### BENEFITING OF SOME PLANT MATERIALS IN IMPROVING LIVER FUNCTIONS FOR HEPATICALLY TOXICATED RATS WITH CCL4

M. M. El-Sayed, Nehad R. El-Tahan, Asmaa M. A. Lasheen Nutrition and Food Science, Faculty of Home Economics, Minufiya University

(Received: Mar. 15, 2015)

**ABSTRACT:** The present study investigated the effects of carrot (Daucus carota, L.), prickly pear (Opuntia ficus-indica, L.) and chicory (Cichorium intybus, L.) on CCl<sub>4</sub> induced hepatotoxicity in rats .Forty five male albino rats were divided into 9 groups (five rats in each group). Two groups were controls, one fed on basal diet only (negative control) and the other fed on basal diet after injection with CCl<sub>4</sub> (positive control). The other groups were injected by CCl<sub>4</sub> then received basal diet containing carrot, chicory and prickly pear at the levels 10, 15% and 15% mixture of the tested plants. Liver damage was assessed by estimation for the plasma concentration of enzymes activities of aspartate amino transferase (AST), alanine amino transferase (ALT), lipid fraction (total cholesterol and triglyceride), cholesterol fraction (HDL-c, LDL-c, VLDL-c), Uric acid, Urea nitrogen and glucose. Results showed an improvement in case of prickly pear followed by carrot and chicory for the above parameters. The best level was 15% of these plants followed by 10% of tested plants. So, this study concluded that CCl<sub>4</sub> induced liver damage in rats can be ameliorated by administration of 15% prickly pear, carrot and chicory.

Key words: Carrot - prickly pear - chicory- liver damage - Cholesterol fraction- glucose.

#### INTRODUCTION

Liver diseases are among the most serious aliment. They may be classified as acute or chronic hepatitis (inflammatory liver diseases), hepatosis (non inflammatory diseases) and cirrhosis (degenerative disorder resulting in fibrosis of the liver) (Kumar et al., 2011).

Carbon tetrachloride (CCI<sub>4</sub>) is a wellknown hepatotoxic agent (llavarasan et al., 2003). A single dose of CCl<sub>4</sub> (20 micro 1/kg) induced hepatotoxicity, manifested biochemically by significant elevation of serum enzymes activities, such as alanine transaminase (ALT). aspartate aminotransferase (AST), and lactate dehydrogenase (LDH) (Mansour, 2000).

Phytotherapy is the treatment and prevention of diseases using plants or plant parts, such as leaves, flowers, roots, fruits, seeds, and rhizomes. Preparation made from them called medicinal plants, or herbs (Weiss and Fintelmann, 2000). Medicinal plants have very important place as they not only maintain the health and vitality of human beings and animals, but also cure

several disease, including liver disorders without causing any toxicity (Govind and Madhuri, 2010).

Daucus carota, Linn. (Family: Umberliferae) is an annual or biennial herb, whose roots are eaten raw and also cooked in many parts of the world (Muralidharan et al., 2008). Carrot polyacetylense possess allelopathic activity which may explain the historical health benefits of carrots since studies investigating  $\beta$ - carotene doesn't seem to adequately explain the reduced risk of certain types of cancer (Brandon and David, 2012).

Carrot could afford a significant protective action in the alleviation of CCI<sub>4</sub> induced hepatocellular injury (Bishayee et al., 1995). Oral administration of carrot produced significant hepatoprotection against lindane induced hepatotoxicity in rats. The increase levels of serum enzymes namely aspartate transaminase. transaminase. alanine alkaline phosphatase and the levels of thiobrabituric acid reactive substances, cholesterol, triglycerides and LDL

cholesterol in ilndane administered rates were observed to be significantly decreased in the lindane (+) carrot extract group. The carrot extract also restored the depressed antioxidants and HDL- cholesterol levels to near normal (Balasubramaniam *et al.*, 1998).

Cactus pear or prickly pear, a member of the cactacea family, originated from arid and semi- arid regions of Mexico. Cactus pear fruit containing betalain pigments is a good potential for the use as a natural food colorant. This fruit contains the red-violet betacyanins in addition to the yellow betaxanthins (Salim et al., 2009). The juice of prickly pear in nutritionally interesting and its dietary intake could provide protection against oxidative damage (Galati et al., 2003). The cactus pear fruit extract were analyzed for determined constituents: ascorbic flavonoids (quercetin, acid, isorhamnetin, myricetin, kaempferol and luteolin), betalains, taurine, total carotenoids and total phenolics. Opuntia ficus indica fruit extract had strong antioxidant capacity and taurine content (Fernandez- lopez et al., 2010).

Opuntia ficus indica fruit juice administration exerts protective and curative effects against the CCI<sub>4</sub>induced degenerative process in rat liver (Galati et al., 2005). The pears of Opuntia have been discovered to contain a plethora of biologically active compounds. Owing to their high nutritional value, in terms of dietary fiber, polyunsaturated fatty acid-rich oil, minerals, protein, and an assortment of other phytochemicals, the pears are gaining popularity as exotic, gourmet diet (Patel, 2013).

Chicory (Cichorium intybus, L.) is a perennial herb in the Asteraceae family with many commercial uses. Historically, chicory was grown by the ancient Egyptians as a medicinal plant, vegetable crop, and for animal forage (Judzntiene and Budiene, Inulin from chicory 2008). roots considered a functional food ingredient as it affects physiological and biochemical processes resulting in better health and reduction of the risk of many diseases (Kaur and Gupta, 2002).

Esculetin, aphenolic compound found in *Cichorium intybus* investigated the possible protective effect against poracetamol and CCl<sub>4</sub>- induced hepatic damage (Gilani *et al.*, 1998). Cichorium root extract therapy led to normalization of some morphofunctional liver features (decreases glycogen content and cell of necrosis and increases the number of cells with pronounced protein synthesis activity) in rats with CCl<sub>4</sub> -induced hepatitis (Krylova *et al.*, 2006) .The present study was carried out to investigate biological effects of carrot, chicory, prickly pear and mixture of the tested plants on serum parameters of liver intoxication in rats.

### MATERIALS AND METHODS Materials Plants

The tested plants were chicory (Cichorium intybus, L.), carrot (Daucus Carota, L.) and prickly pear (Oputia Ficus indica, L.). Chicory was purchased from herbalist of Cairo, Egypt. Carrot and prickly pear were purchased from the local markets of Shebin El-kom, Menofia Governorate, Egypt.

Carbon tetra choloride (CCl<sub>4</sub>) was used as an inducer for liver cirrhosis. It was purchased from El-Gomhorya Company, Cairo, Egypt as 10% liquid solution.

#### Chemical reagents

Reagent kits were purchased from Diamond Diagnostics (Egypt).

#### **Experimental animals**

Forty five white male albino rats weighing about 200  $\pm$  5g were used as experimental animals in the present investigation. They were obtained from the animal house of Research Institute of Ophthalmology, El-Giza, Egypt. They were kept under observation for one week (as adapted period) before the onset of the experiment. The animals were housed in stainless steel cages at normal atmospheric temperature (25  $\pm$  5°C) and had a 12 h light-dark cycle. Food and water were consumed *ad libitum*.

#### Methods:

#### Induction of liver intoxication in rats

Forty rats were treated subcutaneous injection of carbon tetra chloride in paraffin oil 50% V/V (2ml/kg b.wt) twice a week for two weeks (Jayaserkar *et al.*, 1997).

#### Preparation of plant powder

These plants were washed and dried in drying oven at 50°C for 3 days, then crushed and milled as a dried powder.

#### **Animals diet**

The basal diet was prepared according to AIN (1993). The vitamin mixture was prepared according to Campbell (1963), while salt mixture was prepared according to Hegsted *et al.* (1941).

#### **Experimental design**

Forty five male albino rats (200 ± 5g) were randomly divided into 9 equal groups (five rats each). All rats were fed on basal diet for one week before starting the experiment for acclimatization. After the adapted period, the initial weight was 205 ± 5g. Groups of rats were as the follows:

Group (1): Rats (n=5) were fed on basal diet only as control negative group.

Group (2): Rats (n=5) were kept without any treatment as positive control group and fed on basal diet after injection with CCl<sub>4</sub>.

Group (3): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing 10% chicory.

Group (4): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing 15% chicory.

Group (5): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing 10% carrot.

Group (6): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing 15% carrot

Group (7): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing10% prickly pear.

Group (8): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing 15% prickly pear.

Group (9): Rats (n=5) were injected by CCl<sub>4</sub> then fed on basal diet containing %15

of mixture chicory, carrot and prickly pear (1:1:1).

At the end of the experimental periods (35 days), rats were scarified using diethyl ether anesthesia at fasting state. Part of the blood was taken to determine the level of serum glucose and other portion of blood samples was collected and allowed to coagulate at room temperature; other portion of blood was added to it, EDTA (ethylene diamine tetracetic acid) and centrifuged at 3000 r.p.m for 15 minutes. Serum was carefully aspirated and transferred into clean covet tubes and stored frozen at -20°C until the time of analysis.

#### **Biochemical analysis:**

Serum Alkaline phosphatase (ALP) was determined according to the procedure of (IFCC., 1983). Aspartate aminotransferase (AST) or (GOT) glutamic -oxaloacetic transaminase and glutamic pyruvic (GPT) transaminase or Alanine aminotransferase (ALT) were carried out according to the method of Henry (1974) and Yound (1975). Serum uric acid was determined according to the method described by Fossati et al. (1980). Serum urea in plasma was determined according to the enzymatic method of Patton and Crouch Glucose was determined by (1977).enzymatic test according to Tietz (1976) and Yound (1975). Enzymatic colorimetric determination of triglycerides was carried out according to Fassati and Prencipe (1982). Total Cholesterol was determined by colorimetric method according to Allain (1974). The determination of HDL was carried out according to the method of Fnedewaid (1972) and Gordon and Amer (1977). The determination of VLDL (very low density lipoproteins) and LDL (low density lipoproteins) was carried out according to the method of Lee and Nieman (1996).

#### Statistical analysis

Statistical analysis were done using the Statistical Package for the Social Sciences (SPSS for WINDOWS, version 11.0; SPSS Inc, Chicago). Comparative analyses were conducted using the general linear models procedure (SPSS Inc). Values of P≤0.05 were considered statistically significant (Snedecor and Cochran, 1967).

#### RESULTS

# 1-Effect of feeding different levels of chicory, carrot and prickly pear on ALP, AST and ALT levels of CCl<sub>4</sub>-intoxicated rats.

The results in Table 1 indicated that mean value of ALP enzyme, rats injected with CCl4 (C +ve group) was 230.7±3.47U\L while in normal rats (C -ve) was 99.3±1.32U\L. These results denote that there was a significant increase in the mean value of ALP enzyme of rats poisoned by CCl<sub>4</sub> as compared to normal rats. The mean values of ALP of diets from groups 3, 4, 5, 6, 7, 8 and 9 were significantly higher than control negative group. Also, it could be noticed that there is no significant differences between the values of ALP enzyme of groups 4, 5 and 7. Meanwhile, rats given CCI4 then fed on diet of group 6 (rats fed on basal diet with 15% carrot) showed the lowest mean value in ALP enzyme level in the serum which was 199.4±1.35 U\L as compared to control positive group and recorded the best result of all treatment. It could be observed that due to intoxicated rats the serum levels of AST in Table 1 showed a significant increase in control positive group as compared to normal rats represents 37.25±5.82 and 22.06±1.07 respectively. There is non significant difference between groups 5, 7 and control positive group. Also, there is no significant differences between groups 3 and 9. Meanwhile, group 6 (rats fed on basal diet with 15% carrot) showed the lowest level in serum AST and recorded the best results as compared to all treatments. The ALT, in rats given CCl<sub>4</sub> then fed on all treatments groups 3, 4, 5, 6, 7, 8, and 9, was significantly higher than control negative group. Also, there is no significant difference between group 5 and control positive group. Groups 3, 4 and 8 showed similar (P>0.05) mean values of ALT. The obtained results showed that there is no significant difference in serum levels of ALT in group 6 as compared to normal rats and the best treatment was recorded for group 6 (rats fed on basal diet with 15% carrot).

Table (1): Effect of feeding different levels of chicory, carrot and prickly pear on ALP, AST and ALT levels of CCI<sub>4</sub>-intoxicated rats.

| Liver function                        | ALP (U\L)               | AST (U\L)               | ALT (U\L)               |  |
|---------------------------------------|-------------------------|-------------------------|-------------------------|--|
| Animal Groups                         | Mean± SD                | Mean ± SD               | Mean ± SD               |  |
| Group (1)<br>Control – ve             | 99.3±1.32 <sup>E</sup>  | 22.06±1.07 <sup>D</sup> | 9.51±0.94 <sup>D</sup>  |  |
| Group (2)<br>Control + ve             | 230.7±3.47 <sup>A</sup> | 37.25±5.82 <sup>A</sup> | 16.10±1.10 <sup>A</sup> |  |
| Group (3)<br>10% chicory              | 227.3±5.06 <sup>A</sup> | 28.6±61.76 <sup>B</sup> | 12.29±0.26 <sup>c</sup> |  |
| Group (4)<br>15% chicory              | 222.4±3.13 <sup>B</sup> | 24.87±0.42 <sup>c</sup> | 11.79±2.28 <sup>c</sup> |  |
| Group (5)<br>10% carrot               | 222.3±7.5 <sup>B</sup>  | 37.52±7.22 <sup>A</sup> | 15.93±1.25 <sup>A</sup> |  |
| Group (6)<br>15% carrot               | 199.4±1.35 <sup>D</sup> | 22.31±1.80 <sup>D</sup> | 10.44±0.79 <sup>D</sup> |  |
| Group (7)<br>10% prickly pear         | 219.7±2.05 <sup>B</sup> | 35.10±4.92 <sup>A</sup> | 14.43±1.21 <sup>B</sup> |  |
| Group (8)<br>15% prickly pear         | 206.3±1.35 <sup>c</sup> | 26.12±0.51 <sup>c</sup> | 12.80±0.48 <sup>C</sup> |  |
| Group (9)<br>15% mixture of all plant | 207.1±3.11 <sup>c</sup> | 31.12±0.51 <sup>B</sup> | 14.80±0.48 <sup>B</sup> |  |

<sup>\*</sup>Non significant differences between the values had the same letter. Significant at p≤0.05.

#### 2- Effect of feeding different levels of chicory, carrot and prickly pear on total cholesterol and triglyceride levels (mg/dl) of CCI<sub>4</sub> intoxicated rats.

Data in Table 2 revealed that injection of CCl<sub>4</sub> led to significant (P≤0.05) increase in serum total cholesterol level in hepatotoxic rats. The mean value ± SD of serum cholesterol in hepatotoxic control +ve group was 172.55±12.38 mg/dl as compared to  $89.78 \pm 5.25$  mg/dl in the control - ve group. The mean values of total cholesterol in rats given CCl4 then fed on all diets of groups 3, 4, 5, 6, 7, 8 and 9 were significantly lower than positive control group .There is no significant differences in total cholesterol between groups 3 and 7, as well as between groups 5, 6 and 8. Groups 6 and 8 showed the lowest levels in total cholesterol as compared to all groups. Concerning triglycerides (Table 2), data revealed that rats injected with CCI4 (control positive group) had higher (P≤0.05) value of serum levels triglycerides compared to normal rats negative group. There were nonsignificant differences between groups 3 and 7 as well as between groups 5 and 9. Meanwhile, group 8 (rats fed on diet contained 15% prickly pear) showed the lowest level in the mean value of serum triglycerides which showed 54.10 ± 4.72 mg/dl as compared to all treatment and recorded the best result.

Table (2): Effect of feeding different levels of chicory, carrot and prickly pear on total

cholesterol and triglyceride levels (mg/dl) of CCl4 intoxicated rats.

| Animal Groups                         | Lipid Fraction                         |                                  |  |
|---------------------------------------|--|----------------------------------|--|
|                                       | Total cholesterol (mg∖dl)<br>Mean ± SD | Triglyceride (mg\dl)<br>Mean± SD |  |
| Group (1)<br>Control – ve             | 89.78±5.25 <sup>F</sup>                | 39.40±0.96 <sup>G</sup>          |  |
| Group (2)<br>Control + ve             | 172.55±12.38 <sup>A</sup>              | 110.70±3.11 <sup>A</sup>         |  |
| Group (3)<br>10% chicory              | 151.29±6.92 <sup>C</sup>               | 91.30±1.44 <sup>c</sup>          |  |
| Group (4)<br>15% chicory              | 165.78±1.72 <sup>B</sup>               | 103.40±2.04 <sup>B</sup>         |  |
| Group (5)<br>10% carrot               | 124.12±4.71 <sup>E</sup>               | 70.00±3.82 <sup>D</sup>          |  |
| Group (6)<br>15% carrot               | 119.46±2.62 <sup>E</sup>               | 60.10±6.74 <sup>E</sup>          |  |
| Group (7)<br>10% prickly pear         | 154.31±6.15 <sup>C</sup>               | 96.00±2.85 <sup>c</sup>          |  |
| Group (8)<br>15% prickly pear         | 117.48±9.24 <sup>E</sup>               | 54.10±4.72 <sup>F</sup>          |  |
| Group (9)<br>15% mixture of all plant | 137.08 ±1.04 <sup>D</sup>              | 74.10±0.92 <sup>D</sup>          |  |

<sup>\*</sup>Non significant differences between the values had the same letter. Significant at p≤0.05.

# 3- Effect of feeding different levels of chicory, carrot and prickly pear on HDL-c, LDL-c, VLDL-c and the ratio between LDL-c/HDL-c levels (mg/dl) of CCl4-intoxicated rats.

It is obvious that rats injected with CCl4 (control+ ve) had serum levels HDL-c value 28.38±5.33 mg/dl. In normal rats (control-ve) the mean value of serum levels HDL-c was 60.58±3.62 mg/dl (Table 3). These finding denote that there was a significant decrease in HDL-c in the serum of rats poisoned by CCl<sub>4</sub> as compared to normal rats (Table 3). There were no significant differences among rats given CCl<sub>4</sub> and fed with diet of groups 3. 5, 6, 7 and 9. Finally group 8 (rats fed on diet contained 15% prickly pear) showed the highest increase in serum level of HDL-c and recorded the best treatments. It could be noticed that the data in table 3 evidence that, LDL-c levels was significantly elevated in control positive group to 104.03±8.07 from 20.86±2.74 mg/dl in control negative group. All rats intoxicated with CCl4 then fed on all

tested plant materials showed significant decrease in LDL-c as compared to control positive group. Group 8 (rats fed on diet contained 15% prickly pear) showed the lowest value of serum LDL-c and recorded the best results as compared to all treatments. Data presented in table 3 also indicated the effect of feeding CCI4 intoxicated rats with chicory, carrot, and prickly pear on the serum levels of VLDL-c. nonsignificant were differences between normal rats (negative control group) and rats fed with diet of group 7, 8, 9, Group 8 (rats fed on diet contained 15% prickly pear) showed the lowest decrease in serum level of VLDL-c and recorded the best compared to all groups. results as Regarding rats injected with CCI<sub>4</sub> without treatment (control positive ), the serum LDLc/HDL-c increase dramatically from 0.34 ± 0.04 for control negative group to 3.67±1.03 for control positive group. Rats fed on basel diet contained 15% prickly pear showed the lowest level in the serum LDL-c/HDL-c and recorded the best results as compared to all treatments.

Table (3): Effect of feeding different levels of chicory, carrot and prickly pear on HDL-c, LDL-c, VLDL-c and the ratio between LDL-c/ HDL-c levels (mg/dl) of CCl<sub>4</sub>-intoxicated rats.

| Lipid fraction<br>Animal Groups       | HDL-c (mg\dl)<br>Mean±SD | LDL-c<br>(mg\dl)<br>Mean± SD | VLDL-c (mg\dl)<br>Mean±SD | LDL-C/HDL-c<br>(mg\dl)<br>Mean±SD |
|---------------------------------------|--------------------------|------------------------------|---------------------------|-----------------------------------|
| Group (1)<br>Control – ve             | 60.58±3.62 <sup>A</sup>  | 20.86±2.74 <sup>H</sup>      | 7.92±0.19 <sup>E</sup>    | 0.34±0.04 <sup>G</sup>            |
| Group (2)<br>Control + ve             | 28.38±5.33 <sup>E</sup>  | 104.03±8.07 <sup>A</sup>     | 22.20±0.64 <sup>A</sup>   | 3.67±1.03 <sup>A</sup>            |
| Group (3)<br>10% chicory              | 50.05±4.08 <sup>C</sup>  | 62.97±8.48 <sup>D</sup>      | 11.42±0.94 <sup>D</sup>   | 1.26±0.22 <sup>C</sup>            |
| Group (4)<br>15% chicory              | 43.38±4.36 <sup>D</sup>  | 74.85±6.59 <sup>C</sup>      | 15.00±0.76 <sup>B</sup>   | 1.73±0.19 <sup>B</sup>            |
| Group (5)<br>10% carrot               | 49.55±1.64 <sup>C</sup>  | 83.30±3.71 <sup>B</sup>      | 13.16±0.76 <sup>C</sup>   | 1.68±0.13 <sup>B</sup>            |
| Group (6)<br>15% carrot               | 52.81±7.63 <sup>C</sup>  | 46.07±6.61 <sup>F</sup>      | 10.87±0.58 <sup>D</sup>   | 0.87±0.57 <sup>E</sup>            |
| Group (7)<br>10% prickly pear         | 52.03±4.16 <sup>C</sup>  | 57.88±8.58 <sup>E</sup>      | 8.86±0.28 <sup>E</sup>    | 1.11±0.26 <sup>D</sup>            |
| Group (8)<br>15% prickly pear         | 56.46±1.94 <sup>B</sup>  | 35.67±6.61 <sup>G</sup>      | 7.88±0.58 <sup>E</sup>    | 0.63±0.52 <sup>F</sup>            |
| Group (9)<br>15% mixture of all plant | 52.46±1.04 <sup>c</sup>  | 45.67±0.61 <sup>F</sup>      | 8.08±0.38 <sup>E</sup>    | 0.86±0.12 <sup>E</sup>            |

<sup>\*</sup>Non significant differences between the values had the same letter. Significant at p≤0.05.

# 4- Effect of feeding different levels of chicory, carrot and prickly pear on glucose levels (mg/dl) of CCI-4intoxicated rats.

Control positive group had significantly higher mean value ±SD of glucose than normal rats, it was being 140.14±0.02 and 80.05±2.11 mg/dl, respectively (Table 4). In rats given CCl4 then fed on all treatments, there were a significant increases in the glucose levels as compared to normal group. There is no significant difference between groups 3, 6 and 8, as well as between groups 4, 5, 7 and 9. Finally, group 8 (rats fed on diet contained 15% prickly pear) showed the lowest increase in glucose level which were 111.9±0.12 and recorded the best treatment.

# 5- Effect of feeding different levels of chicory, carrot and prickly pear on Uric acid and Urea nitrogen levels (mg/dl) of CCl<sub>4</sub> intoxicated rats.

Results revealed that, treated rats with CCl₄ -intoxicated diet control positive group led to a significant increase (P≤0.05) in serum uric acid when compared with control negative group. The mean values of uric acid of groups 4, 5, 6, 7, 8 and 9 were significantly lower than positive control group (Table 5). Nonsignificant differences were observed between groups 3 and control positive group. Meanwhile, group 9 (rats fed on diet contained 15% mixture of all plant materials) showed the lowest level in serum uric acid among all treatment and recorded the best results compared to normal group.

For urea nitrogen, there is nonsignificant difference between group 3 and control positive group. Also, group 4 was similar (P>0.05) to normal group. Groups 5, 6, 7, 8 and 9 showed lower (P≤0.05) urea nitrogen than both control groups. Finally, group 9 (rats fed on basal diet with 15% mixture of all plant materials) showed the lowest level of urea nitrogen among all treatment groups.

Table (4): Effect of feeding different levels of chicory, carrot and prickly pear on glucose levels (mg/dl) of CCI-4intoxicated rats.

| levels (mg/di) or col-4mtoxicated rats. |                             |  |  |
|---|-----------------------------|--|--|
| Animal Groups                           | Glucose (mg\dl)<br>Mean± SD |  |  |
| Group (1)<br>Control – ve               | 80.05±2.11 <sup>D</sup>     |  |  |
| Group (2)<br>Control + ve               | 140.14±0.02 <sup>A</sup>    |  |  |
| Group (3)<br>10% chicory                | 117.14±0.59 <sup>C</sup>    |  |  |
| Group (4)<br>15% chicory                | 130.13±0.21 <sup>B</sup>    |  |  |
| Group (5)<br>10% carrot                 | 130.12±0.85 <sup>B</sup>    |  |  |
| Group (6)<br>15% carrot                 | 116.11±0.14 <sup>C</sup>    |  |  |
| Group (7)<br>10% prickly pear           | 129.11±0.20 <sup>B</sup>    |  |  |
| Group (8)<br>15% prickly pear           | 111.9±0.12 <sup>c</sup>     |  |  |
| Group (9)<br>15% mixture of all plant   | 125.79±0.72 <sup>B</sup>    |  |  |

<sup>\*\*</sup>Non significant differences between the values had the same letter. Significant at p≤0.05

Table (5): Effect of feeding different levels of chicory, carrot and prickly pear on Uric acid and Urea nitrogen levels (mg/dl) of CCl₄ intoxicated rats.

| and orea introgen levels (ingral) or oois intexted rates. |                                |                                   |  |
|---|--------------------------------|-----------------------------------|--|
| Kidney function Animal Groups                             | Uric acid (mg\dl)<br>Mean ± SD | Urea Nitrogen (mg∖dl)<br>Mean ±SD |  |
| Group (1)<br>Control – ve                                 | 1.05±0.11 <sup>c</sup>         | 14.7±0.9 <sup>B</sup>             |  |
| Group (2)<br>Control + ve                                 | 1.14±0.12 <sup>A</sup>         | 15.6±2.2 <sup>A</sup>             |  |
| Group (3)<br>10% chicory                                  | 1.13±0.11 <sup>A</sup>         | 15.6±1.4 <sup>A</sup>             |  |
| Group (4)<br>15% chicory                                  | 1.14±0.09 <sup>A</sup>         | 14.9±0.6 <sup>B</sup>             |  |
| Group (5)<br>10% carrot                                   | 1.12±0.05 <sup>B</sup>         | 13.6±1.9 <sup>c</sup>             |  |
| Group (6)<br>15% carrot                                   | 1.11±0.15 <sup>B</sup>         | 14.1±0.8 <sup>C</sup>             |  |
| Group (7)<br>10% prickly pear                             | 1.11±0.10 <sup>B</sup>         | 13.8±0.9 <sup>c</sup>             |  |
| Group (8)<br>15% prickly pear                             | 0.9±0.72 <sup>D</sup>          | 13.1±1.2 <sup>D</sup>             |  |
| Group (9)<br>15% mixture of all plant                     | 0.79±0.72 <sup>E</sup>         | 10.1±1.2 <sup>E</sup>             |  |

<sup>\*\*</sup>Non significant differences between the values had the same letter. Significant at p≤0.05

#### **Discussion:**

The reactive electrons species from CCl<sub>4</sub> induces rat liver cirrhosis that resembles the human disease, and it can serve as a suitable animal model for studying human liver cirrhosis (An *et al.*, 2006).

Toxicity experienced by the liver during CCl<sub>4</sub> poisoning results from the production of a metabolite, CCl<sub>4</sub> which is a direct hepatotoxin responsible for change in cell permeability and it inhibits mitochondrial activity followed by cell death (Ambrose *et al.*, 2009). It has also been reported that chronic CCl<sub>4</sub> exposure produced cirrhosis in rats (Chieli and Malvadi, 2008).

An obvious sign of hepatic injury is the leakage of cellular enzyme into plasma (Schmidt *et al.*, 1975). When the liver cell plasma membrane is damaged, a variety of enzymes normally located in the cytosol are released into blood stream. Their estimation in the serum is a useful quantitative marker for the extent and type of hepatocellular damage (Ansari *et al.*, 1991). ALT and AST

are the most often used and most specific indicators of hepatic injury and represent markers of hepatocellular necrosis. These liver enzymes catalyze transfer of alphaamino group aspartate and alanine to the alpha-ketoglutaric acid. Whereas ALT is primarily localized to the liver, AST is present in a wide variety of tissue, including heart, skeletal, kidney, brain, and liver. AST is present in both the mitochondria and cytosol of hepatocytes, but ALT is found only in the cytosol. In an asymptomatic person with isolated elevation of AST or ALT level, diagnostic clues can be garnered from the degree of elevation (Rosen and Keeffe, 1998).

Results of the current study revealed that administration of CCl<sub>4</sub> caused significant increases in the levels of aspartate aminotransferase, alanine aminotransferase, glucose levels, lipid profile and kidney enzymes and these are in agreement with Túnez et al. (2005). On the other hand, the current study demonstrated that the

treatment with Cactus pear extract caused marked ameliorations of transaminase enzymes activity (ALT and AST). The results are in accordance with Tapiero et al. (2002) who showed the effect of carrot extracts on carbon tetrachloride—induced hepatotoxicity in rats.

The mechanism by which the cactus pear fruit induces its hepatoprotective activity is not certain. However, it is possible that βsitosterol, a constituent of cactus pear, which is at least partly responsible for the protective activity against hepatotoxicity (Tesoriere et al., 2004). An additional and important factor in the hepatoprotective activity of any drug is the ability of its constituents to inhibit the aromatase activity of cytochrome P-450, thereby favoring liver regeneration. On that basis, it is suggested that flavonoids in cactus pear could be a factor contributing to its hepatoprotective ability through inhibition of cytochrome P-450 aromatase (Kowalska et al., 1990). In addition, the recorded content of vitamin C in the chicory (35 -38 mg per 100 g) may also play a role in hepatoprotection. Previous in vivo studies indicate that hepatic microsomal drug metabolism decreases in ascorbic acid deficiency and is augmented when high supplements of the vitamin are given to guinea pigs (Burtis and Ashwood, 2001).

Blood glucose concentration is known to depend on the ability of the liver to absorb or produce glucose. The liver performs its glucostatic function owing to its ability to synthesize or degrade glycogen according to the needs of the organism, as well as via gluconeogenesis (Ahmed *et al.*, 2006). The blood sugar level after overnight fasting in cirrhotic patients is believed to decrease only in severe hepatic failure (Kruszynska and McIntyre, 1991). This is confirmed by our data that indicate that glucose levels in cirrhosis decreased.

#### Conclusions

The study clearly demonstrates that 15% of tested plants and its mixture have potential for treatment and prevention of CCl<sub>4</sub> -induced hepatic cytotoxicity. This study, along with other research, targets

cactus pear as a potentially safe and effective fruit that has important medicinal values and benefits.

#### REFERENCES

- Ahmed, O.M., H. Abdel Hamid, M. Bastway and N.A. Hasona (2006). Antihyperglycemic effects of Cactus pear and carrots aqueous extract in diabetic and hypercholesterolemic rats. J. Egypt. Ger. Soc. Zool., 51A: 371-393.
- AIN, (1993). American Institute of purified Diet for Laboratory Rodent, Final Report. J. Nutr., 123: 1939- 1951.
- Allain, C. (1974). Cholesterol Enzymatic colorimetric Method. J. Clin. Chem., 20:470.
- Ambrose, A. M., F. DeEds and L. J. Rather (2009). Further studies on toxicity of CCl<sub>4</sub> in rats. *Proceedings of Society of Experimental and Biological* Medicine, 74: 134-140.
- An, J. H., J. Seong, H. Oh, W. Kim, K. H. Han and Y. H. Paik (2006). Protein expression profiles in a rat cirrhotic model induced by CCl<sub>4</sub>. *Korean J.* Hepatol., 12: 93-102.
- Ansari, R. A., S. C. Tripathi, G. K. Patnaik and B. N. Dhawan (1991). Antihepatotoxic properties of picroliv, an active fraction from rhizomes of Picrorhiza kurroa. J. Ethnopharmacol., 34: 61-68.
- Balasubramaniam, P., L. Pari and V. P. Menon (1998). Protective effect of carrot (*Daucus Carota, L.*) against lindane-induced hepatotoxicity in rats. Phytother. Res., 12(6): 434-436.
- Bishayee, A., A. Sarkar and M. Chatterjee (1995). Hepatoprotective activity of carrot (*Daucus carota, L.*) against carbon tetrachloride intoxication in mouse liver. J. Ethnopharmacology, 47(2): 69-74.
- Brandon, T. M. and M. B. David (2012). Carrot Bisacetylenic oxylipins-Phytochemicals behind the mask of the super food. J. Agric. Food Chem., (10): 167-184.
- Burtis, C.A. and E.R. Ashwood (2001). Tietz's Fundamentals of Clinical Chemistry. WB Saunders, Philadelphia. 565.

- Campbell, J. A. (1963). Methodology of protein evaluation R. A. G. Nutr. Document R. 10 Led. 37: June Meeting. New York.
- Chieli, E. and G. Malvadi (2008). Role of Cyt P-450 dependent and FAA containing mono oxygenases in the bioactivation of Thioacetamide and CCl<sub>4</sub>, thiobezamide and their sulphoxides. Biochem. Pharmacol., 34: 395-396.
- Fassati, P. and L. Prencipe (1982). Triglyceride Enzymatic Colorimetric Method. J. Clin. Chem., 28:2077.
- Fernandez Lopez, J. A., L. Almela, J. M. Obon and R. Castellar (2010). Determination of antioxidant constituents in cactus pear fruits. Plant Foods Hum. Nutr., 65(3): 253-259.
- Fnedewaid, W.T. (1972). Determination of HDL. Clin. Chem., 8:499.
- Fossati, P., L. Prencipe and G. Berti (1980).

  Use of 3.5- dichloro- z-hydroxybenzenesulfonic acid/4 aminophenazone chromogenic systems in direct enzymic assay of uric acid in serum and urine. Clin. Chem., 26: 227-231.
- Galati, E. M., M. R. Mondello, D. Giuffrida, G. Dugo, N. Miceli, S. Pergolizzi and M. F. Taviano (2003). Chemical characterization and biological effects of Sicilian *Opuntia ficus indica, (L.) Mill*. Fruit Juice: antioxidant and antiulcerogenic activity. J. Agric. Food Chem., 51(17): 4903-4908.
- Galati, E. M., M. R. Mondello, E. R. Lauriano, M. F. Taviano, M. Galluazzo and N. Miceli (2005). *Opuntia ficus indica, (L.) Mill.* fruit juice protects liver from carbon tetrachloride- induced injury, phytother Res., 19(9); 796-800.
- Gilani, A. H., K. H. Janbaz and B. H. Shah (1998). Esculetin prevents liver damage induced by paracetamol and CCI<sub>4</sub>. Pharmacol Res., 37 (1):31-35.
- Gordon, T. and M. Amer (1977). Determination of HDL. J. Med., 62:707.
- Govind, P. and S. Madhuri (2010). Significance of Fruits and Vegetables In Malnutrition Cancer. PL Arch 10(2): 517-522.
- Hegsted, D., R. Mills and E. Perkins (1941). Salt Mixture. J. Biol. Chem., 138:459.

- Henry, R. J. (1974). Clinical Chemistry Principles and Techniques. 2<sup>nd</sup>ED., Harper and publisher, New York, Philadelphia.
- IFCC. (1983). Methods for measurement of catalytic concentration of Enzymes, parts
  5: IFCC, methods for alkaline hosphatase. J. Clin. Chem. Clin Biochem., 21:731-748.
- Ilavarasan, R., M. Vasudevan, S. Anbazhagan and S. Venkataraman (2003). Antioxidant activity ofthespesia populnea bark extracts against carbon tetrachloride-induced liver injury in rats. J. Ethnopharmacol., 87: 227-230.
- Jayasekhar, P., P. V. Mohanan and K. Rahinam (1997). Hepatoprotective activity of ethyl acetate extract of *Acacia catechu*. Indian. J. Pharmacology, 29:426-428.
- Judzentiene, A. and J. Budiene (2008). Volatile constituent from aerial part and roots of *Cichorium intybes*, *L.* (Chicory) grown in Lithuania. Chemija, 19(25): 25-28.
- Kaur, N. and A. K. Gupta (2002). Applications of inulin and oligofructose in health nutrition. Journal of Biosciences, 27 (7): 703 714.
- Kowalska, D.P., R.M. Feeley and D.P. Jones (1990). Use of exogenous glutathione for metabolism of peroxidized methyl linoleate in rat small intestine. J. Nutr., 120: 1115-1121.
- Kruszynska, Y.T. and N. McIntyre (1991). Carbohydrate Metabolism. In: McIntyre, N.; Benhamou, P.J.; Bircher, J.; Rizzetto, M. and Rodes, J. (Eds.) Oxford Textbook of Clinical Hepatology. Oxford University Press, Oxford. 129-143.
- Krylova, S. G., L. A. Efimova, Z. K. Vymiatina and E. P. Zueve (2006). The effect of cichorium root extract on the morphofunctional state of liver in rats with carbon tetrachloride induced hepatitis model. EKSP Klin Farmakol, 69(6): 34-36
- Kumar, C. H., A. Ramesh, J. N. Kumar and I. B. Mohammed (2011). A Review on hepatoprotective activity of medicinal plants, IJPSR, 2 (3): 501-515.

- Lee, R. D. and D. C. Nieman (1996). Nutritional Assessment. 2<sup>nd</sup> ED. Mosby, Missoun, USA.
- Mansour, M. A. (2000). Protective effects of thymoquinone and deferrio-xamine against hepatotoxicity of carbon tetrachloride in mice. Life Sci., 66(26): 2583-2591.
- Muralidharan, P., G. Balamurugan and P. Kumar (2008). Inotropic and Cardioprotective effects of *Daucus carota*, *Linn*. on isoproterenol- induced myocardial infarction. Bangladesh J. Pharmacol.,(3): 74-79.
- Patel, S. (2013). Reviewing the prospects of Opuntia pears as low cost functional Foods. Reviews in Environmental Science and Bio/Technology, 12(3):223-234.
- Patton, C. and S. Crouch (1977).

  Determination of serum urea

  Enzymatically Anal. Chem., 49.
- Rosen, H. R. and E. B. Keeffe (1998). Laboratory evaluation of the patient with signs and symptoms of liver disease. In: Brandt LJ. (Ed.) Clinical Practice of Gastroenterology. Vol 2, Churchill Livingstone, Philadelphia, 812-20.
- Salim, N., C. Abdel Wahab, C. Rabah and B. Ahcene (2009). Chemical Composition of *Opuntia ficus- Indica, (L.)* fruit. African J. Biotechnology; 8(8): 1623-1624.
- Schmidt, E., F. W. Schmidt, J. Mohr, P. Otto, I. Vido, K. Wrogeman and C. Herfarth (1975). Liver Morphology and Enzyme Release. Further studies in the isolated

- perfused rat liver. In: Keppler (Ed.) Pathogenesis and Mechanism of Liver Cell Necrosis. Medical and Technical Publishing Co. Ltd., Lancaster. 147.
- Snedecor, G. W. and W. C. Cochran (1967). Statistical Methods. 6<sup>th</sup> ED. Iowa State University press. Ames. Lowa. USA.
- Tapiero, H., K. D. Tew, G. N. Ba and G. Mathe (2002). Polyphenols: Do they play a role in the prevention of human pathologies? Biomed Pharmacother., 56: 200–207.
- Tesoriere, L., D. Butera, A. M. Pintaudi, M. Allegra and M. A. Livrea (2004). Supplementation with cactus pear (*Opuntia ficus-indica*) fruit decreases oxidative stress in healthy humans: a comparative study with vitamin C. Am. J. Clin. Nutri., 80: 391-395.
- Tietz, N.W. (1976). Fundamentals of Clinical Chemistry. Philadelphia, W.B. Saunders. P243.
- Túnez, I., M. Carmen Muñoz, M. A. Villavicencio, F. J. Medina, E. P. de Prado, I. Espejo, M. Barcos, M. Salcedo, M. Feijóo and P. Montilla (2005). Hepatoand neurotoxicity induced by Thioacetamide and CCl<sub>4</sub> Protective effects of melatonin and dimethylsulfoxide. Pharmacol. Res., 52: 223-228.
- Weiss, R. F. and V. Fintelmann (2000). Herbal Medicine 2<sup>nd</sup> ED., Georg Thieme Verlag, New York.
- Yound, D. S. (1975). Determination of GOT. Clin. Chem., 21.

## الاستفادة من بعض المواد النباتية في تحسين وظائف الكبد لدى الفئران المصابة بالتسمم الكبدى برابع كلوربد الكربون

محمد مصطفى السيد ، نهاد رشاد الطحان ، أسماء محمد عبدالمجيد الشين قسم التغذية وعلوم الأطعمة . كلية الأقتصاد المنزلي . جامعة المنوفية

#### الملخص العربي

تهدف هذه الدراسة إلى معرفة تأثير الجزر والتين الشوكى والهندباء على الفئران المصابة بالتسمم الكبدى. قسمت الفئران (45 فار ذكور بيضاء) إلى 9 مجموعات بكل مجموعة 5 فئران. تم استخدام مجموعتان ضابطتان غذيت المجموعة الأولى على الوجبة الأساسية فقط كمجموعة ضابطة سالبة والمجموعة الثانية فتغذيت على الوجبة الأساسية بعد حقنها برابع كلوريد الكربون كمجموعة ضابطة موجبة ، أما المجموعات الأخرى فتم حقنها برابع كلوريد الكربون ثم تغذيت على الوجبة الأساسية محتوية على نسبة 10%, 15 % من الجزر والتين الشوكى والهندباء و15% خليط من النباتات المستخدمة . تم تقدير وجود تلف الكبد عن طريق تقدير نشاط وتركيز الإنزيمات في البلازما وهي انزيم اسبرتات ترانس امينيز والانين امينو ترانسفيريز والكوليستيرول الكلي والتراي جلسريد والليبوبروتينات منها الليبوبروتين عالى الكثافة والليبوبروتين منخفض الكثافة و الليبوبروتين منخفض الكثافة جدا وحمض اليوريك واليوريا والجلوكوز . أظهرت النتائج وجود تحسن لهذه القياسات باستخدام الجزر والتين الشوكي جدا وحمض النتائج باستخدام نسبة 15% يليها نسبة 10% من هذه النباتات. لذلك نستخلص من هذه الدراسة ان تلف الكبد بالفئران الناجم عن رابع كلوريد الكربون يمكن تحسنه باستخدام 15% من التين الشوكي يليه الجزر والهندباء .