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Improvements in the neurodevelopmental toxicity of heavy metals for *Dicentrarchus labrax* by an algal mixture of *Spirulina platensis* and *Chlorella vulgaris*

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In response to the rising demand for aquatic products for human consumption, aquaculture has developed significantly as an agronomic activity worldwide. Proteins and amino acids are significant macromolecules that govern vital metabolic processes and act as building blocks for the creation of compounds with biological significance. Fish play a significant function in human nutrition and are a significant dietary source of high-quality animal proteins and amino acids. The effect of toxic metals (cadmium and lead) on total central amino acids compositions (Essential, Conditionally Essential, and Non-Essential), and Crude protein concentration were examined in the current study for the juvenile European sea Dicentrarchus labrax, a stress-sensitive species, and evaluation the protective effect of the dietary Spirulina platensis and Chlorella vulgaris algal mixture as a supplement. Fish fed the control diet and kept in freshly filtered seawater without the addition of any pollutants (Control). Total metals (lead nitrate and cadmium chloride (0.006 mg/l) in the ratio (1:1) were added to rearing fresh seawater and fish were fed as the following diets in the other groups: control with metal group, 3% algal mixture supplemented diet (D1), 5% algal mixture supplemented diet (D2), and 7% algal mixture supplemented diet (D3) for 5 weeks. The results showed a significant decrease in the total amino acids in the control with the metal group compared to the control and a significant increase in essential amino acids, conditionally essential amino acids, non-essential amino acids, and crude protein concentration in the other 3 diet groups. By concentrating on the contributions towards our understanding of the role these metals have on adverse neurological outcomes in affected populations, this study demonstrated the impact of the mixed mixture of algae Spirulina platensis and Chlorella vulgaris on the treatment of the effect of cadmium (Cd) and lead (Pb) on neurodevelopment outcomes.

ABSTRACT

INTRODUCTION

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Aquaculture is an important contributor to sustainable food and nutritional stability and is the fastest-growing variable in productivity and profitability. Marine fish are sensitive animals and their survival in captivity is full of challenges. Understanding the relationship between industrial fish farming and fish welfare science is essential to

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provide optimal survival conditions for fish and management systems. Despite the phenomenal growth of its production, aquaculture is experiencing various problems, and presents many difficulties, particularly when it comes to stress (Schreck and Tort 2016).

The European sea bass *Dicentrarchus labrax* is one of the main species of industrial marine fish farming, and the uncontrolled phenomenon of stress can have a major impact on aquaculture producers, leading to increased mortality and economic losses FSBI (2002). Instead of using medications to treat sick fish, there is a clear trend toward enhancing fish health and welfare to stop disease outbreaks by using dietary supplements or probiotics (Reverter et al. 2020; El-Saadony et al. 2021). The antioxidant and immune systems of fish are stimulated by dietary supplements, which play important roles in fish health and welfare Ahmadifar et al. (2021). Aquafeed production uses a variety of feed supplements, such as herbal materials, vitamins, minerals, and amino acids that enhance fish welfare and health Lee et al. (2015). Amino acids (AAs) play crucial roles in physiological processes, as well as contributing to fish growth as the building blocks of proteins and as metabolic pathway intermediaries. They act as building blocks for the synthesis of numerous biologically significant compounds, such as nucleotides, peptides, hormones, and neurotransmitters. Additionally, (AAs) control the expression of genes and the protein phosphorylation cascade, both of which are crucial for cell signaling (Wu, 2010; Zhu et al., 2013; Webster and Thompson, 2015), in addition, the innate and cell-mediated immune responses, as well as the transport and metabolism of nutrients in animal cells Wang et al. (2013). It is widely recognized that the amount of protein and dietary (AA) an animal consumes significantly impacts their growth, reproduction, health, and feed costs (Boshra et al., 2006; Watanuki et al., 2009; Wu et al., 2017). Animal AAs are categorized as either Essential (EAAs) or Nonessential (NEAAs) depending on the growth or nitrogen balance or Conditionally Essential Amino Acids (CEAAs) (Rahimnejad and Lee 2013; Zhu et al., 2013). For human nutrition, the EAAs are arginine, cystine, histidine, leucine, lysine, methionine, threonine, tryptophan, tyrosine, and valine; the CEAAs are glutamine, glutamic acid, glycine, proline, and taurine; and the NEAA are aspartic acid, serine, and alanine. However, the idea of Functional Amino Acids (FAAs) has recently been put forth. FAAs are those that take part in and control important metabolic pathways to enhance an organism's health, survival, growth, development, lactation, and reproduction Wu (2013). The FAAs also show tremendous potential in the prevention and treatment of viral diseases, intrauterine growth restriction, infertility, intestinal and neurological dysfunction, and metabolic diseases (such as obesity, diabetes, and cardiovascular disorders). FAAs in human nutrition include taurine, arginine, cystine, leucine, methionine, tryptophan, tyrosine, aspartate, glutamic acid, glycine, and proline **Wu** (2013). Determining the dietary AA requirements for various fish species is necessary due to the significant effects of AAs on animal growth, feed costs, and nitrogen pollution (Rahimnejad and Lee 2013). The dietary requirements of AA depend on animal species, developmental stage,

physiological status, intestine microbiota, and environmental factors **Zhu** *et al.* (2013). The majority of amino acids in the diet come from proteins, and the ratio of essential to non-essential amino acids is used to assess the quality of dietary protein. The dietary essential amino acids (EAA) in high-quality proteins are readily digested and present in amounts that meet human needs (WHO, 2007).

Global health concerns include environmental exposure to neurotoxic metals and metalloids like arsenic, cadmium, lead, mercury, or manganese. Environmental metals can affect neurodevelopment, neurobehavioral, and cognition, as well as lead to neurodegeneration, depending on the length of exposure over a lifetime. There is growing evidence that links early-life neurological diseases to environmental exposure to metal contaminants **Carmona** *et al.* (2021). Harmful heavy metals are those that demonstrate highly toxic qualities when ingested by animals, including humans. Cadmium, lead, and mercury are among the top 10 substances that the World Health Organization considers to be particularly dangerous for human health. Exposure to these metals has also been related to many neurodegenerative problems in humans **Von Stackelberg** *et al.* (2015).

Microalgae have a significant role in various industries around the world, including food production, medicine, biofuels, cosmetics, and agriculture. Spirulina platensis and Chlorella vulgaris are two of the most well-known microalgae genera, both of which have bioactive substances like protein, vitamins, pigments, long-chain polyunsaturated fatty acids, sterols, and other substances that make them particularly interesting from the perspective of their potential health benefits. Spirulina has been chemically examined and has been demonstrated to be an excellent source of proteins, vitamins, lipids, minerals, nucleic acids, enzymes, and pigments Andrade et al. (2018). Spirulina is an important source of protein for human nutrition, with the largest concentrations of leucine, valine, isoleucine, and tryptophan Demelash (2018). Neurodevelopmental disorders (NDDs) are caused by genetic or environmental factors and can include intellectual difficulties, communication problems, traumatic brain injuries, autism spectrum disorders, epilepsies, and motor and coordination problems Cordeiro et al. (2015). Rodents and fish are model organisms for many of these human illnesses, allowing experiments to clarify the molecular underpinnings of their origins (Ricceri et al., 2007; Kisby et al., 2013; Green and Planchart 2018).

This study aimed to illustrate the effect of the mixed mixture of the algal of *Spirulina platensis* and *Chlorella vulgaris* on the treatment of the toxic effect of cadmium (Cd), and lead (Pb) on the brain central amino acids for European sea bass *Dicentrarchus labrax* by specifically focusing on the contributions toward our understanding of the role these metals have on adverse neurological outcomes in affected populations.

MATERIALS AND METHODS

Materials and methods

The experimental diets

Spirulina platensis algae was prepared, according to the technique developed by Abou El-Kheir *et al.* (2008) Spirulina platensis was grown and harvested at the Hydrobiology Lab of the National Institute of Oceanography and Fisheries in El-Qanater El-Khayria. At the marine hatchery at the National Institute of Oceanography and Fisheries in Alexandria, Egypt, *Chlorella vulgaris* cultures were grown Hasanein *et al.* (2018). The fundamental practical diet was designed to meet the nutritional needs of juvenile European sea bass and contained about 47% crude protein and 12% crude fat. Three concentrations of dry extract were used to create the experimental diets. The basal diet served as the control and a mixture of Spirulina platensis and Chlorella vulgaris (AM, 1:1) were introduced at 30, 50, and 70g kg-1 diet.

The experimental design

The European sea bass *Dicentrarchus labra* was acquired from the National Institute of Oceanography and Fisheries' marine hatchery in Alexandria and acclimated for a week prior to beginning the experimental circumstances. Following fish acclimation, the initial fish weight of approximately around (IW = 5.0 g), at a stocking density of 15 fish per aquarium, were placed in 15 glass aquariums, each measuring 100 L, and were supplied with filtered natural seawater that was kept at a temperature of 23°C. These were the five experimental groups. The first group was fed a control diet and kept in fresh, filtered seawater without the addition of any pollutants (Control). As a contaminant, lead nitrate and cadmium chloride (0.006 mg/l) in a ratio (1:1) were introduced to the rising fresh filtered seawater and fish were fed as the following diets in the other groups: control with mixed metal, 3% algal mixture supplemented diet (D1), 5% algal mixture supplemented diet (D2), and 7% algal mixture supplemented diet (D3). Three times per day, fish were fed 5% of their body weight. The water was changed daily to avoid waste buildup, and the feces were drained before the first feeding. Filtered seawater was supplied with a salinity of 37.5 ppt and a temperature of 23.4 \pm 1.2 °C. The oxygen provided by aeration was present during the study at a level of 5.93 mg L-1. For the duration of the trial, water quality parameters were recorded. Total ammonia and nitrite levels were measured throughout the feeding experiment, the PH was 7.6 ± 0.31 , ammonia levels were 0.05 \pm 0.01 and nitrite concentrations were 1.4 mg/l. The natural photoperiod was 10 h light: and 14 h dark. To maintain a steady concentration of heavy metals in each aquarium, freshly prepared stock solutions of heavy metals were supplied daily to rearing water following the residual siphon. Five weeks were spent on the experiment.

Ethical Clearance

The author declares that the sampling of this species under investigation was done following the guidelines of the international conventions for the use of animals in scientific research.

Tissue preparation

The fish was rapidly killed by a sharp knife blow to the head and the entire brain was swiftly removed, fast rinsed in iso-osmolar NaCl salt water to remove blood, and then weighed **Hegazi** *et al.* (2010).

Amino acid compositions:

Three whole brain samples were pooled and homogenized to exactly three times their weight in ice-cold distilled water. Crude homogenates (0.3 g) were deproteinized by the addition of 10 mL g⁻¹ sulfosalicylic acid (3.5%). The supernatant was collected after centrifugation at 3500 rpm for 15 min and stored at -16° C for amino acid analysis. The brain's free amino acids were determined using a Beckman amino acid analyzer (model 157 CL).

Estimation of amino-acids:

The determination of amino acids was accomplished according to the method adopted by **Sánchez-Machado** *et al.* (2003). First, the amino acids were hydrolyzed by hydrochloric acid (HCl) and then derivatized then analyzed on the HPLC.

Derivatization with PITC:

The derivatization procedure was a modification of the method of **González-Castro et al.** (**1997**). Amino acid standard solution or sample hydrolysate (30μ L) was placed in a tube and dried in an oven for 20 min at 42°C. Methanol–water–TEA (2:2:1, v/v; 50 µL) was then added to the residue, and the resulting solution was vacuum-dried for 20 min at room temperature (25 °C). Methanol–water–TEA–PITC (triethylamine(TEA) and phenylisothiocyanate (PITC)) (7:1:1:1, v/v; 50 µL) was then added, and the tubes were vortex for 15 s, then left for 20 min at room temperature. The resulting solution was dried for 100 min at room temperature (25 °C). After derivatization, Na₂HPO₄ (5 mM containing 5% acetonitrile; 100 µL) was added as the diluent, with vortex mixing for 15 s. The mobile phase was prepared from two solutions, A and B. Solution A was 0.14 M ammonium acetate buffer containing 0.05% (v/v) TEA (pH adjusted to 6.4 with glacial acetic acid). Solution B was acetonitrile-water 60:40 (v/v). The relative abundance of

each peak and the comparison to amino acid standards allow for the quantification of each amino acid in the sample.

Statistical analysis

The mean \pm SD of each reading parameter is used to represent it. One-way analysis of variance (ANOVA) and multiple comparisons were used to analyze the data by Dunnett's experiment with a computer program (GraphPad 6 In Stat Softeware, Inc.

RESULTS

Fish were exposed to mixed of lead nitrate and cadmium chloride (0.006 mg/l) in a ratio (1:1) for 5 weeks. In (Fig. 1), the content of Essential Amino Acids (EAAs) in the whole brain tissues arginine(Arg) and histidine (His), showed a significant decrease in them in the control with metal group at (P < 0.05) in comparison with the control. Diets with an algal mixture of Spirulina platensis and Chlorella vulgaris (AM, 1:1) at 3% (D1), 5% (D2), and 7% (D3) showed a significant increase in both amino acids composition between the control and the other 3 diets group. No difference appeared within D1, D2, and D3 in (Arg) but in (His) D2 showed the highest value. In amino acids iso leucine (Iso) and leucine (Leu)There was a significant decrease in the control with metal group at (P < 0.05) in comparison with the control and a significant increase between the control and the D1, D2, and D3 groups. There was no significant difference between D1, D2, and D3 in both of them. In amino acids methionine(Met), lysine(Lys), and threonine (Thr) There was a significant decrease in control with the metal group at (P < 0.05) in comparison with control and a significant increase between the control and the D1, D2, and D3 groups. There was no significant difference within D1, D2, and D3 in 3 amino In phenylalanine (Phe), tyrosine (tyr), Valine (Val), and acid contents. tryptophan(Try)There was a significant decrease in control with metal group at (P < 0.05) in comparison with the control and a significant increase between the control and the D1, D2, and D3 groups. There was no significant difference within D1, D2, and D3 in 3 amino acid contents (Phe), (tyr), and (Val) but in tryptophan (Try) D2 and D3 values were greater than D1. In amino acid cysteine (cyst), there was no value in group control with metal and there was a significant increase between the control group and the D1, D2, and D3 groups. D2 showed the highest value among the 3 diet groups. In general, D1, D2, and D3 showed amino acid content greater than the control and control with metal.

In **Conditionally Essential Amino Acids (CEAA)**, in fig. (2), There was a significant decrease in control with the metal group at (P < 0.05) in total values of amino acids glutamic (Glu), glycine(Gly), and proline (Pro) in comparison with the control group and a significant increase between the control group and the groups D1, D2, and D3. There was no significant difference within D1, D2, and D3 in (Glu) and (Gly), amino acid

contents but in (Pro) D2 showed the highest value among the 3 diet groups. Three diet groups showed amino acid content greater than the control and control with metal

In **Nonessential Amino Acids (NEAAs)** composition in the whole brain tissues (Fig. 3). There was a significant decrease in total amino acid alanine(Ala), aspartic acid(Asp), and serine(Ser) in the control with the metal group at (P < 0.05) in comparison with control and significant increase between the control and the D1, D2, and D3 groups. There was no significant difference within D1, D2, and D3 in (Ala) contents but in (Asp), and (Ser) D2, D3 were greater than D1. The three diet groups showed amino acid content greater than the control and control with metal.

The percentage of crude protein appeared a significant decrease in the control with the metal group at (P < 0.05) in comparison with the control and a significant increase between the control and the D1, D2, and D3 groups in (Fig. 3). There was no significant difference within D1, D2, and D3.











Fig. (1): The effects of an algal mixture of *Spirulina platensis* and *Chlorella vulgaris* on the central Essential Amino Acids (EAAs) composition in the whole brain tissues (mg/100g crude Protein) of *Dicentrarchus labrax* exposed to lead nitrate and cadmium chloride (0.006 mg/l) in the ratio (1:1) for 5week. Each reading represents the Mean \pm SD of 15 samples. The difference was significant at $\Box P \leq 0.05$.



Fig. (2): The effects of an algal mixture of *Spirulina platensis* and *Chlorella vulgaris* on Conditionally Essential Amino Acids (CEAAs) composition in the whole brain tissues (mg/100g crude Protein) of *Dicentrarchus labrax* exposed to lead nitrate and cadmium chloride (0.006 mg/l) in the ratio (1:1) for 5week. Each reading represents the Mean \pm SD of 15 samples. The difference was significant at $\Box P \leq 0.05$.



Fig. (3): The effects of an algal mixture of *Spirulina platensis* and *Chlorella vulgaris* on the Nonessential Amino Acids (NEAAs) composition in the whole brain tissues (mg/100g crude Protein) of *Dicentrarchus labrax* exposed to lead nitrate and cadmium chloride (0.006 mg/l) in the ratio (1:1) for 5week and percentage of crude protein. Each reading represents Mean± SD of 15 samples. The difference was significant at $\Box P \leq 0.05$.

DISCUSSION

In the current study, it could be observed that fish in group control with metal showed reduced content of Essential (EAAs), Conditionally Essential Amino Acids (CEAAs), and Nonessential Amino Acids (NEAAs) in the whole brain of Dicentrarchus labrax. An algal mixture of Spirulina platensis and Chlorella vulgaris (AM, 1:1) at 3%, 5%, and 7% as a supplement showed significantly increased values of (EAAs), (CEAA) and (NEAAs) in comparison with control. In this study, the algal mixture showed repairing the harmful effect of toxic metals (lead and cadmium) on the central amino acids and the total crude protein content in the brain tissues of *Dicentrarchus labrax*. These supplemented diets in the three examined ratios can be easily and affordably included in a daily diet for seabass (Dicentrarchus labrax) and provide central amino acids that are necessary for improvements and repairing brain toxicity and consequently improving growth. This is illustrated and agreed by our previous studies, Hasanein et al. (2018) demonstrated the beneficial effects on the growth and feed utilization of Dicentrarchus labrax when Spiruling platensis and Chlorella vulgaris were combined at 3%, 5%, and 7% supplemented diet in the presence of a 0.006 mg/l Cd and Pb concentration, the three supplemented diet reduced the accumulation of cadmium and lead in the white muscles and stimulate ROS scavenging in it. The present results are also suitable and agreed with our previous study (Hasanein and Helal, 2021) reported that at concentrations of 3%, 5% and 7%, the dietary supplement of S. platensis and C. vulgaris for seabass (Dicentrarchus labrax) has natural chelation properties for the mixture of lead and cadmium in the liver toxicity and gill toxicity that contain high levels of vital metals that have anti-Cd and anti-Pb effects. These dietary supplements increased the antioxidant activities in the liver, which is responsible for scavenging ROS, and repaired the ionic disturbance in the gills (caused by exposure to cadmium and lead). In the present study, the content of brain amino acid Arginine (Arg) showed (2.5 control, 1.8 control with metal, 3.1 (D1), 3.3 (D2), and 3.28 (D3) mg/100g crude protein). The three dietary supplements raised arginine content which has enormous properties in repairing physiological conditions. This finding agreed with several studies that explained the role of the addition of AAs as a supplement for fish diets in healthy growth such as Han et al., 2018), a considerable rise in serum levels of insulin (INS) and insulin-like growth factor-I (IGF-I) has also been linked to dietary arginine. Wang et al. (2015) reported that arginine promotes the target of the rapamycin (TOR) signaling pathway to encourage protein synthesis and myogenesis. Arginine also activates adenosine 5 -monophosphate (AMP)activated protein kinase (AMPK) to assist the body in conserving and using the available energy rather than producing lipids. Additionally, arginine has been shown to have a variety of immuno-modulatory effects, including controlling fish's innate and adaptive immune responses, preventing leukocyte apoptosis, and regulating their susceptibility to bacterial infection Hoseini et al. (2020). Cell division, wound healing, ammonia elimination, immunological function, and hormone release are all significantly influenced by arginine. Additionally, it serves as a precursor for the biological synthesis of nitric oxide, which is crucial for maintaining blood pressure, clotting, and neurotransmission. Numerous illnesses, including sepsis, preeclampsia, hypertension, erectile dysfunction, anxiety, and others, benefit from its additional treatment in humans. Mohanty et al. (2014).

Because mature fish have low activity levels of arginine biosynthesis enzymes such pyrroline-5-carboxylate (P5C) synthase, ornithine transcarboxylase (OTC), and carbamoyl phosphate synthase (CPS) III, arginine is a necessary amino acid for fish Andersen et al. (2016). Only N-acetylglutamate synthase (NAGS) in zebrafish (Danio rerio) has been thoroughly studied and reported to be an intermediate form from microbial to mammalian NAGS on the evolutionary path. Other urea cycle enzymes such as argininosuccinate synthetase (ASS) and argininosuccinate lyase (ASL) have rarely been evaluated in fish Caldovic et al. (2014). Although adult fish do not require much adenosine triphosphate (ATP) for the urea cycle because they mostly excrete ammonia through their gills, the urea cycle enzymes are quite active during the early stages of fish development Wright (2011). Numerous physiologically active metabolites, such as nitric oxide (NO), creatine, and polyamines, are produced from arginine in fish (Han et al., 2018; Zheng et al., 2019). However, depending on the dosage, the fish's health, and the environment, arginine supplementation's effects on fish development and immunological responses may differ Azeredo et al. (2015). Reduced fish development rate has been linked to dietary arginine deficiency a lack of dietary arginine has been linked to slower fish growth rates. Appropriate arginine supplementation can enhance fish growth and immunological responses, but an excess of dietary arginine can have the opposite effect Hoseini et al. (2019). The amount of arginine needed by fish is not constant and fluctuates depending on the species. Overall, carnivorous species require more arginine in their diets than omnivorous fish Wang et al. (2021).

In this study, the content of brain amino acid Histidine (His) showed (3.7 control, 2.8 control with metal, 4.68 (D1), 5.37 (D2), and 4.89 (D3) mg/100g crude protein). The algal mixture supplied His which recovered the depletion in group control with metal. This is agreed with (Liao et al., 2013) who revealed that Histidine aids in the removal of heavy metals from the body's myelin sheath maintaining tissue growth and repairing it. His is a precursor to histamine and has a variety of functions in protein interaction. In addition, Jiang et al.(2016), reported according to the growth performance of young grass carp fed a histidine-supplemented diet for two weeks the dietary histidine requirement was 7.63 g/kg. In addition, Michelato et al. (2017) reported the final weight, weight gain, feed conversion ratio, protein efficiency ratio, and net protein utilization in Nile tilapia juveniles fed diets containing graded levels of histidine. Unlike animals and birds, Fish can continually add new muscle fibers to their bodies. Histidine cannot be produced in the body by itself and must be received through nutrition, histidine is an essential amino acid in mammals, fish, and poultry. Histidine deficiency causes a reduction in body weight in some organisms. Additionally, nitrogen balances cannot be kept with a diet lacking histidine. Histidine insufficiency may not impair nitrogen balance if total protein intake is higher than the current standard (0.75 g/kg) according to a study by Kriengsinvos et al. (2002). However, (Moro et al. (2020) illustrated histidine shortage results in a reduction in protein turnover and amino acid oxidation. Given these additional aspects of protein metabolism, histidine is regarded as a necessary amino acid in healthy people. Several studies have also noted histidine's necessity for ruminants. In this study, 5% of the algal mixture showed a higher value of histidine with no significant differential effects between the ratios 5%, 3% and 7%.

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In this study, the content of brain amino acid **Isoleucine (Iso)** showed (4.9 control, 3.2 control with metal, 5.93 (D1), 6.44 (D2), and 6.11 (D3) mg/100g crude protein). The content of brain amino acid leucine (Ieu) showed (6.7 control, 5.17 control with metal, 7.58 (D1), 8.70 (D2), and 7.79 (D3) mg/100g crude protein). The algal mixture supplied Ieu and Iso which are important for protein production and immunity. This agrees with Wu. et al. (2017), fish require the branched-chain amino acid leucine (Leu) to survive. However, the main focus of the following studies was on how Leu affected growth, protein retention, and immunity as **Deng** et al. (2014) on juvenile golden pompano (Trachinotus ovatus), Tan et al. (2016) on rainbow trout (Oncorhynchus mykiss), Choo et al. (1991) on Black carp Juvenile hybrid grouper Epinephelus fuscoguttatus $\mathcal{Q} \times Epinephelus$ lanceolatus \mathcal{O} , Zhou et al. (2019) on fingerling channel catfish Ictalurus punctatus, Wilson et al. (1980) on blunt snout bream Megalobrama amblycephala, Ren et al. (2015) on Mylopharyngodon piceus, Catla catla Zehra, et al. (2015) on *Heteropneustes fossilis*, and (Farhat and Khan 2014) on stinging catfish. Leu promoted proliferation and differentiation, according to studies done recently on primary preterm rat satellite cells and porcine myoblasts (Jie-Min et al., 2015; Zhang et al., **2018**). Leu is the only dietary amino acid that may increase the production of muscle protein and it plays a significant therapeutic function in the treatment of stress-related illnesses such as burns, trauma, and sepsis in humans. Leu has been discovered to decrease the breakdown of muscle tissue when taken as a dietary supplement by enhancing the synthesis of muscle proteins. Marine fishes had unusually high levels of leucine Mohanty et al. (2014). Leu is one of these EAA and is crucial for healthy growth and appropriate development in both humans and animals Yamamoto et al. (2004). Although these studies have only been conducted on fish, an ideal dietary leucine level has been shown to increase the secretion of growth hormone and regulate relative gene expression for animal growth **Deng** et al.(2014). Leu overload or deficit, however, may impair animal growth and decrease diet conversion Choo et al.(1991).

Li, et al. (2010), revealed that leu deficiency causes decreased growth rate, low feed efficiency, and poor protein retention according to research on large yellow croaker, Pseudosciaena crocea, grass carp Ctenopharyngodon idellus. In general, leucine absorption and digestion, which are correlated with the functions of digestive enzymes, determine the rate of fish growth. The capacity of nutrients, such as leucine, to be digested and absorbed is generally correlated with the activity of digestive enzymes (Wu et al. (2016). Few studies have been done to look into the effects of dietary leucine on the activities of fish liver enzymes, although many studies have shown that dietary AA plays a significant role in digestive and absorptive enzymes (Tan et al., 2016; Wu et al. 2016). Leucine and its metabolite, hydroxy-methylbutyrate (HMB), are also crucial for the proper functioning of the immune system in animals. Adding leucine and/or HMB to the diet can increase the quantity of NK cells and liver-associated lymphocytes, which in turn lowers animal mortality (Giri et al., 2015; Jiang et al., 2015). In the head kidney of Labeo rohita fingerlings, appropriate leucine supplementation could improve immunological parameters such as the expression levels of antioxidant enzymes, microglobulin, and immunoglobulin-M. Additionally, adequate dietary leucine controls the tight junction transcript abundance, intestinal immune status, and immune-related signaling molecules according to (**Wu** *et al.*, **2017**). Isoleucine is a branched-chain amino acid that is necessary for the development of muscles and healthy growth (**Charlton 2006**). In this study, the content of amino acid **Valine** (**Val**) showed (4.7 control, 2.0 control with metal, 5.23 (D1), 5.19 (D2), and 5.48 (D3) mg/100g crude protein. Valine has been identified as an essential amino acid of fish **Nose** (**1979**). It is involved in various metabolic pathways and is essential for growth, tissue repair, and maintaining nitrogen balance in the body. Valine deficiency resulted in reduced growth performance in mrigal carp (*Cirrhinus mrigala*) (**Ahmed and Khan, 2006**). In this study, 3%, 5%, and 7% of the algal mixture showed no different effects between leucine, iso leucine, lysine, tyrosine, and valine.

In this study, the content of brain amino acid **Methionine** (**Met**) showed (2.9 control, 1.63 control with metal, 3.18 (D1), 3.78 (D2), and 3.69 (D3) mg/100g crude protein. Methionine in mammals is also used to treat depression, alcoholism, allergies, asthma, copper poisoning, radiation side effects, schizophrenia, drug withdrawal, Parkinson's disease, and liver diseases (**Mischoulon and Fava., 2002**).

In this study, the content of brain amino acid **Threonine** (**Thr**) showed (3.5 control, 2.7 control with metal, 4.75 (D1), 5.6 (D2), and 4.83 (D3) mg/100g crude protein. Threonine is important for numerous nervous system disorders. In this study fish in the group control with metal showed a reduced amount of threonine in comparison with control. An algal mixture supplement showed significant increases in the values of threonine. The ratios of 3%, 5%, and 7% of the algal mixture showed no different effects between them. So The ratios of 3%, 5%, and 7% of the algal mixture showed a repairing effect for threonine. This is agreed with (**Hyland, 2007**) reported, numerous nervous system disorders, such as spinal spasticity, multiple sclerosis, familial spastic paraparesis, and amyotrophic lateral sclerosis, are treated with threonine.

In this study, the content of brain amino acid Phenylalanine (Phe) showed (3.8 control, 2.5 control with metal, 4.5 (D1), 5.2 (D2), and 5.1(D3) mg/100g crude protein. The content of brain amino acid Tyrosine (Tyr) showed (1.2 control, 0.9 control with metal, 1.6 (D1), 1.7 (D2), and 1.6 (D3) mg/100g crude protein. The content of brain amino acid Tryptophan (Try) showed (0.7 control, 0.3 control with metal, 0.9 (D1), 1.3 (D2), and 1.26(D3) mg/100g crude protein. In the three diet groups, the content of brain **Phe**, **Tyr**, **and Try** reduced the stress response resulting from heavy metal toxicity this agrees with the study of Salamanca et al. (2020) on gilthead seabream (Sparus aurata) and meagre (Argyrosomus regius) to study fish welfare in sea farms food supplemented with (phenylalanine and tyrosine) improved the stress response in both species by lowering several stress indicators. (Phe) is an important amino acid, and diets are the only way to consume it. This amino acid is broken down by The two mechanisms that make up the traditional metabolic pathways oxidation to the Tyr and transamination to phenylpyruvate Shafik et al. (2014). Tyr affects the synthesis of the thyroid hormones (triiodothyronine and thyroxine) and is a direct precursor of catecholamine hormones the content of plasma thyroid hormones helps identify stress in Rutilus rutilus caspicus Hoseini et al. (2014). Tyr can also be converted into hydroxyphenyl pyruvate and enter

the energy metabolism. A Phe/Tyr-enriched diet in mammals reduces the effects of an acute stressor **Lehnert** *et al.* (1984). A (Phe) metabolite called hydroxyphenyl pyruvate has been demonstrated by **Cotoia** *et al.*(2014) to enhance cellular viability in mice under stressful circumstances. Both AAs (Phe/Tyr) reduced incidences of bone deformation and death in white seabream larvae (*Diplodus sargus*) **Saavedra** *et al.* (2010) while (Herrera *et al.* (2016) confirmed the moderating effect of Phe on stressed cods in (*Gadus morhua*).

Tryptophan promotes brain Serotonin,5HT synthesis, which may stimulate 5HT release and function. It is a natural amino acid precursor in 5HT biosynthesis. The brain's monoaminergic systems, which include serotonin (5-hydroxytryptamine; 5-HT), dopamine DA, and noradrenaline NA as key neurotransmitters, are activated as part of the acute stress response. The neuronal responses in certain regions of the brain, primarily the telencephalon, hypothalamus, and brain stem, are modulated by these monoamines depending on the type of stressor, its duration, and the severity of its occurrence (Kaslin and Panula, 2001; Barton, 2002; and Martorell-Ribera et al., 2020). Tryptophan should have low toxicity and few negative effects because it is a natural part of the human diet. Due to these benefits, dietary tryptophan supplementation has been utilized with varying degrees of effectiveness in the treatment of neuropsychiatric diseases (Demelash, 2018). The central nervous system's diffuse network of the serotonergic system plays a critical role in the control of mood and behavior cognition. The gut-brain axis is a two-way communication pathway that connects the brain's emotional and cognitive regions with the digestive system's peripheral functions. It is becoming more and more clear that the gut microbiota affects behavior, and this influence has been extended to tryptophan and serotonin, raising the potential that changes in the gut may play a role in the pathophysiology of illnesses of the human central nervous system (Jenkins et al., 2016). Depression is frequently accompanied by low levels of certain brain chemicals or neurotransmitters, such as serotonin, dopamine, noradrenaline, and gamma-aminobutyric acid in humans (GABA) (Van Praag 1983; Diehl and Gershon 1992; Stockmeier 1997; Rush 2007). The amino acids tryptophan, tyrosine, phenylalanine, and methionine are frequently helpful in treating a variety of mood disorders, including depression, according to several studies (Petty 1995; Leonard, **1997**).

In this study, the content of brain amino acid **Glutamic acid** (**Glu**) showed (11.8 control, 9.6 control with metal, 12.6 (D1), 14.2 (D2), and 13.5 (D3) mg/100g crude protein. In this study, the content of brain amino acid **Glycine** (**Gly**) showed (12.7control, 7.6 control with metal, 13.2 (D1), 13.8 (D2), and 13.5 (D3) mg/100g crude protein. The content of **Proline** (**Pro**) in the brain showed (0.4 control, 0.3 control with metal, 0.7 (D1), 0.9 (D2), and 0.6 (D3) mg/100g crude protein. The dietary supplementation of the algal mixture increased the content of three (CEAAs) which improves growth performance, antioxidant, and immune responses. Glutamic acid plays a significant role in the metabolism of amino acids, due to its function in transamination reactions and the need to synthesize essential molecules like glutathione, which is needed for the elimination of highly toxic peroxides, and the polyglutamate folate cofactors **Mohanty** *et al.* (**2014**). The dietary glycine supplementation improves growth performance, antioxidant, and immune responses in some fish and shellfish species, including Nile tilapia, *Oreochromis niloticus*, largemouth bass, *Micropterus salmoides*, and white-leg shrimp, *Litopenaeus vannamei.* glycine is a non-essential amino acid used in fish

nutrition (Xie *et al.*, 2014). It has been discovered to increase the activity of many antioxidant enzymes and is a part of the antioxidant molecule glutathione (Xie *et al.*, 2016; Rossi *et al.*, 2021). Additionally, supplementing with glutathione has been shown to enhance immune responses in the mitten crab, *Eriocheir sinensis* according to Liu *et al.* (2019), suggesting that dietary glycine supplementation may enhance fish immune systems by promoting glutathione synthesis (Liu *et al.*, 2019).

In this study, the content of brain amino acid Alanine (Ala) showed (5.2 control, 2.8 control with metal, 5.6 (D1), 5.8 (D2), and 5.45 (D3) mg/100g crude protein. In this study, the content of brain amino acid Aspartic acid (Asp) showed (9.4 control, 6.5 control with metal, 10.1 (D1), 13.8 (D2), and 12.7 (D3) mg/100g crude protein. the content of brain amino acid Serine (Ser) showed (5.9 control, 3.3 control with metal, 6.2 (D1), 7.4 (D2), and 6.8 (D3) mg/100g crude protein. The dietary supplementation of the algal mixture increased the content of three (NEAAs). NEAA is produced by the body on its own, some of the nutritionally significant NEAA is essential for controlling gene expression, micro-RNA levels, cell signaling, blood flow, nutrient transport, and metabolism in animal cells. They also aid in the development of brown adipose tissue, intestinal microbial growth, anti-oxidative responses, and innate and cell-mediated immune responses. Aspartic acid (FAA) controls the release of critical hormones and is a precursor to AAs methionine, threonine, isoleucine, and lysine. Similar to glycine, cysteine, and tryptophan, serine is the building block for all of these amino acids and is crucial for cell signaling. Schizophrenia can also be treated with serine in humans (Mohanty et al., 2014).

CONCLUSION

The algal mixture of of *Spirulina platensis* and *Chlorella vulgaris* at concentrations of 3%, 5% and 7% as a supplement in the diet of *Dicentrarchus labrax* revealed improvements in the brain toxicity of cadmium and lead due to increased central amino acids content in the whole brain tissues which consequently improve all physiological functions of the central amino acids.

REFERENCES

Abou El-kheir, W. S.; Ibrahim, E. A. and Helal, A. M. (2008). Large-Scale Production of economically important cyanobacterium (*Spirulina platensis*) by using commercial fertilizers. Journal of Environmental Science., 17(3): 1 – 19.

Ahmed, I. and Khan, M.A. (2006). Dietary branched-chain amino acid valine, isoleucine and leucine requirements of fingerling Indian major carp, *Cirrhinus mrigala* (Hamilton). Br J Nutr. 96(3):450-60.

Ahmadifar, E.; Yousefi, M.; Karimi, M.; Fadaei, R.; Raieni, Dadar, M.S.; Yilmaz, S.; Dawood, M.O. and Abdel-Latif, H.M. (2021). Benefits of dietary polyphenols and polyphenol-rich additives to aquatic animal health: an overview Rev. Fish. Sci. Aquacult., 29: 478-511.

Andersen, S. M.; Waagbo, R. and Espe (2016). Functional amino acids in fish health and welfare. Front Biosci; pp. 143-69.

Andrade, L.M.; Andrade, C.J.; Dias, M.; Nascimento, C.A.O. and Mendes, M.A. (2018). Chlorella and Spirulina Microalgae as Sources of Functional Foods, Nutraceuticals, and Food Supplements; an Overview. MOJ Food Process Technol 6(2). Aquaculture, 532, Article 736031.

Azeredo, R.; Pérez-Sánchez, J.; Sitjà-Bobadilla, A.; Fouz, B. and Tort, L. (2015). Aragão C. European sea bass (*Dicentrarchus labrax*) immune status and disease resistance are impaired by arginine dietary supplementation. *PloS One.* ;10.

Barton, B. A. (2002). Stress in fishes: a diversity of responses with particular reference to changes in circulating corticosteroids. Integr. Comp. Biol. 42: 517–525.

Boshra, H.; Li, J. and Sunyer, J.O. (2006).Recent advances on the complement system of teleost fishFish. Shellfish Immunol. Feb;20(2):239-62.

Caldovic, L.; Haskins, N.; Mumo, A.; Majumdar, H.; Pinter, M. and Tuchman, M. (2014). Expression pattern and biochemical properties of zebrafish N-acetylglutamate synthase. *PLoS One.*; 9(1)

Carmona, A.; Roudeau, S. and Ortega, R. (2021). Molecular Mechanisms of Environmental Metal Neurotoxicity: A Focus on the Interactions of Metals with Synapse Structure and Function. Toxics, 9: 198.

Choo, P.; Smith, T.K.; Cho, C.Y. and Ferguson, H.W. (1991). Dietary excesses of leucine influence growth and body composition of rainbow trout. *J. Nutr.*, *121*, 1932–1939.

Cordeiro, C.N.; Tsimis, M. and Burd, I. (2015). Infections and Brain Development. Obstet Gynecol Surv. 70:644–655.

Cotoia, A.; Scrima, R.; Gefter, J.V.; Piccoli, C.; Cinnella, G.; Dambrosio, M.; Fink, M.P. and Capitanio, N. (2014). p-Hydroxyphenylpyruvate, an intermediate of the Phe/Tyr catabolism, improves mitochondrial oxidative metabolism under stressing conditions and prolongs survival in rats subjected to profound hemorrhagic shock. PLoS ONE, 9, e90917.

Demelash, S. (2018). Spirulina as a main source of tryptophan for mental illness: Improving level of serotonin through tryptophan supplementation. GLOBAL J. OF MEDICINE AND PUBLIC HEALTH. Vol. 7, No. 2.

Deng, Y.; Jiang, W.; Liu, Y.; Jiang, J.; Kuang, S.; Tang, L.; Wu, P.; Zhang, Y.; Feng, L. and Zhou, X. (2014). Differential growth performance, intestinal antioxidant status and relative expression of Nrf2 and its target genes in young grass carp (*Ctenopharyngodon idella*) fed with graded levels of leucine. *Aquaculture, 434, 66–73.* **Diehl, D.J. and Gershon, S.** (1992). The role of dopamine in mood disorders. Comp Psychiatry. 1992; 33:115–20.

| 1311 | Improvements in the neurodevelopmental toxicity of heavy metals for Dicentrarchus labrax by |
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| | an algal mixture of Spirulina platensis and Chlorella vulgaris |

El Saadony,M.T.; Alagawany, M.; Patra, A.K.; Kar, I.;Tiwari, R.; Dawood M.A.O.; Dhama, K. and Abdel-Latif, H.M.R. (2021). The functionality of probiotics in aquaculture: An overview Fish. Shellfish Immunol., 117: 36-52.

Farhat and Khan, **M.A.** (2014). Response of fingerling stinging catfish, *Heteropneustes fossilis* (Bloch) to varying levels of dietary L-leucine in relation to growth, feed conversion, protein utilization, leucine retention, and blood parameters. *Aquacult. Nutr.*, 20: 291–302.

FSBI. (2002). Briefing Paper 2, Fisheries Society of the British Isles, Granta Information Systems. In Fish Welfare; FSBI: Cambridge, UK, 2002; Volume 25.

Green, A.J. and Planchart, A. (2018). The neurological toxicity of heavy metals: A fish perspective. Comp Biochem Physiol C Toxicol Pharmacol. 208: 12–19.

Giri, S.S.; Sen. S.S; Chi, C.; Kim, H.J.; Yun, S.; Park, S.C. and Sukumaran, V.(2015). Effect of dietary leucine on the growth parameters and expression of antioxidant, immune, and inflammatory genes in the head kidney of *Labeo rohita* fingerlings. Vet Immunol Immunopathol. 15;167(1-2):36-43.

González-Castro, M.J.; Lopéz-Hernández, J.; Simal-Lozano, J. and Oruňa-Concha, M.J. (1997). Determination of Amino Acids in Green Beans by Derivatization with Phenylisothiocianate and High-Performance Liquid Chromatography with Ultraviolet DetectionJ Chromatogr. Sci., 35:181–185.

Han, F.; Chi, S.; Tan, B.; Dong, X.; Yang, Q. and Liu, H. (2018). Metabolic and immune effects of orange-spotted grouper, *Epinephelus coioides* induced by dietary arginine. *Aquacult Rep.*; 10:8–16.

Hasanein, S.S; Saleh, E. N.; El-Sayed, S. H. and Helal, A. M. (2018). The Effect of dietary supplementation of *Spirulina platensis* and *Chlorella vulgaris* algae on the growth and disease resistance of the sea bass (*Dicentrarchus labrax*). Egyptian Journal of Aquatic Biology & Fisheries.Vol. 22(4): 249-262.

Hasanein, S. S. and Helal, A. M. (2021). Suppressive of hepatotoxicity caused by lead and cadmium in the sea bass *Dicentrarchus labrax* by using an algal mixture of *Spirulina platensis* and *Chlorella vulgaris*, Egyptian Journal of Aquatic Biology & Fisheries Vol. 25(6): 343 – 360.

Hegazi, M. M.; Attia, Z.I.; Hegazi, M. A. M. and Hasanein, S. S. (2010). Metabolic consequences of chronic sublethal ammonia exposure at cellular and subcellular levels in Nile tilapia brain Aquaculture 299, 149–156.

Herrera, M.; Herves, M.A.; Giráldez, I.; Skar, K.; Mogren, H.; Mortensen, A. and Puvanendran, V. (2016). Effects of amino acid supplementations on metabolic and physiological parameters in Atlantic cod (*Gadus morhua*) under stress. Fish Physiol. Biochem., 43: 591–602.

Hoseini, S. M.; Ahmad Khan M.; Yousefi, M. and Costas, B. (2020). Roles of arginine in fish nutrition and health: insights for future researches. *Rev Aquacult.*; 12:2091–2108.

Hoseini, S.M.; Yousefi, M.; Hoseinifar, S. H. and VanDoan, H. (2019). Effects of dietary arginine supplementation on growth, biochemical, and immunological responses of common carp (*Cyprinus carpio* L.), stressed by stocking density. *Aquaculture.;* 503:452–459.

Hoseini, S.M.; Hedayati, A.and Ghelichpour, M. (2014). Plasma metabolites, ions, and thyroid hormones levels, and hepatic enzymes activity in Caspian roach (*Rutilus rutilus caspicus*) exposed to waterborne manganese. Ecotoxicol. Environ. Saf., 107: 84–89.

Hyland, K. (2007). Inherited disorders affecting dopamine and serotonin: critical neurotransmitters derived from aromatic amino acids. *Journal of Nutrition*. 137(6):1568S–1572S.

Jenkins, T.A.; Nguyen, J.C.; Polglaze, K.E. and Bertrand, P.P. (2016). Influence of Tryptophan and Serotonin on Mood and Cognition with a Possible Role of the Gut-Brain Axis. 20: 8(1).

Jiang, W.-D; Deng, Y.-P.; Liu, Y.; Biao, Q.; Jiang, J.; Kuang, S-Y.; Tang, W.P.; Zhang, Y-A.; Zhou, X-Q. and Feng, L.(2015). Dietary leucine regulates the intestinal immune status, immune-related signalling molecules and tight junction transcript abundance in grass carp (*Ctenopharyngodon idella*) Aquaculture. 444: 134-142.

Jiang,W.-D.; Qu, B.; Feng, L.; Jiang, J.; Kuang, S.-Y.; Wu, P.; Tang, L.; Tang,W.-N.; Zhang, Y.-A. and Zhou, X.-Q.; et al. (2016). Histidine prevents cu-induced oxidative stress and the associated decreases in mrna from encoding tight junction proteins in the intestine of grass carp (*Ctenopharyngodon idella*). PLoS ONE 2016, 11, e0157001.

Jie-Min, D.; Mu-Xue, Y.; Zhen-Yu, S.; Chu-Yi, G.; Si-Qi, Z. and Xiao-Shan, Q. (2015). Leucine promotes proliferation and differentiation of primary preterm rat satellite cells in part through mTORC1 signaling pathway. *Nutrients* 2015, *7*, 3387–3400.

Kaslin, J. and Panula, P. (2001). Comparative anatomy of the histaminergic and other aminergic systems in zebrafish (*Danio rerio*). J. Comp. Neurol. 440, 342–377.

Kisby, G. E.; Moore, H. and Spencer, P. S. (2013). Animal models of brain maldevelopment induced by cycad plant genotoxins. Birth Defects Res Part C Embryo Today Rev. 99:247–255.

Kriengsinyos, W.; Rafii, M.; Wykes, L.J.; Ball, R.O. and Pencharz, P.B. (2002).Long-term effects of histidine depletion on whole-body protein metabolism in healthy adults. J. Nutr. 132, 3340–3348.

Lee, C.S.; Lim, C. and Webster, C.D. (2015). Dietary Nutrients, Additives, and Fish Health Wiley-Blackwell, NJ, USA.

Lehnert, H.; Reinstein, D.K.; Strowbridge, B.W. and Wurtman, R.J. (1984). Neurochemical and behavioral consequences of acute, uncontrollable stress: Effects of dietary tyrosine. Brain Res., 303: 215–223.

Leonard, B.E. (1997). The role of noradrenaline in depression: A review. J Psychopharmacol. ;11: S39–47.

Li, Y.; Ai, Q.; Mai, K.; Xu, W.; Cheng, Z. and He, Z. (2010). Dietary leucine requirement for juvenile large yellow croaker Pseudosciaena crocea (Richardson, 1846). *J. Ocean Univ. China* **2010**, *9*: 371–375.

Liao, S.M.; Du, Q.S.; Meng, J.Z. ;Pang, Z.W. and Huang, R.B. (2013). The multiple roles of histidine in protein interactions. Chem Cent J. 1;7(1):44.

Liu, J.-D.; Chi, C.; Zheng, X.-C.; Xu, C.-Y.; Zhang, C.-Y.; Ye, M.-W. and Liu, W.-B.(2019).Effect of dietary glutathione supplementation on the immune responses and the fatty acid and amino acid composition in Chinese mitten crab, *Eriocheir sinensis* Aquacult. Rep., 15, Article 100217.

Lõrincz, M. L. and Adamantidis, A. R. (2017). Monoaminergic control of brain states and sensory processing: existing knowledge and recent insights obtained with optogenetics. Prog. Neurobiol. 151, 237–253.

Martorell-Ribera, J.; Venuto, M.T.;Otten, W.; Brunner, R.M.; Goldammer, T.; Rebl, A. and Gimsa, U. (2020) Time-Dependent Effects of Acute Handling on the Brain Monoamine System of the Salmonid *Coregonus maraena*. Front. Neurosci. 14:591738.

Michelato, M.; Zaminhan, M.; Boscolo, W.R.; Nogaroto, V.; Vicari, M.; Artoni, R.F.; Furuya, V.R.B. and Furuya, W. M. (2017). Dietary histidine requirement of Nile tilapia juveniles based on growth performance, expression of muscle-growth-related genes and haematological responses. Aquaculture 2017, 467, 63–70.

Mischoulon, D. and Fava M. (2002). Role of S-adenosyl-L- methionine in the treatment of depression: a review of the evidence. *American Journal of Clinical Nutrition*. 76(5).

Mohanty, B.; Mahanty, A. ;Ganguly,S. ; Sankar, T. V.; Kajal Chakraborty, k. Rangasamy, A.; Paul, B.; Sarma, D.; Mathew, S.; Kunnath Asha, K . and Behera, K. (2014). Amino Acid Compositions of 27 Food Fishes and Their Importance in Clinical Nutrition . J .Amino Acids 2014; 2014: 269797.

Moro, J.; Tomé, D.; Schmidely, P.; Demersay, T-C. and Azzout-Marniche, D. (2020). Histidine: A Systematic Review on Metabolism and Physiological Effects in Human and Different Animal Species.Nutrients, 12: 1414.

Nose, T. (1979). Summary report on the requirements of essential amino acids for carp. In Finfish Nutrition and Fishfeed Technology, 145–156 [Halver, JE and Tiews, K, editors]. Berlin: Heinemann.

Petty, F. (1995). GABA and mood disorders: A brief review and hypothesis. J Affect Disord. ; 34:275–81.

Ren, M.; Habte-Tsion, H.; Liu, B.; Miao, L.; Ge, X.; Xie, J.; Liang, H.; Zhou, Q.and Pan, L.(2015). Dietary leucine level affects growth performance, whole body composition, plasma parameters and relative expression of TOR and TNF-a in juvenile blunt snout bream, *Megalobrama amblycephala*. *Aquaculture*, 448: 162–168.

Rahimnejad, S. and Lee, K.-J.(2013). Dietary valine requirement of juvenile red sea bream Pagrus major Aquaculture 416-417:212-218.

Reverter, M.; TapissierBontemps,N.; Sarter, S.; Sasal, P. and Caruso, D. (2020). Moving towards more sustainable aquaculture practices: a meta- analysis on the potential of plant- enriched diets to improve fish growth, immunity and disease resistance Rev. Aquacult., 13, 537-555.

Ricceri, L.; Moles, A. and Crawley, J. (2007). Behavioral phenotyping of mouse models of neurodevelopmental disorders relevant social behavior patterns across the life span. *Behav Brain Res.* 2007;176:40–52.

Rossi, W. ; Allen,K.M. ;Habte-Tsion, H.M. andMeesala,K.M.(2021). Supplementation of glycine, prebiotic, and nucleotides in soybean meal-based diets for largemouth bass (*Micropterus salmoides*): Effects on production performance, whole-body nutrient composition and retention, and intestinal histopathology Aquaculture, 532, Article 736031.

Rush, A. J. (2007). The varied clinical presentations of major depressive disorder. J Clin Psychiatry. 2007; 68:4–10.

Saavedra, M.; Conceição, L.E.; Barr, Y.; Helland, S.; Ferreira, P.P.; Yúfera, M. and Dinis, M.T. (2010). Tyrosine and phenylalanine supplementation onDiplodus sarguslarvae: Effect on growth and quality. Aquac. Res., 41: 1523–1532.

Salamanca, N.; Giráldez, I.; Morales, E.; de La Rosa, I. and Herrera, M. (2020). Phenylalanine and Tyrosine as Feed Additives for Reducing Stress and Enhancing Welfare in Gilthead Seabream and Meagre. Animals (Basel). Dec 29;11(1):45.

Sánchez-Machado, D.I.; López- Cervantes, J.; López-Hernández, J.; Paseiro-Losada, P. and Simal-Lozano, J. (2003). High-Performance Liquid Chromatographic Analysis of Amino Acids in Edible Seaweeds after Derivatization with Phenyl Isothiocyanate. Chromatographia, 58 (3/4): 159–163.

Schreck, C.B. and Tort, L. (2016.). The Concept of Stress in Fish. In Fish Physiology; Elsevier: Amsterdam, The Netherlands, pp. 1–34.

Shafik, M.; Ibrahime, H.; Elyazeid, I.A.; Abass, O. and Saad, H.M. (2014). The stress of phenylalanine on rats to study the phenylketonuria at biochemical and molecular level. J. Appl. Pharm. Sci., 4, 24–29.

Stockmeier, C. A. (1997). Neurobiology of serotonin in depression and suicide. Ann N Y Acad Sci. 1997; 836:220–32.

Tan, X.; Lin, H.; Huang, Z.; Zhou, C.; Wang, A.; Qi, C. and Zhao, S. (2016). Effects of dietary leucine on growth performance, feed utilization, non-specific immune responses and gut morphology of juvenile golden pompano *Trachinotus ovatus*. *Aquaculture*, *465*, 100–107.

Van Praag, H.M. (1983). Depression, suicide and the metabolism of serotonin in the brain. J Affect Disord.; 4:275–90.

von Stackelberg, K.; Guzy, E.; Chu, T. and Claus Henn, B. (2015). Exposure to Mixtures of Metals and Neurodevelopmental Outcomes: A Multidisciplinary Review Using an Adverse Outcome Pathway Framework. Risk Anal Off Publ Soc Risk Anal. 2015; 35:971–1016.

35:971–1016.

Wang, B.; Liu, Y.; Feng. L.; Jiang, W.D.; Kuang, S.Y. and Jiang, J. (2015). Effects of dietary arginine supplementation on growth performance, flesh quality, muscle antioxidant capacity and antioxidant-related signalling molecule expression in young grass carp (*Ctenopharyngodon idella*) *Food Chem.* ;167:91–99.

Wang, Q.; Xu, Z. and Ai, Q. (2021). Arginine metabolism and its functions in growth, nutrient utilization, and immunonutrition of fish. Anim Nutr. 7(3):716-727.

Wang, W.; Wu, Z.; Dai, Z.; Yang, Y.; Wang J. and Wu, G. (2013). Glycine metabolism in animals and humans: implications for nutrition and health. *Amino Acids*. 2013;45(3):463–477.

Watanuki, H.; Chakraborty, G.; Korenaga, H.; Kono, T.; Shivappa, R.B. and Sakai, M. (2009). Immunostimulatory effects of natural human interferon-alpha (huIFN-alpha) on carps Cyprinus carpio L. Vet Immunol Immunopathol. 15;131(3-4):273-7.

WHO. (2007). World Health Organization. Protein and amino acid requirements in human nutrition. *WHO Technical Report.* series 935. Geneva, Switzerland:

Wilson, R.P.; Poe, W.E. and Robinson, E.H. (1980). Leucine, isoleucine, valine and histidine requirements of fingerling channel catfish. J. Nutr. 110, 627–633.

Wright, P.A. (2011). Nitrogenous-waste balance|Ureotelism *Encycl Fish Physiol*. 2011:1444–1449.

Wu, C.; Ye, J.; Gao, J.; Chen, L. and Lu, Z. (2016). The effects of dietary carbohydrate on the growth, antioxidant capacities, innate immune responses and pathogen resistance of juvenile Black carp Mylopharyngodon piceus. Fish Shellfish Immunol. 49:132-42.

Wu, G. (2013). Functional amino acids in nutrition and health. *Amino Acids*. 2013;45(3):407–411.

Wu, G. (2010). Functional amino acids in growth, reproduction, and health. *Advances in Nutrition*. 1(1):31–37.

Wu, C.; Chen, L.; Lu, Z.; Gao, J.E.; Chu, Y.; Li, L.; Wang, M.; Zhang, G.; Zhang, M.and Ye, J. (2017). The effects of dietary leucine on the growth performances, body composition, metabolic abilities and innate immune responses in black carp *Mylopharyngodon piceus*. *Fish Shellfish Immun.*, 67, 419–428.

Xie, S.-W. ; **Tian, L.-X.**; **Jin, Y.** ;**Yang, H.-J.** ;**Liang, G.-Y. and Liu, Y.-J.** (2014). Effect of glycine supplementation on growth performance, body composition and salinity stress of juvenile Pacific white shrimp, *Litopenaeus vannamei* fed low fishmeal diet Aquaculture, 418–419: 159-164.

Xie, S. W. ;Zhou, W.; Tian, L.; Niu, J. and Liu, Y. (2016). Effect of N-acetyl cysteine and glycine supplementation on growth performance, glutathione synthesis, anti-oxidative and immune ability of Nile tilapia, *Oreochromis niloticus* Fish. Shellfish Immunol., 55: 233-241.

Yamamoto, T.; Shima, T. and Furuita, H. (2004). Antagonistic effects of branchedchain amino acids induced by excess protein-bound leucine in diets for rainbow trout (*Oncorhynchus mykiss*) Aquaculture, pp. 539-550.

Zehra, S. and Khan, M. A. (2015). Dietary leucine requirement of fingerling Catla *catla* (Hamilton) based on growth, feed conversion ratio, RNA/DNA ratio, leucine gain, blood indices and carcass composition. *Aquacult. Int.*, 23: 577–595.

Zhang, S.; Chen, X.; Huang, Z.; Chen, D.W.; Yu, B.; Chen, H.; Luo, J.; He, J.; Zheng, P. and Yu, J. (2018). Leucine promotes differentiation of porcine myoblasts through the protein kinase B (Akt)/Forkhead box O1 signalling pathway. *Brit. J. Nutr.*, *119*: 727–733.

Zheng, H.; Guo, Q.; Duan, X.; Xu, Z. And Wang, Q. (2019). L-arginine inhibited apoptosis of fish leukocytes via regulation of NFkB-mediated inflammation, NO synthesis, and anti-oxidant capacity. *Biochimie*. 158:62–72

Zhou, Z.; Wang, X.; Wu, X.; Gao, Y.; Li, X.; Dong, Y. and Yao, W. (2019). Effects of dietary leucine levels on growth, feed utilization, neuro-endocrine growth axis and TOR-related signaling genes expression of juvenile hybrid grouper (*Epinephelus fuscoguttatus* $\stackrel{\frown}{} \times$ *Epinephelus lanceolatus* $\stackrel{\frown}{}$). Aquaculture, 504: 172–181.

Zhu, L.Y.; Nie, L.; Zhu, G.; Xiang, L. X. and Shao, J. Z. (2013). Advances in research of fish immune-relevant genes: a comparative overview of innate and adaptive immunity in teleosts. Dev Comp Immunol. 39(1-2):39-62.