

Growth, as well as Leaf and Stem Anatomy in Periwinkle Plant as Affected by Certain Biotic and Abiotic Elicitors

Naglaa H. El- Sheikh; S. Farouk; Z. E. A. Mohamed and A. A. Arafa
Agriculture Botany Dept., Fac. of Agric., Mansoura Univ., 35516, El- Mansoura, Egypt.



ABSTRACT

Catharanthus roseus (L.) G. Don is a potential source of several pharmaceutically materials used in cancer treatment. There is a growing demand for its alkaloids that are biosynthesized naturally in a very low quantity. The current study aimed to enhance plant growth and improved internal structure by using certain elicitors. The elicitors tested were calcium chloride (CaCl_2) at 4.4 g/l^{-1} , sodium nitroprusside (SNP) at 0.149 g/l^{-1} , salicylic acid (SA) at 0.138 g/l^{-1} singly or in combination with yeast extract (YE) at either 5 or 10 g/l^{-1} . Pot experiments were done at the experimental farm of Agric. Bot. Dep., Mansoura Univ., Egypt; during the two growing seasons of 2015 and 2016. Results indicated that, tested elicitors positively affected on studied growth attributes, and the magnitude of effect was elicitor- dependent. Stem length as well as stem dry weight and number of leaves were increased in response to YE at 5 and 10 g/l^{-1} , respectively. Under the influence of abiotic elicitors, most growth parameters were increased due to SA and SNP. An additive effects were detected in the combination treatments between either SA or SNP and YE. Stem dry weight was highest in plants treated with SA in combination with YE at 5 g/l^{-1} . In addition, applied elicitors, generally increased proportions of the photosynthesizing tissues in the leaf and the conductive tissues in the stem. The data suggested to using a combined treatment of SA with YE at 5 g/l^{-1} to increase *C. roseus* biomass, consequently its yield from alkaloids.

Keywords: Anatomical studies; Calcium chloride; *Catharanthus roseus*; Elicitors; Salicylic acid; Sodium nitroprusside; Yeast extract.

INTRODUCTION

Periwinkle (*Catharanthus roseus* (L.) G. Don; Apocynaceae, is an important perennial tropical herb that produces over 100 monoterpenoid indole alkaloid including two commercially vital cytotoxic dimeric alkaloids used in cancer chemotherapy (Shukla *et al.*, 2012). Nevertheless, tons of its materials are required to extract small quantity of the alkaloid vinblastine (VLB) or vincristine (VCR) (Loyola-Vargas *et al.*, 2007). The latter authors added that, the cost of 1 kg of VLB in the market is around one million Dollars and the world annual production in near about 12 kg. Alternately, VCR cost about 3.5 million dollars per kg and its annual production is 1 kg. The low yield of indole alkaloids, consequently their high cost have stimulated numerous efforts to develop alternative strategies for their production (Ebrahimzadeh *et al.*, 1996). Accumulation of secondary metabolites in plant tissues can be enhanced by the treatments of various kind of elicitors (Angelova *et al.*, 2006; Dipti *et al.*, 2016). Elicitation classified as biotic and abiotic is a renewable alternative for secondary metabolite production (Maqsood *et al.*, 2017).

Yeast extract (YE) is an autolysate of the cell of baker's yeast (*Saccharomyces cerevisiae*). It has been recognized to be a rich resource of phytohormone, vitamins, enzymes, amino acids and nutrients (Amer, 2004). These constituents elicit plant defense responses by triggering metabolite biosynthesis, (Cai *et al.*, 2012), and enhance alkaloid assimilation in *C. roseus*, (Samar *et al.*, 2015; Dipti *et al.*, 2016). In addition, foliar YE improved growth, and productivity of several plants (Mohamed, Manal *et al.*, 2018; Xi *et al.*, 2019).

Exogenous application of salicylic acid (SA) enhanced *C. roseus* monomeric and dimeric alkaloids, (Khan *et al.*, 2007). SA that induced plant growth under normal or stressed conditions participates in the regulation of multiple physiological processes in plant (Ghassemi-Golezani *et al.*, 2017; Kareem *et al.*, 2017). Furthermore, Farouk *et al.*, (2018) found that salicylic acid enhance leaf area, plant dry mass and plant height in maize.

Sodium nitroprusside (SNP), is a tiny diffusible bioactive signaling molecule, plays multifunctional roles in plant develop and productivity under normal or stressed conditions (Ahmad *et al.*, 2018; Hanafy *et al.*, 2018). Treatment of SNP enhanced the production of terpenoid indole alkaloids of *C. roseus* cells (Xu *et al.*, 2005). SNP can mediate stomatal movement (Garcia-Mata and Lamattina, 2007), modulate the expression of cell cycle genes (Correa-Aragunde *et al.*, 2006), regulate plant maturation and senescence and the cytokinin signalling pathway (Mishina *et al.*, 2007).

Calcium (Ca^{+2}) has been occupied as a second messenger of plant responses to a diversity of external stimuli (Kiegle *et al.*, 2000). Supplementation with calcium improved root elongation (Kurth *et al.*, 1986) and shoot growth (Amer, 2004). Additionally, Ca^{+2} increased the build- up of *C. roseus* alkaloids in cell cultures (Zhao *et al.*, 2001).

The aim of the current study was to assess the enhancing effect of biotic elicitor (yeast extract 'YE' at 0, 5 or 10 g/l^{-1}), and abiotic elicitors (salicylic acid 'SA' at 0.138 g/l^{-1} , calcium chloride ' CaCl_2 ' at 4.4 g/l^{-1} , and sodium nitroprusside 'SNP' at 0.149 g/l^{-1} , beside water) as a foliar application on growth as well as internal structure of the stem and the leaf of *C. roseus*.

MATERIALS AND METHODS

Two pot experiments were done in the experimental farm and laboratories of the Agric. Bot. Dept., Faculty of Agric, Mansoura University, Egypt through the two (2015 and 2016) seasons. Seeds were obtained from the Horticulture Research Institute, ARC, Giza, Egypt, disinfected with 0.01% HgCl_2 solution for 3 minutes, washed with distilled water and sown on 14th March in both growing seasons at the rate of 20 seeds/ pot in plastic pots (25 cm inner diameter and 30 cm depth), each pot containing 6 kg air-dried clay loam soil. The physiochemical characteristics of the experimental soil were assessment following the method of Motsara and Roy (2008) and summarized in Table (1).

Table 1. The physiochemical characters of the experimental soil

Soil properties	1 st season	2 nd Season	Soil properties	1 st season (meq/L)	2 nd Season (meq/L)
Clay %	40.3	42.8	Cations	Calcium	1.2
Silt%	26.5	25.6		Magnesium	1.1
Fine Sand%	24.7	23.7		Sodium	2.1
Coarse Sand%	8.5	7.9		Potassium	0.2
Hygroscopic Water%	5.3	5.2	Anions	Carbonate	----
SSP%	56	58		Bicarbonate	1.3
EC dSm	0.54	0.56		Chloride	2.1
pH (1:1.5, soil: water)	7.53	7.45		Sulphate	1.2

Phosphorous as calcium superphosphate (15.5% P₂O₅) at the rate of 2 g/pot (333kg/fed), and 2 g/pot (333 kg/ fed) nitrogen as ammonium nitrate (33.5%N) and 1 g/pot (166 kg/ fed) potassium fertilizer as potassium sulphate (48% K₂O) were mixed into the upper 3 cm of the soil of all pots before planting. Pots were divided into three sets consisting of 20 pots for each, arranged in a randomized complete block design with five replicates. Plants were thinned to leave five plants/ pot at 30 days after sowing (DAS). Plant sprayed with YE, CaCl₂, SNP, and SA using hand pressure sprayer. Tween 20 (0.05%) was added as a wetting agent. Control plants were sprayed with tap water. Spraying was take place twice till dripping, the first one at 45 DAS, and whereas the second after two weeks from the first one.

Samples from each treatment were taken randomly after 75 DAS for morphological and anatomical characteristics. Root length and plant height (cm), root and shoot fresh and dry weight (g), numbers of branches and leaves per plant were recorded. For the anatomical studies specimens from the 3rd upper leaf and the 2nd upper internode were taken, killed and fixed for at least 48 hr.in F.A.A. (10 ml formalin, 5 ml glacial acetic acid, and 85 ethyl alcohol 70%). They were washed in 50% ethyl alcohol, dehydrated in a normal butyl alcohol series, embedded in paraffin wax (melting point 56 C°), sectioned by rotary microtome at a thickness of 20 µ, double stained with crystal violet- erythrosine, cleared in xylene and mounted in Canada balsam. Anatomical quantitative data were obtained using calibrate of eyepiece micrometer according to Gerlash (1977).

Statistical analysis:

Data were subjected to analysis of variance according to Norman and Streiner (2003), least significant differences (L.S.D) at 0.05, were used for comparison between the control and the other experimental treatments.

RESULTS AND DISCUSSION

Vegetative growth characters:

Treatment with YE at 5 g/l⁻¹ increased plant height and stem dry weight in both seasons whereas decreased number of branches during the second season. It had no significant effect on these parameters during the first season throughout both seasons. At 10 g/l⁻¹, YE increased leaf number during the first season whereas decreased branches number during the second season. Additionally it had no significant effect on plant height (Fig. 1)

Treatment with CaCl₂ increased stem fresh weight during the first season whereas decreased root length, numbers of leaves and branches during the second season. Other parameters were not significantly. Treatment with SNP increased leaf fresh and dry weight throughout the

experimental period as well as number of leaves, stem fresh weight and root fresh weight, only during the first season. However, SNP decreased number of branches, during the second season only.

The effect of SA on growth parameters was predominant with regard to leaf growth. Number of leaves, as well as leaf fresh and dry weight was also increased. Moreover, SA increased stem fresh weight, and stem length during the second season as well as root fresh and dry weight during the first and the second season, respectively.

The interaction between biotic and abiotic elicitors was significant for most of the studied growth parameters, only during the second season (Table 2, 3). In this case, stem length, number of leaves as well as branches, stem fresh weight, and leaf fresh weight were highest in SA treated plants with 5 g/l⁻¹ YE. In addition, the maximum stem dry weight was recorded when treated with SA and combined with YE at 5 g/l⁻¹. Positive interaction effect on growth attributes resulted from the interaction between SNP and YE. Stem fresh weight and leaf fresh weight were also increased due to SNP combined with YE at 5 and 10 g/l⁻¹, respectively. On the other hand, stem dry weight was notably increased in plants received the combination between CaCl₂ and YE at 10 g/l⁻¹.

Numerous investigations indicated that spraying plants with YE improved plant growth (Mohamed Manal *et al.*, 2018; Xi *et al.*, 2019). This enhancement may be attributed to ability of YE to increase the biosynthesis of plant growth regulators, specially, gibberellin that stimulate cell division and enlargement as well as synthesis of proteins, nucleic acid and chlorophyll (George *et al.*, 2008; Sarhan 2008). Likewise, Bevilacqua *et al.* (2008) demonstrated that yeast extract contains growth factors, free amino acids which enhance vegetative growth of plants.

Calcium (Ca⁺²) is an important nutrient that is extremely compartmentalized in plant tissues, and this partitioning is a defining constituents of its biochemical functions. In its structural role, calcium is a key component of the cell wall middle lamella where it helps to join adjacent cells together and strengthen general construction (Marschner, 1995). In addition Ca⁺² influences membrane function, stabilizes membranes and influences permeability by birding phosphate and carboxylate groups of membrane phospholipids and protein (Davies and Monk-Talbot, 1990).

SNP plays extremely vital roles in improving plant development of several plants by modulating a diversity of metabolic pathways including: (1) scavenging ROS through activating antioxidant system (Rai *et al.*, 2018); (2) acting as a signal molecule induces alteration in the expression of stress-responsive genes (Gill *et al.*, 2013); (3) inhibition of ethylene biosynthesis (Zhu *et al.*, 2008).

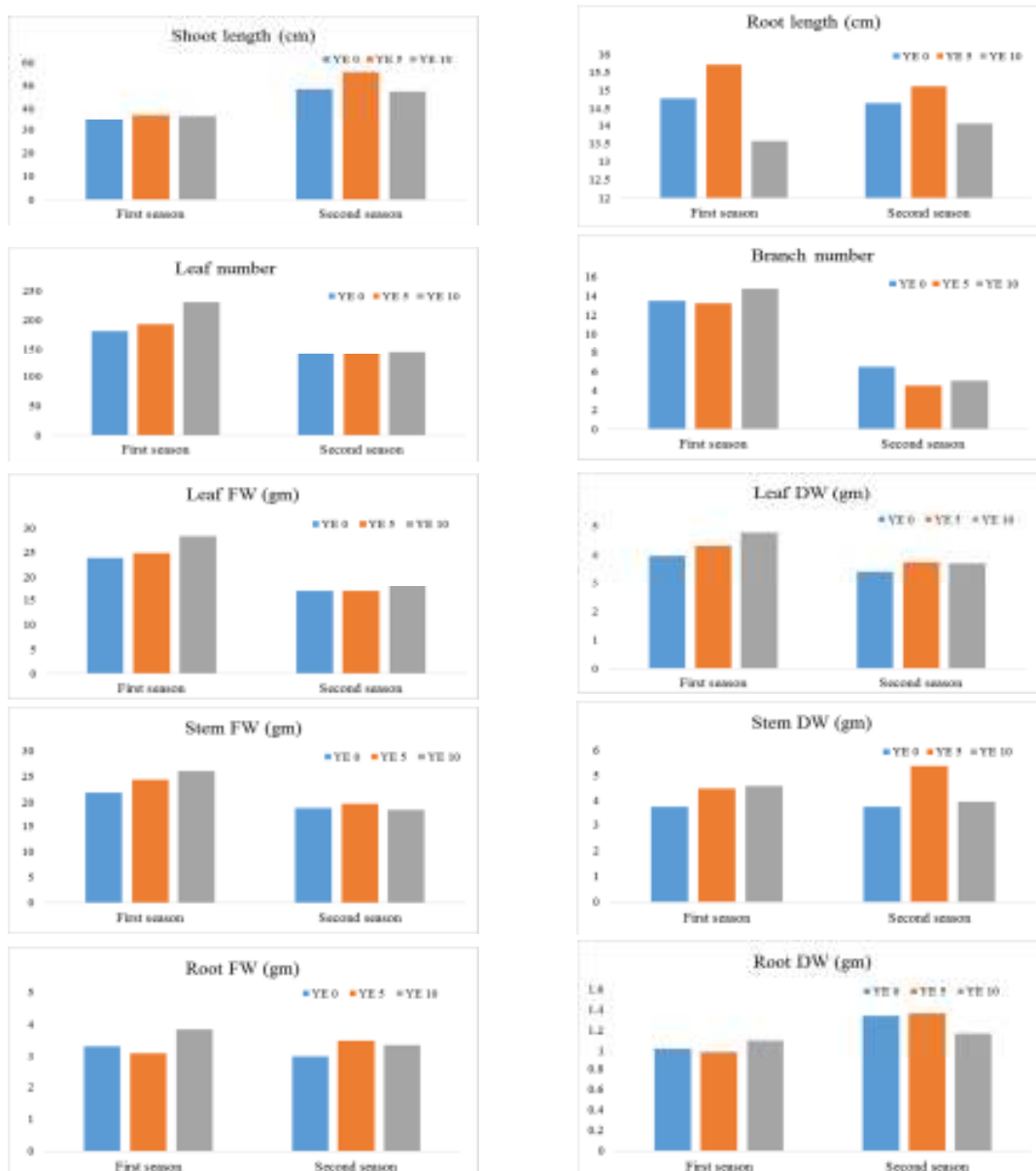


Fig. 1. Growth attributes of *C. roseus* as affected by yeast extract (YE) at either 5 or 10 g/ L⁻¹ at 75 days after sowing in both seasons.

Table 2. Plant height, root length as well as numbers of leaves and branches of *C. roseus* plants as affected by elicitors at 75 days after sowing during the two growing seasons (2015-2016).

Treatment g/l		Plant height (cm)		Root Length (cm)		No. of leaves		No. of branches	
		1 st season	2 nd season	1 st season	2 nd season	1 st season	2 nd season	1 st season	2 nd season
Interactions									
YE 0	W	29.3	39.3	12.3	14.3	102	115	10	8
	CaCl ₂	36.3	44.0	17.0	13.0	197	91	13	4
	SNP	38.0	51.0	14.3	17.3	208	128	14	6
	SA	36.6	59.3	16.3	14.0	219	237	19	8
YE 5	W	34.3	52.6	16.6	15.3	176	118	15	4
	CaCl ₂	36.0	51.3	14.0	12.6	166	69	12	4
	SNP	37.0	59.6	16.3	17.3	218	196	15	4
	SA	42.3	60.3	16.0	15.3	212	175	11	6
YE 10	W	37.3	51.0	13.0	15.6	164	207	12	7
	CaCl ₂	36.3	48.3	13.0	13.3	242	108	16	4
	SNP	38.0	42.6	13.0	13.3	251	113	14	4
	SA	35.6	48.3	15.3	14.0	268	156	17	5
LSD at 5%	Ns	8.51	Ns	Ns	Ns	35.5	Ns	Ns	2.157

W=water, CaCl₂= calcium chloride, SNP= sodium nitroprusside, SA= salicylic acid, YE= yeast extract.

Table 3. Fresh and dry weight of *C. roseus* plant organs as affected by used elicitors at 75 days after sowing during the two growing seasons (2015-2016).

Treatment g/l		Leaf FW(g)		Stem FW(g)		Root FW(g)		Leaf DW(g)		Stem DW(g)		Root DW(g)	
		1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd	1 st	2 nd
		season	season	season	season	season	season	season	season	season	season	season	season
Interactions:													
YE 0	W	11.43	10.81	8.20	13.22	1.73	3.47	2.143	2.68	1.500	2.773	0.700	1.073
	CaCl ₂	22.73	11.80	23.46	12.84	3.71	1.93	3.643	2.507	4.123	2.880	1.073	1.010
	SNP	28.07	18.32	22.49	11.98	3.92	2.79	4.930	4.083	4.207	4.190	1.193	1.673
	SA	33.40	26.83	33.58	37.80	3.89	3.81	5.163	4.397	5.170	5.097	1.090	1.603
YE 5	W	19.99	15.55	16.23	16.06	2.92	3.06	3.747	3.310	3.900	3.810	0.967	1.100
	CaCl ₂	24.32	14.76	23.05	17.87	3.00	3.17	4.173	2.780	3.923	3.970	1.030	1.263
	SNP	27.88	18.73	29.30	22.47	3.45	3.82	4.650	4.683	4.883	6.650	0.877	1.050
	SA	27.26	18.29	29.12	22.42	3.05	3.92	4.667	4.18	5.177	7.057	1.060	2.047
YE 10	W	24.27	14.09	24.73	17.41	3.47	3.64	3.833	3.810	4.920	4.217	0.910	1.137
	CaCl ₂	28.98	18.10	24.57	18.23	3.76	3.20	4.750	2.83	4.023	5.970	1.127	1.160
	SNP	31.29	20.62	28.51	19.64	4.11	3.92	5.427	4.853	4.587	2.613	1.133	1.117
	SA	28.76	18.55	26.49	18.74	4.05	2.65	5.027	3.300	4.720	2.953	1.21	1.243
LSD at 5%		Ns	5.78	Ns	9.437	Ns	Ns	Ns	Ns	Ns	1.963	Ns	Ns

W=water, CaCl₂= calcium chloride, SNP= sodium nitroprusside, SA= salicylic acid, YE= yeast extract

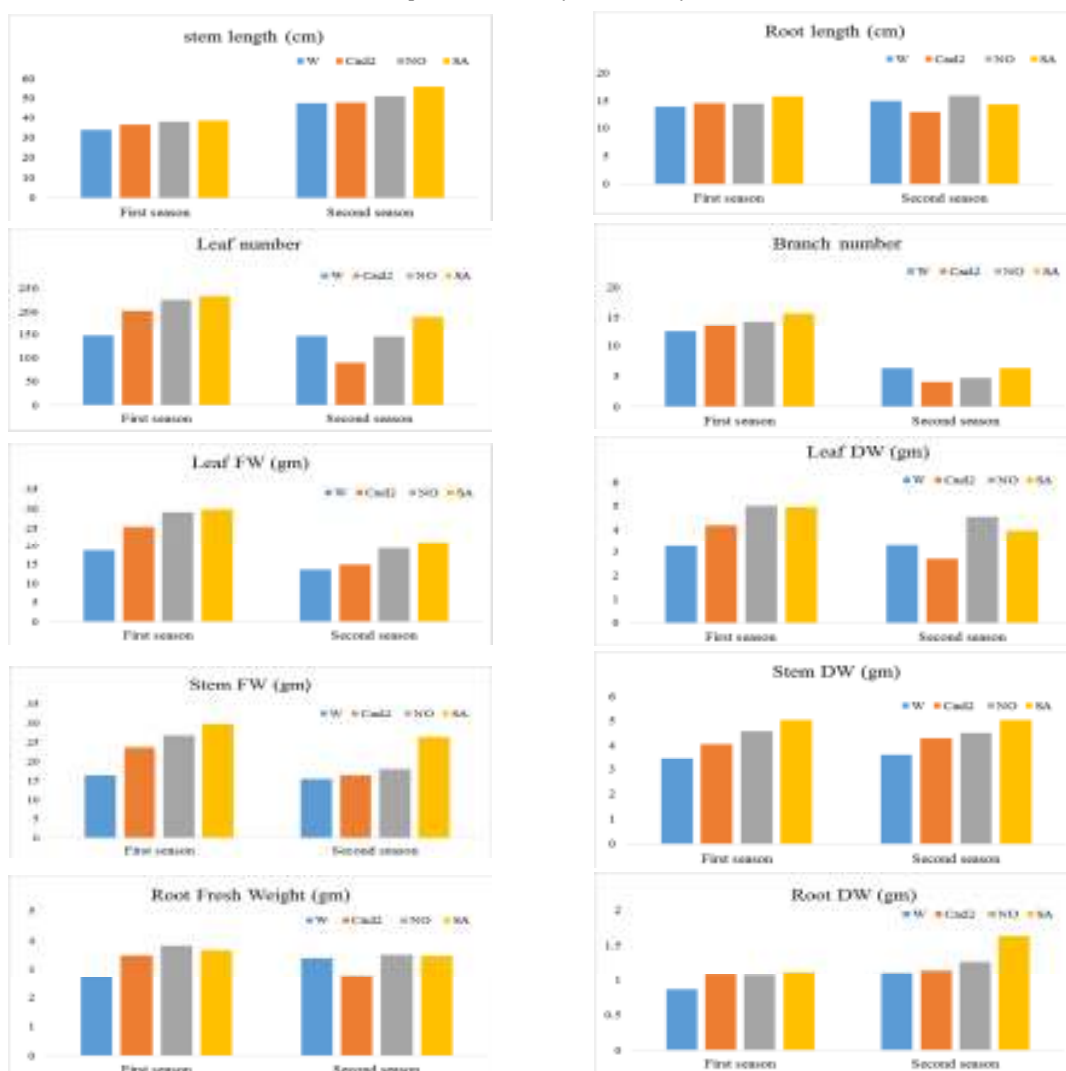


Fig. 2. Growth attributes of *C. roseus* as affected by treatment with calcium chloride (CaCl₂), sodium nitroprusside (SNP) and salicylic acid (SA) at 75 days after sowing in both seasons

Plant growth enhancement under SA application might be due to inducing biochemical or physiological processes, including cell division, cell differentiation, and morphogenesis (Fardus *et al.*, 2017; Kareem *et al.*, 2017). Additionally, the biosynthesis of many enzymes involved in metabolic pathway like glyoxylate cycle, the pentose phosphate pathway and glycolysis was strongly activated by SA, documented that SA enhances the release from a quiescence state to the establishment of a vigorous plant (Rajjou *et al.*, 2006).

The enhancing impact of SA application on plant growth may be resulted from SA- induced physiological processes i.e. improved carbon assimilates, raised biosynthesis of metabolites and maintenance of plant water status (Habibi, 2012). Mitosis and cell enlargement within the apical meristems were enhanced by SA (Sakhabutdinova *et al.*, 2003). Moreover, SA maintained RWC and photosynthetic (Mathur and Vyas, 2007). Moreover, SA spraying accelerating total soluble sugar build up that served as a substrate for enhancing initiation of leaf primordia (Munns *et al.*, 1979). It had inhibit impact on ethylene production by preventative accumulation of ethylene precursor, 1- aminocyclopropane 1- carboxylic acid (ACC) (Li *et al.*, 1992) and blocking ACC oxidase (Leslie and Romani, 1986). Salicylic acid boost leaves auxin and zeatin, in addition gibberellin (Shehata *et al.*, 2000); raised cytokinin level in plant (Sano *et al.*, 1994), additionally raised chlorophyll (Zayed, 1986).

In general, SA may regulate plant growth by at least one or more of the following mechanisms: (1) decreasing ethylene production hence, increasing chlorophyll as well as preventing the formation of abscission zone (Leslie and Romani, 1986); (2) protecting

plant cell against hazards of oxygen free radical under stress conditions by activating a new isoenzyme of ascorbate peroxidase, a H₂O₂ scavenging enzyme (Janda *et al.*, 1999); (3) increasing the rate of photosynthetic electron transport and/or the photochemical quenching (Tari *et al.*, 2002); (4) decreasing transpiration rate, as recorded in *Phaseolus vulgaris* and *Commelina communis* (Larque-Saavedra, 1978).

Anatomy of the Leaf:

Without YE application, foliar application of SNP markedly increased leaf blade thickness (25%) due to increasing spongy parenchyma thickness (50%) without any alteration in palisade parenchyma thickness (Table.4 and Fig. 3). The greatest leaf dimension at midrib region was obtained under the application of SNP. Application of CaCl₂ gave greatest phloem tissue thickness (33%), and xylem tissue thickness (33%) relative to untreated plants or other elicitors. The same Table and Figure postulated that, application of YE at both concentration obviously declined in most cases all studied anatomical characteristics except the thickness of xylem tissue which increased by 33% and 16% due to application of YE at 5 or 10 g/l⁻¹ respectively.

The interaction between YE and used abiotic elicitors, gave highest leaf blade thickness, palisade, spongy parenchyma thickness, dimension of leaf at midrib region and xylem tissue thickness. Similarly the highest palisade parenchyma thickness (50%) due to the application of SA supplemented with YE at 10 g/l⁻¹; and the xylem tissue thickness (50%) was obtained with application of SA with 5 g/l⁻¹ YE. However, application of SNP with 5 or 10 g/l⁻¹ YE increased phloem tissue thickness (33, 16%) respectively relative to the control.

Table 4. Anatomical characteristics of *C. roseus* leaf as affected by biotic and abiotic elicitors used as well as their interactions at 75 days from sowing during the 2nd growing season.

Parameters	Blade – Thickness		Palisade Parenchyma Thickness		Spongy – Parenchyma Thickness		Dimension of leaf at midrib region				Xylem Tissue Thickness		Phloem Tissue Thickness		
	µm	% ±	µm	% ±	µm	% ±	Length		Width		µm	% ±	µm	% ±	
							µm	% ±	µm	% ±					
Treatments (g/l)															
Water	15.84	0	5.93	0	7.41	0	8.91	0	23.76	0	5.94	0	2.97	0	
YE 0	CaCl ₂	+13	5.93	0	9.26	+25	13.77	+55	32.4	+36	7.92	+33	3.96	+33	
	SNP	+25	5.93	0	11.11	+50	13.77	+55	28.08	+18	5.94	0	2.97	0	
	SA	-13	4.44	-25	7.41	0	14.17	+59	37.8	+59	8.91	+50	2.97	0	
YE 5	Water	-25	5.93	0	3.70	-50	11.34	+27	27	+14	7.92	+33	1.98	-33	
	CaCl ₂	-13	4.44	-25	7.41	0	8.91	0	27	+14	5.94	0	4.95	+53	
	SNP	+38	7.41	+25	11.11	+50	15.79	+77	33.48	+50	7.92	+33	2.97	0	
SA	15.84	0	5.93	0	7.41	0	9.71	+9	31.32	+32	8.91	+50	3.96	+33	
YE 10	Water	-25	4.44	-25	5.55	-25	12.55	+41	37.8	+59	6.93	+16	2.97	0	
	CaCl ₂	-13	5.93	0	5.55	-25	8.50	-5	20.52	-14	5.94	0	3.96	+33	
	SNP	+25	7.41	+25	9.26	+25	9.31	+5	23.76	0	6.93	+16	4.95	+53	
SA	17.83	+13	8.89	+50	5.55	-25	8.91	0	16.2	-32	5.94	0	4.95	+53	

W = water, CaCl₂= calcium chloride, SNP = sodium nitroprusside, SA = salicylic acid, YE = yeast extract.

Anatomy of the Stem:

The data in Table (5) and Figure (4) recognized that, application of either 5 or 10 g/l⁻¹ YE increased stem diameter, epidermis, cortex thickness, vascular cylinder dimension, and phloem thickness regarding YE- free plants. The interaction between biotic and abiotic elicitors, in most cases showed an additive effect on the studied anatomical characteristics. The greatest stem diameter (29%), epidermis, cortex thickness (75%), vascular

cylinder dimension (135%), xylem thickness (20%), and phloem thickness (100%) were obtained by Ca foliar application under 5 g/l⁻¹ YE, meanwhile the greatest pith diameter (8%) and the big metaxylem vessels diameter (80%) were obtained due to application of Ca with 10 g/l⁻¹ YE or SA with 5 g/l⁻¹ YE.

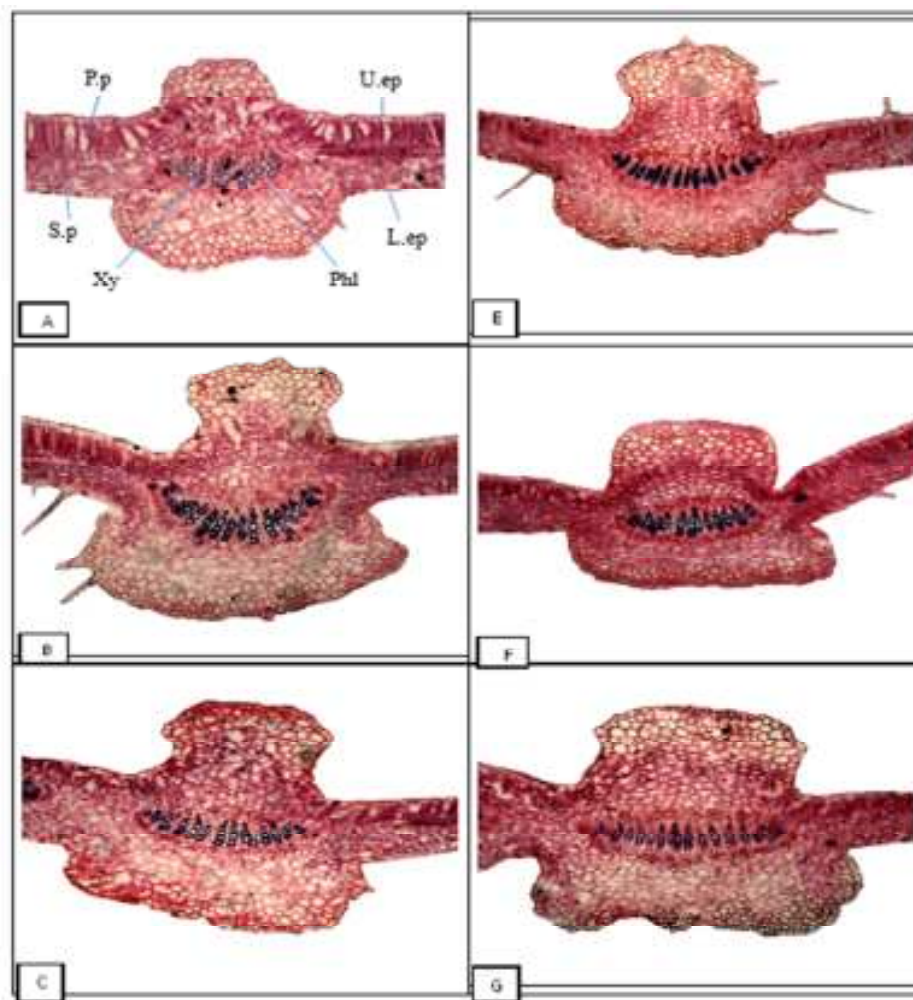


Fig. 3. Cross sections of Periwinkle leaf showing the effect of CaCl₂, SNP, SA, yeast extract with or without application.

A = Control B = CaCl₂ at 4.4 g/l⁻¹ C = SNP at 0.149 g/l⁻¹ D = SA at 0.138 g/l⁻¹
 E = YE at 5 g/l⁻¹ F = YE at 5 g/l⁻¹+ CaCl₂ at 4.4 g/l⁻¹ G = YE at 5 g/l⁻¹+ SNP at 0.149 g/l⁻¹ H = YE at 5 g/l⁻¹+ SA at 0.138 g/l⁻¹
 U. ep= upper epidermis, L. ep= lower epidermis, P.P= palisade parenchyma, S.P= spongy parenchyma, Xy= xylem, Phl= phloem

Table 5. Anatomical characteristics of *C. roseus* stem as affected by biotic and abiotic elicitors used as well as their interactions at 75 days from sowing during the 2nd growing season.

Parameters	Stem – diameter		Epidermis, Cortex Thickness		Vascular Cylinder-Diameter		Pith – diameter		Xylem-Thickness		Phloem Thickness		Big metaxylem Vessels diameter		
	µm	100 % ±	µm	100 % ±	µm	100 % ±	µm	100 % ±	µm	100 % ±	µm	100 % ±	µm	100 % ±	
Treatments (g/l)															
YE 0	Water	164.50	0	52.50	0	59.50	0	47.25	0	8.75	0	3.50	0	1.265	0
	CaCl ₂	198.77	+21	39.37	-25	143.50	+141	43.61	-8	8.75	0	5.25	+50	2.024	+60
	SNP	171.35	+4	52.5	0	108.50	+82	47.25	0	7.00	-20	5.25	+50	1.265	0
	SA	260.45	+58	78.75	+50	159.25	+166	72.69	+54	10.50	+20	8.75	+150	1.265	0
YE 5	Water	198.77	+20	65.62	+25	106.75	+79	43.61	-8	8.75	0	7.00	+100	2.530	+100
	CaCl ₂	212.47	+29	91.86	+75	140.00	+135	47.25	0	10.50	+20	7.00	+100	1.771	+40
	SNP	198.77	+20	39.37	-25	108.50	+82	47.25	0	7.00	-20	7.00	+100	1.518	+20
	SA	205.62	+25	52.49	0	126.00	+112	47.25	0	12.25	+40	5.25	+50	2.277	+80
YE 10	Water	205.62	+25	65.61	+25	103.25	+74	50.88	+8	5.25	-40	5.25	+50	1.012	-20
	CaCl ₂	205.62	+25	65.61	+25	91.00	+53	50.88	+8	8.75	0	3.50	0	1.518	+20
	SNP	212.47	+29	78.74	+50	91.00	+53	43.61	-8	7.00	-20	7.00	+100	1.518	+20
	SA	185.06	+12	65.61	+25	84.00	+41	36.34	-23	7.00	-20	5.25	+50	1.710	+35

W= water, CaCl₂= calcium chloride, SNP= sodium nitroprusside, SA= salicylic acid, YE= yeast extract

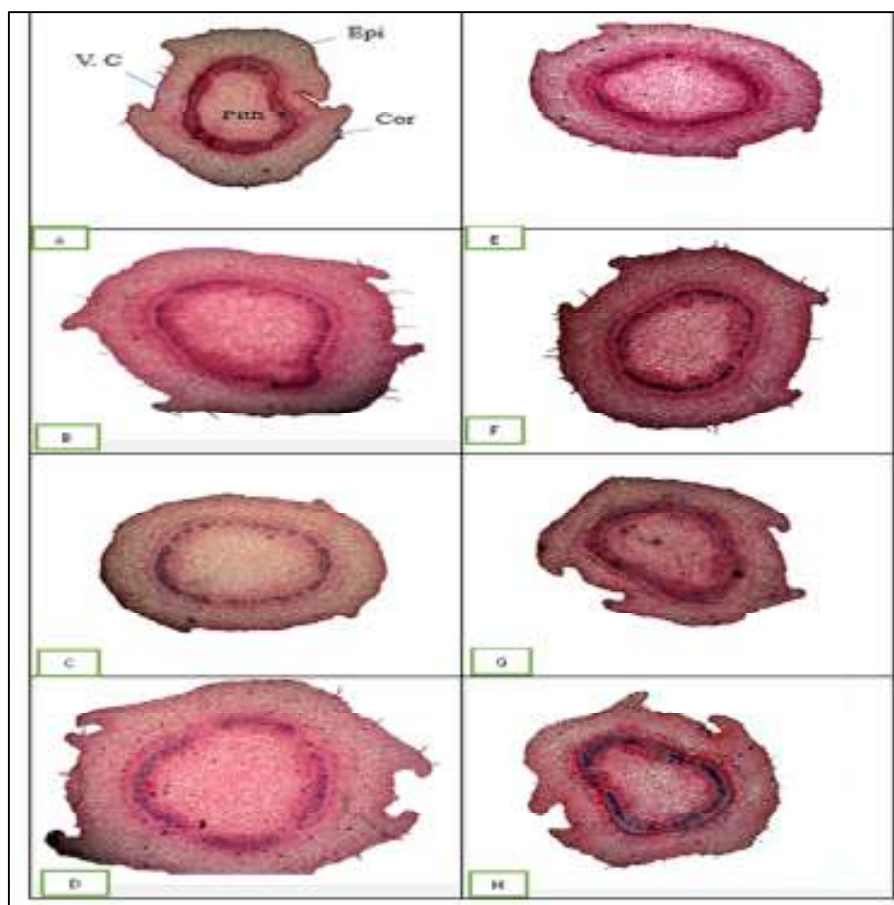


Fig. 4. Cross sections of periwinkle stem showing the effect of CaCl₂, SNP, SA, yeast extract with or without application.

A = Control B = CaCl₂ at 4.4 g/l⁻¹ C = SNP at 0.149 g/l⁻¹ D = SA at 0.138 g/l⁻¹
 E = YE at 5 g/l⁻¹ F = YE at 5 g/l⁻¹+ CaCl₂ at 4.4 g/l⁻¹ G = YE at 5 g/l⁻¹+ SNP at 0.149 g/l⁻¹ H = YE at 5 g/l⁻¹+ SA at 0.138 g/l⁻¹
 Epi= epidermis, Cor= cortex, V. C= vascular cylinder

Leaves are principle sites of essential biochemical process, like photosynthetic and transpiration. Anatomical alteration induced by elicitors in higher plants are better observed indicators; they can be directly used in agriculture and handled (Shao *et al.*, 2008). Application of both elicitors used improved in most cases all anatomical characteristics of periwinkle plants. Accordingly, foliar spraying with SNP considerably raised all leaf anatomical characters of leaves, i.e. dimension of midrib region due to increasing the dimensions of main vascular bundles (Farouk and Arafá Sally, 2018). Moreover, Farouk (2005) reported that application of either calcium chloride or salicylic acid raised the thickness of the pea leaf blade, by increasing the thickness of mesophyll tissue and the main midrib main vascular bundle. All these promotive effects of elicitors, generally increased the photo assimilate production and increased plant growth.

CONCLUSION

The great periwinkle plant growth can be obtained in this investigation by foliar application of 0.138 g/l⁻¹ salicylic acid with 5 g/l⁻¹ yeast extract.

REFERENCES

- Amer, S.S.A. (2004). Growth, green pods yield and seeds yield of common bean (*Phaseolus vulgaris* L) as affected by active dry yeast, salicylic acid, and their interaction. *J. Agric. Sci. Mansoura. Univ.*, 29 (3): 1407-1422.
- Angelova, Z.; Georgk, v. S and Roos, W. (2006). Elicitation of plants. *Biotech. Biotechnological Eciuiip.* 20, 72- 83.
- Bevilacqua, A.; Corbo, M.R.; Mastromatteo, M and Sinigaglia, M. (2008). Combined effects of pH, yeast extract, carbohydrates and di-ammonium hydrogen citrate on the biomass production and acidifying ability to a probiotic lactobacillus plantarum strain, isolated from table olives, in a batch system. *World J. Microbiol Biotechnol.*, 24: 1721-1729.
- Cai, Z.; Kastell, A.; Mewis, I.; Knorr, D and Smetanska, I. (2012). Polysaccharide elicitors enhance anthocyanin and phenolic acid accumulation in cell suspension cultures of *Vitis vinifera*. *Plant Cell Tissue Org. Cult.*, 108, 401–409.
- Correa-Aragunde, N.; Graziano, M., Chevalier, C and Lamattina, L. (2006). Nitric oxide modulates the expression of cell cycle regulatory genes during lateral root formation in tomato. *J. Exp. Bot.*, 57: 581-588.

- Davies, H.W and Monk-Talbot, L.S (1990): Permeability characteristics and membrane lipid composition of potato tuber cultivars in relation to Ca²⁺ deficiency. *Phytochemistry*, 29:2833-2835.
- Dipti, A.; Mujib, A.; Maqsood, M.; Ali, M and Zafar, N.; (2016). *Aspergillus flavus* fungus elicitation improves vincristine and vinblastine yield by augmenting callus biomass growth in *Catharanthus roseus*. *Plant Cell Tissue Org. Cult.* 126, 291– 303.
- Ebrahimzadeh, H.; Ataei-Azimi, A and Noori- Daloi, MR. (1996). The distribution of indole alkaloids in different organs of *Catharanthus roseus* G. Don. (*Vinca rosea* L, Daru, J. Sch. Pharm. 6(1&2): 11-24.
- Fardus, J.; Abdul Matin, M.d.; Hasanuzzaman, M.d. S.; Hossain, S.D.; Hossain, Md. A.; Rohman, M and Hasanuzzaman, M. (2017). Exogenous salicylic acid-mediated physiological responses and improvement in yield by modulating antioxidant defense system of wheat under salinity. *Not. Sci. Biol.* 9(2):219-232. DOI: 10.15835/nsb929998.
- Farouk S. (2005). Response of *Pisum sativum* L to some osmoregulators and plant growth substances under salt stress. PhD thesis, fac. of Agric. Mansoura Univ.
- Farouk, S.; Arafa, S. A. (2018). Mitigation of salinity stress in canola plants by sodium nitroprusside application. *Spanish Journal of Agricultural Research*, Volume 16, Issue 3, e0802.
- Garcia-Mata, C and Lamatina, L. (2007). Abscisic acid (ABA) inhibits light induced stomatal opening through calcium- and nitric oxide-mediated signaling pathways. *Nitric Oxide*, 17: 143-151.
- George, E.F.; Hall, M.A and De- Klerk, G.J. (2008). Micropropagation uses and methods,. *Plant Propagation by Tissue Culture*. Springer. Netherlands, pp. 29-64.
- Gerlash, D (1977). *Botanische mikrotechnik Ein einfuhrny*. Thieme Verlage, Stult gort Bro.
- Ghassemi- Golezani, K. S.; Ghassemi and Yaghoubian, I. (2017). Improving oil and falconoid contents of milk thistle under water stress by salicylic acid. *Adv. Hortic. Sci.*, 31(1):19-23.
- Gill, S.S.; Hasanuzzaman, M.; Nahar, K.; Macovei, A and Tuteja, N. (2013). Importance of nitric oxide in cadmium stress tolerance in crop plants. *Plant Physiol Biochem.*, 63: 254- 261.
- Habibi, G. (2012). Exogenous salicylic acid alleviates oxidative damage of barley plants under drought stress. *Acta Biol. Szeged.*, 56, 57–63.
- Hanafy, A.A.H.; Mohamed Hanaa, F.Y.; Orabi, IOA and EL-Hefny, A.M. (2018). Influence of gamma rays, humic acid and sodium nitroprusside on growth, chemical constituents and fruit quality of snap bean plants under different soil salinity levels. *Biosci Res.*, 15 (2): 575-588.
- Janda, T.; Szalai, G.; Tari, I. and Paldi, E. (1999): Hydroponic treatment with salicylic acid decreases the effects of chilling injury in maize (*Zea mays* L.) plants. *Planta* 208:175-180.
- Kareem, F.; Rihan, H. and Fuller, M. (2017). The effect of exogenous applications of salicylic acid and molybdenum on the tolerance of drought in wheat. *Agri Res. Tech. Open Access J.* 9(4): 555768. DOI: 10.19080/ARTOAJ.2017.09.555768. 009.
- Khan, N.A.; Sarniullah, Singh, S. and Nazar, R. (2007). Activities of antioxidant enzymes, sulphur assimilation, photosynthetic activity and growth of wheat [*Triticum aestivum*] cultivars differing in yield potential under cadmium stress, *J. Agro. Crop Sci.*, 193,435- 444.
- Kiegle, E.; Moore, C.A.; Hasedoff, J.; Tester, M.A. and Knight, M.R. (2000). Cell-type-specific calcium responses to drought, salt and cold in the *Arabidopsis* root. *Plant J.* 23, 267-278.
- Kurth, E.; Cramer, G.R.; Lauchli, A. and Epstein, E. (1986): Effect of NaCl and CaCl₂ on cell enlargement and cell production in cotton root. *Plant Physiology*, 82:1102-1106.
- Larque´-Saavedra, A. (1978). The antitranspirant effect of acetylsalicylic acid on *Phaseolus vulgaris* L. *Physiol. Plant.* 43, 126–128.
- Leslie, C.A. and Romani, R.J. (1986): Salicylic acid a new inhibitor of ethylene biosynthesis. *Plant Cell Rep.* 5(2): 144-146.
- Li, C.J. and Bangerth, F. (1992): The possible role of cytokinins, ethylene and indole acetic acid in apical dominance. IN: *Progress in plant growth regulation* (karssen, C.M.; C. Van Li and D. Vreugdenhil, eds), pp 431-436. Kluwer Academic publ., Netherlands.
- Loyola-Vargas, V.M.; Galaz- Avaios, R.M. and Ku- Cauch, R. (2007). *Catharanthus roseus* biosynthetic enzymes: the road ahead. *Phytochem. Rev.* 6, 307-339.
- Maqsood, M.; Mujib, A. (2017). Yeast extract elicitation increases Vinblastine and vincristine yield in protoplast-derived tissues and plantlets in *Catharanthus roseus*. *Revista Brasileira de Farmacognosia. BJP* 382 1–8.
- Marschner, H. (1995). *Mineral nutrition of high plant*. Academic Press, pp: 330-355.
- Mathur, N. and Vyas, A. (2007). Physiological effect of some bioregulators on vegetative growth, yield and chemical constituents of pearl millet (*Pennisetum typhoides* (Burm) Stapf. and Hubb). *Int. J. Agric. Res.* 2 (3): 238-245.
- Mishina, T.E.; Lamb, C. and Zeier, J. (2007). Expression of a nitric oxide degrading enzyme induces a senescence programme in *Arabidopsis*. *Plant Cell Environ.*, 30: 39-52.
- Mohamed, M.F.; Thaloath, A.T.; Essa, R. E. Y and Gobarah Mirvat, E (2018). The stimulatory effects of tryptophan and yeast on yield and nutrient status of wheat plants (*Triticum aestivum* L.) grown in newly reclaimed soil. *Middle East j. of Agriculct Research.*, 7 (1) 27- 33.
- Motsara, M. R. and Roy, R. N. (2008). *Guide to laboratory establishment for plant nutrient analysis; FAO fertilizer and plant nutrition Bulletin* in No; 19.
- Munns, R.; Brady, C.J. and Barlow, E.W.R. (1979). Solutes accumulation in the apex and leaves of wheat during water stress. *Aust. J. Plant Physiol.*, 6, 379-389.
- Norman, G.R and Streiner, DL (2003). *PDQ statistics*, 3rd Ed. Bc Deckker Inc., London.
- Rai, K.K., Rai, N. and Rai, S.P. (2018). Salicylic acid and nitric oxide alleviate high temperature induced oxidative damage in *Lablab purpureus* L plants by regulating bio-physical processes and DNA methylation. *Plant Physiol Biochem.*, 128: 72-88.
- Rajjou, L.; Belghazi, M.; Huguet, R.; Robin, C.; Moreau, A.; Job, C and Job, D. (2006). Proteomic investigation of the effect of salicylic acid on *Arabidopsis* seed germination and establishment of early defense mechanisms. *Plant Physiology.*, 141, 910–923.
- Sakhabutdinova, A. R.; Fatkhutdinova, D. R.; Bezrukova, M. V and Shakirova, F. M. (2003): Salicylic acid prevents the damaging action of stress factors on wheat plants. *Bulg. j. Plant Physiol.*, special issue, 314–319.

- Samar, F.; Mujib, A and Dipti, T. (2015). NaCl amendment improves vinblastine and vincristine synthesis in *Catharanthus roseus*: a case of stress signaling as evidenced by antioxidant enzymes activities. Plant Cell Tissue Org. Cult. 121, 445–458.
- Sano, H.; Seo, S.; Orudjev, E.; Yosseflan, S.; Ishizuka, K and Ohashi, Y (1994): Expression of the gene for a small GTP binding protein in transgenic tobacco elevates cytokinin levels, abnormally induces salicylic acid in response to wounding and increases resistance to tobacco mosaic virus infection. Proc. Of the Nat. Acad. Sci. of the United State of American. 91(22): 10556-10560.
- Sarhan, T.Z. (2008). Effect of biological fertilizers, animal residues and urea on growth and yield of potato plants cv. Desiree (*Solanum tuberosum* L.) Ph.D. Thesis, Horticulture Sciences and Landscape Design (Vegetable), College of Agriculture and Forestry, University of Mosul.
- Shao Hong- Bo.; Li-Ye Chu.; Jaleel, CA.; Zhao, Chang-Xing. (2008) Water-deficit stress-induced anatomical changes in higher plants. Comptes Rendus Biologies. 331:215–225.
- Shehata, S.A.M.; Saeed, M.A and Abou EL-Nour, M.S (2000): Physiological response of cotton plant to the foliar spray with salicylic acid. Annals Agric. Sci., Ain Shams Univ. 45(1): 1-18.
- Shukla, A.K.; Shasany, A.K.; Khanuja, S.P.S. (2012). cDNA-AFLP-based numerical comparison of leaf and root organ cDNAs in *Catharanthus roseus*. OMICS 16, 397–401.
- Tari, I.; Csiszár, J.; Szalai, G.; Horváth, F.; Pécsvárad, A.; Kiss, G.; Szepesi, A.; Szabó, M., and Erdei, L. (2002): Acclimation of tomato plants to salinity stress after a salicylic acid pre-treatment. Acta. Biol. Szegediensis, 46: 55-56.
- Xi, Q.; Lai, W.; cui, Y.; Wv, H.; Zhao, T (2019). Effect of yeast extract on seedling growth promotion and soil improvement in afforestation in a semiarid chestnut soil area. Forests: 10 (76) doi: 10. 3390/ f10010076.
- Xu, M and Dong, J (2005). Nitric oxide stimulates indole alkaloid production in *Catharanthus roseus* cell vsuspension cultures through a protein kinase-dependent signal pathway. Enzyme Microb. Teehnol 37, 49-53
- Zayed, A. E (1986). Comparative studies on the effect of some auxin transport inhibitors on cucumber (*Cucumis sativa* L.). Angew. Botanik, 60: 23-29.
- Zhao, J.; Zhu, W.H and H.u, Q (2001). Effects of light and plant growth regulators on the biosynthesis of vindoline and other indole alkaloids in *Catharanthus roseus* callus cultures. Plant Growth Regul. 33:43–4
- Zhu, S.H.; Sun, L.N.; Liu, M.C.; Zhou, J (2008). Effect of nitric oxide on reactive oxygen species and antioxidant enzymes in kiwifruit during storage. J. Sci. Food Agric., 88: 2324- 2331.

النمو والتركيبة التشريحي لأوراق وسيقان نبات الونكا تحت تأثير بعض المحفزات الحيوية والغير حيوية نجلاء حسن الشيخ ، سعد فاروق ، زين العابدين عبد الحميد محمد و عرفه أحمد عرفه قسم النبات الزراعي، كلية الزراعة، جامعة المنصورة

تعتبر الونكا مصدرا هاما للعديد من القلويدات ذات الأهمية الدوائية، وبصفة خاصة الفينكريستين والفينبلستين اللذان يستخدمان في علاج السرطان، والتي يزداد عليهما الطلب عالميا برغم تكوينهما بكميات صغيرة جدا، وقد تم إجراء الدراسة الحالية بهدف زيادة إنتاجية النبات وتحسين نموه وتركيبه الداخلي. واستخدم لهذا الغرض بعض المحفزات الحيوية والغير حيوية مثل كلوريد الكالسيوم بتركيز ٤.٤ جم/ لتر، نيتروبروسيد الصوديوم بتركيز ١.٤٩ جم/ لتر، حامض السالسليك بتركيز ٠.١٣٨ جم/ لتر منفردة أو متداخلة مع مستخلص الخميرة بتركيز ٥ أو ١٠ جم/ لتر. وقد أجريت تجارب أصص في المزرعة البحثية ومعامل قسم النبات الزراعي، بكلية الزراعة، جامعة المنصورة، خلال موسمي نمو (٢٠١٥-٢٠١٦). وأوضحت النتائج بصفة عامة أن المحفزات المختبرة قد أثرت إيجابيا علي صفات النمو المدروسه معتمدة علي نوع المحفز حيث زاد طول الساق، الوزن الجاف له بالإضافة إلي عدد الأوراق للنبات استجابة للمعاملة بمستخلص الخميرة بتركيز ٥ أو ١٠ جم/ لتر بالتتابع. وكانت معظم صفات النمو مرتفعة عند المعاملة بحامض السالسليك ونيتروبروسيد الصوديوم بالتتابع، وكان التأثير التحفيزي لإضافة العاملين معا واضحا خاصة عند التداخل ما بين أي من حامض السالسليك، أونيتروبروسيد الصوديوم مع مستخلص الخميرة، فقد زاد الوزن الجاف للساق في النباتات المعاملة بحامض السالسليك مع مستخلص الخميرة بتركيز ٥ جم/ لتر. ومن الناحية التشريحية وجد أن استخدام المحفزات المستخدمة بصفة عامة أدت إلي زيادة نسبة الأنسجة الممتلئة للتخليق الضوئي في أنسجة الورقة، والأنسجة الوعائية في الساق، وعليه فإنه لزيادة الكتلة الحيوية لنبات الونكا وكذلك فإنه يوصي بالمعاملة بحامض السالسليك مع مستخلص الخميرة بتركيز ٥ جم/ لتر.