

Postpartum Fertility in Relation to some Antioxidant Activity and Hormonal Profile in Buffaloes

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ABSTRACT

A total of 30 normally calved buffaloes, on day 14 postpartum, were used to study the relationship between fertility status and each of antioxidant activity, follicles (≥ 5 mm in diameter) populations and hormonal profiles. The reproductive indices in terms of interval to first heat and first service, Days open and services per conception were calculated. The buffaloes were classified into two groups on the basis of Days open whether it was ≤ 90 days (G1, n=20) or from 91 to 120 (G2, n=10). Ultrasonographic (US) examination was conducted on Day 21 postpartum to check cervical and uterine gravid horn diameter as well as every 3 days to estimate follicles populations (≥ 5 m) during two intervals: from 14 to 20 day and from 21 to 29day postpartum. Blood samples were collected on the Day of the first service for estimating serum concentration of antioxidant activity, Zinc, Selenium and estradiol and on Day 10 of the same cycle to assay serum progesterone concentration. While there was an increase ($P<0.01$) in the follicles populations on both ipsi- and contra- lateral ovaries in G1 compared with G2 during the interval: 14-20 day, there was an increase in G2 compared with G1 during the interval 21-29. The anti-oxidant activity and Malonaldehyde as well as Zinc and Selenium serum concentrations were higher ($P<0.01$) in G1 compared with G2. Also serum estradiol concentration on the Day of first service and progesterone on Day 10 of the same cycle were higher in G1 compared with G2. It is concluded that estimating follicular populations beyond Day14 postpartum and antioxidant activity on the day of the first service are predictive for postpartum fertility in buffaloes

Keywords: fertility, antioxidant activity, hormonal profile, buffaloes, postpartum.

INTRODUCTION

Reproductive function is considered a major factor determining the economic importance of buffalo cows (Barile, 2005). Fertility in dairy cows is defined as the ability to ovulate a competent oocyte and provides an oviductal and uterine environment capable of fertilization and complete embryonic and fetal development (Pursley *et al.*, 1997). Reproductive inefficiency in buffaloes such as delayed first service, silent heat, poor conception rates or ovarian inactivity is a major problem facing buffalo production, especially in animals kept in small holder farms (Sharma, 2003 and Ahmed, 2006). Managemental, nutritional and pathological factors are considered the main causes of the reproductive inefficiency (Campanile *et al.*, 1997). Blood profiles might be a potential aid in characterizing the problem. The blood components are mirror, which reflects the healthy condition of animals. So, the biochemical studies are very important for clinicians in the field during interpretation of their findings (Nebel and Jobst, 1998). Biochemical metabolites concentrations in the follicular fluid of the bovine ovary fluctuate considerably with the stage of cycle and follicle size (Tabatabaei and Mamoei 2010). Oxidative stress has a negative effect on animal performance which causes tissue damage and changes in the female biology (Megahed *et al.* 2002). Trace elements as selenium and zinc are among the antioxidants which can positively affect fertility in cattle. Decreased blood concentrations of these elements was associated with infertility in heifers (Baldi *et al.* 2000). Nishimoto *et al.* (2009) found a highly significant positive correlation between the concentration of estradiol and follicular size in follicles of the healthy cattle. Abe *et al.* (2002) reported that high progesterone concentration in the follicular fluid was predictive of subsequent implantation and pregnancy. Also, for thyroid hormones, Suriyathaporn (2000) mentioned that

thyroid hormones are part of the complex hormonal mechanism that regulates steroidogenesis in the ovary.

Controlled and regulated estrous cycle is done by many hormones which affected follicular growth and corpus luteum development and maintenance. Follicle stimulating hormone, luteinizing hormone, estradiol and progesterone are the hormones most commonly mentioned when discussing the estrous cycle. However, there are other substances that play an important role in controlling the estrous cycle in either an autocrine or paracrine type within the ovary (Borromeo *et al.*, 1996). Enzymes such as hydroxyl steroid dehydrogenase (3β - HSD) plays a central role in the biosynthesis of steroid hormone, including androgen inter (Penning, 1997). It catalyzes the final step in biosynthesis of progesterone (P4) in the ovary. Increasing level of P4 during the the ovulatory follicle growth is a key factor to improve fertility rate of lactating dairy cows, hence cows with greater P4 level have greater probability to be pregnant.

The goal of the present study was to test if there is a relationship between Antioxidant activity, follicle populations and hormonal profile in relation to fertility of Egyptian buffaloes.

MATERIALS AND METHODS

This study was carried out at Mehallet Mousa Experimental station belonging to Animal Production Research Institute (APRI), Agricultural Research Center, located in the North Nile Delta, Kafr el sheikh Governorate, Egypt.

Animals.

A total of 30 multiparous Egyptian lactating buffaloes (4-5 lactations), with live body weight ranged between 440 and 600 kg, were enrolled in this study. Body condition score (BCS) was recorded for all buffaloes (2.5-4.5) based on a scale from 1 (emaciated) to 5 (very fat) according to Edmonson *et al.* (1989). Buffaloes were in postpartum period at approximately 14 days after parturition. The buffaloes calved normally under veterinary supervision. They had neither placental

retention nor postpartum infection. Also, they suffered no metabolic disorders during postpartum period. They were housed in semi shaded open pens. They were fed to meet both maintenance and milk production requirements according to APRI (1997). The ration consisted of concentrate feed mixture (CFM), berseem hay (BH), corn silage and rice straw (RS) animals were fed twice a day. They had free access to water and milked twice daily.

Reproductive management:

Assessment of uterine involution:

During postpartum period, the reproductive tract was examined by rectal palpation and trans rectal ultrasonographic (US) scanning on Day 21 postpartum to check cervical and uterine gravid horn diameter as well as once every three days to estimate follicles populations (≥ 5 mm) during two intervals: from 14 to 20 day and from 21 to 29 day postpartum.

Heat detection:

Buffaloes were observed twice daily at an intervals of 12 hrs. by an experienced herd man. Buffaloes stand to be mounted by the teaser bull were considered in estrous. The interval to first estrous was calculated by subtracting day of parturition (Day 0) from the day of estrous.

Breeding:

The buffaloes observed in estrous during service period \geq Day 45 postpartum were taken to fertile bull to be served (hand mate breeding) according to morning/evening system of breeding. These buffaloes were re-observed for estrous signs on Day 21-23 post-breeding. Buffaloes return to estrous had to be bred for at least ≤ 3 times. The interval to 1st service was calculated by subtracting the day of calving from day of first service.

Pregnancy diagnosis:

The non-return buffaloes were examined by trans-rectal ultrasonography on Days 28-30 post-breeding. Conception rate was calculated by dividing conceived buffaloes on total number of naturally-bred buffaloes and multiplied by 100. Days open was calculated by subtracting day of last calving from Day of fertile service. The pregnant buffaloes were classified into two groups: G1 in which buffaloes conceived at ≤ 90 days postpartum and G2 in which buffaloes conceived during the interval between Days 91-120 postpartum.

Detection of follicles (≥ 5 mm) populations:

The follicles populations in terms of number of follicles ≥ 5 mm on both ovaries (ipsi- and contra- lateral to pregravid horn) were detected by transrectal ultrasonographic scanning. The ultra-sonographic examination was done every 3 days starting from Day 14 to 29 postpartum. The means \pm SE of number of follicles ≥ 5 mm counted in the first 3 examinations done on Days 14, 17 and 20 (interval from 14-20 day) were compared with their analogues detected on Days 23, 26 and 29 (interval from 21-29 day postpartum). All ultrasonographic examinations were done using ULTRASCAN 900 using multi-frequency, 6-8 MHz, linear transducer by the same examiner.

Blood sampling and assays:

blood sampling

Blood samples were collected on the day of estrous of the first estrous cycle during service period to assay serum concentrations of total antioxidant capacity, reduced glutathione, malondialdehyde, zinc, selenium, triiodothyronine (T3), thyroxine (T4) and estradiol (E2). Also blood samples were taken on Day 10 of the same cycle to measure serum progesterone concentrations. Ten ml of blood was drawn from the jugular vein of each animal in clean vacutainer tube without anticoagulant, centrifuged at 3000 rpm for 10 minutes. The serum was stored at -20 C until analysis.

Assays:

The antioxidant parameters (reduced glutathione, total antioxidant capacity and Malondialdehyde); trace elements (zinc and selenium) and hormones (triiodothyronin, thyroxin, estradiol and progesterone) profiles were compared between buffaloes which conceived at day ≤ 90 and those conceived in the period between Days 91 to 120 postpartum.

Estimation of antioxidant capacity:

Total antioxidant capacity (TAC) was measured using calorimetric method according to Koracevic *et al.* (2001), while glutathione (GSH) and malondialdehyde (MAD) concentrations were estimated according to Yoshida *et al.* (2005).

Estimation of zinc and selenium.

Trace elements including selenium (Se) and zinc (Zn) serum concentrations were determined using atomic absorption spectrophotometer as outlined by Varley *et al.* (1980).

Hormonal assays.

Direct radioimmunoassay technique was performed for the determination of thyroid hormones (triiodothyronine and thyroxine) concentrations, progesterone, (P4) and estradiol 17 β hormones in representative serum samples. Kits of "Diagnostic Products Corporation, (DCP) Los Angeles, USA" with ready antibody coated tubes were used according to the procedures outlined by the manufacturer.

Statistical analysis:

The Data were statistically analyzed using T-test within SPSS (2008) program.

RESULTS AND DISCUSSION

Reproductive performance:

Twenty out of 30 (CR, 67%) buffaloes conceived on Day ≤ 90 postpartum (G1) and the remaining 10 buffaloes (CR, 33%) conceived throughout the period extending from day 91 to 120 postpartum (G2). It was noted that while there was no difference in the interval to first estrous between G1 and G2, there was difference ($p < 0.05$) in the interval to first service between two groups. Also there were highly significant ($P < 0.01$) increase in both number of services per conception (NSC) and days open (DO) in G2 compared with G1 (Table, 1).

Table 1. Indices of postpartum reproductive performance in buffaloes.

Reproductive parameter	G1(n = 20)	G2 (n = 10)	Significant
Interval from calving to first estrus (days)	31.60±0.86	33.10±0.98	NS
Interval from calving to first service (days)	55.20±0.97	61.00±1.96	**
Number services per conception	1.35±0.10	2.80±0.25	**
Days open (days)	63.45±3.24	104.10±3.92	**

NS: Not significant. * Significant at P<0.05. ** high Significant P < 0.01.

In buffaloes, Hassan *et al.* (2013) found that NSC and DO values in pregnant animals having no dominant follicles on the ovary ipsilateral to the previous gravid uterine horn, were higher (P<0.01) than those having dominant follicles counter parts. Moreover, the pregnant cows with high genetic merits showed longer DO and NSC, even though with an earlier recovery of cyclicity. Also, Khasatiya (2005) reported that values of 1st postpartum estrus interval, service period length and calving interval were significantly lower in fertile than in infertile animals of Surti buffaloes.

postpartum follicles(≥ 5 mm) populations

During the ultrasonographic examination period of 14 to 20 days postpartum, the number of follicles (≥ 5

mm) in either ipsi- and contra- lateral ovaries was higher in G1 compared with G2. On the other hand, the follicle (≥ 5 mm) populations, on either ipsi- and contra- lateral ovary, were higher in G2 compared with G1 during the period from 21-29 days. However there were significant differences in the follicle populations (P < 0.01) between ipsi- and contra- lateral ovaries in either G1 or G2 during either 14-20 or 21-29 examination period (Table, 2). It was observed that while no significant differences was recorded in cervical diameter between G1 and G2 on Day 21 postpartum, there was a highly significant difference (P<0.01) in the uterine gravid horn diameter between two groups.

Table 2. follicle (≥ 5 mm) populations, cervical and uterine diameters on Day 21 postpartum in G1 and G2

Parameter	G1(n = 20)		G2(n = 10)		Sign.
	Ipsilateral Ovary	Contralateral ovary	Ipsilateral Ovary	Contralateral ovary	
No. of follicles ≥ 5 mm at 14 - 20 days p.p.	0.91±0.06 ^a	2.51±0.09 ^a	0.72±0.07 ^b	1.99±0.13 ^b	**
No. of follicles ≥ 5 mm at 21 - 29 days p.p.	1.29±0.06 ^b	2.13±0.07 ^b	1.30±0.10 ^a	2.40±0.14 ^a	**
Cervical diameter (cm)	3.58±0.10		3.61±0.17		NS
Diameter of uterine gravid horn (cm)	3.82±0.08		6.44±0.22		**

NS: Not significant. ** high Significant P < 0.01.

In buffaloes, Hassan *et al.* (2013) found that number of follicles with diameter of ≥ 5 mm on the ovary ipsilateral was leaser than on ovary contralateral to the pregnant uterine horn durin the interval from 14 to 28 day-postpartum in both pregnant and non-pregnant animals. In dairy cattle, Hussein (2005) recorded similar results during fourth weeks postpartum. Usmani (1992) mentioned that follicular activity started 6 days earlier in the ovaries contralateral to the gravid horn (21 days) than in the ipsilateral (27 days) ovaries and was higher in the same ovaries during the first 35 postpartum days (Singh *et al.* 1979). This results may be attributed to the influence of the follicle on the uterine endometrium and/or myometrium. Sheldon *et al.* (2003) mentioned that uterine function can be greatly affected by estradiol synthesized by a follicle ≥ 10 mm diameter. Also, Ireland *et al.*, (1984) found that concentrations of plasma estradiol are greater within the utero- ovarian vein draining the ovary containing the ovulatory follicle.

Similarly in buffaloes, Hassan *et al.* (2013) found insignificant differences in the cervical diameter of involuted cervix between conceived and non-conceived animals. The involution of the cervix completed at earlier interval than involution of the uterus in both groups, but the differences were not significant. Similar results were obtained Shah *et al.* (2004) indicating complete involution of the cervix on 24-39 days postpartum. Also, Hassan *et al.* (2013)

found that the difference between diameter of pregravid uterine horn in both groups was significant (P<0.05).

Antioxidant capacity of buffaloes plasma in relation to estrous cycle:

On day 0 (day of 1st service), antioxidants activity, in term of concentration of glutathione reduced, total antioxidants and concentration of malondialdehyde (MDA), were significantly (P<0.01) higher in G1 than G2 (Table 3). Glutathione reduced is an antioxidant, preventing to important cellular components damage, caused by reactive oxygen species (ROS) such as free radicals and peroxides (Sharma *et al.*, 2011). Increase plasma glutathione reduced might may lead to embryonic cells protection from oxidative stress, which is considered as one of the major reasons for improving rate of conception in pregnant cows (Guerin *et al.*, 2001). Also, Prabaharan *et al.*, (2015) observed higher concentration of reduced glutathione in pregnant than in non-pregnant cows during the interval from day 0 to day 20 and the mean concentration of serum MDA was lower in pregnant and non-pregnant cows in group i.m. injected with vitamin A, C and E for 3 weeks than in control group.

The MDA is a by-product of lipid peroxidation used as an index of the rate of tissue reaction chain. In addition, MDA is used as an indicator of cellular oxidative stress. Prabaharan *et al.*, (2015) reported lower concentration of serum MDA in pregnant than in non-pregnant ones. It was reported that MDA promotes

cross linking bonds in the membranes of the body, leading to changes in permeability of ions and activity of enzymes (Ahmed *et al.*, 2010).

In mare, Abo-El Maaty and El-Shahat (2012) found that total antioxidant capacity activity was significantly higher ($P < 0.05$) in serum obtained from luteal phase than those obtained at estrous one. On the other hand, Castillo *et al.*, (2003) recorded high malondialdehyde concentrations at estrus. They attributed the high level to an increase in the production of reactive oxygen species, which suggested to

originate mainly from steroidogenesis occurring in granulosa cells.

In both goats and sheep, Nawito *et al.*, (2016) found that malondialdehyde level was significantly higher ($P < 0.01$) in pregnant than in non-pregnant group. But, total antioxidant capacity level was lower in pregnant compared with non-pregnant goats. Also, Amer *et al.* (2014) monitored a decreased level of total antioxidant capacity in pregnant ewes that fed traditional or untraditional diets.

Table 3. Blood profiles of antioxidant activity; zinc, selenium and estradiol on the Day of first service and progesterone on Day 10 post breeding in G1 and G2.

Parameter	G1 (n=20)		G2 (n=10)		Sig.
	Estrous cycle day		Estrous cycle day		
	(day 0)	Day 10	(day 0)	Day 10	
Glutathione reduced (mmol/L)	5.66±0.18		4.71±0.30		**
Total antioxidant capacity (mmol/L)	1.64±0.05		0.76±0.05		**
Malondialdehyde (mmol/ml)	1.69±0.07		0.81±0.043		**
Zinc (µg/dl)	131.17±1.35		121.35±1.14		**
Selenium (µg/dl)	136.65±1.47		122.55±1.29		**
Ttriiodothyronine (ng/ml)	95.57±1.87		84.74±9.69		NS
Thyroxine (µg/dl)	3.37±0.16		2.82±0.13		**
Estrogen (ng/ml)	16.47±0.43		20.76±0.55		**
Progesterone (ng/ml)		5.29±0.21		1.49±0.13	**

GSH.R: Glutathione reduced; TAC: Total antioxidant capacity; MDA: Malondialdehyde; Se : Selenium; T3: Ttriiodothyronine; T4: Thyroxine ;P4: Progesterone; ES17: estrogen. NS: Not significant. * Significant at $P < 0.05$. ** high Significant $P < 0.01$.

Concentration of some trace elements in relation to estrous:

Table 3, shows decreased ($P < 0.01$) concentrations of trace element in blood serum of G2 in comparing with G1. In this respect, Graham (1991) reported that Zn or Se deficiency can cause decreased production, especially when a deficiency corresponds to the phases of growth, reproduction, or lactation. Animals having Zn deficiency are usually suffering from infertility (Shuttle, 1986). However, Du Plessis *et al.*, (1999) found small ovaries and decreased ovarian response to FSH-induced superovulation regimen in the molybdenum-induced trace elements depletion. The later may have central effects via the hypothalamus-pituitary axis on LH secretion leading to reduction in ovarian estradiol secretion and acyclicity of animals. Furthermore, LH-induced differentiation of bovine theca cells *in vitro* can be prevented by thiomolybdates and these effects can be ameliorated by trace elements supplementation. In this way, trace elements responsive sub-fertility may result from perturbation of the normal pattern of ovarian steroidogenesis (Kendall *et al.*, 2006). In buffaloes, Alavi-Shoushtari *et al.*, (2015) found that level of Zn in cyclic animals was higher than in acyclic ones. The lowest serum Zn content was observed in pro-and met-estrous, suggesting Zn levels were actively secreted in uterine lumen during the estrous cycle and were not dependent on blood serum. In Nili-Ravi buffaloes, Akhtar *et al.* (2009) showed that serum zinc and selenium levels were significantly lower in anestrus buffaloes compared with cyclic ones, The deficiencies of zinc or/and selenium could be responsible for anestrus condition. Decreasing level of zinc was associated with decreasing steroid hormone concentrations, indicating some correlation between

plasma zinc levels and P4 and estrogen levels for proper reproductive processes. The Se deficiency is also associated with reduced fertility in cattle and sheep (Hidiroglou, 1979) and high Se concentration reduces the incidence of anestrus (Harrison *et al.*, 1984).

Hormonal profile:

Regarding thyroid hormones, it has been observed that while they thyroxine (T4) showed significant increase in G1 compared with G2 there was non-significant variation in G1 compared with G2 on the Day of the first service. Thyroid hormones are part of the complex hormonal mechanism that regulates steroidogenesis in the ovary and Low triiodothyronine concentrations were associated with lower concentrations of estradiol and diminished estrus expression (Jorritsma *et al.* 2003). Deranged thyroid activity is a potential cause for reproductive abnormalities in the females of farm animals (Aggarwal and Singh, 2010). In fact, thyroid hormones have direct stimulatory effects on ovarian function in cattle, acting at the level of granulosa and thecal cells (Mutinati *et al.* 2010).

Furthermore, thyroid hormones play an important role in the pre and post-implantation embryo development in cattle (Ashkar *et al.* 2010). Induced hypothyroidism was known to reduce the fertilization rate in dairy cattle (Bernal *et al.* 1996). Ghuman *et al.*, (2011) found that a subtle thyroid activity in lactating buffaloes may have some impacts on their fertility status. They added that non-cycling and non-pregnant lactating buffaloes showed lower ($P < 0.05$) plasma concentrations of thyroid stimulating hormone and thyroid hormones, respectively, compared to their counterparts. The serum concentrations of estradiol on the day of the first service and progesterone on the 10 of the same cycle were significantly ($P < 0.01$) higher in G1

compared with G2 (Table, 3). The higher estradiol level in G1 may be attributed to higher follicles population on both ipsi- and contra- lateral ovaries during 14-20 days postpartum. Although follicles populations during this period had no direct effect on the serum E2 level on the day of first service but they are considered good indicators of ovarian activity especially steroidogenic one in G1. However high estrogen levels at the time of the first service in G1 had a positive effect on the conception rate (67%). This may be explained in the light of the fact that high E2 level at the Day of estrus (estradiol surge) induces high LH surge with subsequent induction of ovulation. Also, the higher progesterone level on Day 10 post-service in G1 favors the higher conception rate in this group.

These results are in agreement with earlier reports in buffaloes (Pahwa and Pandey, 1983) in terms of minimum level on the day of estrus with gradual rise to the higher levels during luteal phase and then declining to basal level at subsequent estrus. Sankar and Prakash (2008) showed that progesterone concentration was reached the basal level during the peri-oestrus period. The plasma progesterone concentration was the lowest on the day of oestrus and increased to register a peak on day 13 of the cycle. In Holstein cattle, Tabatabaei *et al.*, (2014) showed that the plasma progesterone concentration in late diestrus was higher than in metestrus, proestrus and during estrous ($P < 0.05$), but the plasma concentrations of estradiol-17- β in proestrus and estrous were higher than the other stages of the cycle ($P < 0.05$). Also, in goats, Pang *et al.*, (2010) found that progesterone concentrations began to rise earlier and were higher in the high prolificacy than in the poor prolificacy goats on most days of the luteal phase, but not during the follicular phase of the cycle or after ovariectomy.

CONCLUSION

It is concluded that estimating follicular populations beyond Day 14 postpartum and antioxidant activity on the day of the first service are predictive for postpartum fertility in buffaloes.

REFERENCES

- Abe, H., Yamashita, S., Satoh, T. and Hoshi, H. (2002). Accumulation of cytoplasmatic lipid droplets in bovine embryos and cryotolerance of embryos developed in different culture systems using serum-free or serum-containing media. *Mol. Reprod. Dev.*, 61 :57-66.
- Abo-El maaty, A. M. and El-Shahat, K. H. (2012). Hormonal and biochemical serum assay in relation to the estrous cycle and follicular growth in Arabian mare. *Asian Pacific J. of Rep.* 1(2): 105-110.
- Aggarwal, A. and Singh, M. (2010). Hormonal changes in heat-stressed Murrah buffaloes under two different cooling systems. *Buffalo Bull.* 29, 1-6.
- Ahmed, W.M. (2006). Adverse conditions affecting ovarian activity in large farm animals. *Proceeding of the 3rd International Conference of Veterinary Research Division, National Research Centre, Cairo, Egypt*, 251-253.
- Ahmed, W.M.; El-Khadrawy, H.H.; Abd El Hameed, A.R. and Amer, H.A. (2010). Applied investigations on ovarian inactivity in buffalo heifers. *Inter J. Acad. Res.*, 2: 26- 32.
- Akhtar, M.S.; Farooq, A.A. and Mushtaq, M. (2009). Serum concentrations of copper, iron, zinc and selenium in cyclic and anoestrous Nili-Ravi buffaloes kept under farm conditions. *Pakst. Vet. J.* 29 (1): 47-48.
- Alavi-Shoushtari, S.M.; Asri-Rezai, S.; Amir Khaki; Abulfazl Belbasi; and Hamid Tahmasebian (2015). Copper and zinc concentrations in uterine fluid and blood serum during the estrous cycle and pre-pubertal phase in water buffaloes *Vet. Res. Forum.* 6 (3) 211 – 215.
- Amer, H.; Ibrahim, N.H.; Donia, G.R.; Younis, F.E. and Shaker, Y.M. (2014). Scrutinizing of trace elements and antioxidant enzymes changes in Barki ewes fed salt-tolerant plants under South Sinai conditions. *J. Am. Sci.*, 10(2):241-249.
- APRI. 1997. Monthly report on the herd production and reproduction status of APRI buffalo breeding experimental stations, tune 1997, (personal communication).
- Ashkar, F.A.; Semple, E.; Schmidt, C.H.; St John, E.; Bartlewski, P.M. and King, W.A. (2010). Thyroid hormone supplementation improves bovine embryo development in vitro. *Hum. Reprod.* 25, 334-344.
- Baldi, A., L. Savoini, E. Pinoti, F. Chei and V. Dello-Orto, (2000). Effect of vitamin E and different energy sources on vitamin E status, milk quality and reproduction in transition dairy cows. *J. Vet. Med.*, 47: 599-608.
- Barile, V.L. (2005). Improving reproductive efficiency in female buffaloes. *Livestock Prod. Sci.*, Vol. 92(3): 183–194.
- Bernal, A.; DeMoraes, G.V.; Thrift, T.A.; Willard, C.C.; Keisler, R.D. and Lucy, M.C. (1996). Perception and interpretation of the effects of under nutrition on reproduction. *J. Anim. Sci.* 74, 1-17.
- Borromeo, V.; Bramani, S.; Berrini, A.; Sironi, G.; Finazzi, M.; Cremonesi, F. and Secchi, C. (1996). Growth hormone but not prolactin concentrations in the fluid of bovine ovarian cysts are related to the cystic stage of luteinization. *Theriogenology.* 46:481–489.
- Campanile, G.; Di Palo, R. and D'Angelo, A. (1997). Profilometabolicone buffalo. *Bubalus Bubalis (Suppl. 4):*236-249.
- Castillo, C.; Hernández, J.; López-Alonso, M.; Miranda, M. and Benedito, J.L. (2003). Values of plasma lipid hydroperoxides and total antioxidant status in healthy dairy cows: Preliminary observations. *Arch Tierz.* 46(3): 227-233.

- Du Plessis, S.S.; Van Niekerk, F.E. and Coetzer, W.A. (1999). The effect of dietary molybdenum and sulphate on the sexual activity and plasma progesterone concentrations of ewes. *Small Rumi. Res.*, 33: 71-76.
- Edmonson, A.J.; Lean, I.J.; Weaver, L.D.; Farver, T. and Webster, G. (1989). A body condition scoring chart for Holstein dairy cows. *J. of Dairy Res.*, 72, 68- 78.
- El-Shahat, K.H. and Kandil, M. (2012). Antioxidant capacity of follicular fluid in relation to follicular size and stage of estrous cycle in buffaloes. 77 (8) 1513–1518.
- Ghuman, J.; Singh, M.; Honparkhe, C.S.; Ahuja, D.S.; Dharmi, G.; Nazir and Gandotra, V.K. (2011). Differential fertility in dairy buffalo: Role of thyroid and blood plasma biochemical milieu. *Iranian J. appl. Anim. Sci.*, 1(2), 105-109.
- Graham, T.W. (1991). Trace element deficiencies in cattle. *Vet.*, 7(1): 153-215.
- Guerin, P.; Mouatassim, E.I.S. and Menezo, Y. (2001). Oxidative stress and protection against reactive oxygen species in the pre-implantation embryo and its surroundings. *Human Reprod. Update*, 7: 175-189.
- Harrison, J. H.; Hancock, D. D. and Conrad, H. R. (1984). Vitamin E and selenium for reproduction of the dairy cow. *J. Dairy Sci.*, 67: 123-132.
- Hassan, A. H.; Waleed, S. and Mahmoud, R. A. (2013). Relationship among uterine involution, ovarian activity, blood metabolites and subsequent reproductive performance in Egyptian buffaloes. *Open J. of Anim. Sci.*, 3(1), 59-69.
- Hidiroglou, M. (1979). Trace element deficiencies and fertility in ruminants: A review. *J. Dairy Sci.*, 62: 1195-1206.
- Hussein, H.A. (2005). Ultrasonographic evaluation for the effect of postpartal ovarian activity and uterine involution on subsequent reproductive performance of dairy cows. *Assiut Vet. Med. J.* 51, 262-272.
- Ireland, J.J.; Fogwell, R.L.; Oxender, W.D.; Ames, K. and Cowley, J.L. (1984). Production of estradiol by each ovary during the estrus cycle of cows. *J. of Anim. Sci.*, 59, 764-771.
- Jorritsma, R.; Wensing, T.; Kruij, T.; Vosa, P. and Noordhuizen, J. (2003). Metabolic changes in early lactation and impaired re-productive performance in dairy cows. *Vet. Res.* 34, 11-26.
- Kendall, N.R.; Marsters, P.; Guo, L.; Scaramuzzi, R.J. and Campbell, B.K. (2006). Effect of copper and thiomolybdates on bovine theca cell differentiation in vitro. *J. Endocrinol.*, 189: 455-463.
- Khasatiya, C.T. (2005). Reproductive traits and metabolic profile in postpartum fertile and infertile surti buffaloes. *Ind. Vet. J.* 82, 637-641.
- Koracevic, D. and Koracevic, G. (2001). Colorimetric method for determination of total antioxidant capacity. *J. Clin. Pathol.* 54, 356-361.
- Megahed, G.A.; Anwar, M.M. and El Ballal, S.S. (2002). Superoxide dismutase, nitric oxide and lipid peroxide productions and its relation to apoptotic changes and serum progesterone levels during physiological life span of buffalo corpora lutea. *Minufyia Vet. Med. J.* 2: 99-112.
- Mutinati, M.; Desantis, S.; Rizzo, A.; Zizza, S.; Ventriglia, G.; Pantaleo, M. and Sciorsci, R.L. (2010). Localization of thyrotropin receptor and thyroglobulin in the bovine corpus luteum. *Anim. Reprod. Sci.* 118, 1-6.
- Nawito, M.F.; Abd El Hameed, A.R.; Sosa, A.S.A. and Mahmoud, K.G.M. (2016). Impact of pregnancy and nutrition on oxidant/antioxidant balance in sheep and goats reared in South Sinai, Egypt. *Vet. World*, 9(8): 801-805.
- Nebel, R.L. and Jobst, S.M. (1998). Evaluation of systematic breeding programs for lactating dairy cows: a review. *J. Dairy Sci.*, 81(4): 1169-74.
- Nishimoto, S.; Glen, A.H.; Akio, M. and Safumi, T. (2009). Classification of Bovine follicles based on the concentration of steroid, glucose and lactate in follicular fluid and the status of accompanying follicles. *J. Reprod.*, 2: 55-61.
- Pahwa, G.S. and Pandey, R.S. (1983). Gonadal steroid hormone concentration in blood plasma and milk of primiparous and multiparous pregnant and non pregnant buffaloes. *Theriogenology*, 19: 491-505.
- Pang, X. S.; Wang, Z. Y.; Zhu, T. G.; Yin, D. Z.; Zhang, Y. L.; Meng, L. and Wang, F. (2010). Concentrations of Progesterone and Estradiol in Peripheral Plasma during the Estrous Cycle and after Ovariectomy in Huanghuai Goats of High or Poor Prolificacy. *Asian-Aust. J. Anim. Sci.*, 23(2) : 188 – 196.
- Penning, T.M. (1997). Molecular endocrinology of hydrosteroid dehydrogenases. *Endocr Rev.*, 18: 281-305.
- Prabaharan, V.; Selvaraju, M.; Napoleon, R.E.; Duraisamy, K.A. and Natarajan, A. (2015). Role of oxidative stress biomarkers on embryonic mortality in bovines. *Inter. J. Vet. Sci.* 4(1): 6-9.
- Pursley, J.R.; Wiltbank, M.C.; Stevenson, J.S.; Garverick, H.A. and Anderson, L.L. (1997). Pregnancy rates per artificial insemination for cows and heifers inseminated at a synchronized ovulation or synchronized estrus. *J. of Dairy Sci.*, 80: 295-300.
- Sankar, R. K. and Prakash, B. S. (2008). Plasma progesterone, oestradiol-17 β and total oestrogen profiles in relation to oestrous behaviour during induced ovulation in Murrah buffalo heifers. *J Anim. Physiol. Anim. Nutr.* 93(4): 486-495.
- Shah, R.G.; Dharmi, A.J.; Kharadi, V.B.; Desai, P.M. and Kavani, F.S. (2004). Hormonal profile in fertile and infertile postpartum Surti buffaloes. *Buffalo J.* 20, 29- 42.
- Sharma, N.; Singh, N.K.; Singh, O.P.; Pandey, V. and Verma, P.K. (2011). Oxidative stress and antioxidant status during transition period in dairy cows. *Asian-Aust J. Anim. Sci.*, 4: 479-484.

- Sharma, R.K. (2003). Reproductive problems of buffaloes and their management. Compendium of lectures delivered to the Cent. Institute of Research on Buffalo, Hisar, India, 119-126.
- Sheldon, I.M.; Noakes, D.E.; Rycroft, A.N. and Dobson, H. (2003). The effect of intrauterine administration of estradiol on postpartum uterine involution in cattle. *Theriogenology*, 59, 1357-1371.
- Shuttle, N.F. (1986). Copper deficiency in ruminants, recent developments. *Vet. Record*, 119: 519-522.
- Singh, N.; Chauhan, F.S. and Singh, M. (1979). Postpartum ovarian activity and fertility in buffaloes. *Ind. J. of Dairy Sci.*, 32, 134-139.
- SPSS (2008). Statistical Package for the social sciences. Release 16, SPSS INC, Chicago, USA.
- Suriyasathaporn, W. (2000). Negative energy balance in postpartum dairy cows: its effect on clinical mastitis and reproductive performance. Ph.D. Thesis, Utrecht University, Department of Farm Animal Health.
- Tabatabaei, S. and Mamoei, M. (2010). Biochemical composition of blood plasma and follicular fluid in relation to follicular size in buffalo, *Springer.*, 20 (5): 441.
- Tabatabaei, S.; AsghariMoghadam, M.; Mamoei, M.; Mirzadeh, K. and Aghaei, A. (2014). Hormonal profile of ovarian follicular fluid and blood plasma during different stages of estrous cycle in Holstein cattle. *Iranian J. appl. Anim.Sci.*, 4(2), 263-268.
- Usmani, R.H. (1992). Effect of non-gravid uterine horn on the pattern of resumption of ovarian functions in postpartum Nili Ravi buffaloes. *Buffalo J.* 8, 265-270.
- Varley, H.; Genlock, A.H. and Bell, M. (1980). *Practical Clinical Chemistry. Vol.1. General topics commen test.* 5th ed. William Heinemann Medical books Ltd, London, UK.
- Yoshida, Y.; Itoh, N.; Hayakawa, M.; Piga, R.; Cynshi, O.; Jishage, K, and Niki, E. (2005). Lipid peroxidation induced by carbon tetrachloride and its inhibition by antioxidant as evaluated by an oxidative stress marker, HODE. *Toxicol. Appl. Pharmacol*, 208: 87-97.

الخصوبة وعلاقتها بنشاط بعض مضادات الاكسدة والهرمونات بالدم في فترة ما بعد الولادة في الجاموس.
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اجريت هذه الدراسة على عدد 30 جاموسة بعد 14 يوم من الولادة وكانت ولادتهم طبيعية وذلك لدراسة العلاقة بين حالة الخصوبة وكل من نشاط مضادات الاكسدة وعدد الحويصلات التيزيد قطرها عن او يساوى 5 مم ومستوى بعض الهرمونات في الدم. تم حساب الفترة من الولادة حتى اول شياح و اول تلقيح والفترة من الولادة حتى اول تلقيحه مخصبة (Days open) وعدد التلقيحات اللازمة للأخصاب. تم تقسيم الحيوانات الى مجموعتين على اساس فترة (Days open) سواء كانت اقل من او تساوى 90 يوم (المجموعة الاولى وعددها 20 جاموسة G1) او كانت من 91 الى 120 يوم (المجموعة الثانية وعددها 10 جاموسة G2). تم فحص المبايض بجهاز السونار كل ثلاثة ايام لتقدير عدد الحويصلات المبيضية والتي يزيد قطرها عن أو يساوى 5مم خلال الفترتين: الفترة الاولى من اليوم 14 وحتى اليوم 21 والفترة الثانية من اليوم 21 وحتى اليوم 29 بعد الولادة. وتم قياس قطر عنق الرحم وقطر قرن الرحم الذي حدث به الحمل عند اليوم ال 21 بعد الولادة. تم اخذ عينات دم يوم الشياح من التلقيحه الاولى لتقدير تركيز مضادات الاكسدة والزنك والسيلينيوم والاستيراد يول وتم اخذ عينات الدم يوم 10 من نفس دورة الشياح لتقدير تركيز البروجيستيرون. وكانت النتائج كالاتي: لوحظ انه بالرغم من وجود فروق معنوية عالية ($P < 0.01$) في عدد الحويصلات المبيضية على كل من المبيضين (المقابل او الغير مقابل لقرن الرحم الذي حدث فيه الحمل) فيالمجموعة الاولى بالمقارنة بالمجموعة الثانية اثناء الفترة من 14 الى 20 يوم بعد الولادة، كان هناك زيادة في المجموعة الثانية عن المجموعة الاولى اثناء الفترة من اليوم 21 وحتى اليوم 29 من بعد الولادة. كان تركيز كل من مضادات الاكسدة وانزيم المألونالدهيد وايضا الزنك والسيلينيوم في سيرم الدم اعلى معنويا ($P < 0.01$) في المجموعة الاولى بالمقارنة بالمجموعة الثانية . وايضا تركيز كل منالاستيراد يول(عند يوم الشياح والتلقيح) والبروجيستيرون(عند اليوم 10 من نفس دورة الشياح) كان اعلى في المجموعة الاولى بالمقارنة بالمجموعة الثانية نستخلص من ذلك التقدير العد الحويصلي للبيوضات بعد اليوم 14 من بعد الولادة ومضادات الاكسدة عند يوم الشياح من التلقيحه الاولى ويعتبر ذلك دليلا تنبؤي للخصوبة في الجاموس بعد الولادة.