

## Chemical Composition, Physicochemical and Microbiological Quality Properties of *Cyprinus carpio* and *Euthynnus alletteratus*

Abdelrahman S. Talab<sup>1\*</sup>, Mohamed M. M. Khallaf<sup>2</sup>, Mamdouh H. O. El-Kalyoubi<sup>2</sup>,  
Abdel-Fattah, A. A.<sup>2</sup>, Shimaa M. Abou-Taleb<sup>2</sup>

<sup>1</sup>National Institute of Oceanography and Fisheries (NIOF), Cairo, Egypt

<sup>2</sup>Food Science Department, Faculty of Agriculture, Ain Shams University, Egypt

\*Corresponding Author: [Abdelrahman\\_saidh@yahoo.com](mailto:Abdelrahman_saidh@yahoo.com)

### ARTICLE INFO

#### Article History:

Received: Feb. 10, 2024

Accepted: March 27, 2024

Online: April 29, 2024

#### Keywords:

Fresh fish,  
Chemical composition,  
Physicochemical,  
Microbiological quality

### ABSTRACT

This research was designed to determine the chemical composition, physicochemical and microbiological quality of fresh common carp (*Cyprinus carpio*) and little tuna (*Euthynnus alletteratus*) fish. The results showed that, values of moisture, protein, lipid, ash, and carbohydrates of fresh common carp and little tuna fish samples were as follows: for common carp, 74.45, 17.74, 4.81, 2.11, 0.89% were recorded, while for little tuna, 70.38, 22.07, 6.15, 1.43 and 0.56% were registered, respectively. Whereas, TVB-N, TMA-N, TBA and pH values of fresh common carp and little tuna fish samples were as follows: for common carp, 8.36mg/ 100g, 1.56mg/ 100g, 0.48mg malonadehyde/ kg and 6.15, 46.55; for little tuna, 16.84mg/ 100g, 2.02mg/ 100g, 0.58mg malonadehyde/ kg, 6.35, 47.86, respectively. Referring to microbial quality characteristics, it is well established that spore forming bacterial count, yeast, and mold count, *E. coli* and *Salmonella* were not detected in fish samples, while total plate count and total coliform count of fresh common carp and little tuna fish samples were recorded as 3.15 and 3.07 log cfu/ g for common carp, and as 1.66 and 1.23 log cfu/ g for little tuna, respectively. The microbiological assessment of the fresh fish indicated that they were of high quality and safe for utilization, either as fresh fish or for further processing and use. The present study provided valuable information on the quality and safety of both fresh common carp and little tuna fish which will greatly help in designing the optimum conditions in developing the post-harvest value chain, leading to higher consumer acceptability of this fish species. Moreover, the findings of this study could be used to optimize the post-harvest handling systems of the fish and subsequent processing in order to deliver a wholesome and safe product to the consumer.

### INTRODUCTION

Indeed, information on the proximate composition, physicochemical and microbiological properties of fresh fish from is limited. Moreover, due to the changes in climate conditions, season, and industrial growth, there could be wide differences in the biochemical constituents of the fish (Javaid *et al.*, 1992). The chemical composition of

fish varies greatly by species and individual depending on age, sex, environmental conditions and season. Proximate composition involves the determination of moisture, lipid, protein, and ash contents. Common carp (*Cyprinus carpio*) is one of the most important fish species in aquaculture throughout the world. Annual world production is around 3000000 tons, with the European production accounting for 145000 tons (FAO, 2012). Common carp (*C. carpio*) is one of the most cultured fish in the world. However, its consumer preference is low, leading to a limited market presence due to the presence of intramuscular bones. Hence, there is a need to develop some convenience products from the meat of carps to enhance their consumer acceptability (Vanitha *et al.*, 2013).

Little tuna *Euthynnus alletteratus* is a pelagic species; it is one of the members of the Scombridae with a wide distribution in the world and is predominant in the Mediterranean Sea and Black Sea (Valeiras & Abad, 2007). It occurs in the Mediterranean throughout the year, with higher abundance during the summer months. The Egyptian Mediterranean catch from *E. alletteratus* fluctuated between 1302 and 1003 tons in the period from 2010 to 2018 (GAFRD, 2019). Some studies were conducted to determine the quality of this fish species. Fadi (2014) found that the chemical composition of kapreeta fish flesh did not have remarkable differences in moisture (77.80%), crude protein (76.99 %), ether extract (4.80%), ash (4.46 %) and carbohydrates 13.75%. Genina (2017) stated that, moisture content of fresh kapreeta fish decreased from 71.75 to 53.88% in untreated smoked fish fillet, and to 54.50% and 54.74% in treated samples, with 5% thyme or sage essential oils, respectively. The protein, lipid and ash contents of fresh kapreeta fish samples were 24.54, 2.11 and 1.60%, respectively. These values increased after liquid smoking in both untreated and treated smoked fish fillets. Hizbullah *et al.* (2020) postulated that, water content, ash content, fat content, protein content, and carbohydrate content of little tuna fish were 73.14, 1.32, 0.08, 22.97 and 2.26%, respectively.

## MATERIALS AND METHODS

### Materials

#### Fish samples collection and preparation

Fresh common carp (*Cyprinus carpio*) with  $44 \pm 6.11$ cm total lengths and  $4.45 \pm 0.51$  kg total weights were obtained from Manzala aquatic farm belonging to GAFAD, Egypt, during December 2019. Moreover, fresh little tuna (*Euthynnus alletteratus*) with  $49 \pm 4.01$ cm total lengths and  $6.50 \pm 0.48$ kg total weights were purchased from Alexandria fish market, Egypt, during December 2019. Fish samples were transported immediately using ice box in three hours to Fish Processing and Technology Laboratory, El-Kanater El-Khairia, Fish Research Station, National Institute of Oceanography and Fisheries. Fish samples were washed well with tap water, beheaded, gutted, filleted after the removal of scales, fins, skin and large bones, then rewashed carefully and drained.

### Analytical methods

Moisture, crude protein, lipid and ash were determined according to the guidelines of AOAC (2012). Carbohydrates were calculated by the difference in the sum of the values of fat, ash, moisture, and protein content. The pH value was measured as described Egbert *et al.* (1992). Total volatile basic nitrogen (TVB-N) contents were determined according to the guidelines of AOAC (2012). Moreover, trimethylamine nitrogen (TMA-N) contents were determined as mentioned by Pearson (1976), and thiobarbituric acid (TBA) values were determined according to Siu and Draper (1978). Total mesophilic bacterial count (TMBC) and total psychrophilic bacterial count (TPBC) were determined according to the method described in APHA (2001). For the total plate count (TPC), *E. coli* and *Salmonella* were determined according to ISO (2003) via a nutrient agar medium, as described by Oxoid (2006). Yeast and mold counts (YMC) were determined in accordance with ISO (2008). Data were expressed as the mean values of three replicates, and standard deviations were statistically analyzed by performing the analysis of variance technique (ANOVA), using the statistical analysis system according to SAS (2008). Differences among means were compared using Duncan's multiple range test (1955) at a significant level of 95% ( $P \leq 0.05$ ).

## RESULTS AND DISCUSSION

### Chemical composition

The chemical composition of fish flesh could be affected by several intrinsic and extrinsic factors. The first factors include age, sex and size which are related to the different ratios of viscera, muscles and bones in fish body. The second factors include the environmental factors, mainly water temperature which regulates the rate of fish growth and food composition which could govern the biochemical composition of fish flesh, especially under cultivation (Nortvedt & Tuene, 1998).

**Table 1.** Chemical composition of fresh common carp and little tuna fish (on wet weight basis)

Parameter	Common carp	Little tuna
Moisture %	74.45±0.85	70.38±0.73
Protein %	17.74±0.43	22.07±0.56
Fat %	4.81±0.59	6.15±0.84
Ash %	2.11±0.82	1.43±0.78
Carbohydrate %	0.89±0.19	0.56±0.19

Data are the mean ± SD, n = 10; Carbohydrates percent calculated by difference.

The moisture, protein, lipid, ash and carbohydrates of fresh common carp fish were 74.45, 17.74, 4.81, 2.11 and 0.89%, respectively, while they recorded values of 70.38, 22.07, 6.15, 1.43 and 0.5 % for fresh little tuna (*Euthynnus alletteratus*) fish samples,

respectively (Table 1). In general, the chemical composition of fish differs from one to the other depending on environment, season, sex, and age. The chemical composition of fresh fish is given as 66– 81% water, 16– 21% protein, 0.2– 15% fat, 1.2– 1.5% mineral and 0– 0.5% carbohydrate (Mazumder *et al.*, 2008). The obtained results coincide with those of Muraleedharan *et al.* (1996), Murthy *et al.* (2019) and Badran *et al.* (2020).

### Physicochemical quality

The pH values are an important factor and can be used as a good indicator for evaluation freshness and quality of fish. Moreover, values of thiobarbituric acid reactive substances (TBARS) is considered as an indicator for the amount of malonaldehyde (MDA), which is the most predominant secondary oxidation products of lipid food (Greene & Cumuze, 1982). In addition, the TVB-N content is an indicative of putrefaction and decompositions of proteins constituent's breakdown in fish tissues (Khallaf, 1990).

**Table 2.** Physicochemical quality of fresh common carp and little tuna fish

Parameter	Common carp	Little tuna
TVB-N ( mg/100g sample)	8.36±0.96	16. 84±0.89
TMA-N( mg/100g sample)	1.56±0.33	2.02±0.57
TBA (mg malonadehyde/kg)	0.48±0.82	0.58±0.47
pH value	6.15±0.94	6.35±0.66
Water holding capacity %	46.55±0.77	47.86±0.98

Data are the mean ± SD, n = 10.

Table (2) shows that the TVB-N, TMA-N, TBA, pH value and WHC of fresh common carp were 8.36mg/ 100g, 1.56mg/ 100g, 0.48mg malonadehyde/ kg, 6.15 and 46.55%, respectively. On the other hand, for fresh little tuna, values recorded were 16.84mg/ 100g, 2.02mg/ 100g, 0.58mg malonadehyde/ kg, 6.35 and 47.86%, respectively. These results agree with those of Živković *et al.* (2004), Khidhir (2011), Ćirković *et al.* (2012) and Khanipour *et al.* (2014). These obtained results indicate that tested fresh fish samples were within national and international standards and did not exceed the maximum permissible limits of TVB-N and TMA-N and TBA. Connell (1995) reported that the limit of acceptability of fish is 20 to 30mg N/ 100g. On the other hand, Kirk and Sawyer (1991) suggested values of 30 to 40mg N/ 100g as the upper limit. Ghaly *et al.* (2010) also suggested that a level below 25mg/ 100mg serves as the borderline for the TVB-N content for various fish and fish products. From the data presented in Table (2), the pH value was 6.35 for fresh little tuna, and 6.15 for fresh common carp fish. This result is in accordance with those of Genina (2017) who found that, the pH of fresh kapreeta (like tuna) fish flesh was 6.4. The WHC mean values in little tuna and common carp fish was 47.86 and 46.55%, respectively. Whereas, water holding ability was inversely proportional with the percentage of calculated WHC according to free water lost upon the

determination. The variation in % WHC could be also discussed based on the fat content of fish species.

### Microbiological quality

The quality of fish could be degraded through a complex process, in which the physical, chemical, and microbiological forms of deterioration are involved. The enzymatic and chemical reactions are usually responsible for the initial loss of freshness, whereas microbial activity is responsible for the obvious spoilage, and thereby establishes product shelf- life (Huss, 1997).

**Table 3.** Microbiological quality of fresh common carp and little tuna fish

Parameter	Common carp	Little tuna
Total plate count (log cfu/g)	3.15	3.07
Total coliform count (log cfu/g)	1.66	1.23
Spore forming bacterial count (log cfu/g)	≤1	≤1
Yeast and mold count (log cfu/g)	≤1	≤1
<i>E. coli</i> (log cfu/g)	ND	ND
<i>Salmonella</i> (log cfu/g)	ND	ND

ND: Not Detected.

Table (3) shows that the total plate count (log cfu/g) and total coliform count (log cfu/g) of fresh common carp and little tuna were as follows: for common carp, values were 3.15 and 3.07, while for little tuna, values were 1.66 and 1.23, respectively, while spore forming bacterial count (log cfu/ g), yeast, and mold count (log cfu/ g) were evaluated. Remarkably, *E. coli* (log cfu/ g) and *Salmonella* (log cfu/g) were not detected in any of the fish samples. This results agree with the findings of Talab *et al.* (2014). The microbiological assessment of the fresh fish indicates that they were of a high quality and safe for utilization, either as fresh fish or for further processing and usage.

## CONCLUSION

The present study provides valuable information on proximate composition, physicochemical and microbiological quality of both fresh common carp and little tuna fish, which will greatly help in designing the optimum conditions for developing the post-harvest value chain, leading to higher consumer acceptability of this fish species.

## REFERENCES

- AOAC (2012).** Official Methods of Analysis of the Association of Official Analytical Chemistry International, 19<sup>th</sup> ed., Gaithersburg, Maryland, USA. 1200 p.
- APHA (2001).** American Public Health Association, Compendium of Methods for the Microbiological Examination of Foods. Washington, D.C. U.S A. <https://www.apha.org>.
- Badran, M.; Sulaeman, S. and Ghanem, B. (2020).** Comparison of some biochemical properties of two marine fish species *Euthynnus alletteratus* and *Siganus rivulatus* in Syrian marine waters. Tishreen University Journal for Research and Scientific Studies -Biological Sciences Series, 42 (5): 257-268.
- Ćirković, M.; Jubojević, D.; Đorđević, V.; Novakov, N. and Petronijević, R. (2012).** Chemical composition of body including fatty acids of four cyprinids fish species cultured at the same conditions. Archiva Zootechnica.,15 (2) :37-50.
- Connell, J. J. (1990).** Methods of assessing and selecting for quality, p. 122-150. In J. J. Connell (ed.), Control of fish quality, 3rd ed. Fishing News Books, Oxford, United Kingdom.
- Egbert, W.R.; Huffman, D.L.; Chen, C.M. and Jones, W.R. (1992).** Microbial and oxidative changes in low-fat ground beef during simulated retail distribution. J. Food Sci., 57 (6): 1269-1269.
- Fadl, W. M. (2014).** Chemical and technological studies on Kapretta fish. M.Sc. of Agric. Sci. Kafrelshiekh Univ., Egypt. 230 p.
- FAO (2012).** Food and Agriculture Organization, The state of world fisheriesand aquaculture. Rome: FAO.209 p.
- GAFRD (2019).** Genera Authority for Fisheries Resource Development, Annual fishery statistics report., Ministry of agriculture, Cairo, Egypt.
- Genina, E. M. (2017).** Lipid quality and stability of some processed marine fish products. Ph.D. of Agric. Sci. Alexandria Univ. 235 p.
- Ghaly, A.E.; Dave, D.; Budge, S. and Brooks, M.S. (2010).** Fish spoilage mechanisms and preservation techniques: Review. Am. J. Applied Sci., 7 (7): 859-877.
- Greene, B.A. and Cumuze, T.H. (1982).** Relationship between TBA numbers and in experienced panelists assessment of oxidized flavor in cooked beef. J. Food Sci., 47: 52-58.
- Hizbullah, H. H.; Sari, N. K.; Nurhayati, T. and Nurilmala, M. (2020).** Quality changes of little tuna fillet (*Euthynnus affinis*) during chilling temperature storage. In IOP Conference Series: Earth and Environmental Science (404, No. 1, p. 012015).
- Huss, H.H. (1997).** Control of indigenous pathogenic bacterial in seafood. Food Control 8 (2): 91-98.
- ISO (2003).** International Organization for Standardization. Microbiology of food and animal feeding stuffs - Horizontal method for the enumeration of microorganisms – Colony - count technique at 30 degrees C, 4833.pp 1-6.

- ISO (2008)**. International Organization for Standardization. Microbiology of food and animal feeding stuffs - Horizontal method for the enumeration of yeasts and molds - Part 2: Colony count technique in products with water activity less than or equal to 0, 95, 21527. pp 1-5.
- Javaid, M.Y.; Salam, A.; Khan, M.N. and Naeem, M. (1992)**. Weight-length and condition factor relationship of a fresh water wild Mahaseer (*Tor putitora*) from Islamabad (Pakistan). Proc. Pak. Congr. Zool. 12, 335–340.
- Khallaf, M. F. (1990)**. Properties of smoked sausage processed from common carp fish. J. Agric. Sci. Mansoura Univ., 15 (18), 1288-1299.
- Khanipour, A. A.; Jorjani, S. and Soltani, M. (2014)**. Chemical, sensory and microbial quality changes in breaded kilka (*Clupeonella cultriventris*) with tempura batter in production stage and during frozen storage. Int. Food Res. J., 21 (6):2421-2430.
- Khidhir, Z.K. (2011)**. Comparative Quality Assessments of Five Local Fresh Fish in Sulaimani City markets. Ph. D. Thesis, Fac. Vet. Med., Sulaimani Univ.
- Kirk, R.S. and Sawyer, R. (1991)**. Pearson's Composition and Analysis of Foods (9th Edn). Longman Scientific and Technical, London, P 510 -511.
- Mazumder, M.S.A.; Rahman, M.M.; Ahmed, A.T.A.; Begum, M. and Hossain, M.A. (2008)**. Proximate composition of some small indigenous fish species (SIS) in Bangladesh. Int. J. Sustainable Crop Prod. 3 (3), 18-23.
- Muraleedharan, V.; Antony, K. P.; Perigreen, P. A. and Gopakumar, K. (1996)**. Utilization of unconventional fish resources for surimi preparation. Proceedings of the second workshop on scientific results of FORV SAGAR, New Delhi: Department of Ocean development, 1 (2):539-543.
- Murthy, L.; Narasimha, A.; Jeyakumari, G.; Phadke, U.; Parvathy, S.; Visnuvinayaga, M. and Ravishankar, C.N. (2019)**. Quality assessment and comparison of three different types of fish retail outlets of navi Mumbai, Maharashtra, India. Int.J. Curr. Microbiol.App.Sci 8 (8): 248-259
- Nortvedt, R. and Tuene, S. (1998)**. Body composition and sensory assessment of three weight groups of halibut (*H. hippoglossus*) fed three pellet sizes and three dietary fat levels. Aquacul., (161): 295-313.
- Oxoid (2006)**. The Oxoid Manual of Culture Media and Other Laboratory Services.9th edition. London, England. 626 p.
- Pearson, D. (1976)**. The Chemical Analysis of Foods (7th edition). Churchill Livingstone. 575p.
- SAS Program (2008)**. SAS System for Windows (Statistical Analysis System). Version 9.2. Cary, USA: SAS Institute Inc.986 p.
- Siu, G.M. and Draper, H.H. (1978)**. A survey of the malonaldehyde content of retail meats and fish. J. Food Sci. 43:1147-1149.
- Talab, A.S. (2014)**. Effect of cooking methods and freezing storage on the quality characteristics of fish cutlets. Adv. J. Food Sci. Technol., 6:468-479.

- Tsighe, N.; Wawire, M.; Bereket, A.; Karimi, S. and Wainaina, I. (2018).** Physicochemical and microbiological characteristics of fresh Indian mackerel, spotted sardine and yellowtail scad, from Eritrea Red Sea waters. *Journal of Food Composition and Analysis*, 70, 98-104.
- Valeiras, J. and Abad, E. (2007).** ICCAT Field Manual. Chapter 2. Description of Species. 2.1 Species Directly Covered by the Convention. 2 (1): 2.11.
- Vanitha, M.; Dhanapal, K.; Sravani, K.; Vidya, G. and Sagar, R. (2013).** Quality evaluation of value-added mince-based products from catla (*Catla catla*) during frozen storage. *Int. J. Sci. Environ. Technol.*, 2 (1):487-501.
- Zivković, D.; Perić, V. and Perunović, M. (2004).** Examination of some functional properties of silver carp (*Hypophthalmichthys molitrix*) and carp (*Cyprinus carpio*) Meat. *Joa Sci*, 49 (2):193-203.