



## Factors Influencing Dietary Tannin Inclusion in Dairy Diets: A review

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### ABSTRACT

The objective of this paper was to evaluate factors affecting tannin dietary inclusion on enteric methane emission (CH<sub>4</sub>) and performance in dairy cows. Dairy production contributes to the greenhouse effect as it naturally emits enteric CH<sub>4</sub>. Therefore, this has sparked a need to control enteric methane emissions using anti-methane natural compounds such as tannins. Even at moderate dietary inclusions, tannin use in animal diets can occasionally reduce dairy performance and enteric CH<sub>4</sub>. This is due to the fact that most studies employ tannins to reduce enteric CH<sub>4</sub> in dairy cows excluding other influential factors by focusing on the tannin inclusion effect alone. Therefore, there is a need to study different factors that influence the effect of tannins on enteric CH<sub>4</sub> and dairy performance regardless of dietary tannin inclusion to improve the control of enteric CH<sub>4</sub> at no expense to dairy performance. Hence, there is a need to identify factors that affect dietary tannin inclusion, such as tannin source, diet and animal factors that need consideration to prevent the control of enteric CH<sub>4</sub> by tannins at the expense of animal performance. This approach would inform future studies relevant to the use of tannins in dairy diets to improve the effect of this treatment through *in vivo* and *in vitro* studies to ensure dairy production is harmless to the environment while meeting production targets.

**Keywords:** Animal-environment relationship, Dairy production, Enteric methane emissions, Tannins.

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### INTRODUCTION

The importance of dairy products to human health as a source of proteins remains undisputed. This animal production provides milk, meat, and by-product commodities such as hides to provide food security. The increase in human population necessitates an increase in animal production to meet the nutritional requirements for the survival of humanity. This demand would accelerate the negative effect of animal production on the environment. Dairy and beef production are two major sources of livestock-related enteric methane (CH<sub>4</sub>) emissions (Caro *et al.*, 2016). A by-product of rumen fermentation, which occurs when feed is transformed into nutrients, is CH<sub>4</sub>. However, CH<sub>4</sub> gas is a greenhouse gas with a heating power of 25-folds or more than that of carbon dioxide (Rira *et al.*, 2015) and constitutes about 55 % of gases released by ruminants (Rira *et al.*, 2015). Controlling CH<sub>4</sub> emissions in dairy cattle is a challenge due to the intensification of the dairy industry.

This challenge has sparked interventions into the control of ruminal digestion to discriminate against the production of CH<sub>4</sub> gas to promote animal production that is harmless to the environment (Nawab *et al.*, 2020). Therefore, phytochemicals can potentially restrict the pathway of microorganisms involved in the formation of CH<sub>4</sub> without negatively affecting the environment (Patra and Puchala, 2023). Condensed tannins are natural polyphenolic phytochemicals found in tropical and temperate foliage that can decrease methanogens and fibre digestion (Jayanegara *et al.*, 2015). This lowers the availability of hydrogen molecules, which are needed for the formation of CH<sub>4</sub> during fermentation. Another benefit of condensed tannin inclusion in dairy rations is the promotion of small-intestinal absorption of proteins by forming a protein-tannin complex that shields proteins from premature digestion in the rumen (Anas *et al.*, 2015).

Dietary inclusion of tannins is influential when studying the effect of condensed tannin inclusion on CH<sub>4</sub> emissions and ruminant performance (Denninger *et al.*, 2020). It is claimed that including condensed tannin at a rate of less than 3% in animal diets reduces CH<sub>4</sub> emissions without changing milk parameters (Naumann *et al.*, 2017). Contrarily, a different study found that tannins reduced CH<sub>4</sub> while impairing milk production (Huyen *et al.*, 2016). Milk yield and quality, nutritional digestibility, and feed intake appear to be impaired and exhibit different responses (Henke *et al.*, 2017) when condensed tannins are included in dairy rations to decrease CH<sub>4</sub> production (Cardoso-Gutierrez *et al.*, 2021).

The dietary inclusion of condensed tannins in dairy rations has been the main factor considered for the control of CH<sub>4</sub> emissions without affecting milk parameters (Avila *et al.*, 2020b), but the results produced are conflicting. The conflicting effects of tannin sources on CH<sub>4</sub> and ruminant performance could be credited to the structural chemistry of tannins. Tannins consist of prodelphinidins (PD) or procyanidins (PC) in their structural chemistry and the PD to PC ratio differs in different tannin sources (Quijada *et al.*, 2018). The inclusion level (<2%) of dietary *Acacia mearnsii* condensed tannins demonstrated a decrease in CH<sub>4</sub> and no effect on milk yield or quality, although protein digestibility was decreased (Alves *et al.*, 2017a). The 3% inclusion of *Mimosa tenuiflora* extract failed to decrease CH<sub>4</sub> (Lima *et al.*, 2019). It has been established that *A. mearnsii* condensed tannins (400 g/d) decrease both CH<sub>4</sub> and milk yield (Williams *et al.*, 2020). Other studies have proven that the inclusion level (0–3%) of quebracho or *A. mearnsii* condensed tannins does not affect milk yield even though CH<sub>4</sub> was not measured (Henke *et al.*, 2017; Gerlach *et al.*, 2018). Some studies reported that dietary inclusions of condensed tannins up to 7.5% or 10% could be used without affecting the milk yield or quality (Griffiths *et al.*, 2013; Kapp-Bitter *et al.*, 2020).

These milk performance variations for different dietary tannin inclusions suggest that other factors may need consideration to reduce CH<sub>4</sub> without negatively impacting milk production and feed intake. Therefore, factors that influence the condensed tannin effect on dairy performance and CH<sub>4</sub> need to be identified. This would improve the discovery of a dietary inclusion of tannins that depresses CH<sub>4</sub> without depressing milk or intake performance. The objective of this review was to describe tannin source, dietary and animal factors that affect tannin results on dairy performance and enteric

CH<sub>4</sub>, and the effects of *A. mearnsii* tannins on dairy performance and enteric CH<sub>4</sub>.

## 1. Intrinsic factors affecting tannin activity

The chemical composition influences the effect of tannins on animal performance and CH<sub>4</sub>. This can be attributed to the tannin effect being different depending on tannin sources.

### 1.1. Structural Form

The structural formation of condensed tannins consists of prodelphinidins (PD) or procyanidins (PC) in nature (Quijada *et al.*, 2018). Condensed tannins have both structures, but one of these structures tends to be dominant, resulting in a varying PD/PC ratio per source of condensed tannins. A higher PD/PC ratio of condensed tannins has been found to reduce *in vitro* CH<sub>4</sub> emissions better than an inverse form of the ratio (Niderkorn *et al.*, 2020). Higher PD/PC ratio condensed tannins may possess the rumen-digestion-resistant 5-deoxy flavan-3-ol subunit by stimulating the exertion of persistent anti-methanogenesis (Niderkorn *et al.*, 2020). The explanation offered is that condensed tannins with a low PD/PC ratio may contain the typical 5-hydroxyl group and this hydroxyl group is susceptible to breakdown during ruminal digestion (Naumann *et al.*, 2018).

Available studies have used PD-condensed tannins from sainfoin pellets and PC-condensed tannins from hazelnut pericarps and there have been contrasting results. *In vivo* Sainfoin inclusion decreased CH<sub>4</sub> emissions with increased milk yield without the depression of protein digestibility (Huyen *et al.*, 2016). Hazelnut inclusion *in vivo* decreased CH<sub>4</sub> emissions without a change in milk yield (Terranova *et al.*, 2021). These limited studies demonstrate that condensed tannin sources with a higher PD/PC ratio are more effective in controlling CH<sub>4</sub> emissions and have positive effects on milk performance. These findings imply that studies that use condensed tannins to decrease CH<sub>4</sub> emissions in dairy diets need to consider the PD/PC ratio of the tannin source to improve the incorporation of condensed tannins in animal diets to prevent deleterious effects on animal performance.

### 1.2. Molecular weight

Observations show that high molecular weight (MW) condensed tannins exert a higher depression of CH<sub>4</sub> emission and acetate concentration than low MW condensed tannins (Saminathan *et al.*, 2015; Petlum *et al.*, 2019). High MW condensed tannins can decrease CH<sub>4</sub> and protein digestibility without decreasing dry matter digestibility (Saminathan *et al.*, 2015). The positive association

between reduced protein digestibility and the high MW of condensed tannins suggests that using a moderate MW condensed tannins is recommended. This would prevent tannins from decreasing CH<sub>4</sub> emissions and animal performance parameters. Condensed tannins with higher MW are associated with high protein binding ability which decreases CH<sub>4</sub> but renders proteins unavailable for absorption in the small intestines and depresses milk performance (Saminathan *et al.*, 2015; Petlum *et al.*, 2019).

High MW condensed tannins may lack a consistent effect on methanogens. Higher MW condensed tannins decrease hydrophilic methanogen order (*Methanomicrobiales*, *Methanopyrales*, *Methano-coccales*, *Methanobacteriales*, *Methanocellales* and *Methanosarcinales*) and increase the methylamine using methanogens (*Methanoplasmatales* or *Thermoplasmatales*) (Saminathan *et al.*, 2016). Therefore, the mode of action of high MW condensed tannins may not be broad-spectrum but only based on suppressing a certain methanogen group. Such inconsistent effects of tannins may result in the proliferation of other methanogens. Conversely, tannin effect on methanogens may be coated because condensed tannins may fail to affect protozoa which may benefit methanogens as they depend on protozoa for methanogenesis (Sarnataro *et al.*, 2020). This could be the cause of condensed tannin inclusions being ineffective on CH<sub>4</sub> emissions in some studies (Focant *et al.*, 2019).

## 2. Dietary factors

Tannins influence ruminant performance by modulating crude protein digestion and fibre digestibility. The chemical composition of the diet supplemented with tannins also influences the effect of tannins on ruminant performance and CH<sub>4</sub>.

### 2.1. Crude protein Digestibility

One of the main challenges that result from the addition of condensed tannins to animal diets is the suppression of crude protein digestibility (Zhang *et al.*, 2019). The compromise of crude protein digestibility suppresses milk production parameters (Henke *et al.*, 2017). There is a suggestion that aiming for crude protein content that is excessive in the basal diet could counter the suppression of crude protein digestibility by dietary condensed tannins which affect milk parameters (Denninger *et al.*, 2020). Finding the crude protein inclusion level to use in a condensed tannin-enriched diet without affecting milk production remains less studied. Diets enriched with 20.4% crude protein content and 2.5% *A. mearnsii* tannins supported conditions that increased CH<sub>4</sub> emissions. In that study, there was an

increased acetate-to-propionate ratio and decreased propionate and total tract digestibility (Koenig and Beauchemin, 2018). In a tanniferous diet with low crude protein, there has been an observation that protein synthesis remains unchanged when protein digestibility is also unchanged (Avila *et al.*, 2020b).

Manipulation of tannin sources seems to be part of the solution to increase protein synthesis and offer moderate protein digestibility that can maintain animal performance. Bayberry condensed tannins (3%) depressed protein digestibility more than *A. mangium* condensed tannins (Zhang *et al.*, 2019). A similar trend persisted when *Leucaena leucocephala* decreased protein digestibility more than *A. saligna* condensed tannins in Barki rams (El-Zaiat *et al.*, 2020). Identifying tannin sources with low suppression of protein digestibility could preserve animal performance. High protein content in the diet increases protein digestibility and milk production without influencing CH<sub>4</sub> (Niu *et al.*, 2016). However, findings remain contrasting regarding dietary protein content in relation to CH<sub>4</sub> emissions and protein digestibility.

Higher dietary protein content (15.2-18.5%) increased the digestibility of crude protein but did not affect CH<sub>4</sub> emissions, milk yield and feed intake (Niu *et al.*, 2016). Increasing protein levels (10-19%) in the concentrate sometimes increases CH<sub>4</sub> emissions (Van Dung *et al.*, 2019). Increasing the crude protein content (14.1-18.1%) in the diet did not affect protein digestibility and CH<sub>4</sub> emissions (Hynes *et al.*, 2016), thus, suggesting that the protein content in the diet may need to be below 18% for CH<sub>4</sub> emissions to decrease and increase the impact of condensed tannins addition in animal diets for controlling CH<sub>4</sub>. The addition of quebracho tannin extract (0-2%) in a diet with 16% crude protein decreased protein digestibility and did not affect CH<sub>4</sub> emissions (Beauchemin *et al.*, 2007).

### 2.2. Fiber digestion

Condensed tannins seem to exert their CH<sub>4</sub> depression ability by decreasing fiber-digesting and methanogenic microorganisms (Díaz Carrasco *et al.*, 2017). This is often attributed to condensed tannins increasing propionate formation over that of acetate from the pyruvate (Moats *et al.*, 2018). Fiber digestion promotes the release of H<sub>2</sub> needed for the formation of CH<sub>4</sub>. Promoting the lower acetate-to-propionate ratio by condensed tannins decreases the H<sub>2</sub> needed for the formation of CH<sub>4</sub> (Geneviève *et al.*, 2018). Formation of strategies to ensure that condensed tannins remain effective in stimulating CH<sub>4</sub> suppression by decreasing fiber-digesting microorganisms and methanogens is needed. Exertion

of this effect by condensed tannins on methanogens may depend on the absence of microbes that counter the tannin-protein complex, decreasing the ruminal digestion of crude protein. *Selenomonas ruminantium* bacteria are reported to exert a the premature dissociation of the tannin-protein complex (Díaz Carrasco *et al.*, 2017). The blend of quebracho and chestnut tannins increased *S. ruminantium* population (Díaz Carrasco *et al.*, 2017). There has also been a further demonstration of increasing acetate and gas production in cultures with prior condensed tannins adaptation (Hoehn *et al.*, 2018).

These studies demonstrate that the resistance of rumen microbes to condensed tannins may occur. Therefore, information on the response of fibrolytic microbes to sources of condensed tannins is necessary to improve the control of CH<sub>4</sub> production using condensed tannins.

### 2.3.Substrate

The source of the basal diet remains part of the contributing factors in the conflicting results found when condensed tannins are incorporated in dairy diets to decrease CH<sub>4</sub> without limiting animal performance. Adding condensed tannins in accordance with the basal diet source could assist in decreasing CH<sub>4</sub> emissions without depressing milk production and other performance parameters. Increasing the concentrate-to-forage ratio (40:60 and 60:40) suppressed CH<sub>4</sub> emissions (Na *et al.*, 2013). Increasing forage levels (37.4% or 53.3%) decreased milk yield and feed intake but increased CH<sub>4</sub> emissions per day without affecting protein digestibility (Niu *et al.*, 2016). This suggests that a certain forage-to-concentrate ratio in the total-mixed ration could suppress CH<sub>4</sub> emission and improve milk production parameters. Hence, the addition of condensed tannins in the diet may lack suppression of CH<sub>4</sub> emissions and the improvement of milk production if the concentrate-to-forage ration in the total mixed ration is low. This could explain findings that demonstrated that total mixed ration enriched with condensed tannins (0-2%) did not affect yield, fat and protein of milk, dry matter intake and CH<sub>4</sub> emission (Denninger *et al.*, 2020).

The selective mixing of pastures in pasture-based animals could suppress CH<sub>4</sub> and enhance milk yield in animal diets by condensed tannins in pasture-based dairy cows. Including white clover in ryegrass pasture at a rate of 0-60% increased milk yield and CH<sub>4</sub> suppression (Lee *et al.*, 2004). Feeding less fibrous and highly digestible pastures or tannin-rich pastures can improve the efficacy of tannin sources inclusion level to control methanogenesis without affecting animal parameters.

### 3. Animal factors

The effect of dietary tannins on ruminant performance may be influenced by lactation factors, gut infestations, animal genetics, and how rumination time and digesta passage rate respond to dietary tannins.

#### 3.1. Lactation

The current findings have shown that dietary condensed tannin inclusion levels are suitable for inclusion in diets of dry cows as milk yield may be interrupted. As days in milk increase, CH<sub>4</sub> emissions increase in dairy cows (Wall *et al.*, 2012; Alstrup *et al.*, 2015). The explanation for this observation was that as days in milk increase, the passage rate in cows decreases, causing feed to be retained in the rumen, increasing the digestibility of fibre, which is conducive to CH<sub>4</sub> formation (Alstrup *et al.*, 2015). The increase in parity of the animal has also been associated with the increase in CH<sub>4</sub> (Bittante *et al.*, 2018). Further findings have shown that primiparous cows have a diverse methanogenic population compared to multiparous cows (Kumar *et al.*, 2015). This suggests that as parity increases methanogens of different survival modes persist making the need for a rumen fermentation additive that can control various methanogen archaea at one time essential. This further emphasizes the need to understand the effect of condensed tannins per methanogen type in cows at different parities to control CH<sub>4</sub> production without limiting animal performance.

#### 3.2. Gut infestations

Gastrointestinal nematode infestations have increased CH<sub>4</sub> in ruminants (Fox *et al.*, 2018). The mechanism by which gastrointestinal nematodes increase CH<sub>4</sub> in the rumen is unclear but could be attributed to these parasites contributing to the effects that support CH<sub>4</sub> production such as fibre digestion. Gastrointestinal nematodes have been associated with an increased protozoa population, producing hydrogen that methanogens use to produce CH<sub>4</sub> (Szulc *et al.*, 2020). The effect of gastrointestinal nematodes on CH<sub>4</sub> may influence the tannin effect on CH<sub>4</sub>. It has been reported that nematode infestations render condensed tannins ineffective against CH<sub>4</sub> emission (Lima *et al.*, 2019). Future studies need to investigate the effect of condensed tannins on gastrointestinal nematodes and CH<sub>4</sub> emissions in one experiment to better understand the relationship between condensed tannins inclusion levels and CH<sub>4</sub> per gastrointestinal infestation in animals.

Studies on the control of gastrointestinal nematodes using condensed tannins need to consider investigating the effect of condensed tannins on the different life stages of these parasites. Findings suggest that dietary condensed tannin

supplementation can decrease faecal egg count and larval populations of gastrointestinal nematodes (Pathak, 2013; Costa-Júnior *et al.*, 2014). Drenching condensed tannins according to body weight to control gastrointestinal nematodes is a promising intervention as it can control gastrointestinal nematodes (Ahmed *et al.*, 2014; Mhlongo, 2018). This intervention is more practical for uniformly controlling gastrointestinal nematodes in animals. An *in vitro* study demonstrated that condensed tannins decrease eggs' hatchability and larval development (Molan and Faraj, 2010). In most studies, the response to condensed tannins of gastrointestinal nematodes has been conducted more in small rather than large stock. Liver flukes rather than nematodes being the main parasites affecting cattle limit the extrapolation of the response of nematodes to condensed tannins in cattle. Studying the response of CH<sub>4</sub> emissions in infested cattle fed condensed tannins may improve the development of the inclusion level that decreases CH<sub>4</sub> without disrupting animal performance.

### 3.3. Animal genetics

Understanding the effect of genetics on CH<sub>4</sub> is part of the under-considered factors that could assist in better controlling CH<sub>4</sub> emission and performance in animals using condensed tannins. Crossbred cows emit more CH<sub>4</sub> than purebred cows (Pedreira *et al.*, 2009). The breed also affects CH<sub>4</sub> emissions as Jersey cows were found to emit less CH<sub>4</sub> than the Holstein-Friesian breed (Ricci *et al.*, 2014). The lactation stage influences CH<sub>4</sub> in cows as dry cows have been found to emit less CH<sub>4</sub> than lactating cows (Dall-Orsoletta *et al.*, 2016). Ruminant animal species influence CH<sub>4</sub> emissions in animals as cattle have been found to emit more CH<sub>4</sub> gas than sheep or goats (Giamouri *et al.*, 2023). Animals with a lower feed intake emit fewer CH<sub>4</sub> emissions (Bell *et al.*, 2011). Rumination time has a genetic positive correlation with the yield of milk and protein and a negative genetic correlation with CH<sub>4</sub> yield (López-Paredes *et al.*, 2020).

### 3.4. Digesta passage rate

Tannin dietary additions decrease nutrient digestibility (Ahnert *et al.*, 2015). Tannins decrease fiber digestion which affects the digesta passage rate. This is due to the positive correlation between feed intake and the passage rate of digesta (Al-Kindi *et al.*, 2017). Increased mean retention time (MRT) of digesta has been found to increase CH<sub>4</sub> emission and the digestion of fibre and organic matter which form conditions that favour the production of CH<sub>4</sub> gas (Huhtanen *et al.*, 2016). Low MRT decreases CH<sub>4</sub> emissions by prioritising energy for the improvement of the efficiency of microbial protein synthesis

instead of volatile fatty acids (VFAs) (Tymensen *et al.*, 2012). This implies that this technique removes hydrogen which is needed for the formation of CH<sub>4</sub> gas by decreasing the acetate-to-propionate ratio. Contrary to the rumen, the addition of carob leaf condensed tannins increased MRT of liquid digesta in the intestines, total gastrointestinal tract (GIT), omasum and abomasum, and foregut, but solid digesta MRT increased in the intestines and total GIT (Silanikove *et al.*, 2001).

Quebracho tannin inclusion (2 or 4%) did not affect total MRT, caecum MRT or passage rate in the mixing compartment (Al-Kindi *et al.*, 2017). *A. mearnsii* condensed tannin (0–2.25%) decreased the disappearance rate of rumen solids, digestion rate in the rumen and passage rate of undigested residues (Tseu *et al.*, 2020). The difference in the effect of condensed tannins on the passage rate of digesta may be due to feed intake. These studies demonstrated that feed intake increased when the passage rate was not changed but decreased where the passage rate increased.

### 3.5. Rumination Time

Regarding the feeding behaviour of ruminants, higher rumination time has been associated with high CH<sub>4</sub> emissions (Watt *et al.*, 2015). This finding makes studying the feeding behaviour of animals fed condensed tannin-enriched diets important for improving animal performance and decreasing CH<sub>4</sub> emissions. *A. mearnsii* condensed tannin extract powder inclusion (0–2.25%) (Tseu *et al.*, 2020) and *Mimosa tenuiflora* condensed tannin extract inclusion (0–3%) (Lima *et al.*, 2019) did not affect the total ruminating time but decreased total eating time. Wherein chestnut condensed tannin powder inclusion (0–10%) (Kapp-Bitter *et al.*, 2020) and tanniferous Sainfoin hay inclusion (0–1%) (Scharenberg *et al.*, 2009) lacked an effect on rumination and eating time. *Mimosa tenuiflora* condensed tannins extract (0–7.5%) increased ruminating, eating and chewing time and decreased idling time (Nascimento *et al.*, 2021).

This result suggests that animals fed diets with added condensed tannins tend to adopt strategies to compensate for the decrease in digestibility that is associated with condensed tannin addition. Those strategies may include decreasing eating time by eating the feed in small amounts to deal with the astringent effect in tannin-enriched feeds if the condensed tannins used have high astringency. This also suggests that the astringency of the condensed tannins may disturb the normal breakdown of feed in animals. This results in increased rumination time to break down the feed into smaller particles. Rumination time seems to also be influenced by the

lactation stage of the animal. Cows in early lactation ruminate less due to the smaller rumen size but gradually ruminate more as the rumen increases in size and so does CH<sub>4</sub> emission (López-Paredes et al., 2020). This suggests inclusion level of the effect of condensed tannins on CH<sub>4</sub> emission and milk production needs to correlate with the lactation stage to improve the CH<sub>4</sub>-decreasing while maintaining the milk production effect on condensed tannin-rich diets.

#### 4. *A. mearnsii* tannin effect on dairy performance

*A. Mearnsii* is one of the most commonly used tannin sources to control CH<sub>4</sub> and improve dairy production. However, this tannin source has contrasting dairy performance and CH<sub>4</sub> results (Lazzari et al., 2023). The contrast in results may be due to a low understanding of the effect of this tannin source on nutrient digestibility, feed intake, and rumen fermentation using similar dietary inclusions.

##### 4.1. Rumen volatile fatty acids

Rumen volatile fatty acids (VFAs) influence CH<sub>4</sub> and milk performance. Acetate correlates with milk fat (Seymour et al., 2005) and CH<sub>4</sub>. While propionate negatively correlates with CH<sub>4</sub> and positively correlates with milk performance (Wu et al., 2021), *A. mearnsii* tannins need to be studied for their effect on rumen VFAs. The addition of *A. mearnsii* tannins (0–41 g/d) decreased acetate but increased propionate and the acetate–propionate ratio (ATPR) (Carulla et al., 2005). Dietary inclusions (0–1.49% DM) of *A. mearnsii* tannins have been noted for decreased ATPR and acetate and increased propionate and total VFAs (Krueger et al., 2010). Dietary inclusions of *A. mearnsii* tannins (0–1.5%) did not affect acetate, propionate, butyrate, ATPR or total VFAs (Perna Junior et al., 2022). Adding 0–2.5% of *A. mearnsii* tannins affected ATPR, butyric acid, decreased propionic acid, and did not affect acetic acid (Koenig and Beachemin, 2018). Encapsulated or non-encapsulated *A. mearnsii* tannin did not affect rumen VFAs at 2% dietary inclusions (Ibrahim and Hassen, 2022). Dietary inclusions of *A. mearnsii* tannins have been found to have no effect on rumen VFAs except for decreasing acetate and butyrate (Avila et al., 2020a). These results suggest that tannins have a contrasting effect on rumen VFAs, which may lead to an inconsistent effect on CH<sub>4</sub> and milk performance.

##### 4.2. Milk performance

Tannins benefit milk performance as they bind to proteins in the rumen. The protein-tannin complex reduces rumen crude protein digestion, increasing crude protein digestion in the small intestines. Increased rumen undegradable protein

increases milk yield (Mikolayunas-Sandroek et al., 2009). Adding *A. mearnsii* tannins (2% DM) has been found to not affect milk yield and milk fat or protein percentages (Alves et al., 2017b; Orlandi et al., 2020a). The dietary addition of *A. mearnsii* tannins (0–2% DM) did not affect milk parameters (Avila et al., 2020b). Inclusion at 0–3% of *A. mearnsii* tannins has been reported to have no effect on milk parameters (Gerlach et al., 2018). The addition of 0–1% of *A. mearnsii* tannin in diets has been reported to have no effect on milk parameters (Orlandi et al., 2020). Drenching cows with *A. mearnsii* tannin (0–400 g/day) decreased milk yield and fat but did not affect milk protein (Williams et al., 2020). Similarly, Grainger et al., (2009) drenched cows with *A. mearnsii* tannins and noted decreased milk yield and no effect on milk protein or fat percentages. These results demonstrate that dietary inclusions of *A. mearnsii* of 0–3% are ineffective in modulating milk parameters while drenching 200–400 *A. mearnsii* would negatively affect milk yield.

##### 4.3. Nutrient intake

Dairy performance relies on the optimum nutrient intake to maintain dairy production. *A. mearnsii* tannins need to be evaluated for their effect on the intake to prevent the decrement of CH<sub>4</sub> at the expense of milk performance. Dietary incorporation of *A. mearnsii* tannins (0–2% DM) has been noted to have no effect on the intake of DM, OM and NDF (Avila et al., 2020a). Similarly, the addition of *A. mearnsii* tannins (4.2% DM) did not affect DM, OM, CP or NDF intakes (Adejoro et al., 2020). The addition of lipid-encapsulated or none-encapsulated *A. mearnsii* (0 or 4% DM) tannin did not affect DM, OM, NDF or ADF intake (Adejoro et al., 2019). Intraruminal infusion of *A. mearnsii* tannins (0–6% DM) decreased DM, OM, and NDF intake (Kozloski et al., 2012). Dietary supplementation of *A. mearnsii* tannins (0–8% DM) decreased DM, CP, and ether extract intake (de S. Costa et al., 2021a). These results show that *A. mearnsii* inclusion of up to 4% does not affect the intake of nutrients. However, nutrient intake decreases when the inclusion of *A. mearnsii* tannins increases to 8%. This may be due to the astringent effect of tannins that decreases intake. Interventions such as encapsulation of tannins have been tried to evade the effect of tannin astringency (Adejoro et al., 2019).

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#### 4.4. Nutrient digestibility

*A. Mearnsii* tannins are to be investigated for their effect on nutrient digestibility to prevent the limitation of dairy performance. This is due to tannins' ability to control CH<sub>4</sub> by decreasing fiber and crude protein digestion in the rumen. Supplementation of *A. mearnsii* tannins decreased DM, NDF, ADF and hemicellulose digestibility but did not affect CP digestibility (Carulla *et al.*, 2005). *A. mearnsii* tannins addition in diets decreased only crude protein digestibility (Avila *et al.*, 2020b). The dietary addition of lipid-encapsulated *A. mearnsii* tannins (0–4% DM) increased the digestibility of DM, OM, CP, NDF and ADF (Adejoro *et al.*, 2020). *A. mearnsii* addition in diets (0–8%) decreased the digestibility of DM, OM, CP and NDF (de S. Costa *et al.*, 2021a). *A. mearnsii* tannins additions (0–2% DM) increased DM and decreased NDF digestibility (Avila *et al.*, 2020b). The addition of *A. mearnsii* tannins at a rate of 0.6 % DM (Perna Junior *et al.*, 2022) and 0.77 % DM (Orlandi *et al.*, 2020a) did not affect the digestibility of nutrients. These studies show that moderate inclusions of *A. mearnsii* tannins do not affect nutrient digestibility. However, 8 % tannin inclusion in diets will likely decrease nutrient digestibility. This may be due to the tannin-protein complex not reasonably reversing in the abomasum and small intestines to allow nutrient digestibility. This shows that encapsulation of tannins may be beneficial in preventing the adverse effect of tannins on nutrient digestibility.

#### 4.5. Enteric CH<sub>4</sub>

Dietary inclusions of *A. mearnsii* tannins (0–2% DM) decreased CH<sub>4</sub> (Alves *et al.*, 2017b). Dietary inclusions of *A. mearnsii* tannins (0–20%) decreased *in vitro* CH<sub>4</sub> (Hassanat and Benchaar, 2013). Inclusions of *A. mearnsii* tannins (0–4.2% DM) did not affect CH<sub>4</sub> (Adejoro *et al.*, 2020).

*Acacia mearnsii* addition in diets (0–1.5% DM) decreased CH<sub>4</sub> (Perna Junior *et al.*, 2022). Dietary *A. mearnsii* tannins (0–4% DM) decreased CH<sub>4</sub> but had a better effect when they were not encapsulated (Adejoro *et al.*, 2019). Drenching *A. mearnsii* tannins (0–263 g/d) decreased CH<sub>4</sub> (Grainger *et al.*, 2009). However, 0–0.6% DM of *A. mearnsii* tannins decreased CH<sub>4</sub> (Perna Junior *et al.*, 2022). Condensed tannins control CH<sub>4</sub> by decreasing rumen microbial populations. Tannin decreases CH<sub>4</sub> by reducing methanogens and fibrolytic rumen populations. During fermentation, fibrolytic microbes release H<sub>2</sub>, which methanogens use for methanogenesis. Dietary inclusions of *A. mearnsii* tannins (0–2% DM) decreased protozoa populations (*Entodinium*) (Avila *et al.*, 2020b). Dietary additions of *A. mearnsii* tannins (0–0.6% DM) did not affect protozoa populations (Kozloski *et al.*, 2012). Tannin introduction to animals by drenching rather than diet incorporation may be a more effective way to control CH<sub>4</sub> without affecting feed palatability.

### CONCLUSION

Basal diet source, mode of action of the tannin source, condensed tannin source, crude protein content, lactation stage, animal genetics, gut health, rumination time, molecular weight of tannin source, on-farm applicability, digesta passage rate and structural form of condensed tannins may influence the effect of the inclusion level of condensed tannins on the control of CH<sub>4</sub> production and performance in ruminants. There is a lack of consistent consideration of these factors in some of the available studies when exploring the inclusion of condensed tannins in the control of CH<sub>4</sub> production, which perpetuates the inconsistent results. These variables need to be considered to improve the consistency of the effect of the condensed tannins as a treatment for reversing CH<sub>4</sub> production and improving performance in ruminants.

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#### Conflicts of interest

The authors declare that there is no conflict of interest regarding the research data and tools used in this review.

### REFERENCES

- ADEJORO, F. A., HASSEN, A., and AKANMU, A. M., 2019. Effect of lipid-encapsulated acacia tannin extract on feed intake, nutrient digestibility and methane emission in sheep. *Animals* 9 <https://doi.org/10.3390/ani9110863>

- ADEJORO, F. A., HASSEN, A., and AKANMU, A. M., 2019.** Effect of lipid-encapsulated acacia tannin extract on feed intake, nutrient digestibility and methane emission in sheep. *Animals* 9 <https://doi.org/10.3390/ani9110863>
- ADEJORO, F. A., HASSEN, A., AKANMU, A. M., and MORGAVI, D. P., 2020.** Replacing urea with nitrate as a non-protein nitrogen source increases lambs' growth and reduces methane production, whereas acacia tannin has no effect. *Anim. Feed Sci. Technol.* 259, 114360 <https://doi.org/10.1016/J.ANIFEEDSCI.2019.114360>
- AHMED, M. A., BASHA, N. A., and NS AHLAI, I. V., 2014.** Wattle tannins as control strategy for gastrointestinal nematodes in sheep. *African J. Agric. Res.* 9 <https://doi.org/10.5897/ajar2014.8718>.
- AHNERT, S., DICKHOEFER, U., SCHULZ, F., and SUSENBETH, A., 2015.** Influence of ruminal Quebracho tannin extract infusion on apparent nutrient digestibility, nitrogen balance, and urinary purine derivatives excretion in heifers. *Livest. Sci.* 177 <https://doi.org/10.1016/j.livsci.2015.04.004>.
- AL-KINDI, A., SCHIBORRA, A., BUERKERT, A., and SCHLECHT, E., 2017.** Effects of quebracho tannin extract and activated charcoal on nutrient digestibility, digesta passage and faeces composition in goats. *J. Anim. Physiol. Anim. Nutr. (Berl.)* 101, 576–588 <https://doi.org/10.1111/jpn.12461>.
- ALSTRUP, L., HELLWING, A. L. F., LUND, P., and WEISBJERG, M. R., 2015.** Effect of fat supplementation and stage of lactation on methane production in dairy cows. *Anim. Feed Sci. Technol.* 207 <https://doi.org/10.1016/j.anifeedsci.2015.05.017>.
- ALVES, T. P., DALL-ORSOLETTA, A. C., and NUNESRIBEIRO-FILHO, H. M., 2017a.** The effects of supplementing *Acacia mearnsii* tannin extract on dairy cow dry matter intake, milk production, and methane emission in a tropical pasture. *Trop. Anim. Health Prod.* 49, 1663–1668 <https://doi.org/10.1007/s11250-017-1374-9>.
- ALVES, T. P., DALL-ORSOLETTA, A. C., and RIBEIRO-FILHO, H. M. N., 2017b.** The effects of supplementing *Acacia mearnsii* tannin extract on dairy cow dry matter intake, milk production, and methane emission in a tropical pasture. *Trop. Anim. Health Prod.* 49, 1663–1668.
- ANAS, M. A., YUSIATI, L. M., KURNIAWATI, A., and HANIM, C., 2015.** Evaluation of *Albazia chinensis* as tannins source for in vitro methane production inhibitor agents sheep rumen liquor. In *The 6th International Seminar on Tropical Animal Production: The 6th International Seminar on Tropical Animal Production*.
- ANIMUT, G., PUCHALA, R., GOETSCH, A. L., PATRA, A. K., SAHLU, T., VAREL, V. H., and WELLS, J., 2008.** Methane emission by goats consuming different sources of condensed tannins. *Anim. Feed Sci. Technol.* 144 <https://doi.org/10.1016/j.anifeedsci.2007.10.015>.
- AVILA, A. S., ZAMBOM, M. A., FACCENDA, A., FISCHER, M. L., ANSCHAU, F. A., VENTURINI, T., TININI, R. C. R., DESSBESELL, J. G., and FACIOLA, A. P., 2020a.** Effects of black wattle (*Acacia mearnsii*) condensed tannins on intake, protozoa population, ruminal fermentation, and nutrient digestibility in jersey steers. *Animals* 10 <https://doi.org/10.3390/ani10061011>.
- AVILA, A. S., ZAMBOM, M. A., FACCENDA, A., WERLE, C. H., ALMEIDA, A. R. E., SCHNEIDER, C. R., GRUNEVALD, D. G., and FACIOLA, A. P., 2020b.** Black wattle (*Acacia mearnsii*) condensed tannins as feed additives to lactating dairy cows. *Animals* 10 <https://doi.org/10.3390/ani10040662>.
- BEAUCHEMIN, K. A., MCGINN, S. M., MARTINEZ, T. F., and MCALLISTER, T. A., 2007.** Use of condensed tannin extract from quebracho trees to reduce methane emissions from cattle. *J. Anim. Sci.* 85 <https://doi.org/10.2527/jas.2006-686>.
- BELL, M. J., WALL, E., SIMM, G., and RUSSELL, G., 2011.** Effects of genetic line and feeding system on methane emissions from dairy systems. *Anim. Feed Sci. Technol.* 166, 699–707.
- BITTANTE, G., CECCHINATO, A., and SCHIAVON, S., 2018.** Dairy system, parity, and lactation stage affect enteric methane production, yield, and intensity per kilogram of milk and cheese predicted from gas chromatography fatty acids. *J. Dairy Sci.* 101 <https://doi.org/10.3168/jds.2017-13472>.
- CARDOSO-GUTIERREZ, E., ARANDA-AGUIRRE, E., ROBLES-JIMENEZ, L. E., CASTELÁN-ORTEGA, O. A., CHAY-CANUL, A. J., FOGGI, G., ANGELES-HERNANDEZ, J. C., VARGAS-BELLO-PÉREZ, E., and GONZÁLEZ-RONQUILLO, M., 2021.** Effect of tannins from tropical plants on methane production from ruminants: A systematic review. *Vet. Anim. Sci.* 14 <https://doi.org/10.1016/j.vas.2021.100214>.
- CARO, D., KEBREAB, E., and MITLOEHNER, F. M., 2016.** Mitigation of enteric methane emissions from global livestock systems through nutrition strategies. *Clim. Change* 137 <https://doi.org/10.1007/s10584-016-1686-1>.
- CARULLA, J. E., KREUZER, M., MACHMÜLLER, A., and HESS, H. D., 2005.** Supplementation of *Acacia mearnsii* tannins decreases methanogenesis and urinary nitrogen in forage-fed sheep. *Aust. J. Agric. Res.* 56, 961–970.
- COSTA-JÚNIOR, L. M., COSTA, J. S., LÔBO, Í. C. P. D., SOARES, A. M. S., ABDALA, A. L., CHAVES, D. P., BATISTA, Z. S., and LOUVANDINI, H., 2014.** Long-term effects of drenches with condensed tannins from *Acacia mearnsii* on goats naturally infected with gastrointestinal nematodes. *Vet. Parasitol.* 205 <https://doi.org/10.1016/j.vetpar.2014.07.024>.
- DALL-ORSOLETTA, A. C., ALMEIDA, J. G. R., CARVALHO, P. C. F., SAVIAN, J. V., and RIBEIRO-FILHO, H. M. N., 2016.** Ryegrass pasture combined with partial total mixed ration reduces enteric methane emissions and maintains the performance of dairy cows during mid to late lactation. *J. Dairy Sci.* 99 <https://doi.org/10.3168/jds.2015-10396>.
- DENNINGER, T. M., SCHWARM, A., BIRKINSHAW, A., TERRANOVA, M., DOHME-MEIER, F., MÜNGER, A., EGGERSCHWILER, L., BAPST, B., WEGMANN, S., CLAUSS, M., OTHERS, and KREUZER, M., 2020.** Immediate effect of *Acacia mearnsii* tannins on methane emissions and milk fatty acid profiles of dairy cows. *Anim. Feed Sci. Technol.* 261, 114388



- <https://doi.org/10.1016/j.anifeedsci.2019.114388>.
- DÍAZ CARRASCO, J. M., CABRAL, C., REDONDO, L. M., PIN VISO, N. D., COLOMBATTO, D., FARBER, M. D., and FERNÁNDEZ MIYAKAWA, M. E., 2017. Impact of Chestnut and Quebracho Tannins on Rumen Microbiota of Bovines. *Biomed Res. Int.* 2017 <https://doi.org/10.1155/2017/9610810>.
- VAN DUNG, D., PHUNG, L. D., and ROUBÍK, H., 2019. Performance and estimation of enteric methane emission from fattening Vietnamese yellow cattle fed different crude protein and concentrate levels in the diet. *Adv. Anim. Vet. Sci.* 7 <https://doi.org/10.17582/journal.aavs/2019/7.11.962.968>.
- EL-ZAIAT, H. M., KHOLIF, A. E., MOHARAM, M. S., ATTIA, M. F., ABDALLA, A. L., and SALLAM, S. M. A., 2020. The ability of tanniniferous legumes to reduce methane production and enhance feed utilization in Barki rams: in vitro and in vivo evaluation. *Small Rumin. Res.* 193 <https://doi.org/10.1016/j.smallrumres.2020.106259>.
- FOCANT, M., FROIDMONT, E., ARCHAMBEAU, Q., DANG VAN, Q. C. C., and LARONDELLE, Y., 2019. The effect of oak tannin (*Quercus robur*) and hops (*Humulus lupulus*) on dietary nitrogen efficiency, methane emission, and milk fatty acid composition of dairy cows fed a low-protein diet including linseed. *J. Dairy Sci.* 102, 1144–1159 <https://doi.org/10.3168/jds.2018-15479>.
- FOX, N. J., SMITH, L. A., HOUDIJK, J. G. M., ATHANASIADOU, S., and HUTCHINGS, M. R., 2018. Ubiquitous parasites drive a 33% increase in methane yield from livestock. *Int. J. Parasitol.* 48 <https://doi.org/10.1016/j.ijpara.2018.06.001>.
- GENEVIÈVE, Z., ADAMA, K., BALÉ, B., CORRÊA, P. S., LEMOS, L. N., VINCENT, N., HAMADOU, T. H., HERVÉ, H., HELDER, L., and LUIZ, A. A., 2018. In vitro rumen fermentation characteristics, methane production and rumen microbial community of two major acacia species used in Sahelian region of Burkina Faso. *Trop. Subtrop. Agroecosystems* 21 <https://doi.org/10.56369/tsaes.2527>.
- GERLACH, K., PRIES, M., THOLEN, E., SCHMITHAUSEN, A. J., BÜSCHER, W., and SÜDEKUM, K. H., 2018. Effect of condensed tannins in rations of lactating dairy cows on production variables and nitrogen use efficiency. *Animal* 12 <https://doi.org/10.1017/S1751731117003639>.
- GIAMOURI, E., ZISIS, F., MITSIOPOULOU, C., CHRISTODOULOU, C., PAPPAS, A. C., SIMITZIS, P. E., KAMILARIS, C., GALLIOU, F., MANIOS, T., MAVROMMATIS, A., and TSIPLAKOU, E., 2023. Sustainable Strategies for Greenhouse Gas Emission Reduction in Small Ruminants Farming. *Sustain.* 15 <https://doi.org/10.3390/su15054118>.
- GRAINGER, C., CLARKE, T., AULDIST, M. J., BEAUCHEMIN, K. A., MCGINN, S. M., WAGHORN, G. C., and ECKARD, R. J., 2009. Potential use of *Acacia mearnsii* condensed tannins to reduce methane emissions and nitrogen excretion from grazing dairy cows. *Can. J. Anim. Sci.* 89 <https://doi.org/10.4141/cjas08110>.
- GRIFFITHS, W. M., CLARK, C. E. F., CLARK, D. A., and WAGHORN, G. C., 2013. Supplementing lactating dairy cows fed high-quality pasture with black wattle (*Acacia mearnsii*) tannin. *Animal* 7 <https://doi.org/10.1017/S1751731113001420>.
- HASSANAT, F., and BENCHAAAR, C., 2013. Assessment of the effect of condensed (acacia and quebracho) and hydrolysable (chestnut and valonea) tannins on rumen fermentation and methane production in vitro. *J. Sci. Food Agric.* 93, 332–339 <https://doi.org/10.1002/jsfa.5763>.
- HENKE, A., DICKHOEFER, U., WESTREICHER-KRISTEN, E., KNAPPSTEIN, K., MOLKENTIN, J., HASLER, M., and SUSENBETH, A., 2017. Effect of dietary Quebracho tannin extract on feed intake, digestibility, excretion of urinary purine derivatives and milk production in dairy cows. *Arch. Anim. Nutr.* 71 <https://doi.org/10.1080/1745039X.2016.1250541>.
- HOEHN, A. N., TITGEMEYER, E. C., NAGARAJA, T. G., DROUILLARD, J. S., MIESNER, M. D., and OLSON, K. C., 2018. Effects of high condensed-tannin substrate, prior dietary tannin exposure, antimicrobial inclusion, and animal species on fermentation parameters following a 48 h in vitro incubation. *J. Anim. Sci.* 96, 343–353.
- HUHTANEN, P., RAMIN, M., and CABEZAS-GARCIA, E. H., 2016. Effects of ruminal digesta retention time on methane emissions: A modelling approach. *in Animal Production Science*.
- HUYEN, N. T., DESRUES, O., ALFERINK, S. J. J., ZANDSTRA, T., VERSTEGEN, M. W. A., HENDRIKS, W. H., and PELLIKAAN, W. F., 2016. Inclusion of sainfoin (*Onobrychis viciifolia*) silage in dairy cow rations affects nutrient digestibility, nitrogen utilization, energy balance, and methane emissions. *J. Dairy Sci.* 99, 3566–3577.
- HYNES, D. N., STERGIADIS, S., GORDON, A., and YAN, T., 2016. Effects of concentrate crude protein content on nutrient digestibility, energy utilization, and methane emissions in lactating dairy cows fed fresh-cut perennial grass. *J. Dairy Sci.* 99 <https://doi.org/10.3168/jds.2016-11509>.
- IBRAHIM, S. L., and HASSEN, A., 2022. Effect of non-encapsulated and encapsulated mimosa (*Acacia mearnsii*) tannins on growth performance, nutrient digestibility, methane and rumen fermentation of South African mutton Merino ram lambs. *Anim. Feed Sci. Technol.* 294, 115502 <https://doi.org/https://doi.org/10.1016/j.anifeedsci.2022.115502>.
- JAYANEGARA, A., GOEL, G., MAKKAR, H. P. S., and BECKER, K., 2015. Divergence between purified hydrolysable and condensed tannin effects on methane emission, rumen fermentation and microbial population in vitro. *Anim. Feed Sci. Technol.* 209 <https://doi.org/10.1016/j.anifeedsci.2015.08.002>.
- KAPP-BITTER, A. N., DICKHOEFER, U., SUGLO, E., BAUMGARTNER, L., KREUZER, M., and LEIBER, F., 2020. Graded supplementation of chestnut tannins to dairy cows fed protein-rich spring pasture: Effects on indicators of protein utilization. *J. Anim. Feed Sci.* 29 <https://doi.org/10.22358/jafs/121053/2020>.
- KOENIG, K. M., and BEAUCHEMIN, K. A., 2018. Effect of feeding condensed tannins in high protein finishing

- diets containing corn distillers grains on ruminal fermentation, nutrient digestibility, and route of nitrogen excretion in beef cattle. *J. Anim. Sci.* 96 <https://doi.org/10.1093/jas/sky273>.
- KOZLOSKI, G. V., HÄRTER, C. J., HENTZ, F., DE ÁVILA, S. C., ORLANDI, T., and STEFANELLO, C. M., 2012.** Intake, digestibility and nutrients supply to wethers fed ryegrass and intraruminally infused with levels of *Acacia mearnsii* tannin extract. *Small Rumin. Res.* 106, 125–130 <https://doi.org/https://doi.org/10.1016/j.smallrumres.2012.06.005>.
- KRUEGER, W. K., GUTIERREZ-BAÑUELOS, H., CARSTENS, G. E., MIN, B. R., PINCHAK, W. E., GOMEZ, R. R., ANDERSON, R. C., KRUEGER, N. A., and FORBES, T. D. A., 2010.** Effects of dietary tannin source on performance, feed efficiency, ruminal fermentation, and carcass and non-carcass traits in steers fed a high-grain diet. *Anim. Feed Sci. Technol.* 159 <https://doi.org/10.1016/j.anifeedsci.2010.05.003>.
- KUMAR, S., INDUGU, N., VECCHIARELLI, B., and PITTA, D. W., 2015.** Associative patterns among anaerobic fungi, methanogenic archaea, and bacterial communities in response to changes in diet and age in the rumen of dairy cows. *Front. Microbiol.* 6 <https://doi.org/10.3389/fmicb.2015.00781>.
- LAZZARI, G., MÜNGER, A., EGGERSCHWILER, L., BORDA-MOLINA, D., SEIFERT, J., CAMARINHA-SILVA, A., SCHRADER, S., ZÄHNER, M., ZEYER, K., KREUZER, M., and DOHME-MEIER, F., 2023.** Effects of *Acacia mearnsii* added to silages differing in nutrient composition and condensed tannins on ruminal and manure-derived methane emissions of dairy cows. *J. Dairy Sci.* <https://doi.org/10.3168/jds.2022-22901>.
- LEE, J. M., WOODWARD, S. L., WAGHORN, G. C., and CLARK, D. A., 2004.** Methane emissions by dairy cows fed increasing proportions of white clover (*Trifolium repens*) in pasture. *Proc. New Zeal. Grassl. Assoc.* 66.
- LIMA, P. R., APDINI, T. A., FREIRE, A. S., SANTANA, A. S., MOURA, M. L. M., NASCIMENTO, J. C. S., RODRIGUES, R. T. S., DIJKSTRA, J., GARCEZ NETO, A. F., QUEIROZ, M. A. A., and MENEZES, R. D., 2019a.** Dietary supplementation with tannin and soybean oil on intake, digestibility, feeding behavior, ruminal protozoa and methane emission in sheep. *Anim. Feed Sci. Technol.* 249 <https://doi.org/10.1016/j.anifeedsci.2019.01.017>.
- LIMA, P. DE M. T., CROUZOUON, P., SANCHES, T. P., ZABRÉ, G., KABORE, A., NIDERKORN, V., HOSTE, H., DO AMARANTE, A. F. T., COSTA-JÚNIOR, L. M., ABDALLA, A. L., and OTHER, S., 2019b.** Effects of *Acacia mearnsii* supplementation on nutrition, parasitological, blood parameters and methane emissions in Santa Inês sheep infected with *Trichostrongylus colubriformis* and *Haemonchus contortus*. *Exp. Parasitol.* 207, 107777.
- LÓPEZ-PAREDES, J., GOIRI, I., ATXAERANDIO, R., GARCÍA-RODRÍGUEZ, A., UGARTE, E., JIMÉNEZ-MONTERO, J. A., ALENDA, R., and GONZÁLEZ-RECIO, O., 2020.** Mitigation of greenhouse gases in dairy cattle via genetic selection: 1. Genetic parameters of direct methane using noninvasive methods and proxies of methane. *J. Dairy Sci.* 103 <https://doi.org/10.3168/jds.2019-17597>.
- MHLONGO, L. C. 2018.** In vitro assessment of selected ethno-medicinal plants as potential alternatives for the control of gastrointestinal nematodes in sheep and goats.
- MHLONGO, L. C., KENYON, P., and NSAHLAI, I. V., 2023.** Effect of dietary inclusions of different types of *Acacia mearnsii* on milk performance and nutrient intake of dairy cows. *Vet. Anim. Sci.* 21 <https://doi.org/10.1016/j.vas.2023.100299>.
- MIKOLAYUNAS-SANDROCK, C., ARMENTANO, L. E., THOMAS, D. L., and BERGER, Y. M., 2009.** Effect of protein degradability on milk production of dairy ewes. *J. Dairy Sci.* 92 <https://doi.org/10.3168/jds.2008-1983>.
- MOATS, J., MUTSVANGWA, T., REFAT, B., and CHRISTENSEN, D. A., 2018.** Evaluation of whole flaxseed and the use of tannin-containing fava beans as an alternative to peas in a co-extruded flaxseed product on ruminal fermentation, selected milk fatty acids, and production in dairy cows. *Prof. Anim. Sci.* 34 <https://doi.org/10.15232/pas.2018-01726>.
- MOLAN, A. L., and FARAJ, A. M., 2010.** The effects of condensed tannins extracted from different plant species on egg hatching and larval development of *Teladorsagia circumcincta* (nematoda: Trichostrongylidae). *Folia Parasitol. (Praha)*. 57 <https://doi.org/10.14411/fp.2010.008>.
- NA, R., DONG, H., ZHU, Z., CHEN, Y., and XIN, H., 2013.** Effects of forage type and dietary concentrate to forage ratio on methane emissions and rumen fermentation characteristics of dairy cows in China. *Trans. ASABE* 56 <https://doi.org/10.13031/trans.56.9972>.
- NASCIMENTO, C. O., PINA, D. S., CIRNE, L. G. A., SANTOS, S. A., ARAÚJO, M. L., RODRIGUES, T. C. G. C., SILVA, W. P., SOUZA, M. N. S., ALBA, H. D. R., and DE CARVALHO, G. G. P., 2021.** Effects of whole corn germ, a source of linoleic acid, on carcass characteristics and meat quality of feedlot lambs. *Animals* 11, 267 <https://doi.org/https://doi.org/10.3390/ani11020267>.
- NAUMANN, H., SEPELA, R., REZAIRE, A., MASIH, S. E., ZELLER, W. E., REINHARDT, L. A., ROBE, J. T., SULLIVAN, M. L., and HAGERMAN, A. E., 2018.** Relationships between structures of condensed tannins from Texas legumes and methane production during in vitro rumen digestion. *Molecules* 23, 2123.
- NAUMANN, H. D., TEDESCHI, L. O., ZELLER, W. E., and HUNTLEY, N. F. 2017.** The role of condensed tannins in ruminant animal production: Advances, limitations and future directions. *Rev. Bras. Zootec.* 46 <https://doi.org/10.1590/S1806-92902017001200009>.
- NAWAB, A., LI, G., AN, L., NAWAB, Y., ZHAO, Y., XIAO, M., TANG, S., and SUN, C., 2020.** The Potential Effect of Dietary Tannins on Enteric Methane Emission and Ruminant Production, as an Alternative to Antibiotic Feed Additives-A Review. *Ann. Anim. Sci.* 20 <https://doi.org/10.2478/aoas-2020-0005>.

- NIDERKORN, V., BARBIER, E., MACHEBOEUF, D., TORRENT, A., MUELLER-HARVEY, I., and HOSTE, H., 2020.** In vitro rumen fermentation of diets with different types of condensed tannins derived from sainfoin (*Onobrychis viciifolia* Scop.) pellets and hazelnut (*Corylus avellana* L.) pericarps. *Anim. Feed Sci. Technol.* 259, 114357.
- NIU, M., APPUHAMY, J. A. D. R. N., LEYTEM, A. B., DUNGAN, R. S., and KEBREAB, E., 2016.** Effect of dietary crude protein and forage contents on enteric methane emissions and nitrogen excretion from dairy cows simultaneously. *in Animal Production Science.*
- ORLANDI, T., POZO, C. A., SCHIAVO, J., OLIVEIRA, L., and KOZLOSKI, G. V., 2020a.** Effect of using Acacia mearnsii tannin extract as a feed additive on nutritional variables and productive performance in dairy cows grazing a temperate pasture. *Anim. Sci. J.* 91 <https://doi.org/10.1111/asj.13407>.
- ORLANDI, T., POZO, C. A., SCHIAVO, J., OLIVEIRA, L., and KOZLOSKI, G. V., 2020b.** Impact of a tannin extract on animal performance and nitrogen excretion of dairy cows grazing a tropical pasture. *Anim. Prod. Sci.* 60, 1183–1188.
- PATHAK, A. K. 2013.** Potential of Using Condensed Tannins to Control Gastrointestinal Nematodes and Improve Small Ruminant Performance. *Int. J. Mol. Vet. Res.* <https://doi.org/10.5376/ijmvr.2013.03.0008>.
- PATRA, A. K., and PUCHALA, R., 2023.** Methane mitigation in ruminants with structural analogues and other chemical compounds targeting archaeal methanogenesis pathways. *Biotechnol. Adv.*, 108268.
- PEDREIRA, M. DOS S., PRIMAVESI, O., LIMA, M. A., FRIGHETTO, R., OLIVEIRA, S. G. DE, and BERTI, T. T., 2009.** Ruminal methane emission by dairy cattle in Southeast Brazil. *Sci. Agric.* 66, 742–750.
- PERNA JUNIOR, F., GALBIATTI SANDOVAL NOGUEIRA, R., FERREIRA CARVALHO, R., CUELLAR ORLANDI CASSIANO, E., and MAZZA RODRIGUES, P. H., 2022.** Use of tannin extract as a strategy to reduce methane in Nellore and Holstein cattle and its effect on intake, digestibility, microbial efficiency and ruminal fermentation. *J. Anim. Physiol. Anim. Nutr. (Berl.)*
- PETLUM, A., PAENKOU, P., LIANG, J. B., VASUPEN, K., and PAENKOU, S., 2019.** Molecular weight of condensed tannins of some tropical feed-leaves and their effect on in vitro gas and methane production. *Anim. Prod. Sci.* 59 <https://doi.org/10.1071/AN17749>.
- QUIJADA, J., DRAKE, C., GAUDIN, E., EL-KORSO, R., HOSTE, H., and MUELLER-HARVEY, I., 2018.** Condensed tannin changes along the digestive tract in lambs fed with sainfoin pellets or hazelnut skins. *J. Agric. Food Chem.* 66, 2136–2142.
- RICCI, P., CHAGUNDA, M. G. G., ROOKE, J., M. HOUDJIK, J. G., DUTHIE, C.-A., HYSLOP, J., ROEHE, R., and WATERHOUSE, A., 2014.** Evaluation of the laser methane detector to estimate methane emissions from ewes and steers. *J. Anim. Sci.* 92, 5239–5250.
- RIRA, M., CHENTLI, A., BOUFENERA, S., and BOUSSEBOUA, H. 2015.** Effects of Plants Containing Secondary Metabolites on Ruminal Methanogenesis of Sheep in vitro. *in Energy Procedia.*
- DE S. COSTA, E. I., RIBIERO, C. V. D. M., SILVA, T. M., RIBEIRO, R. D. X., VIEIRA, J. F., DE O. LIMA, A. G. V., BARBOSA, A. M., DA SILVA JÚNIOR, J. M., BEZERRA, L. R., and OLIVEIRA, R. L., 2021a.** Intake, nutrient digestibility, nitrogen balance, serum metabolites and growth performance of lambs supplemented with Acacia mearnsii condensed tannin extract. *Anim. Feed Sci. Technol.* 272, 114744 <https://doi.org/https://doi.org/10.1016/j.anifeedsci.2020.114744>.
- DE S. COSTA, E. I., RIBIERO, C. V. D. M. V. D. M., SILVA, T. M., RIBEIRO, R. D. X. X., VIEIRA, J. F., DE O. LIMA, A. G. V., BARBOSA, A. M., DA SILVA JÚNIOR, J. M., BEZERRA, L. R., OLIVEIRA, R. L., DE, E. I., RIBIERO, C. V. D. M. V. D. M., SILVA, T. M., RIBEIRO, R. D. X. X., VIEIRA, J. F., DE, A. G. V., BARBOSA, A. M., SILVA JÚNIOR, J. M. D., BEZERRA, L. R., and OLIVEIRA, R. L., 2021b.** Intake, nutrient digestibility, nitrogen balance, serum metabolites and growth performance of lambs supplemented with Acacia mearnsii condensed tannin extract. *Anim. Feed Sci. Technol.* 272, 114744 <https://doi.org/https://doi.org/10.1016/j.anifeedsci.2020.114744>.
- SAMINATHAN, M., SIEO, C. C., ABDULLAH, N., WONG, C. M. V. L., and HO, Y. W., 2015.** Effects of condensed tannin fractions of different molecular weights from a *Leucaena leucocephala* hybrid on in vitro methane production and rumen fermentation. *J. Sci. Food Agric.* 95 <https://doi.org/10.1002/jsfa.7016>.
- SAMINATHAN, M., SIEO, C. C., GAN, H. M., RAVI, S., VENKATACHALAM, K., ABDULLAH, N., WONG, C. M. V. L., and HO, Y. W., 2016.** Modulatory effects of condensed tannin fractions of different molecular weights from a *Leucaena leucocephala* hybrid on the bovine rumen bacterial community in vitro. *J. Sci. Food Agric.* 96, 4565–4574.
- SARNATARO, C., SPANGHERO, M., and LAVRENČIČ, A., 2020.** Supplementation of diets with tannins from Chestnut wood or an extract from *Stevia rebaudiana* Bertoni and effects on in vitro rumen fermentation, protozoa count and methane production. *J. Anim. Physiol. Anim. Nutr. (Berl.)* 104 <https://doi.org/10.1111/jpn.13414>.
- SCHARENBERG, A., KREUZER, M., and DOHME, F., 2009.** Suitability of sainfoin (*Onobrychis viciifolia*) hay as a supplement to fresh grass in dairy cows. *Asian-Australasian J. Anim. Sci.* 22 <https://doi.org/10.5713/ajas.2009.80675>.
- SEYMOUR, W. M., CAMPBELL, D. R., and JOHNSON, Z. B., 2005.** Relationships between rumen volatile fatty acid concentrations and milk production in dairy cows: A literature study. *Anim. Feed Sci. Technol.* 119 <https://doi.org/10.1016/j.anifeedsci.2004.10.001>.
- SILANIKOVE, N., GILBOA, N., and NITSAN, Z., 2001.** Effect of polyethylene glycol on rumen volume and retention time of liquid and particulate matter along the digestive tract in goats fed tannin-rich carob leaves (*Ceratonia siliqua*). *Small Rumin. Res.* 40 [https://doi.org/10.1016/S0921-4488\(00\)00209-1](https://doi.org/10.1016/S0921-4488(00)00209-1).
- SZULC, P., MRAVČÁKOVÁ, D., SZUMACHER-STRABEL, M., VÁRADYOVÁ, Z., VÁRADY, M., ČOBANOVÁ, K., SYAHRULAWAL, L., PATRA, A. K., and CIESLAK, A.,**

2020. Ruminant fermentation, microbial population and lipid metabolism in gastrointestinal nematode-infected lambs fed a diet supplemented with herbal mixtures. *PLoS One* 15 <https://doi.org/10.1371/journal.pone.0231516>.
- TERRANOVA, M., EGGERSCHWILER, L., ORTMANN, S., CLAUSS, M., KREUZER, M., and SCHWARM, A., 2021.** Increasing the proportion of hazel leaves in the diet of dairy cows reduced methane yield and excretion of nitrogen in volatile form, but not milk yield. *Anim. Feed Sci. Technol.* 276, 114790 <https://doi.org/10.1016/j.anifeedsci.2020.114790>.
- TSEU, R. J., PERNA JUNIOR, F., CARVALHO, R. F., SENE, G. A., TROPALDI, C. B., PERES, A. H., and RODRIGUES, P. H. M., 2020.** Effect of tannins and monensin on feeding behaviour, feed intake, digestive parameters and microbial efficiency of nellore cows. *Ital. J. Anim. Sci.* 19 <https://doi.org/10.1080/1828051X.2020.1729667>.
- TYMENSEN, L. D., BEAUCHEMIN, K. A., and MCALLISTER, T. A., 2012.** Structures of free-living and protozoa-associated methanogen communities in the bovine rumen differ according to comparative analysis of 16S rRNA and mcrA genes. *Microbiol. (United Kingdom)* 158 <https://doi.org/10.1099/mic.0.057984-0>.
- WALL, E., COFFEY, M. P., and POLLOTT, G. E. 2012.** The effect of lactation length on greenhouse gas emissions from the national dairy herd. *Animal* 6 <https://doi.org/10.1017/S1751731112000936>.
- WATT, L. J., CLARK, C. E. F., KREBS, G. L., PETZEL, C. E., NIELSEN, S., and UTSUMI, S. A., 2015.** Differential rumination, intake, and enteric methane production of dairy cows in a pasture-based automatic milking system. *J. Dairy Sci.* 98 <https://doi.org/10.3168/jds.2015-9463>.
- WILLIAMS, S. R. O. O., HANNAH, M. C., ECKARD, R. J., WALES, W. J., and MOATE, P. J., 2020.** Supplementing the diet of dairy cows with fat or tannin reduces methane yield, and additively when fed in combination. *Animal* 14, s464–s472.
- WU, X., HUANG, S., HUANG, J., PENG, P., LIU, Y., HAN, B., and SUN, D., 2021.** Identification of the Potential Role of the Rumen Microbiome in Milk Protein and Fat Synthesis in Dairy Cows Using Metagenomic Sequencing. *Animals* 11, 1247.
- ZHANG, J., XU, X., CAO, Z., WANG, Y., YANG, H., AZARFAR, A., and LI, S., 2019.** Effect of different tannin sources on nutrient intake, digestibility, performance, nitrogen utilization, and blood parameters in dairy cows. *Animals* 9 <https://doi.org/10.3390/ani9080507>.

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