1).

Supplementation of basal diet with sodium butyrate (500 g/ton feed) and *Origanium* 

*vulgare* (500 g/ton feed) alone or in combination significantly increased albumin (Alb), total proteins (TP), globulin (Glb) and

albumin/globulin ratio on day 35 of age of broiler chickens as shown in Table (4).



Fig (1): Effect of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination on serum level of malondialdehyde (MDA) in broiler chickens.

**Table (4):** Effect of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination on serum concentrations of total proteins (TP), albumin (Alb), globulin (Glb) and albumin/globulin ratio in serum of broilers on day 35 of age of broiler chickens.

Parameters	ТР	Alb	Glb	Alb/Glb
Groups	(g/mL)	(g/mL)	(g/mL)	Ratio
_		_	-	
G1	$32.2 \pm 1.08^{d}$	$13.0 \pm 0.9^{d}$	$20.2 \pm 0.9^{d}$	0.643
G2	$34.1 \pm 0.53^{\circ}$	$15.9 \pm 0.3^{c}$	$23.1 \pm 0.7^{\circ}$	0.688
G3	$38.8 \pm 0.23^{b}$	$19.0\pm0.4^{b}$	$27.2 \pm 0.6^{b}$	0.698
G4	$39.4 \pm 0.23^{b}$	$20.5 \pm 0.2^{b}$	$28.2 \pm 0.5^{b}$	0.726
G5	$41.1 \pm 1.06^{a}$	$24.4 \pm 0.3^{a}$	$32.5 \pm 0.3^{a}$	0.750

Mean  $\pm$  SD in each column with non-similar superscript letters are significant at  $P \leq 0.05$ .

G1 = Basal diet (BD)

G2= BD containing 500 g/ton of Sodium butyrate (SB)

G3= BD containing 500 g/ton of Origanium vulgare (OV)

G4= BD containing 250g/ton of SB plus OV G5= BD containing 500 g/ton of SB plus OV

As demonstrated in Fig (2), addition of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination caused an elevation in serum immunoglobulins IgG and IgM concentrations in broiler chickens on day 35 of age.



**Fig (2):** Effect of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination on serum concentration of immunoglobulins IgG and IgM on day 35 of age of broiler chickens.

As demonstrated in Table (5), addition of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination resulted in an increased immunity

parameters phagocytic activity (PA), phagocytic index (PI), lysozyme activity (LA) and nitric oxide (NO) concentration in broiler chickens.

**Table (5):** Effect of sodium butyrate (SB) and *Origanium vulgare* (OV) alone or in a mixture on phagocytic activity (PA), phagocytic index (PI), lysozyme activity (LA) and nitric oxide (NO) concentration on day 35 of age of broiler chickens.

Parameters	PA	PI	LA	NO
Groups	(%)	(%)	(µg / ml)	(µM)
G1	$17.2 \pm 0.05^{d}$	$55.3 \pm 1.5^{d}$	$240.3 \pm 4.5^{d}$	$66.7 \pm 1.4^{d}$
G2	$18.4 \pm 0.02^{\circ}$	$58.9 \pm 1.3^{c}$	$245.9 \pm 33^{\circ}$	$69.9 \pm 2.3^{c}$
G3	$19.2 \pm 0.04^{b}$	$60.2 \pm 1.4^{b}$	$258.2\pm4.4^{b}$	$742 \pm 1.3^{b}$
G4	$20.5\pm0.06^{b}$	$61.3 \pm 1.6^{b}$	$259.3\pm3.6^{b}$	$75.5 \pm 1.6^{b}$
G5	$22.6 \pm 0.09^{a}$	$62.8 \pm 1.4^{a}$	$262.8 \pm 5.4^{a}$	$78.9 \pm 2.4^{a}$

Mean  $\pm$  SD in each column with dissimilar superscript letters are significant at  $P \leq 0.05$ .

G1 = Basal diet (BD)

G2= BD containing 500 g/ton of Sodium butyrate (SB)

G3= BD containing 500 g/ton of *Origanium vulgare* (OV)

G4= BD containing 250 g/ton of SB plus OV

G5= BD containing 500 g/ton of SB plus OV

As recorded in Table (6) addition of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination resulted in an increase in live weight and carcass weight. The dressing percent (DP %) was 71.5%. There was a significant increase in weights of bursa and a decrease in abdominal fat. Non-significant changes in weights of spleen, thymus, liver, heart, and gizzard were reported.

Carcass traits	G1	G2	G3	G4	G5
$\mathbf{L}$ is a set $(\mathbf{a})$	1996.21	2075.24	2094.51	2184.21	2190.68
Live wi.(g)	±1.20 <sup>a</sup>	±1.70 <sup>c</sup>	±1.80°	$\pm 1.10^{b}$	$\pm 1.85^{b}$
Corooss wit (g)	1427.29	1483.80	1497.57	1561.71	1577.03
Carcass wt. (g)	$\pm 1.82^{a}$	±1.78 <sup>c</sup>	±1.99°	$\pm 1.17^{b}$	$\pm 1.45^{b}$
Dressing (%)	71.5	71.5	71.5	71.5	71.5
Spleen (g)	$1.15 \pm 1.45^{a}$	1.20	1 45 +1 61 <sup>a</sup>	1 55 +1 74 <sup>a</sup>	$1.55 \pm 1.52^{a}$
		±1.35 <sup>a</sup>	1.45 ±1.01	1.55 ±1.74	
Thymus (a)	$1.75 \pm 1.5/a$	1.82	$1.01 \pm 1.15^{a}$	$1.02 \pm 1.12^{a}$	$1.04 \pm 1.13^{a}$
Thymus (g)	1.75 ±1.54	±1.14 <sup>a</sup>	1.71 ±1.15	$1.72 \pm 1.12$	1.74 ±1.13
Bursa	$2.55 \pm 0.12^{d}$	2.72	$2.86 \pm 0.10^{b}$	$2.88 \pm 0.10^{b}$	$2.95 \pm 0.11^{a}$
Fabricius (g)		±0.15 <sup>c</sup>			

**Table (6):** Effect of sodium butyrate (SB) and *Origanium vulgare* (OV) alone and in combination on carcass traits at 35 days of age of broiler chickens. (5 replicates)

Liver (g)	48.28	48.50	48.35	48.15	48.28
Liver (g)	±1.83 <sup>a</sup>	±1.71 <sup>a</sup>	±1.95 <sup>a</sup>	±1.35 <sup>a</sup>	$\pm 1.45^{a}$
Heart (g)	$2.35 \pm 1.62^{a}$	$2.50 \pm 1.15^{a}$	$2.45 \pm 1.71^{a}$	$2.65 \pm 1.14^{a}$	2.55 ±1.31 <sup>a</sup>
Gizzard (g)	$4.45 \pm 1.52^{a}$	$4.50 \pm 1.84^{a}$	$4.45 \pm 1.61^{a}$	$4.65 \pm 1.23^{a}$	$4.55 \pm 1.10^{a}$
Abdominal fa	ut 2.21±0.92 <sup>a</sup>	1.95±0.29 <sup>b</sup>	1.90±0.83 <sup>b</sup>	1.85±0.74°	1.80±0.63 <sup>d</sup>

Mean values  $\pm$ SE with different letter superscript in each raw are significant at *P*  $\leq$ 0.05,

G1 = Basal diet (BD)

G2= BD containing 500 g/ton of Sodium butyrate (SB)

G3= BD containing 500 g/ton of *Origanium vulgare* (OV)

G4= BD containing 250g/ton of SB plus OV

G5=BD containing 500 g/ton of SB plus OV

DP= Dressing percent,

Dressing % = Carcass weight /Live weight X 100

# **DISSCUSION:**

In the present study, effects of feed additives sodium butyrate (SB) and *Origanium vulgare* (OV) separately or combined on growth performance, serum biochemical indexes, immune status and carcass traits of broiler chickens were evaluated.

Our findings showed that supplementing basal diet with SB and OV alone and in combination led to significant improvements in BWG and FCR in broiler chickens at 35 days of age. These findings agreed with those of Abd El-Ghaney (2017); Alagawany *et al.* (2018); Lan *et al.* (2020); Alagawany *et al.* (2021) and Fathi *et al.* (2023). The previous studies specified that addition of SB and OV improved the growth performance. Moreover, other phytogenic feed additives, vitamins, and minerals when added to basal diets, produced excellent growth promoting effects in animals and broiler chickens. However, Wang *et al.* (2017) mentioned that nutrition is a crucial factor in preserving the broiler health, ensuring high-quality eggs in laying hens and body growth and immune status of broilers.

The current results indicated that supplementation of basal diet with SB and OV alone and in combination significantly lowered serum liver enzymes AST and ALT activities and TC, TG and MDA levels. It significantly increased serum activities of SOD and CAT of broilers on day 35 of age. These outcomes were partly consistent with the results reported by Sikandar et al. (2017); Ri et al. (2017); Alagawany et al. (2018); Lan et al. (2020) and Zhang et al. (2021).

The obtained results indicated that supplementation of basal diet SB and OV alone and in combination led to decreased TC and TG levels and mitigated lipid peroxidation and oxidative stress as it lowered MDA level and increased SOD and CAT activities (Ri et al., 2017 and Lan et al., 2020). However, Parvizi et al., (2020) and Zaazaa et al., (2022) concluded that thyme and Oregano improved using biochemical indexes (AST, ALT, TC and TG) of broilers.

Regarding immunity parameters, this study indicated that supplementation of basal diet SB and OV alone or in combination significantly PA, PI, LA, and NO concentration in broiler chickens. These results agreed with Ri et al., (2017); Alagawany et al., (2018); Lan et al., (2020) and Parvizi et al., (2020) who discovered that addition of SB and OV alone or in combination to basal diet enhanced immunity in broiler chickens.

Concerning carcass traits, the present results showed that supplementation of basal diet SB and OV alone or in combination significantly improved carcass characteristics. It increased live weight and carcass weight and the dressing % was 71.5%. Consequently they increased the weight of bursa of Fabricius and decreased abdominal fat Non- significant changes in the weight of other organs and were reported. These results agreed with those of Alagawany et al. (2018); Lan et al. (2020) and Parvizi et al. (2020) who found that SB and OV alone and in combination resulted in a good carcass traits of broiler chickens.

# **CONCLUSION**

It could be concluded that supplementation of basal diet with sodium butyrate (SB) and Origanium vulgare (OV) alone and in combination can be used as a useful and effective strategy for enhancing growth performance and biochemical parameters and enhance immune status and improve carcass traits of broiler chickens. It has hepatoprotective. antioxidant and immunostimulant effects. The mechanisms of action underlying these effects require further study in broiler chickens.

# REFERENCES

- Abdelli, N.; Sola-Oriol, D. and Perez, F. (2021): Phytogenic Feed Additives Poultry: Achievements, in Prospective and Challenges. Animals (Basel): 11(12): 3471-4381.Doi: 10.3390/ani11123471.
- Abd El-Ghaney, D.; El-Far, A.; Sadek, K.; El-Sayed, Y. and Abdel-Latif, M.A. (2017): Impact of dietary thyme (Thymus vulgaris) on broiler chickens concerning immunity, antioxidant status, and performance. Alex. J. Vet. Sci.;55(1): 169-179. Doi: /10.5455/ajvs.275352
- El-Hack, M. E.; Mahgoub, Abd S. A.; Alagawany, M. and Ashour, E. (2017):Improvingproductive A. performance and mitigating harmful emissions from laying hen excreta via feeding on graded levels of corn DDGS with or without Bacillus subtilis Probiotic .J. Anim. Physiol. Anim. Nutri.; 101(5): 904-913.
- Aebi, H.E. (1984): Catalase in vitro. Methods Enzymol.; 105:121-126. Doi:10.1016/s00766879(84)05016-3.
- Adhikari, S. (2018): Supplementation of Toxin Binders in Broiler to Study its Performance. Acta Sci. Agric.; 2(7): 83-87.
- Al-Khalaifah, H.S. (2018): Benefits of probiotics and/or prebiotics for

antibiotic- reducedpoultry. Poult. Sci.; 97: 3807-3815.

- Alagawany, M.; El-Hack, M.E.A.; Farag, M.R.; Shaheen, H.M.; Abdel-Latif, M.A.; Noreldin,A.E. and Patra, A.K. (2018).The usefulness of Oregano and its derivatives in poultry
- nutrition. Worlds Poult. Sci. J.; 74(3): 463- 474.
- Alagawany, M.; Shaaban, S.E.; Mayada, R.F.; Tiwari, R.; Yatoo, M.I.; Karthik, K.; Michalak, I. and Dhamah, K. (2021): Nutritional significance of amino acids, vitamins
- and minerals as nutraceuticals in poultry production and health: A comprehensive review. Vet. Qaurt. ; 41(1):1-29. Doi: 10.1080/01652176.2020.18578
- Allain, C. C.; Poon, L. S. and Chan, C. S. (1974): Enzymatic determination of serum totalcholesterol. Clin. Chem.; 20: 470-475. Doi.org/10.1093/clinchem/20.4.470

87

- Bergmeyer, H.U., Schreiber, P. and Wahlefeld, A.W. (1978): Optimization of methods foraspartate and alanine aminotransferases. Clin. Chem.; 24: 58-61. PMID: 2240
- Bos, H. and de Souza, W. (2000): Phagocytosis of yeast: a method for concurrent quantification of binding and internalization using differential interference contrast microscopy. J. Immunol. Methods; 238(1-2):29-43.
- Doi: 10.1016/s0022-1759(00)00132-0.
- Debnath, B.C.; Biswas, P. and Roy, B. (2019): The effects of supplemental threonine onperformance, carcass characteristics, immune response and gut health of broilers insubtropics during pre-starter and starter period. J. Anim. Physiol. Anim. Nutri.; 03(1):29-40.

- Engvall, E. and Perlmann, P. (1971): Enzyme-linked immunosorbent assay (ELISA). Quantitative assay of immunoglobulin G. Immunochem.; 8(9):871-874. Doi: 10.1016/0019-2791(71)90454-x.
- M.; Hosayni, M.: Alizadeh. S.: Fathi, Zandi, R.; Rahmati, S. and Rezaee, V. (2023): Effects of black cumin (Nigella Sativa) seed meal on performance, growth blood andbiochemical indices, meat quality and cecal microbial load in broiler chickens. Livestock Sci.; Available online 8 June 2023, 105272. Doi: org/10.1016/j.livsci.2023.105272
- Hashemi, S.R. and Davoodi, H.(2011):Herbal plants and their derivatives as growth and health promoters in animal nutrition. Vet. Res. Commun.; 35:169-180. Doi: 10.1007/s11259-010-9458-2.
- Kidane, Z.; Mengistu, A.; and Singh, H. (2017): Effect of oyster mushroom, garlic and ginger as feed additives on feed intake, growth performance, and economic efficiency of broilers.Br. J. of Poult. Sci., 6(1): 7-15. Doi: https://www.doi.org/10.5829/idosi.bj ps.2017.07.15
- Lan, R.; Zhao, Z.; Siqi, Li S. and An, L. (2020): Sodium butyrate as an effective feed additive to improve performance, liver function, and meat quality in broilers under hot
- climatic conditions. Poult. Sci; 99(11):5491-5500. Doi: 10.1016/j.psj.2020.06.042
- Nishikimi, M., Appaji, N. and Yagi, K. (1972): The occurrence of superoxide anion in thereaction of reduced phenazine methosulphate and molecular oxygen. Biochem. Biophys. Res. Commun.; 46(2): 849-854.Doi:org/10.1016/S0006-291X (72)80218-3.

- Ohkawa, H.; Ohishi, N. and Yagi, K. (1979): Assay for lipid peroxides in animal tissues bythiobarbituric acid reaction. Analyt. Biochem.; 95 (2): 351-358.
- Olivera, A.A.; Keller, K.M.; Devesa, M.V.; Keller, L.A.M.; Dias, E.O.; Martini-Santos, B.J.Leitãoa, D.F.; Cavaglierid, D.F. and Rosa, C.A.R. (2015): Effect of three differentantimycotoxin additives on broiler chickens exposed to aflatoxin B1. ; Arch. Med. Vet.; 47:175-183
- Parvizi. O.; Taherkhani, R. and Abouzari, M. (2020): Evaluation the effect of using thyme and oregano powder in comparison to the antibiotic and probiotic supplementation on
- growth, some immune responses and intestinal morphology of broiler chicks. Ukrainian J. Vet. Agri. Sci., 3(1): 3-8.
- Peeters, T.L. and G.R. Vantrappen. (1977): Factors influencing lysozyme determinations by the lysoplate method. Clinica. Chimica. Acta; 74: 217-225.
- Ri, C.S.; Jiang, X.R.; Kim, M.H.; Wang, J.;
  Zhang, H.J.; Wu, S.G.; Bontempo,
  V. and Qi, G.H. (2017): Effect of
  dietary oregano powder
  supplementation on the growth
  Performance, antioxidant status and
  meat quality of broiler chicks. Ital. J.
  Anim. Sci.; 16(2):264-252.
- Ricke, S.C.; Dittoe, D.K.; Richardson, K.E. (2020): Formic acid as an antimicrobial for poultry production: A review. Front. Vet. Sci.; 7: 1-13. Doi: 10.3389/fvets.2020.00563
- Rosa P.S.; Filho DE, F.; Dahlke, F.; Vieira, B.S.; Macari, M. and Furlan R.L. (2007):Performance and carcass characteristics of broiler chickens with different growth potential and submitted to heat stress. Braz. J. Poult. Sci.: 9 (3). 12-18.

Doi.org/10.1590/S1516-635X2007000300007

- Sikandar, A.; Zaneb, H.; Younus, M.; Masood, S.; Aslam, A.; Farina, K.; Ashraf, S.; Yousa, M.S. and Habib Rehman, H. (2017): Effect of sodium butyrate on performance, immune status, microarchitecture of small intestinal mucosa and Lymphoid organs in broiler chickens. Asian-Australas J. Anim. Sci.; 30(5): 690-699. Doi: 10.5713/ajas.16.0824
- Snedecor, G.W. and Cochran W.G. (1986): "Statistical Methods". The 6th Ed. Iowa StateUniversity Press, Ames, Iowa, USA. Pages 381-418.
- Song, B.; Li, H.; Wu, Y.; Zhen, W.; Z. Wang, Z.; Xia, Z. and Guo, Y. (2017): Effect ofmicroencapsulated sodium butyrate dietary supplementation on growth performance and intestinal barrier function of broiler chickens infected with necrotic enteritis. Anim. Feed.

Sci. Tech.; 232: 6-12.

- Sun, J.; Zhang, X.; Broderick, M. and Harry Fein, H. (2003): Measurement of nitric oxideproduction in biological systems by using Griess Reaction Assay. Sensors; 3(8): 276-284.
- Doi.org/10.3390/s30800276
- Tóthová, C.; Sesztáková, E.; Bielik, B. and Nagy, O. (2019): Changes of total protein andprotein fractions in broiler chickens during the fattening period. Vet. World. ; 12(4): 598-604. Doi: 10.14202/vetworld.2019.598-604
- Wahlefeld. A.W. (1974): Triglycerides determination after enzymatic hydrolysis, Methods In: of Enzymatic Analysis. Ed. H. U. Bergmeyer, 2nd English Ed. Academic Press, New York (USA); pp. 18-31.

- Wang, J.; Yue, H.; Wu, S.; Zhang, H. and Qi, G. (2017). Nutritional modulation of health, egg quality and environmental pollution of the layers. Anim. Nutri.; 3(2):91-96.
- Zaazaa, A.; Mudalal, S.; Alzuheir, I.; Samara, M.; Jalboush, N.; Fayyad, A. and Petracci, M. (2022): The impact of thyme and oregano essential oils dietary supplementation on broiler health, growth performance and prevalence of growth – related breast muscle abnormalities. Anim.; (12(21): 3065-
  - 3070

- Zhang, L.Y.; Peng, Q.Y.; Liu, Y.R.; Ma, Q.G; Zhang, J.Y.; Guo, Y.P., Xue, Z. and Zhao, L.H(2021): Effects of oregano essential oil as an antibiotic growth promoter alternative on growth performance, antioxidant status, and intestinal health of broilers.
- Poult Sci.; 2021 Jul; 100(7): 101163. Doi: 10.1016/j.psj.2021.101163
- Zheng K.; Wu L.; He, Z.; Yang, B. and Yang, Y. (2017): Measurement of the total protein in serum by biuret method with uncertainty evaluation. Measurement;112:16-21.