



The Growth Rates of the Nile Tilapia (*Oreochromis niloticus*) and the Common Carp (*Cyprinus carpio* L.) Under Biofloc Technology by Using Different Carbon Sources

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ABSTRACT

The study aimed to evaluate the impact of different carbon sources on biofloc generation of Nile tilapia, *Oreochromis niloticus* and common carp, *Cyprinus carpio*. 360 healthfully Nile tilapia and common carp were subject to an experiment for the current research work. The Nile tilapia had a starting body weight of 6.5 ± 0.4 g, while the common carp had a starting body weight of 6.9 ± 0.1 g. The samples were divided into six equal groups, with the initial three groups designated for the Nile tilapia and the remaining groups for the common carp. Each group was duplicated three times with 20 fish in each replication. Four diets were adjusted for the experiment. The first and fourth groups, serving as control groups, were given a baseline control diet containing 25% crude protein (CP). Groups two, three, five, and six were given a basic diet with 25% CP, along with additional molas and starch for carbohydrate supplies. The trial extended for 60 days. Both the Nile tilapia and common carp exhibited the highest feed conversion ratio and a superior weight gain when provided with starch and molasses compared to the control group. There was no notable change in water temperature across the groups. The biofloc groups had higher oxygen levels compared to the control group. Ammonia levels decreased in all treated groups in comparison with the control groups. The Nile tilapia and the common carp experienced an improvement in both growth performance and water quality in a biofloc system utilizing different carbon sources.

INTRODUCTION

In 2020, the aquaculture industry made a substantial contribution by producing 122 million tons of food to global food security (FAO, 2022; Khanjani *et al.*, 2023a). Since the 1980s, aquaculture has become more important in supplying fish for human consumption, as shown by FAO (2018) and Tinh *et al.* (2021). Thus, future growth in the aquaculture industry should focus on maximizing resource efficiency, as suggested by Crab *et al.* (2012) and the World Bank (2013).

Enhancing the effective use of essential natural resources including water, land, and fish feed is vital for aquaculture operations (Tinh *et al.*, 2021). Sustainable growth is necessary to protect the environment, conserve natural resources, and maintain the profitability of the aquaculture industry. Aquaculturists globally are keen in sustainable production techniques viz. biofloc technology (Bossier & Ekasari, 2017; Salama *et al.*, 2021).

Biofloc technology (BFT) is a method created to address these issues (**Ahmad et al., 2017; El-Sayed et al., 2021; Mabroke et al., 2021; Khanjani et al. (2023b). Ekasari and Crab (2015)** advocated using aquaculture biofloc technology systems as an efficient alternative to traditional fish raising systems to solve water conservation and minimize water circulation. Biofloc technology is hailed as the upcoming "blue revolution" in aquaculture because to its capacity to stimulate the growth of microorganisms by the addition of carbohydrates, including sucrose, glucose, cassava, starch, molasses, or cellulose to the water in order to regulate the carbon-to-nitrogen ratio (C:N ratio). Higher C:N ratio promotes bacterial development, resulting in rapid proliferation (**Khanjani et al., 2023b**).

BFT is a potential strategy for improving water quality and decreasing nutrient-rich effluents in the environment because of its low water exchange, as demonstrated by **Martínez-Cordova et al. (2015)** and **Walker et al. (2019)**. Biofloc technology utilizes a consortium of microorganisms, including bacteria, microalgae, fungus, and zooplankton, to enhance water quality through the formation of protein-rich microbial bioflocs. This approach decreases the feed conversion ratio (FCR) of aquatic species, enabling the utilization of low-protein feeds, and resulting in cost reduction and economic benefits in aquaculture. In order to achieve low or zero-water exchange in BFT, it is essential to use cost-effective or easily accessible waste carbon sources to decrease expenses and improve overall profits (**Zaki et al., 2020**).

Carbohydrates are vital nutrition for animals. Sufficient carbohydrate levels in farmed fish diets enhance the quality of extruded feed pellets and support the protein sparing effect for energy requirements. Carbohydrates are a cost-efficient and ideal energy source in diets. However, fish species vary in their ability to use carbohydrates depending on their feeding habits, the quantity of dietary carbohydrates, and the complexity of their diet. Studies have shown that animals maintained in a controlled setting benefit from ingesting biofloc by experiencing an improved growth rate, a decreased feed conversion ratio, and reduced feed costs (**Burford et al., 2004; Wasielesky et al., 2006**). Bioflocs including bioactive compounds that improve survival and defense mechanisms, offer nutrients, and control water quality. This suggests a novel approach to health management in aquaculture by boosting the innate immune system of cultivated animals (**Ahmad et al., 2017**).

Catfish and tilapia, freshwater finfish species cultivated using the BFT technique, were the predominant aquaculture species globally in 2016, accounting for more than half of total aquaculture production, as reported by **FAO (2018)**. The Nile tilapia is a warm-water omnivorous fish specimen that is widely farmed for its fast growth, high fillet yield, and strong resistance to diseases (**Wang et al., 2005**). The Nile tilapia can efficiently metabolize up to 40% of digestible food carbohydrates, making it an ideal fish species for investigating the effects of dietary carbohydrates in long-term studies.

There is a growing interest in cultivating freshwater fish from the Cyprinidae family using the BFT technique. Minimal attempts have been made to evaluate the effectiveness of the system with various species in this family (**Kamilya et al., 2017**). These fish exhibit suspension-feeding behavior and possess a greater tolerance to increased quantities of suspended solids, making them ideal for thriving in the BFT system (**Romano et al., 2018**). Using different carbon sources in a biofloc system significantly affects water quality indicators, microbial community composition, and the growth and survival of common crop fry (**Mishra et al., 2024**). The amount of organic carbon significantly impacts the water quality and the development of the Nile tilapia in biofloc systems (**AbouelFadl et al., 2022**).

This study intended to assess the effects of different carbon sources on water quality indicators and the growth rates of the Nile tilapia and common carp.

MATERIALS AND METHODS

Experimental site and duration

The research was carried out in El-hag Bahy El-deen Monir Farm in Kafr El-Sheikh Governorate, Egypt, and spanned a duration of 60 days.

Study design and dietary treatments

360 healthy Nile tilapia (*Oreochromis niloticus*) and common carp (*Cyprinus carpio*) were obtained from a private farm in Kafr El-Sheikh, Egypt. The initial body weight was 6.5 ± 0.4 g for the Nile tilapia and 6.9 ± 0.1 g for the common carp. The fish were kept in a water circulation system to maintain ideal circumstances. The research was conducted in eighteen plastic containers, each with a capacity of 35 liters and measuring 48 x 35 x 20 cm. The fish were acclimated for 20 days before starting the experiment. Before the experiment started, the participants were divided into six equal groups, with three groups per species, and each group was reproduced three times with 20 fish in each replication. Four experimental diets were presented for discussion: The first and fourth groups were assigned as controls and received the baseline control diet containing 25% crude protein (CP). Groups two, three, five, and six were given a baseline diet with 25% CP, supplemented with molasses and starch for carbohydrates. The fish were given a daily ration equivalent to 5% of their body weight, split into two feedings every day. Every 20 days, fish in each replicated tank were weighed, and adjustments were made to the daily meal and carbon source quantities according to the fish's weight.

Supervising the quality of water

The study involved daily and weekly measurements of water temperature, dissolved oxygen levels, pH, and ammonia concentrations using thermometers, portable digital pH meters (Martini Instruments type 201/digital), and waterproof portable BOD and DO meters (type Hanna waterproof IP67). TAN was measured using **El Sayed's (2006)** technique.

Growth performance parameters

Measurements of growth parameters were taken at 20-day intervals. After the feeding trial, growth parameters such as initial weight, weight gain (WG), survival rate (SR), specific growth rate (SGR), and feed conversion rate (FCR) were computed using the formulas outlined by **Castell and Tiews (1980)**.

$$\text{Weight gain (WG)} = \text{Final fish weight (g)} - \text{Initial fish weight (g)}$$

$$\text{Survival rate (\%)} = \text{Number of live fishes} * 100 / \text{Total initial number of fish}$$

$$\text{Specific growth rate (SGR \%)} = \frac{\log \text{FW} - \log \text{IW}}{t} * 100$$

$$\text{Feed conversion ratio (FCR)} = \text{Feed intake (g)} / \text{Weight gain (g)}$$

FW denotes the final weight of the fish in grams. The initial weight of fish is labeled as IW in grams, and t signifies the total experimental days.

Statistical analysis

Excel 2013 was used to create a database for the initial investigation, employing descriptive statistics. The data were presented as the mean value plus or minus the standard deviation. The data were subjected to one-way analysis of variance (ANOVA) to assess the effect of treatment inclusion on fish performance. The data were analyzed using SPSS version 16 software (SPSS, 1997). Mean differences were analyzed using Duncan's multiple range test with a significance level of $P < 0.05$.

RESULTS

Water quality

Table (1) displays the water quality parameter measurements for the Nile tilapia and common carp. The average water temperature fluctuated between 26.6 and 26.8°C. The biofloc groups had higher dissolved oxygen levels compared to the control group in both the Nile tilapia and common carp treatments, ranging from 6.5±0.4 to 7.5±0.5 and 6.8±0.5 to 8.0±0.5 mg/L, respectively. Ammonia levels were reduced in all treatment groups in comparison with the control group. Concentrations in tilapia treatments ranged from 1.06±0.9 to 0.8±0.7, whereas in common carp treatments, concentrations varied from 1.02±0.7 to 0.6±0.4. The pH levels for the tilapia treatment ranged from 7.3±0.6 to 7.8±0.5, whereas for the common carp treatment, it ranged from 7.9±0.4 to 8.1±0.5.

Table 1. Water quality values of aquaria used in rearing the Nile tilapia (*O. niloticus*) and common carp (*C. carpio*) after 60 days of culture with different diets in the biofloc system

Item	The Nile tilapia (<i>O. niloticus</i>)			Common carp (<i>C. carpio</i> L.)		
	Control	Starch	Molas	Control	Starch	Molas
Temperature (o C)	26.8±2.3	26.6±2.5	26.8±2.5	26.8±2.3	26.6±2.5	26.8±2.5
DO (mg/L)	6.5±0.4	7.3±0.5	7.8±0.6	6.8±0.5	7.9±0.4	8.0±0.5
pH	7.3±0.6	7.6±0.4	7.8±0.5	7.9±0.4	8.1±0.5	7.9±0.4
Ammonia (mg/L)	1.06±0.9	0.9±0.3	0.8±0.7	1.02±0.7	0.6±0.4	0.9±0.5

Growth performance parameters

Table (2) shows the growth rates of the Nile tilapia in the control, molas, and starch groups. Weight gain and the feed conversion ratio (FCR) were somewhat higher in fish fed starch compared to those fed molas. The control group exhibited less significance in comparison with both starch and molasses. Survival rates did not differ significantly across the starch, molas, and control groups, with the highest survival rate shown in the starch group. SGR levels were somewhat higher in fish that received molas compared to the starch treatment, and both were moderately raised compared to the control group.

Table 2. Growth performance of the Nile tilapia (*O. niloticus*) after 60 days of culture using different diets in the biofloc system

Item	Control	Starch	Molas
Initial weight (g/fish)	6.5±0.4	6.5±0.4	6.6±0.7
Final weight (g/fish)	23.4±0.8 ^a	24.6±0.4 ^a	24.3±1.0 ^a
Weight gain (g/fish)	16.9±1.3 ^b	18.1±0.4 ^a	17.7±0.8 ^a
Survival rate (%)	96.3±1.3 ^b	97.4±0.5 ^a	96±1.3 ^b
SGR (%/d)	3.4±0.2 ^b	4.4±0.4 ^a	4.5±0.5 ^a
FCR	1.4±0.1 ^a	1.1±0.1 ^b	1.2±0.1 ^b

Values with different superscripts in the same row indicated significant differences ($P \leq 0.05$)

Table (3) displays the growth rates and survival rate parameters findings. The starch groups exhibited the greatest weight growth in the common carp, followed by the molas group, and lastly the control group. Molas had the highest survival rate, and there was no significant difference in survival rates between the starch and control groups. The starch groups had the most significant FCR values, whilst the molas and control groups did not demonstrate any significant difference.

Table 3. Growth performance of common carp (*C. carpio*) after 60 days of culture using different diets in the biofloc system

Item	Control	Starch	Molas
Initial weight (g/fish)	7.2±0.08	6.9±0.1	7.1±0.2
Final weight (g/fish)	±0.544.2 ^b	±0.849.1 ^a	±2.346.5 ^b
Weight gain (g/fish)	37±0.5 ^c	42.2±0.3 ^a	39.4±0.1 ^b
Survival rate (%)	95.2±0.6 ^b	95.8±0.5 ^b	96.8±0.4 ^a
SGR (%/d)	2.8±0.6 ^a	2.9±0.2 ^a	3.1±0.3 ^a
FCR	1.5±0.1 ^a	1.1±0.2 ^b	1.3±0.1 ^a

Values with different superscripts in the same raw indicated significant differences ($P \leq 0.05$)

DISCUSSION

Water quality characteristics are crucial in aquaculture since they have a direct impact on fish development and weight gain in addition to playing a key role in sustaining a healthy aquatic environment (Sunny *et al.*, 2017; Khanjani *et al.*, 2020, 2021; Osman *et al.*, 2021). The study found that water quality metrics for the Nile tilapia and common carp in a biofloc system with various carbon sources matched the standard range recommended by numerous authors (Popma & Lovshin, 1995; Awad *et al.*, 2021; Abouel-Fadl *et al.*, 2022). Biofloc technology improves water quality by converting ammonium into simpler chemicals more quickly than nitrification (Kumar *et al.*, 2019). Waste products and leftover nutrients from feed are converted into bacterial biomass, creating biofloc which can serve as extra food for aquatic animals (Avnimelech, 2006).

The examination found that the water temperature ranged from 26.6 to 26.8 °C in all treatments, aligning with the optimal temperature range for fish development and survival, as shown in other studies (Essa, 1993; Boyd, 1998; Henish, 2016). Hwihy *et al.* (2021) found that *Oreochromis niloticus* grown in biofloc systems at various stocking densities showed better outcomes than the control group of fish (28.5± 0.53, 7.8± 0.33). On the other hand, dissolved oxygen is an essential factor for differentiating between different types of water bodies (Ibrahim & Ramzy, 2013; Osman *et al.*, 2021). The oxygen requirements of fish in the culture system are frequently influenced by the levels of nitrite and ammonia (Tilak *et al.*, 2007; Remen *et al.*, 2008).

The two selected carbon sources in the research exhibited significantly higher levels of dissolved oxygen compared to the control. Several writers have corroborated this discovery (Thilakan *et al.*, 2019; Tabarrok *et al.*, 2020; AbouelFadl *et al.*, 2022; Liu *et al.*, 2023). The dissolved oxygen (DO) levels varied between 6 and 7.8 mg L⁻¹ throughout the experiment. Both the tilapia and common carp biofloc groups showed elevated oxygen levels in comparison with the control group. In their study, Hwihy *et al.* (2021) found a decrease in the dissolved oxygen levels in the biofloc in comparison with the control group. Maintaining optimal pH levels is crucial for balancing the presence of dangerous and innocuous ammonia in aquatic settings. Notably, ammonia poisoning can be lethal to fish in ponds. The pH levels were suitable for tilapia, grass carp, and common carp in the BFT system, utilizing various

carbon sources (Tabarrok *et al.*, 2020; Awad *et al.*, 2021; AbouelFadl *et al.*, 2022; Liu *et al.*, 2023).

Suita *et al.* (2015) and Wang *et al.* (2016) found that bacteria in biofloc contribute to food consumption, water quality control, and the breakdown of nitrogen compounds, specifically ammonia. All experimental groups exhibited significantly reduced levels of ammonia compared to the control group, reaching optimal values for fish rearing. The group that consumed molasses had the lowest amount, followed by starch in the Nile tilapia, and vice versa in the common carp. The quicker decrease in ammonia levels observed in the molasses and starch groups, which use simple carbon sources, is likely because these sources are more easily absorbed and used by heterotrophic bacteria. The bacteria metabolize ammonia, improving water quality according to several studies (Khanjani *et al.*, 2017; El-Shafiey *et al.*, 2018; Tabarrok *et al.*, 2020; Awad *et al.*, 2021; Khanjani *et al.*, 2021; Liu *et al.*, 2023). *Oreochromis niloticus* and *Cyprinus carpio* had increased final weight, weight growth, and SGR in bioflocs treatments with meals including molasses and starch compared to the control group. This finding is consistent with previous studies (Avnimelech, 1999, 2007; AbouelFadl *et al.*, 2022) which centered on *Oreochromis mossambicus*. The growth of tilapia was not affected by the use of various carbon sources in biofloc production, as reported by Silva *et al.* (2017). Mishra *et al.* (2024) found that the growth rate of different carbon sources in the BFT system increased when raising common carp fry. Multiple authors have shown that the growth of shrimp or tilapia was not impacted by the addition of different carbon sources in the biofloc system. The growth of tilapia in a biofloc system was not affected by the inclusion of molasses, sugar, or cassava starch. Both common carp and the Nile tilapia demonstrated a good adaptation to altered nutritional circumstances in a biofloc system, leading to an improved growth. Microbial clusters raised the production and effectiveness of digestive enzymes, leading to a greater nutrient absorption in the fish's gut. This likely enhanced the fish's growth and effectiveness of feed consumption (Moss *et al.*, 2001; Xu *et al.*, 2012; Xu & Pan, 2012).

The development of the Nile tilapia and common carp fish cultivated in BFT was unaffected by the carbon source. The current research validated the finding of Mishra *et al.* (2024) determining that the survival rate was greater in the experimental tanks with different carbon sources in the BFT system than in the control group. The biofloc groups exhibited the most notable feed conversion ratio (FCR) values in comparison with the control groups in the study. Previous studies have linked growth, indicating good health and a lower feed conversion ratio (FCR), to biofloc technology (BFT) systems' capacity to supply necessary nutrients like unsaturated fatty acids, crude protein, vitamins, and minerals (Azim *et al.*, 2008; Bakhshi *et al.*, 2018). Several research have shown that biofloc can lower production costs and can be used as a food source for the Nile tilapia and common carp (Crab *et al.*, 2010; Xu & Pan, 2014; Luna-González *et al.*, 2017; Adineh *et al.*, 2019; Zhao *et al.*, 2022). Furthermore, Zhao *et al.* (2012) showed that biofloc enhanced carp production in a polyculture system.

CONCLUSION

The study showed that employing biofloc as a substitute carbon source may improve water quality and boost development in the Nile tilapia and the common carp farming, without requiring water exchange.

REFERENCES

- AbouelFadl, K.Y.; Ahmed, S.A. and Badrey, A.E.A.** (2022). Effects of Biofloc on Water Quality and Growth Performance of the Nile Tilapia (*Oreochromis niloticus*) in a Zero-Water Exchange System. *Egyptian Journal of Aquatic Biology and Fisheries*, **26**(6): 765-779.
- Adineh, H.; Naderi, M.; Khademi Hamidi, M. and Harsij, M.** (2019). Biofloc technology improves growth, innate immune responses, oxidative status, and resistance to acute stress in common carp (*Cyprinus carpio*) under high stocking density. *Fish and Shellfish Immunology*, **95**: 440–448. <https://doi.org/10.1016/j.fsi.2019.10.057>
- Ahmad, I.; Babitha Rani, A.M.; Verma, A.K. and Maqsood, M.** (2017). Biofloc technology: an emerging avenue in aquatic animal healthcare and nutrition. *Aquaculture international*, **25**(3): 1215-1226.
- Avnimelech, Y.** (1999). Carbon/nitrogen ratio as a control element in aquaculture systems. *Aquaculture*, **176**: 227–235.
- Avnimelech, Y.** (2006). Bio-filters: the need for a new comprehensive approach. *Aquacultural engineering*, **34**(3): 172-178.
- Avnimelech, Y.** (2007). Feeding with microbial flocs by tilapia in minimal discharge bioflocs technology ponds. *Aquaculture*, **264**: 140–147.
- Awad, A.M.; Badrey, A.E.; AbouelFadl, K.Y. and Osman, A.G.** (2021). The impact of dietary carbon sources on the blood characteristics of grass carp (*Ctenopharyngodon Idella*) cultured in biofloc system. *Aquaculture, Aquarium, Conservation & Legislation*, **14**(6): 3178-3188.
- Azim, M.E.; Little, D.C. and Bron, J.E.** (2008). Microbial protein production in activated suspension tanks manipulating C:N ratio in feed and the implications for fish culture. *Bioresource Technology*, **99**: 3590-3599.
- Bakhsh E.M.; Khan S.A.; Marwani H.M.; Danish E.Y.; Asiri A.M. and Khan S.B.** (2018). Performance of cellulose acetate-ferric oxide nanocomposite supported metal catalysts toward the reduction of environmental pollutants. *International journal of biological macromolecules*, **107**: 668-677.
- Bossier, P. and Ekasari, J.** (2017). "Biofloc technology application in aquaculture to support sustainable development goals." *Microbial biotechnology*, **10**(5): 1012-1016.
- Boyd, C.E.** (1998). Pond water aeration systems. *Aquacultural engineering*, **18**(1): 9-40.
- Burford, M.A.; Thompson, P.J.; McIntosh, R.P.; Bauman, R.H. and Pearson, D.C.** (2004). The contribution of flocculated material to shrimp (*Litopenaeus vannamei*) nutrition in a high-intensity, zero-exchange system. *Aquaculture*, **232**(1-4): 525-537.
- Castell, J.D. and Tiewes, K.** (1980). Report of the EIFAC, IUNS and ICES Working Group on Standardization of Methodology in Fish Research, Hamburg, FRG, 21- 23 March 1979. IFAC Tech. Pap. (3) 24.
- Crab, R.; Chielens, B.; Wille, M.; Bossier, P. and Verstraete, W.** (2010). The effect of different carbon sources on the nutritional value of bioflocs, a feed for *Macrobrachium rosenbergii* postlarvae. *Aquaculture Research*, **41**: 559–567.
- Crab, R.; Defoirdt, T.; Bossier, P. and Verstraete, W.** (2012). Biofloc technology in aquaculture: Beneficial effects and future challenges. *Aquaculture*, **356**: 351–356.

- Ekasari, J. and Crab, R.** (2015). Verstraete W. Primary nutritional content of bio-flocs cultured with different organic carbon sources and salinity. *Hayati Journal of Biosciences*, **17**:125–130.
- El-Sayed, A.F.M.** (2006). *Tilapia Culture*. CABI Publishing, CAB International, Wallingford, Oxfordshire, UK, 277 pp.
- El-Sayed, S.; Friedman, M.; Anan, T.; Faris, M.A. and Sallam, H.** (2021). Diverse marine fish assemblages inhabited the paleotropics during the Paleocene-Eocene thermal maximum. *Geology*, **49**(8): 993-998.
- El-Shafiey, M.H.M.; Mabroke, R.S.; Mola, H.R.A.; Hassaan, M.S. and Suloma, A.** (2018). Assessing the suitability of different carbon sources for Nile tilapia, *Oreochromis niloticus* culture in BFT system. *AACL Bioflux*, **11**(3): 782-795.
- Essa, M.A.** (1993): Studies on the mass production on Nile tilapia (*Oreochromis niloticus*) fry in concrete basins. *Delta Journal Science*, **17**: 183-195.
- FAO** (2018). Fishery and aquaculture statistics. *FAO yearbook. Fishery and Aquaculture Statistics= FAO Annuaire. Statistiques des Peches et de l'Aquaculture= FAO Anuario. Estadisticas de Pesca y Acuicultura*, I-82.
- FAO (2022)**. Aquaculture in SOFIA 2022. *FAO Aquaculture Newsletter*, (66), 7-8.
- Henish, S.A.A.** (2016): The effect of Biofloc and aquaculture systems on growth and production of *Liza carinata* PH D Thesis, Faculty of Science, Ain Shams University, Egypt.
- Hwihi, H.; Zeina, A.; Abu Husien, M. and El-Damhougy, Kh.** (2021). Impact of Biofloc technology on growth performance and biochemical parameters of *Oreochromis niloticus*. *Egyptian Journal of Aquatic Biology and Fisheries*, **25**(1): 761-774.
- Ibrahim L.A. and Ramzy, E.M.** (2013). Water quality and its impact on *Tilapia zilli* (case study) Qarun Lake-Egypt. *International Water Technology Journal*, **3**(4): 170-191.
- Kamalam, B.S.; Medale, F. and Panserat, S.** (2017). Utilization of dietary carbohydrates in farmed fishes: new insights on influencing factors, biological limitations and future strategies. *Aquaculture*, **467**: 3-27.
- Kamilya, D.; Debbarma, M.; Pal, P.; Kheti, B.; Sarkar, S. and Singh, S.** (2017). Biofloc technology application in indoor culture of *Labeo rohita* (Hamilton, 1822) fingerlings: The effects on inorganic nitrogen control, growth and immunity. *Chemosphere*, **182**: 8-14.
- Khanjani, M.H.; Alizadeh, M. and Sharifinia, M.** (2020). Rearing of the Pacific white shrimp, *Litopenaeus vannamei* in a biofloc system: The effects of different food sources and salinity levels. *Aquaculture Nutrition*, **26**(2): 328-337.
- Khanjani, M.H.; Alizadeh, M. and Sharifinia, M.** (2021). Effects of different carbon sources on water quality, biofloc quality, and growth performance of Nile tilapia (*Oreochromis niloticus*) fingerlings in a heterotrophic culture system. *Aquaculture International*, **29**(1): 307-321.
- Khanjani, M.H.; Mozanzadeh, M.T.; Sharifinia, M. and Emerenciano, M.G.C.** (2023a). Biofloc: A sustainable dietary supplement, nutritional value and functional properties. *Aquaculture*, **562**: <https://doi.org/10.1016/j.aquaculture.2022.738757>
- Khanjani, M.H.; Mozanzadeh, M.T.; Sharifinia, M. and Emerenciano, M.G.C.** (2023b). Broodstock and seed production in biofloc technology (BFT): An updated review focused

- on fish and penaeid shrimp. *Aquaculture*, **579**: <https://doi.org/10.1016/j.aquaculture.2023.740278>
- Khanjani, M.H.; Sajjadi, M.M.; Alizadeh, M. and Sourinejad, I.** (2017). Nursery performance of Pacific white shrimp (*Litopenaeus vannamei* Boone, 1931) cultivated in a biofloc system: the effect of adding different carbon sources. *Aquaculture Research*, **48**: 1491-1501.
- Kumar, A.; Reddy, A.; Rani, A., Rathore, G.; Lakra, W. and Jayant, M.** (2019). Water quality and nutrient dynamics of biofloc with different C/N ratios in inland saline water. *Indian journal of animal research*, **9**(5): 783–791.
- Liu, Y.; He, Y.; Jia, X.; Fan, Y.; Yuan, L.; Shen, Y. and Li, J.** (2023). Effects of biological flocculation technology (BFT) on water quality dynamics and immune response of grass carp (*Ctenopharyngodon idella*). *Aquaculture and Fisheries*. DOI:10.1016/j.aaf.2023.04.005
- Luna-Gonzalez, A.; AvilaLeal, J.; Fierro Coronado, J.; Alvarez Ruiz, P.; Esparza Leal, H.; Escamilla Montes, R. and Lopez Alvarez, E.** (2017). Effects of bacilli, molasses, and reducing feeding rate on biofloc formation, growth, and gene expression in *Litopenaeus vannamei* cultured with zero water exchange. *Latin American Journal of Aquatic Research*, **45**: 900–907. <https://doi.org/10.3856/vol45-issue5-fulltext-4>
- Mabroke, R.S.; Zidan, A.E.N.F.; Tahoun, A.A.; Mola, H.R.; Abo-State, H. and Suloma, A.** (2021). Feeding frequency affect feed utilization of tilapia under biofloc system condition during nursery phase. *Aquaculture Reports*, **19**:100625.
- Martínez-Córdova, L.R.; Emerenciano, M.; Miranda-Baeza, A. and Martínez-Porchas, M.** (2015). Microbial-based systems for aquaculture of fish and shrimp: an updated review. *Reviews in Aquaculture*, **7**(2): 131-148.
- Mishra, R.; Tamot, P. and Vyas, V.** (2024). Effect of Biofloc system using several carbon sources on survival rate and weight gain of (*Cyprinus carpio L.*) Fry. *International Journal of Fauna and Biological Studies*, **11**(1): 7-11.
- Moss, S.M.; Divakaran, S. and Kim, B.G.** (2001). Stimulating effects of pond water on digestive enzyme activity in the Pacific white shrimp, *Litopenaeus vannamei* (Boone). *Aquaculture Research*, **32**: 125-132.
- Osman, A.G.M.; Farrag, M.M.S.; Badrey, A.E.A.; Khedr, Z.M.A. and Kloas, W.** (2021). Water quality and health status of monosex Nile Tilapia, *Oreochromis niloticus* cultured in aquaponics system (ASTAF-PRO). *Egyptian Journal of Aquatic Biology & Fisheries*, **25**(2): 785–802.
- Popma, T.J. and Lovshin, L.L.** (1995). Worldwide prospects for commercial production of Tilapia. *Aquaculture production manual*. Auburn University, Alabama 36849 International Center for Aquaculture and Aquatic Environments, p 42.
- Remen, M.; Imstrand, A.K.; Stefansson, S.O.; Jonassen, T.M. and Foss, A.** (2008). Interactive effects of ammonia and oxygen on growth and physiological status of juvenile Atlantic cod (*Gadus morhua*). *Aquaculture*, **274**: 292–299.
- Romano, N.; Dauda, A.B.; Ikhsan, N.; Karim, M. and Kamarudin, M.S.** (2018). Fermenting rice bran as a carbon source for biofloc technology improved the water quality, growth, feeding efficiencies, and biochemical composition of African catfish *Clarias gariepinus* juveniles. *Aquac. Res.*, **49**: 3691–3701.

- Salama, H.K.; Henish, S.; Mohamed, K.A.; Nassif, M.G.; El-Naggar, M.M.; Khalil, M.T. and Suloma, A.** (2021). The effect of dietary protein level and amino acid supplementation on the Nile tilapia (*Oreochromis niloticus*) nursering performance under biofloc system conditions at cold suboptimal water temperature. *Egyptian Journal of Aquatic Biology & Fisheries*, **26**(3): 917-936.
- Silva, U.L.; Dario, R.F.; Maurício, N.D.C.P. and Eudes, D.S.C.** (2017). Carbon sources and C:N ratios on water quality for Nile tilapia farming in biofloc system. *Rev. Caatinga, Mossoró.*, **30**: 1017-1027.
- SPSS** (1997). Statistical package for the social sciences, Versions16, SPSS in Ch, Chi-USA.
- Suita, K.; Fujita, T.; Hasegawa, N.; Cai, W.; Jin, H.; Hidaka, Y.; ... & Ishikawa, Y.** (2015). Norepinephrine-induced adrenergic activation strikingly increased the atrial fibrillation duration through β 1- and α 1-adrenergic receptor-mediated signaling in mice. *PLoS One*, **10**(7), e0133664.
- Sunny, N.E.; Bril, F. and Cusi, K.** (2017). Mitochondrial adaptation in nonalcoholic fatty liver disease: novel mechanisms and treatment strategies. *Trends in Endocrinology & Metabolism*, **28**(4): 250-260.
- Tabarrok, M.; Seyfabadi, J.; Salehi Jouzani, G. and Younesi, H.** (2020). Comparison between recirculating aquaculture and biofloc systems for rearing juvenile common carp (*Cyprinus carpio*): Growth performance, haemato-immunological indices, water quality and microbial communities. *Aquaculture research*, **51**(12): 4881-4892.
- Thilakan, A.P.; Kundan Kumar, K.K.; Shukla, S.P.; Sanath Kumar, S.K.; Neelam Saharan, N.S. and Sutanu Karmakar, S.K.** (2019). Occurrences of triclosan in Versova creek of Mumbai, India and its toxicity on selected aquatic organisms. *Journal of Experimental Zoology, India*, **22**(2): 737-742.
- Tilak, K.S.; Veeraiyah, K. and Raju, J.M.P.** (2007). Effects of ammonia, nitrite and nitrate on hemoglobin content and oxygen consumption of freshwater fish, *Cyprinus carpio* (Linnaeus). *Journal of Environmental Biology*, **28**(1): 45-47.
- Tinh, D.T.; Thuy, N.T. and Ngoc Huy, D.T.** (2021). Doing Business Research and Teaching Methodology for Undergraduate, Postgraduate and Doctoral Students-Case in Various Markets Including Vietnam. *Ilkogretim Online- Elementary Education Online*, **20**(1): 1414-1418.
- Walker, T.R.; Adebambo, O.; Del Aguila Feijoo, M.C.; Elhaimer, E.; Hossain, T.; Edwards, S.J.; Morrison, C.E.; Romo, J.; Sharma, N.; Taylor, S. and Zomorodi, S.** (2019). Chapter 27: Environmental effects of marine transportation. In: Sheppard, C. (Ed.), *World Seas: An Environmental Evaluation*, 2nd edn. Academic Press, pp.: 505-530.
- Wang, J.T.; Liu, Y.J.; Tian, L.X.; Mai, K.S.; Du, Z.Y.; Wang, Y. and Yang, H.J.** (2005). Effect of dietary lipid level on growth performance, lipid deposition, hepatic lipogenesis in juvenile cobia (*Rachycentron canadum*). *Aquaculture*, **249**(1-4): 439-447.
- Wang, Q.; Wang, Z.; Awasthi, M.K.; Jiang, Y.; Li, R.; Ren, X. and Zhang, Z.** (2016). Evaluation of medical stone amendment for the reduction of nitrogen loss and bioavailability of heavy metals during pig manure composting. *Bioresource Technology*, **220**: 297-304.

- Wasielisky, J.R.W.; Atwood, H.; Stokes, A. and Browdy, C.L.** (2006). Effect of natural production in a zero-exchange suspended microbial floc based super-intensive culture system for white shrimp *Litopenaeus vannamei*. *Aquaculture*, **258**(1-4): 396-403.
- Xu, W.J. and Pan, L.Q.** (2012). Effects of bioflocs on growth performance, digestive enzyme activity and body composition of juvenile *Litopenaeus vannamei* in zero-water exchange tanks manipulating C/N ratio in feed. *Aquaculture*, **356-357**: 147-152.
- Xu, W.J. and Pan, L.Q.** (2014). Dietary protein level and C/N ratio manipulation in zero-exchange culture of *Litopenaeus vannamei*: Evaluation of inorganic nitrogen control, biofloc composition and shrimp performance. *Aquaculture Research*, **45**(11): 1842-1851.
- Xu, W.J.; Pan, L.Q.; Zhao, D.H. and Huang, J.** (2012). Preliminary investigation into the contribution of bioflocs on protein nutrition of *Litopenaeus vannamei* fed with different dietary protein levels in zero-water exchange culture tanks. *Aquaculture*, **350**: 147-153.
- Zaki, M.A.; Alabssawy, A.N.; Nour, A.E.; El Basuini, M.F.; Dawood, M.A.; Alkahtani, S. and Abdel-Daim, M.M.** (2020). The impact of stocking density and dietary carbon sources on the growth, oxidative status and stress markers of Nile tilapia (*Oreochromis niloticus*) reared under biofloc conditions. *Aquaculture Reports*, **16**: 100282.
- Zhao, P.; Huang, J.; Wang, X.; Song, X.; Yang, C.; Zhang, X. and Wang, G.** (2012). The application of bioflocs technology in high-intensive, zero exchange farming systems of *Marsupenaeus japonicus*. *Aquaculture*, **354-355**: 97-106.
- Zhao, P.; Wu, C.C.; Limbu, S.M.; Li, R.; Chen, L.; Qiao, F.; Luo, Y.; Zhang, M.; Han, T. and Du, Z.** (2022). More simple more worse: Simple carbohydrate diets cause alterations in glucose and lipid metabolism in Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, **550**: <https://doi.org/10.1016/j.aquaculture.2021.737857>