

Citation: Egypt.Acad.J.Biolog.Sci. (C.Physiology and Molecular biology) Vol. 15(2) pp419-433 (2023) DOI: 10.21608/EAJBSC.2023.320708



Egypt. Acad. J. Biolog. Sci., 15(2):419-433 (2023) Egyptian Academic Journal of Biological Sciences C. Physiology & Molecular Biology ISSN 2090-0767 <u>www.eajbsc.journals.ekb.eg</u>



Genotypic Detection of New-Delhi Metallo-β-Lactamase Producing Carbapenem Resistant *Escherichia coli* in Holy Karbala Province-Iraq

Rasol A. Taqi and Oday M. Hadi

Medical Laboratory Techniques Department, College of Health and Medical Techniques-Kufa, Al-Furat Al-Awsat Technical University, Najaf, Iraq ***E-mail:** <u>sayyedrasool6@gmail.com</u> ; <u>odayzwain@gmail.com</u>

ARTICLE INFO

Article History Received: 29/8/2023 Accepted: 10/10/2023 Available: 14/10/2023

Keywords:

New Delhi metallo beta lactamase, carbapenem, antibiotic resistance, *Escherichia coli*.

ABSTRACT

Background: Carbapenem is a beta-lactam antibiotic that works similarly to Penicillins and Cephalosporins by binding to Penicillin-binding sites and subsequently inhibiting cell wall synthesis. Due to their high potency and extraordinarily broad antibacterial action, Carbapenem antibiotics, which belong to the most recent generation of β -lactam antibiotics, are frequently used in clinics to treat bacterial infections. The current study aimed to shed light on *Escherichia coli*'s resistance to β -lactam antibiotics, especially Carbapenem, and the potential link between Carbapenem resistance and the production of β-lactamase variants bla (NDM-1, NDM-2, NDM-3, NDM-4, NDM-5, GIM-1, VIM-1, SPM-1, and IMP-1) in Karbala province, Iraq. Materials and methods: Four hundred-eight isolates have been identified according to traditional methods (colonial morphology on MacConkey and Eosin methylene blue (EMB) agar, and biochemical reactions), and confirmed by the Vitek2® system. Carbapenem susceptibility was assayed by using the Vitek2[®] system. Isolates were subjected to multiplex PCR targeting βlactamase variants genes. Results: Out of a total of 408 samples randomly collected from different clinical sources, 24 (19.51%) Escherichia coli isolates were diagnosed. Four (16.67%) of the Escherichia coli isolates tested positive for imipenem and meropenem resistance. PCR experiments indicated that only two (50%) isolates contained the blaNDM-1 gene and four (100%) carried the blaNDM-3 gene; no additional beta-lactamases were found.

INTRODUCTION

Resistance to carbapenems in *Enterobacteriaceae* is a serious problem, as these medications are often the last line of battle against infections caused by these bacteria. There are three types of β -lactamases that inactivate carbapenems (also known as carbapenemases): KPCs, metallo- β -lactamases (MBLs), and Oxacillinases (Peirano *et al.*, 2011). The VIM and IMP forms of MBL are the most prevalent types discovered in *Enterobacteriaceae* (Cornaglia *et al.*, 2007). The phenomenon in which the microorganisms grow in the presence of therapeutic agents that earlier negatively affected them, this known as antimicrobial resistance (Dadgostar, 2019). As a result of developing microorganisms resistant to antibiotics, it annually generates significant rates of mortality, morbidity, and economic costs. Identification and comprehension of antibiotic resistance are critical for medical treatments for managing disorders caused by antibiotic-resistant bacteria and for public health measures to stop the development of resistance (Boolchandani *et al.*, 2019).

Citation: Egypt.Acad.J.Biolog.Sci. (C.Physiology and Molecular biology) Vol. 15(2) pp419-433 (2023) DOI: 10.21608/EAJBSC.2023.320708

Antibiotic-resistant bacteria have emerged globally as a result of the improper and excessive usage of antibiotics in the medical, and agricultural industries. This wide trend affects а variety of microorganisms with a high prevalence, endangering human health (Pulingam et al., 2022).Due to their high potency and extraordinarily broad antibacterial action, carbapenem antibiotics, which belong to the recent generation of β-lactam most antibiotics, are frequently used in clinics to treat bacterial infection (Coulthurst et al., 2005; Cornaglia et al., 2011). Carbapenem is a beta-lactam antibiotic that works similarly to penicillins and cephalosporins by binding to penicillin-binding sites and subsequently inhibiting cell wall synthesis (Zhanel et al., 2007; Meletis, 2016). However, in contrast to penicillins and cephalosporins, carbapenem is believed to have a broader spectrum. Carbapenem is one of the newest beta-lactam antibiotics to be constructed. It has an unusual structure that provides high stability against β-lactamases, particularly extended-spectrum β-lactamases such as ceftazidime, ceftriaxone, and cefepime (INGGRAINI et al., 2021). Carbapenemase is split into three types based on functional structure, including β -lactamase class A, B, and D (Whitley et al., 2020). Serine β -lactamases are the name given to Class A and D β -lactamases, which have serine residues at the active site. At the active site of Class B β -lactamase, there are metal ions (zinc) (Queenan and Bush, 2007). The important carbapenem-resistant most components that hydrolyze practically all βlactam antibiotics are known as New Delhi metallo-beta-lactamases (NDMs), and they are extensively spread throughout the world (Van Duin and Doi, 2017; Logan and Weinstein, 2017).

MATERIALS AND METHODS

A total of 408 samples were randomly collected from the two main hospitals (Imam Al-Hussein, peace be upon him, Medical City and Imam Al-Hassan Al-Mujtaba, peace be upon him, Hospital) in the holy Karbala province from September to December 2022. Those samples were obtained from several clinical sources (i.e. urine, stool, blood, sputum, CSF, body fluid, seminal fluid, abscess, and swabs from different regions of the body).

Identification of the Escherichia coli :

Escherichia coli were presumptively identified by traditional method (colonial morphology on MacConkey and Eosin methylene blue (EMB) agar, biochemical reactions) (as mentioned by Tille, 2015). The Vitek2® technology was used for confirmatory identification.

Antimicrobial Susceptibility Testing:

The antibiotic susceptibility of all *Escherichia coli*-positive isolates has been evaluated via the Vitek2® technology, which has been approved by the Clinical and Laboratory Standards Institute (CLSI) (Clinical and Laboratory Standards Institute, 2017).

The antimicrobial agents used in antimicrobial susceptibility testing include amikacin, ampicillin, aztreonam, cefepime, ceftazidime, ceftriaxone, ciprofloxacin, gentamicin, imipenem, meropenem, minocycline, piperacillin, piperacillin/ tazobactam, ticarcillin/clavulanic acid. ticarcillin, tobramycin, and trimethoprim /sulfamethoxazole.

Phenotypic Detection of Carbapenem Resistance Strains:

All *Escherichia coli* isolates were tested for carbapenem antibiotics (imipenem and meropenem) by using the Vitek2® system in accordance with global Clinical and Laboratory Standards Institute (CLSI) guidelines(Clinical and Laboratory Standards Institute, 2017).

Genotypic Detection of Carbapenem Resistance Genes:

MBLs were detected in *E. coli* by bla PCR amplification (NDM-1, NDM-2, NDM-3, NDM-4, NDM-5, GIM-1, VIM-1, IMP-1, and SPM-1) as previously described (Hornsey *et al.*, 2011; Kaase *et al.*, 2011; Qin *et al.*, 2016; Vural *et al.*, 2020; Thapa *et al.*, 2022).

RESULTS

Growth Rate of Microorganisms Among Collected Samples:

From a total of the 408 collected samples, there were 260 (63.72%) female and 148 (36.28%) male. The bacterial growth rate was 30.15% in all cultivated samples

(123/408). Of these 13.24 % (54/123) were recorded to be Gram-positive bacterial isolates and 16.91% (69/123) were Gram-negative isolates. Table 1, illustrates ten age groups that ranged from 1-98 years and the growth rate with sex distribution for each group.

	Recorded	-Sample's	Grow			
Age in years	Male No. (% to Male total)	Female No. (% to female total)	Positive bacterial growth rate for both sex samples No. (%)	Negative bacteri- al growth rate for both sex samples No. (%)	Total No. (%)	
1-10	11 (7.43)	14 (5.38)	8 (6.5)	17 (68)	25 (6.13)	
10-20	13 (8.78)	32 (12.31)	12 (9.76)	33 (73.34)	45 (11.03)	
21-30	25 (16.89)	75 (28.85)	19 (15.45)	81 (81)	100 (24.51)	
31-40	20 (13.51)	57 (21.92)	28 (22.76)	49 (63.64)	77 (18.87)	
41-50	27 (18.24)	40 (15.38)	20 (16.26)	47 (70.1)	67 (16.42)	
51-60	28 (18.92)	21 (8.08)	18 (14.63)	31 (63.26)	49 (12.01)	
61-70	14 (9.46)	13 (5)	12 (9.76)	15 (55.56)	27 (6.62)	
71-80	10 (6.76)	6 (2.31)	6 (4.88)	10 (62.5)	16 (3.92)	
81-90	0 (0)	1 (0.38)	0 (0)	1 (100)	1 (0.25)	
91-100	0 (0)	1 (0.38)	0 (0)	1 (100)	1 (0.25)	

Table 1: Sample distribution by sex, age, positive and negative bacterial growth.

Escherichia coli, Staphylococcus species, *Candida* species, *Enterobacter* species, *Pseudomonas* species, *Klebsiella* species, *Streptococcus* species, *Proteus* species, and unidentified bacteria have been identified in 24 (19.51%), 22 (17.88%), 18 (14.63%), 16 (13.01%), 15 (12.2%), 11 (8.95%), 10 (8.13%), 2 (1.62%), and 5 (4.07%), respectively. According to the statistics above, the most common isolate was *E. coli*.

Distribution of *Escherichia coli* Isolates Among Positive Samples:

Escherichia coli isolates were distributed among sex as 18(75%) female and 6(25%) male, with female to male ratio 3:1, and among sample type as 13(54.17%) urine, 4(16.67%) body fluid, 4(16.67%) swab, 2(8.33%) sputum, and 1(4.16%) stool. Table 2, shows the distribution of *Escherichia coli* among sex and sample type.

The age of patients who carry *Escherichia coli* ranged from 4-80 years and are separated into 4 age groups, Table 3, will demonstrate the number (%) and the sex ratio of each age group.

	E. coli distri	Total No. (%)			
Sample type	Male No. (% to Male total)	Female No. (% to female total)	of <i>E. coli</i> iso- lates		
Urine	3 (50%)	10 (55.56%)	13 (54.17%)		
Body fluid	0 (0%)	4 (22.22%)	4 (16.67%)		
Swabs	2 (33.33%)	1 (5.55%)	3 (12.5%)		
Sputum	1 (16.67%)	1 (5.55%)	2 (8.33%)		
Wound swab	0 (0%)	1 (5.55%)	1 (4.16%)		
Stool	0 (0%)	1 (5.55%)	1 (4.16%)		

Table 2: Distribution of *Escherichia coli* isolates among patient's sex and sample type.

Age group (years)	Sample's No. (%)	Male (%)	Female (%)
1-20	6 (25%)	2 (33.33%)	4 (22.22%)
21-40	4 (16.67%)	0 (0%)	4(22.22%)
41-60	11 (45.83%)	3 (50%)	8 (44.44%)
61-80	3 (12.5%)	1 (16.67%)	2 (11.11%)
Total	24	6	18

Table 3: Patient's age and sex distribution of *Escherichia coli*-positive isolates.

Antimicrobial Resistance of *Escherichia* coli Positive Isolates:

Antimicrobials have been employed for treating bacterial infections for over seventy years, although these low-molecularweight bioactive chemicals also have a variety of other medical uses (Uddin *et al.*, 2021). One of the most serious global health challenges, antibiotic resistance affects food security, human, and animal health, and results in significant economic losses. It is primarily brought on by the inappropriate use of antibiotics in agriculture, the environment, and animal and human medicine (Caniça et al., 2019). The following antimicrobial agents were tested Escherichia *coli* isolates: amikacin, ampicillin, aztreonam, cefepime, ceftazidime, ceftriaxone, ciprofloxacin, gentamicin, imipenem, meropenem, piperacillin, minocycline, piperacillin/tazobactam, ticarcillin/clavulanic acid, ticarcillin, tobramycin, and trimethoprim/sulfamethoxazole. Table 4. presents the outcomes of antimicrobial susceptibility among sex.

Table 4: Antimicrobial susceptibility profile of *Escherichia coli* isolates among sex.

Antimicrobial classes		A	Sensitiv	e No. (%)	Intermed	liate No. (%)	Resistant No. (%)		
A	ntimicropial classes	Antimicrobial agents	Male	Female	Male	Female	Male	Female	
	Cephalosporins	Cefepime	4(16.67%)	12(54.17%)	0 (0%)	0 (0%)	2 (8.33%)	6 (20.83%)	
		Ceftazidime	2 (8.33%)	7 (29.17%)	0 (0%)	0 (0%)	4(16.67%)	11(45.83%)	
	Penicillin	Ceftriaxone	2 (8.33%)	5 (20.83%)	0 (0%)	0 (0%)	4(16.67%)	13(54.17%)	
	Peniciliii	Ampicillin	0 (0%)	0 (0%)	0 (0%)	0 (0%)	6 (25%)	18 (75%)	
E E		Piperacillin	0 (0%)	0 (0%)	0 (0%)	0 (0%)	6 (25%)	18 (75%)	
<i>β</i> -lactam	Carbonanama	Ticarcillin	0 (0%)	0 (0%)	0 (0%)	0 (0%)	6 (25%)	18 (75%)	
8-1	Carbapenems	Imipenem	5(20.83%)	15 (62.5%)	0 (0%)	0 (0%)	1 (4.17%)	3 (12.5%)	
		Meropenem	5(20.83%)	15 (62.5%)	0 (0%)	0 (0%)	1 (4.17%)	3 (12.5%)	
	β -lactam/ β -lactamase	Piperacillin/ tazobactam	3 (12.5%)	11(45.83%)	0 (0%)	0 (0%)	3 (12.5%)	7 (29.17%)	
	inhibitor	Ticarcillin /clavulanic acid	1 (4.17%)	7 (29.17%)	2(8.33%)	1 (4.17%)	3 (12.5%)	10(41.66%)	
	Aminoglycosides	Aztreonam	1 (4.17%)	4 (16.67%)	0 (0%)	0 (0%)	5(20.83%)	14(58.33%)	
		Amikacin	4(16.67%)	11(45.83%)	0 (0%)	3 (12.5%)	2 (8.33%)	4 (16.67%)	
l u	Fluoroquinolones	Gentamicin	4(16.67%)	5 (20.83%)	0 (0%)	0 (0%)	2 (8.33%)	13(54.17%)	
8-lactam	Fluoroquinoiones	Tobramycin	4(16.67%)	5 (20.83%)	0 (0%)	0 (0%)	2 (8.33%)	13(54.17%)	
β-1	Tetracycline	Ciprofloxacin	2 (8.33%)	4 (16.67%)	0 (0%)	0 (0%)	4(16.67%)	14(58.33%)	
-	Sulfonamide	Minocycline	1 (4.17%)	8 (33.33%)	0 (0%)	0 (0%)	5(20.83%)	10(41.67%)	
Non	Cephalosporins	Trimethoprim/ sulfamethoxazole	3 (12.5%)	5 (20.83%)	0 (0%)	0 (%)	3 (12.5%)	13(54.17%)	

Distribution of Beta-Lactamase Variants Among Carbapenem-Resistant *Escherichia coli*:

Due to their wide variety of therapeutic indications, β -lactam antibiotics are one of the most commonly recommended drug groups. Since its debut in the 1930s, the fight against bacterial infectious diseases has undergone a significant change (Pandey and Cascella, 2022). Several β -lactam-

inactivating-lactamases have developed as a result of the widespread usage of β -lactam antibiotics as medications during the past 90 years (Bush and Bradford, 2020). The widespread usage of β -lactams has facilitated the development and spread of resistance, much like with other antimicrobial groups (Tooke *et al.*, 2019). The beta lactamases used in the present study are *bla*(NDM-1, NDM-2, NDM-3, NDM-4, NDM-5, GIM-1, VIM-1, IMP-1, and SPM-1), discovered by PCR

technique and gel electrophoresis system. Table 5, contains information regarding β lactamase variants found in individuals infected with *Escherichia coli* in the current study.

Isolate code	Hospital types in		Age	Sample	e Beta lactamase variants								
	Karbala province Sex	Sex	in years	type	NDM-1	NDM-2	NDM-3	NDM-4	NDM-5	GIM-1	VIM-1	IMP-1	SPM-1
67	Imam A1-Hasan	F	6	Urine	-	-	-	-	-	-	-	-	-
99	Imam Al-Hasan	F	17	Urine	-	-	-	-	-	-	-	-	-
119	Imam A1-Hussein	F	19	Fluid	-	-	+	-	-	-	-	-	-
216	Imam A1-Hussein	F	20	Urine	-	-	-	-	-	-	-	-	-
202	Imam A1-Hussein	F	26	Fluid	-	-	-	-	-	-	-	-	-
70	Imam A1-Hussein	F	31	Urine	-	-	-	-	-	-	-	-	-
37	Imam A1-Hussein	F	35	Fluid	-	-	-	-	-	-	-	-	-
95	Imam A1-Hussein	F	38	Urine	-	-	-	-	-	-	-	-	-
14	Imam Al-Hasan	F	45	Urine	-	-	-	-	-	-	-	-	-
146	Imam A1-Hussein	F	45	Fluid	-	-	-	-	-	-	-	-	-
223	Imam A1-Hussein	F	47	Swab	-	-	-	-	-	-	-	-	-
200	Imam A1-Hussein	F	49	Urine	-	-	-	-	-	-	-	-	-
147	Imam Al-Hussein	F	51	Swab	-	-	-	-	-	-	-	-	-
66	Imam Al-Hussein	F	52	Urine	-	-	-	-	-	-	-	-	-
162	Imam A1-Hussein	F	52	Urine	-	-	+	-	-	-	-	-	-
40	Imam Al-Hasan	F	53	Stoo1	-	-	-	-	-	-	-	-	-
154	Imam Al-Hussein	F	63	Sputum	-	-	-	-	-	-	-	-	-
29	Imam A1-Hussein	F	80	Urine	+	-	+	-	-	-	-	-	-
78	Imam Al-Hussein	М	4	Swab	+	-	+	-	-	-	-	-	-
40	Imam A1-Hussein	М	15	Urine	-	-	-	-	-	-	-	-	-
163	Imam A1-Hussein	М	50	Urine	-	-	-	-	-	-	-	-	-
189	Imam Al-Hussein	М	60	Urine	-	-	-	-	-	-	-	-	-
196	Imam A1-Hussein	Μ	60	Sputum	-	-	-	-	-	-	-	-	-
170	Imam A1-Hussein	М	70	Swab	-	-	-	-	-	-	-	-	-

Table 5: Beta-lactamase variants profile for all *Escherichia coli* isolates.

As demonstrated in Table 5, four patients (16.67%) carried carbapenem-resistant *Escherichia coli*, which was caused by β -lactamases (specifically New Delhi metallo beta-lactamase). Those four NDM-positive *Escherichia coli* isolates were collected from three females and one male. There was no relationship between sex and infection with NDM-positive *Escherichia coli*, with males (16.67%) and females (16.67%) having the same percentage (one of six isolates harbored NDM in both males and females).

Escherichia coli-positive-NDM was found in individuals of varying ages, with four isolates collected from patients aged four, nineteen, fifty, and eighty years.

 β -lactamases found in Karbala by the current investigation are exclusively NDM, particularly NDM-1, and NDM-3, with NDM-3 having the highest incidence. NDM-1 was detected in one urine sample and one swab, whereas NDM-3 was detected in two urine samples, one swab, and one fluid.

Two patients were infected with *Escherichia coli* that expressed both *bla*NDM-1 and *bla*NDM-3, as shown in Figure 1A, 1C and Table 5, while the other two patients were infected with *Escherichia coli* that expressed only *bla*NDM-3, as shown in figure 1C and table 5; no additional beta-lactamases were detected (Figs. 1B, 1D, 1E, 1F, 1G, 1H, and 1I).

F: females; M: males; NDM: New Delhi metallo beta lactamase; GIM: German-imipenemase; VIM: Verona Integron-Borne Metallo-Lactamase; IMP: Imipenemase; SPM: São Paulo metallo-beta-lactamase.

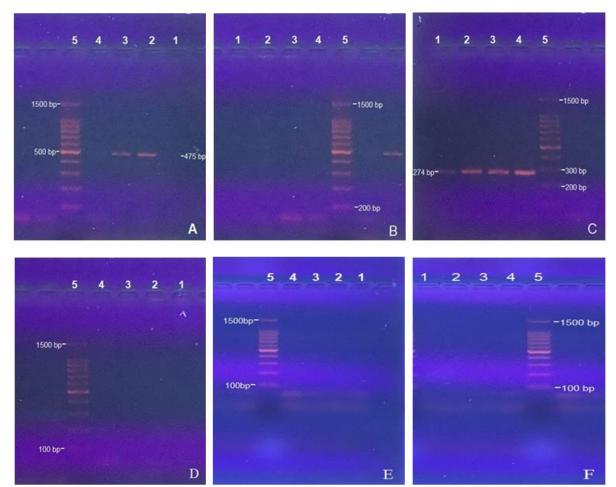


Fig. 1: Gel electrophoresis of PCR amplicon of (A) *bla*NDM-1, (B) *bla*NDM-2, (C) *bla*NDM-3, (D)*bla*NDM-4, (E)*bla*NDM-5, (F)*bla*GIM-1, (G)*bla*VIM-1, (H)*bla*IMP-1, and (I) *bla*SPM-1. Lane 1: Sample no. 119; Lane 2: sample no. 29; lane 3: sample no. 78; lane 4: sample no. 162; lane 5: marker.

DISCUSSION

Globally, AMR issues are becoming worse, even with carbapenems, the final line of defense. It has been demonstrated that widespread inappropriate use and misuse of antibiotics is the major source of drug resistance among microorganisms in countries with low or middle incomes (LMICs), where appropriate infection control and antibiotic stewardship are severely lacking (Pokharel and Adhikari, 2020). The development of carbapenem-resistant Enterobacteriaceae is the result of selective pressure from improper carbapenem usage. The majority of pathogens that are found in almost all common diseases brought on by Gram-negative bacteria (GNB) belong to the family Enterobacteriaceae. More precisely, the primary agents identified from infections caused by GNB are *Enterobacterales*, such as E. coli and K. pneumoniae (Thapa et al., 2022). This investigation was carried out to discover carbapenem-resistant E. coli as well as the potential acquisition of MBL carbapenemase genes (NDM variants, GIM-1, VIM-1, SPM-1, and IMP-1) among such strains in Karbala province, Iraq, in order for this study to become a useful reference in the study region to determine the overall prevalence of drug resistance. In this present study, about 30.15 percent of the cultured samples tested positive for considerable bacterial growth. Similar findings were seen in earlier studies carried out in several Iraqi districts (Sabry et al., 2021), while some other studies reported higher growth rates (Alzaidi and Mohammed, 2022; Al-Asady et al., 2022). This ratio of no growth results from

those patients who probably uptake antibiotics, suffering from a viral infection, immunological disorder, or other conditions.

The sex ratio of the current study was higher in females and this corresponds with the results published in Italy that showed females were more infected by *Escherichia coli* than males (Magliano *et al.*, 2012), and in Kirkuk City, Iraq that illustrates *Escherichia coli* strains were isolated from 76.4% females and 23.5% males (Aljebory and Mohammad, 2019). The body physiology and lifestyle of females make them most susceptible to disease and infection, this reason may be the most acceptable cause pushing females to visit hospitals more than males.

The infection rate was higher than 30% in children under the age of ten and in patients over the age of 51, because the immunity in children is low and in the growth stage in addition the children's hygiene is low especially when they meet their peers and have fun playing, whereas in the elderly, immunity is gradually weakened and become slower in responding to the pathogen in addition to many changes that happen for the body physiology such as menstrual cycle stopping in female. Age group 31-40 also have an infection rate of more than 30%. Comparable with the results of previous studies include the infection rate among age groups was 1.3% for 10-20 years, 8.1% for 20-30 years, 8.8% for 30-40 years, 5% for 40-50 years, and 1.3% for 50-60 years (Ranjit et al., 2018). Another research in Karbala province, Iraq reported that the greatest occurrence (33.33%) of bacterial infection among patients occurred in the age group 31-45 years (Abdul-Husin, 2021), while in Shahrekord, Iran recorded that the age group 30-39 years occupy the highest rate 54.78% of infection (Tajbakhsh et al., 2015). This age group of patients is considered a productive age group, as they have passed the stage of adolescence and have become more mature and interested in life. They realize that age is progressing and time is passing, so it must be used properly, and its perseverance is often the use of time in order to develop in all aspects of life, for example, increasing wealth and building social relationships, and therefore all these interests and goals can make a person forget his health or ignore it. On the other hand, achieving these goals often requires communication with many people, some of whom may be infected with a specific pathogen, and thus the rate of infection transmission increases for them. The infection rate for the age groups 81-90 years and 91-100 years cannot be evaluated since the sample size was insufficient to be depended on.

Escherichia coli was the most prevalent isolate among positive samples with a 19.51% ratio, according to the data of the current study. Many studies support these results and have similar results that E. coli were the predominant isolates in various provinces in Iraq (Karbala, Baghdad, and Basra), Iran, Syria, and Nepal (Thapa et al., 2022; and Mahdi et al., 2020; Salman et al., 2022; Jasim et al., 2022; Hassanzadeh et al., 2009; Abbood et al., 2022). This high percentage of Escherichia coli infection is because Escherichia coli are the normal bowel flora, therefore, it can easily spread from person to person in case of low personal hygiene in both males and females and in females this ratio becomes higher which in addition to the previous reason the body physiology of reproductive system of female (shorter urinary tract and a closer distance between the anus and the urethral opening) simplify the self-infection.

In the present study, E. coli were mostly infecting females and this corresponds with the results of several previous studies (Magliano et al., 2012; Aljebory and Mohammad, 2019). According to the findings of the present investigation, urinary tract infections 13(54.17%) were the most prevalent source of Escherichia coli, similar to many previous studies (Abdul-Husin, 2021; Ronald, 2002; Naqid et al., 2020; Karam et al., 2019). Body fluid infections are mostly caused by Escherichia coli bacteria findings and these findings are also reported in many published research (Rouf and Nazir, 2019). Swabs obtained from different clinical sources bear the same percentage of body

fluid and occupy the second stage of infections in humans after the urine samples. Results of the study in Kurdistan, Iraq showed that Escherichia coli was most detected after urine at 92.2% in the wound 3.9% and cervical 1.5% swabs (Nagid et al., 2020). Another study in Najaf province, Iraq, illustrated that the occurrence of E. coli infections in the urinary tract was 91%, wounds 7%, and burns 2% (Najm and Hussein, 2023), while research published in Egypt showed that 28.7% of all Escherichia coli isolates obtained during their study were swabbed from wounds, throat, and vagina (El-Baz et al., 2022). During the present study from all Escherichia coli isolates 8.33% were obtained from the respiratory tract as sputum. Results of a study done in Egypt demonstrate that the respiratory tract has 3.3% of Escherichia coli isolates (El-Baz et al., 2022). In accordance with the prior research, acceptable respiratory tract infections include Streptococcus pneumoniae the most usually identified, followed by Moraxella catarrhalis and Haemophilus influenza, whereas the nonwere Escherichia acceptable coli. Staphylococcus aureus, and Pseudomonas aeruginosa (Popova et al., 2019). Escherichia coli is normal bowel flora, but it is opportunistic and can cause infection in many conditions in addition to several strains that can cause intestinal and extra-intestinal infections such as EPEC, ETEC, etc.

The age group 41-60 years had the highest rate of Escherichia coli infection, accounting for 45.83% of all Escherichia coli isolates, followed by the 1-20 years with 25%, 21-40 years with 16.67%, and 61-80 years with 12.5%. The males (33.33%) infection rate of the 1-20 years age group was higher than females (22.22%) and these results were parallel to the previous study (Hossain et al., 2020), and differ from others (Yadav and Prakash, 2017). Escherichia coli primarily affects females (22.22%) in patients aged 21 to 40. The research performed in Bangladesh (Hossain et al., 2020) and in Nepal (Yadav and Prakash, 2017), showed that infection rates were in females 32.4%, males 17.7%, 48.45%, and females males 43.80%, respectively. The prevalence rate of the age group 41-60 years reveals that males (50%) are mostly infected by *Escherichia coli*, which agrees with the findings prior study (Yadav and Prakash, 2017). The rate of infection in the age group 61-80 years demonstrates that males (16.67%) are the majority infected by *Escherichia coli*, which is supported by the results published in many recent research (Hossain *et al.*, 2020).

In this current study, a majority of the E. coli isolates displayed resistance against frequently suggested medicines. All Escherichia coli isolates were resistant to ticarcillin, piperacillin, and ampicillin, and they were resistant to at least three distinct antibiotic classes, in this situation, the bacteria termed multidrug resistance. These findings concord with many previous reports (Mohammed et al., 2021; Abdelmongy et al., 2022). Multidrug resistance is described as resistance to at least three medications from different antibacterial classes, primarily cephalosporins, penicillins, carbapenems, β lactam/β-lactamase inhibitors, monobactam, aminoglycosides, fluoroquinolones, tetracycline, and sulfonamides (Saderi and Owlia, 2015). Escherichia coli isolates intriguingly displayed worrisome penicillin resistance, with a 24 (100%) resistance rate for all penicillin antibiotics. The outcome is identical to the prior study (Puvača and de, 2021). Aztreonam 19 (79.16%)and ciprofloxacin 18 (75%) were both less effective against Escherichia coli which was highly resistant to those antibiotics, followed by ceftriaxone 17 (70.84%), trimethoprim/ sulfamethoxazole 16 (66.67%), 15 (62.5%) for each ceftazidime, gentamicin, tobramycin, and minocycline, while ticarcillin/clavulanic acid resistant rate was 13 (54.16%), and piperacillin/tazobactam 10(41.67 %). Previous studies showed slightly different rates of resistance to aztreonam (85%), and minocycline (60%) (Biagi et al., 2022; Popova et al., 2019). Ciprofloxacin (93.81%),trimethoprim/sulfamethoxazole (87%), ceftazidime (95.88%), and showed Piperacillin/Tazobactam (90%). higher resistance rates in previous reports

(Mohammed et al., 2021; Ahmad et al., 2022; and Ruaa, 2023). The resistance rate to ceftriaxone (58%), gentamycin (28%),tobramycin (20%), and ticarcillin/clavulanic acid (27.27%), increased when compared to previously published research (Mohammed et al., 2021; Ali and Al-Dahmoshi, 2021; and Abdul-Jabar et al., 2020). The lowest rate of resistance was to cefepime, amikacin, imipenem, and meropenem with 8(29.16%), 6(25%), 4(16.67%), and 4(16.67%) resistant rate, respectively. Previous studies found a higher rate of resistance to cefepime (65%), while a lower rate of amikacin (11%), imipenem (5%), and meropenem (0%) were reported (Mohammed et al., 2021; Ahmad et al., 2022). However, there was the lowest degree of resistance to carbapenems, which are regarded as one of the most strong and effective β -lactams.

Resistance to carbapenems can emerge through three different processes: efflux pump-over activity, porin loss (mutation), and carbapenemase enzyme production. Nonetheless, the production of this enzyme (or variations of this enzyme) is considered to be the fundamental mechanism of resistance in carbapenem-resistant E. coli (Hammoudi and Ayoub, 2020). In the present study among 24 E. coli positive isolates, four (16.67%) were carried carbapenem-resistant Escherichia coli, which was caused by β lactamases (specifically New Delhi metallo beta-lactamase). Those four NDM-positive Escherichia coli isolates were collected from three females and one male. There was no relationship between sex and infection with NDM-positive Escherichia coli, with males (16.67%) and females (16.67%) having the same percentage (one of six isolates harbored both NDM in males and females). Escherichia coli-positive-NDM was found in individuals of varying ages, with four isolates collected from patients aged four, nineteen, fifty, and eighty years. The findings aligned with prior research done by Ismail and Mahmoud, 2018, in which three blaNDM-1 positive isolates and one blaNDM-2 positive six carbapenem-resistant isolate among isolates in Baghdad, Iraq. Another study in

Najaf, discovered 12 Iraq among phenotypically MBL-producing Р. aeruginosa isolates, 4 (33.3%) and 3 (25%), respectively, carried blaVIM and blaIMP genes. Furthermore, two isolates had the blaSPM and blaSIM genes, while one had the gene (Alkhudhairy blaNDM and Al-Shammari, 2020). In Al Jouf, Saudi Arabia, the researcher reported that 71 (74%) had blaNDM-1, and 24 (25%) carried blaNDM gene variants (Junaid, 2021. NDM-producing Escherichia coli isolates were reported to make up 18% (22/122) and 20% (14/71), respectively. of the carbapenemaseproducing Enterobacteriaceae identified in Switzerland and the Hesse area of Germany. Additionally, in Switzerland and the Hesse region, respectively, the overall numbers of NDM-like producing Escherichia coli isolates were 77% (17/22) and 93% (13/14) for NDM-5 producers (Chakraborty et al., 2021).

Metallo-βlactamases (MBLs) were encountered from clinical isolates globally with increasing frequency during the past several years (Thapa *et al.*, 2022).

CONCLUSIONS

According to the present study's findings, the age group 61-70 years had the greatest percentage of infection in general, at 44.4 %, when compared to other age groups. Escherichia coli is the most common bacterium in the province of Karbala, per the findings. Although Escherichia coli is the most prevalent cause of urinary tract infection, it can also cause respiratory tract wound infection, infection. and burn infection. Escherichia *coli* isolates are resistant to the majority of antimicrobial agents. Throughout this study, Escherichia coli showed 100% resistance to ampicillin ticarcillin. Antibiotics including and ciprofloxacin, gentamicin, minocycline, and trimethoprim/ sulfamethoxazole all have significant rates of antibiotic resistance. The percentages are lowest in amikacin, cefepime, imipenem, meropenem, and piperacillin/tazobactam. All carbapenemresistant Escherichia coli in Karbala province produce NDM-3, and 50% only produce

NDM-1, with no other NDM variants observed.

Declaration of Conflicting Interests:

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Funding:

The author (s) received no financial support for the research, authorship, grants and/or publication of this article.

Author Contributions:

Researcher Rasol Abdulameer Taqi took the lead in writing the manuscript. A comprehensive search was performed by Dr. Oday Mitib Hadi and Rasol Abdulameer Taqi. All authors contributed to the article revision, read, and approved the submitted version.

REFERENCES

- Abbood, A., Malek, Z., & Thallaj, N. (2022). Antibiotic resistance of urinary tract pathogens in Syrian children. *Research Journal of Pharmacy and Technology*, 15(11), 4935-4939. https://www.indianjournals.com/ijor .aspx?target
- Abdelmongy, M., Zaki, M., & Hamouda, R. (2022). Antibacterial activity of multidrug resistance Escherichia coli by some natural products. *Research Journal of Applied Biotechnology*, 8(2), 5-12. https://journals.ekb.eg/ article_272175.html
- Abdul-Husin, F. (2021). Plasmid-Mediated Mechanism of Quinolone Resistance on E. coli Isolates from Different Clinical Samples. *Archives of Razi Institute*, 76(3), 561. https://www. ncbi.nlm.nih.gov/pmc/articles/
- Abdul-Jabar, H. H., Abd, A. H., & Abdulamir, A. S. (2020). Efficacy of Piperacilline/ Combinations of Tazobactam, Ceftazidime, Amikacin Bacteriophage and against Enterobacteriaceae Sepsis in Neonates: In Vitro Study. Systematic Reviews in Pharmacy, 11(10). https://web.p.ebscohost.com

- Ahmad, S., Ali, F., Qureshi, S. A., Uzma, B., Shakeela, Q., Sabir, M. S., ... & Farooq, M. (2022). The evaluation of antibiotic susceptibility pattern and associated risk factors of UTI in tertiary care hospital of Peshawar. *Pakistan journal of pharmaceutical sciences*, 35(3), 897-903. www. researchgate.net/profile/Farooq
- Al-Asady, F. M., Al-Saray, D. A., & Al-Araji,
 A. E. (2022, January). Screening of urinary tract bacterial infections and their antibiogram among nonpregnant women admitted to Al-Sadiq hospital, *Iraq. In AIP Conference Proceedings* (Vol. 2386, No. 1). AIP Publishing. https:// pubs.aip.org/aip/acp/article-abstract/ 2386/1/020006/2820880/
- Ali, S. A., & Al-Dahmoshi, H. O. (2021). Antibiotic Resistance Profile of Escherichia coli and Klebsiella pneumoniae Isolated from Patients with Cystitis. Annals of the Romanian Society for Cell Biology, 1336-1347. http://www. annalsofrscb.ro/index.php
- Aljebory, I. S., & Mohammad, K. A. (2019). Molecular Detection of Some Virulence Genes of *Escherichia coli* Isolated from UTI Patients in Kirkuk City, Iraq. *Journal of Global Pharma Technology*, 11(3), 349-355. https://www.researchgate. net/profile/Ibraheem- Aljebory/ publication/ 334282659
- Alkhudhairy, M. K., & Al-Shammari, M. M. M. (2020). Prevalence of metallo-βlactamase–producing Pseudomonas aeruginosa isolated from diabetic foot infections in Iraq. *New microbes and new infections*, 35, 100661. https://www.sciencedirect. com/ science/ article/pii/ S2052297520300135
- Alzaidi, J. R., & Mohammed, A. S. (2022). First Record of Dissemination of BLBLI-Resistant from Public Hospitals in Baghdad, Iraq. *The Open Microbiology Journal*, 16(1).

https://openmicrobiologyjournal.co m

- Badulla, W. F., Alshakka, M., & Mohamed Ibrahim, M. I. (2020). Antimicrobial resistance profiles for different isolates in Aden, Yemen: a crosssectional study in a resource-poor setting. *BioMed Research International*, 2020. https://www. hindawi.com/ journals/bmri/2020/ 1810290/
- Biagi, M., Lee, M., Wu, T., Shajee, A., Patel, S., Deshpande, L. M., ... & Wenzler, E. (2022).Aztreonam in combination with imipenemrelebactam against clinical and isogenic strains of serine and metallo-β-lactamase-producing enterobacterales. Diagnostic Microbiology Infectious and Disease, 103(2), 115674. www. sciencedirect. com/science/article/ abs/pii/
- Boolchandani, M., D'Souza, A. W., & Dantas, G. (2019). Sequencingbased methods and resources to study antimicrobial resistance. *Nature Reviews Genetics*, 20(6), 356-370. https://www.nature.com/ articles/s41576-019-0108-4
- Bush, K., & Bradford, P. A. (2020). Epidemiology of β-lactamaseproducing pathogens. *Clinical microbiology reviews*, 33(2), e00047-19. https://journals.asm. org/doi/full/10. 1128/cmr.00047-19
- Caniça, M., Manageiro, V., Abriouel, H., Moran-Gilad, J., & Franz, C. M. (2019). Antibiotic resistance in foodborne bacteria. *Trends in Food Science & Technology*, 84, 41-44. https://www. sciencedirect.com
- Chakraborty, T., Sadek, M., Yao, Y., Imirzalioglu, C., Stephan, R., Poirel, L., & Nordmann, P. (2021). Crossborder emergence of Escherichia coli producing the carbapenemase NDM-5 in Switzerland and Germany. Journal of Clinical Microbiology, 59(3), e02238-20.

https://journals.asm.org/doi/full/10. 1128.

- Clinical and Laboratory Standards Institute. (2017). Performance standards for antimicrobial susceptibility testing. CLSI supplement M100, 106-112.
- Cornaglia, G., Akova, M., Amicosante, G., Cantón, R., Cauda, R., Docquier, J. D., Edelstein, M., Frère, J. M., Fuzi, M., Galleni, M., Giamarellou, H., Gniadkowski, M., Koncan, R., Libisch, B., Luzzaro, F., Miriagou, V., Navarro, F., Nordmann, P., Pagani, L., Peixe, L., ... ESCMID Study Group for Antimicrobial Resistance Surveillance (ESGARS) (2007). Metallo-beta-lactamases as emerging resistance determinants in Gram-negative pathogens: open issues. International journal of agents, 29(4), 380antimicrobial 388. https://doi.org/10.1016/j. ijantimicag.2006.10.008
- Cornaglia, G., Giamarellou, H., & Rossolini, G. M. (2011). Metallo-β-lactamases: a last frontier for β-lactams?. *The Lancet infectious diseases*, 11(5), 381-393. https://www.thelancet. com/journals/lancet/article/
- Coulthurst, S. J., Barnard, A. M., & Salmond, G. P. (2005). Regulation and biosynthesis of carbapenem antibiotics in bacteria. *Nature Reviews Microbiology*, 3(4), 295-306. https://www.nature.com/ articles/nrmicro
- Dadgostar, P. (2019). Antimicrobial resistance: implications and costs. *Infection and drug resistance*, 3903-3910. https://www.tandfonline. com/doi/full/10.2147/IDR.S234610
- El-Baz, R., Said, H. S., Abdelmegeed, E. S., & Barwa, R. (2022).virulence Characterization of determinants phylogenetic and background multiple of and extensively drug resistant Escherichia coli isolated from different clinical sources in Egypt. Applied Microbiology and

Biotechnology, 106(3), 1279-1298. El-Baz, R., Said, H. S., Abdelmegeed, E. S., & Barwa, R. .(2022)

- Hammoudi Halat, D., & Ayoub Moubareck,
 C. (2020). The current burden of carbapenemases: Review of significant properties and dissemination among gram-negative bacteria. *Antibiotics*, 9(4), 186. https://www.mdpi.com/2079-6382/9/4/186
- Hassanzadeh, P., Motamedifar, M., & Hadi, N. (2009).Prevalent bacterial infections in intensive care units of University medical Shiraz of sciences teaching hospitals, Shiraz, Iran. Japanese Journal of Infectious Diseases, 62(4), 249-53. https:// www.jstage. jst.go.jp/article/yoken/ 62/4/62_JJID.2009.249/_article/char / ja/
- Hornsey, M., Phee, L., & Wareham, D. W. (2011). A novel variant, NDM-5, of the New Delhi metallo-β-lactamase in a multidrug-resistant *Escherichia coli* ST648 isolate recovered from a patient in the United Kingdom. *Antimicrobial agents and chemotherapy*, 55(12), 5952-5954. https://journals.asm.org/doi/full/
- Hossain, A., Hossain, S. A., Fatema, A. N., Wahab, A., Alam, M. M., Islam, M. N., ... & Ahsan, G. U. (2020). Age and sex-specific antibiotic resistance patterns among Bangladeshi patients with urinary tract infection caused by Escherichia coli. *Heliyon*, 6(6), e04161. https://www.cell.com/ heliyon/pdf/S2405
- Inggraini, M., Nurfajriah, S., Priyanto, J. A., & Ilsan, N. A. (2021). Antimicrobial susceptibility and molecular species identification of clinical carbapenem -resistant bacteria. *Biodiversitas Journal Of Biological Diversity*, 22(2). https://v2.smujo. id/biodiv/ article/view/7189
- Ismail, S. J., & Mahmoud, S. S. (2018). First detection of New Delhi metallo-β-

lactamases variants (NDM-1, NDM-2) among Pseudomonas aeruginosa isolated from Iraqi hospitals. *Iranian journal of microbiology*, 10(2), 98. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6039455/

- Jasim, H. A., Ibrahim, R. S., Abbas, B. A., Ali, B., Najim, Q., & Ali, A. (2022, January). Isolation and identification of some bacterial species from dialysis unit in Basrah. In *AIP Conference Proceedings* (Vol. 2386, No. 1, p. 020007). AIP Publishing LLC. https://pubs.aip.org/aip/ acp/article-abstract/2386/
- Junaid, K. (2021). Molecular Diversity of NDM-1, NDM-5, NDM-6, and NDM-7 Variants of New Delhi Metallo-β-Lactamases and Their Impact on Drug Resistance. *Clinical Laboratory*, (8). https://web.s. ebscohost.com
- Kaase, M., Nordmann, P., Wichelhaus, T. A., Gatermann, S. G., Bonnin, R. A., & Poirel, L. (2011). NDM-2 carbapenemase in Acinetobacter baumannii from Egypt. *Journal of Antimicrobial Chemotherapy*, 66(6), 1260-1262. https://academic.oup. com/ jac/ article/66/6/1260/719696
- Karam, M. R. A., Habibi, M., & Bouzari, S. (2019). Urinary tract infection: Pathogenicity, antibiotic resistance development effective and of vaccines against Uropathogenic Escherichia coli. Molecular immunology, 108, 56-67. https:// www. sciencedirect. com/science/ article/abs/pii/S0161589018306357
- Kareem Raheem, Z., & Abdulhamid Said, L. (2023). Antibiotic Susceptibility Profile of Bacteria Causing Aerobic Vaginitis in Women in Iraq. *Archives of Razi Institute*, 78(1), 31-43. https://archrazi.areeo.ac.ir/article
- Logan, L. K., & Weinstein, R. A. (2017). The epidemiology of carbapenemresistant Enterobacteriaceae: the impact and evolution of a global menace. *The Journal of infectious*

diseases, 215(suppl_1), S28-S36. https://academic.oup.com/jid/article /215/ suppl_1/S28/3092084

- Magliano, E., Grazioli, V., Deflorio, L., Leuci, A. I., Mattina, R., Ro-mano, P., & Cocuzza, C. E. (2012). Sex and age-dependent etiology of community-acquired urinary tract infections. *The scientific world journal*, 2012. https://www.hindawi. com/journals/tswj/2012/349597/
- Mahdi, M. S., Mohammed, S. H., Ahmed, M. M., Al-Deen, A. N., Fadhil, N., Zaid, N. Y. A., & Alkhafaji, I. H. J. (2020).Resistance Rates of the most Isolated Bacteria from different Clinical Samples, Kerbala, Iraq. *International Journal of Drug Delivery Technology (IJDDT)*, Vol. 10(3): Page 427?http://impactfactor. org/PDF/IJDDT/ 10/IJDDT, Vol.10, Issue3,Article21.pdf
- Meletis, G. (2016). Carbapenem resistance: overview of the problem and future perspectives. *Therapeutic advances in infectious disease*, 3(1), 15-21. https://journals.sagepub. com/doi/ full/10.1177/2049936115621709
- Mohammed, E. J., Allami, M., Sharifmoghadam, M. R.. & Bahreini, M. (2021). Relationship between antibiotic resistance O-serogroups patterns and in uropathogenic Escherichia coli strains isolated from Iraqi patients. Jundishapur Journal of Microbiology, 14(8). https:// brieflands.com/articles/jjm-118833. html
- Najm, R., & Hussein, J. M. (2023). Isolation and Identification of Pathogenic Escherichia coli from Different Sources in Najaf Hospital. Journal of Population Therapeutics and Clinical Pharmacology, 30(8), 459-464. https://www.jptcp.com/ index.php/ jptcp/article/view/1706
- Naqid I A, Balatay A A, Hussein N R, Saeed K A, Ahmed H A, et al. Antibiotic Susceptibility Pattern of Escherichia

coli Isolated from Various Clinical Samples in Duhok City, Kurdistan Region of Iraq. *International Journal of Infectious Diseases*, 2020;7(3):e103740. https://doi.org/ 10. 5812/iji.103740. https:// brieflands.com/ articles/iji-103740. html

- Pandey, N., & Cascella, M. (2022). Beta lactam antibiotics. In StatPearls [Internet]. StatPearls Publishing. https://europepmc.org/article/nbk/nb k545311
- Peirano, G., Schreckenberger, P. C., & Pitout, J. D. (2011). Characteristics of NDM-1-producing *Escherichia coli* isolates that belong to the successful and virulent clone ST131. *Antimicrobial agents and chemotherapy*, 55(6), 2986–2988. https://doi.org/10.1128/AAC.01763 -10
- Pokharel, S., & Adhikari, B. (2020). Antimicrobial resistance and over the counter use of drugs in Nepal. *Journal of Global Health*, 10(1). https://www.ncbi.nlm.nih.gov/ pmc/ articles/PMC7296207/
- Popova, G., Boskovska, K., Arnaudova-Danevska, I., Smilevska-Spasova, O., & Jakovska, T. (2019). Sputum quality assessment regarding sputum diagnosing culture for lower respiratory tract infections in children. Open Access Macedonian Journal of Medical Sciences, 7(12), 1926. https://www.ncbi.nlm. nih.gov/pmc/articles/
- Pulingam, T., Parumasivam, T., Gazzali, A. M., Sulaiman, A. M., Chee, J. Y., Lakshmanan, M., ... & Sudesh, K. (2022).Antimicrobial resistance: Prevalence, economic burden, of resistance mechanisms and strategies to overcome. European Journal of Pharmaceutical Sciences, 170. 106103. https://www. sciencedirect.com/ science/ article/pii/S0928098721004048

- Puvača, N., & de Llanos Frutos, R. (2021). Antimicrobial resistance in escherichia coli strains isolated from humans and Pet animals. *Antibiotics*, 10(1), 69. https://www.mdpi. com/2079-6382/10/1/69
- Qin, S., Zhou, M., Zhang, Q., Tao, H., Ye, Y., Chen, H., ... & Feng, X. (2016). First identification of NDM-4-producing *Escherichia coli* ST410 in China. *Emerging microbes & infections*, 5(1), 1-3. https://www. tandfonline.com/ doi/full/10. 1038/ emi.2016.117
- Queenan, A. M., & Bush, K. (2007). Carbapenemases: the versatile βlactamases. *Clinical microbiology reviews*, 20(3), 440-458. https:// journals.asm.org/doi/full/10. 1128/cmr.00001-07
- Ranjit, E., Raghubanshi, B. R., Maskey, S., & Parajuli, P. (2018). Prevalence of bacterial vaginosis and its association with risk factors among nonpregnant women: A hospital based study. *International journal of microbiology*, 2018. https://www. hindawi. com/journals/ ijmicro/ 2018/8349601/
- Ronald, A. (2002). The etiology of urinary tract infection: traditional and emerging pathogens. *The American journal of medicine*, 113(1), 14-19. https://www.sciencedirect. com/ science/article/abs/pii/S0002934302 010550
- Rouf, M., & Nazir, A. (2019). Aerobic profile bacteriological and antimicrobial sensitivity pattern of bacteria isolated from sterile body fluids: a study from a tertiary care hospital North India. in Microbiology Research Journal International, 28(1), 1-10. http:// impactarchive.uk/id/eprint/1978/
- Ruaa, S. H. (2023). Screening of some bacterial and fungal infections in neutropenic cancer patients in Al-Najaf governorate-Iraq. *Iranian Journal of War and Public Health*,

15(2), 1001-1007. https://ijwph. ir/browse.php?a

- Sabry, N. N., Owaied, Y. H., & Mahmoud, A. (2021.J. November). Epidemiological Study of Common Bacterial and Parasitic Infections in Some Areas of Salah Al-Din Governorate. Iraq. In IOP Conference Series: Earth and Environmental Science (Vol. 923, No. 1, p. 012081). IOP Publishing. https://iopscience.iop.org/article/10. 1088/1755-1315/923/1/012081/
- Saderi, H., & Owlia, P. (2015). Detection of multidrug resistant (MDR) and extremely drug resistant (XDR) P. aeruginosa isolated from patients in Tehran, Iran. *Iranian journal of pathology*, 10(4), 265. https://www. ncbi.nlm.nih.gov/pmc/articles/PMC 4539747/
- Salman, H. A., kamil Alhameedawi, A., Muhamad, S. M. S. G., & Taha, Z. (2022).Prevalence of multiantibiotic resistant bacteria isolated from children with urinary tract infection from Baghdad, Iraq. https://www.mbl.or.kr/journal/view. html? doi=10.48022/ mbl.2110. 10011
- Tajbakhsh, E., Tajbakhsh, S., & Khamesipour, F. (2015). Isolation and molecular detection of Gram negative bacteria causing urinary tract infection in patients referred to Shahrekord hospitals, Iran. *Iranian Red Crescent Medical Journal*, 17(5). https:// www.ncbi.nlm.nih.gov/pmc/articles /PMC4464383/
- Thapa, A., Upreti, M. K., Bimali, N. K., Shrestha, B., Sah, A. K., Nepal, K., ... & Rijal, K. R. (2022). Detection of NDM Variants (bla NDM-1, bla NDM-2, bla NDM-3) from Carbapenem-Resistant *Escherichia coli* and Klebsiella pneumoniae: First Report from Nepal. *Infection and Drug Resistance*, 4419-4434. https://www.tandfonline.com/ doi/ full/10.2147/IDR.S369934

- Tille, P. (2015). Bailey & Scott's diagnostic microbiology-E-Book. Elsevier Health Sciences. https://books. google.iq/books
- Tooke, C. L., Hinchliffe, P., Bragginton, E. C., Colenso, C. K., Hirvo-nen, V. H., Takebayashi, Y., & Spencer, J. (2019). β-Lactamases and β-Lactamase Inhibitors in the 21st Century. *Journal of molecular biology*, 431(18), 3472-3500. https: //www. sciencedirect.com/ science/article/pii/S0022283619301 822
- Uddin, T. M., Chakraborty, A. J., Khusro, A., Zidan, B. R. M., Mitra, S., Emran, T. B., ... & Koirala, N. (2021). Antibiotic resistance in microbes: History, mechanisms, therapeutic strategies and future prospects. *Journal of infection and public health*, 14(12), 1750-1766. https:// www.sciencedirect.com/science/arti cle/
- Van Duin, D., & Doi, Y. (2017). The global epidemiology of carbapenemase producing Enterobacteriaceae. *Virulence* 8: 460–469. https:// www.tandfonline.com/doi/ full/10. 1080/21505594.2016.1222343
- Vural, E., Delialioğlu, N., Ulger, S. T., Emekdas, G., & Serin, M. S. (2020). Phenotypic and molecular detection

of the metallo-Beta-lactamases in carbapenem-resistant Pseudomonas aeruginosa isolates from clinical samples. *Jundishapur Journal of Microbiology*, 13(2). https:// brieflands.com/articles/jjm-90034.html

- Whitley, V., Kircher, S., Gill, T., Hindler, J. A., O'Rourke, S., Cooper, C., ... & Denys, G. A. (2020). Multicenter evaluation of the BD Phoenix CPO detect test for detection and classification of carbapenemaseproducing organisms in clinical isolates. Journal of Clinical 58(5), e01752-19. *Microbiology*, https://journals.asm.org/ doi/full/10. 1128/jcm.01752-19
- Yadav, K., & Prakash, S. (2017). Screening of ESBL producing multi-drug resistant E. coli from urinary tract infection suspected cases in southern Terai of Nepal. *Journal of Infectious Diseases and Diagnosis*, 2(2), 116. https://d1wqtxts1xzle7.cloudfront. net/82339635/
- Zhanel, G. G., Wiebe, R., Dilay, L., Thomson, K., Rubinstein, E., Hoban, D. J., ... & Karlowsky, J. A. (2007). Comparative review of the carbapenems. *Drugs*, 67, 1027-1052. https://link.springer.com/article/10.2 165/00003495-200767070-00006