

SEROLOGICAL AND MOLECULAR DETECTION OF TOXOPLASMA GONDII IN SHEEP IN DIYALA PROVINCE, IRAQ

MOHAMMED K. BAWI AND IBRAHIM A.H. AL-ZUBAIDI

Unit. Zoonotic Diseases, College of Veterinary Medicine, University of Baghdad

Received: 10 July 2024; Accepted: 2 August 2024

ABSTRACT

Toxoplasma gondii is the common cause of toxoplasmosis in both humans and animals. Sheep toxoplasmosis represents an important role in public health, causing reproductive and economic losses due to abortion and neonatal mortality worldwide. The seroprevalence and molecular detection of sheep toxoplasmosis in Diyala province, Iraq, were the aim of this study. One hundred male slaughterhouses sheep and 100 grassing dairy ewes were examined and divided into groups according to age, season, breed, importation, contact with cats, pregnancy status and abortion. *Toxoplasma gondii* infection was investigated using a *Toxoplasma* rapid test and polymerase chain reaction (PCR) test. The overall seroprevalence of toxoplasmosis in rams was 18%, while it was 14% in grassing dairy ewes. The PCR test confirmed the seropositive infection in 50% and 35.7% of rams and dairy ewes, respectively. Adult dairy ewes and sheep from slaughterhouses had a statistically insignificantly greater seroprevalence of toxoplasmosis than juveniles. The prevalence of the disease was slightly higher in the winter groups in either rams or dairy ewes. Sheep imported from Syria and Iran had twice as high an infection rate as native sheep from Iraq. The infection rate among dairy ewes from the Karadi and Hamdani breeds was not considerably greater than those of other breeds. Sheep with a history of abortion showed higher seroprevalence, and also toxoplasmosis was more prevalent in previously integrated sheep with cats. Therefore, continuous surveillance of sheep toxoplasmosis detection is highly recommended for further prevention.

Keywords: toxoplasmosis, Iraq, sheep, seroprevalence, *bl* gene.

INTRODUCTION

Toxoplasmosis is a common foodborne illness caused by *Toxoplasma gondii* (*T. gondii*), a common protozoan, considered a member of the phylum *Apicomplexa* (viera da silva *et al.*, 2005; Pereira *et al.*, 2010; and Darayani *et al.*, 2014). Toxoplasmosis is a common infectious agent that results in reproductive

failure in sheep. It causes placentitis, abortions, and stillbirths, which over time cause significant financial losses (Ali *et al.*, 2019; Clune *et al.*, 2021). Lambs are mostly affected by eating *T. gondii* oocysts while grazing in a contaminated environment (Hassanain *et al.*, 2011; Motoi *et al.*, 2022 and Iqbal *et al.*, 2023). Only three clonally distinct forms of *T. gondii* isolates-types I, II, and III-have been identified in sheep flesh (Duby, 2002; Elfadaly *et al.*, 2017; Hill and Aleem *et al.*, 2018). It was possible for the same beef sample to contain multiple infections. The biological traits of the less virulent strains were concealed by

Corresponding author: MOHAMMED K. BAWI

E-mail address:

mohammed.bawi2204m@covm.uobaghdad.edu.iq

Present address: Unit. Zoonotic Diseases, College of Veterinary Medicine, University of Baghdad

the more virulent strains (Shaapan *et al.*, 2015; Dubey, 2016; and Assim and Saheb, 2018). Sheep have higher global seroprevalences of *T. gondii* than other agricultural animals due to their higher infection rates (Stelzer *et al.*, 2019; Daher *et al.*, 2021; and Gharban and A-kaabi, 2022).

Mutton is a popular meat source, and because it has more nutrients than other red meats, it is consumed more often around the world, particularly in less developed nations, giving people sufficient dietary protein (Weiss and Dubey, 2009; Mahmoud *et al.*, 2021). *T. gondii* tissue cysts, which are present in edible animal flesh, especially sheep mutton, are the main source of human toxoplasmosis because they can survive in undercooked tissues (Toalebetal, 2013; Aguirre *et al.*, 2019; and Segbedzi *et al.*, 2023). In addition to mutton, *T. gondii* tachyzoites have been found in the milk of a number of food animal species, such as sheep, goats, and cows. The infectious tachyzoites can live in fresh cheese and are expelled in milk (Munoz-Zanzi *et al.*, 2012; Dawson *et al.*, 2020; and Almeria and Dubey, 2021).

The high prevalence of toxoplasmosis observed in humid regions of the world affects one-third of the global population (Dubey and Beattie, 1988; Studeničová *et al.*, 2006; and WHO, 2015). The significance of this disease made the global epidemiological research crucial as its seroprevalence rated 40% in the USA, 30% in the UK, and 50% in southern Europe (Punda-Polić *et al.*, 2000; Diza *et al.*, 2005), while in Serbia it was 20.5% (Stopić *et al.*, 2022), Albania 48.6% (Maggi *et al.*, 2009), Turkey 28.57% (Karakavuk *et al.*, 2024), Egypt 57.3% (Elaadli *et al.*, 2023), Qatar 29/8% (Abu-Madi *et al.*, 2008), Syria 56.01% (Adi and Alkhaled, 2011), Iran 30.4%, and in Pakistan 34.5%.

Serological assays that identify anti-*T. gondii* specific immunoglobulins IgM and IgG antibodies in sera samples are the

primary methods used to diagnose toxoplasmosis in sheep (Shaapan *et al.*, 2008; Hassanain *et al.*, 2018; and McCall *et al.*, 2022).

Research on toxoplasmosis in Iraq has not been conducted nationwide. Several research focus on individual governorates and provide information on the seroprevalence of infection in those governorates (Al-Taie and Abdulla, 2011; Mahmood *et al.*, 2013; Al-Hindawi and Al-Shanawi, 2015; Al-Maamuri *et al.*, 2015; and Asal and AL-zubaidi, 2016; Al-Khafagi and Zainab, 2016; Ali and Al-Warid, 2021; Musa *et al.*, 2021; Alkubaisi and AL-zubaidi, 2023; and Bawi and AL-zubaidi, 2024). Therefore, this study was designed to clarify the occurrence of toxoplasmosis in sheep in Diyala province in Iraq, using serological and molecular methods, to determine the prevalence of toxoplasmosis in sheep (rams and dairy ewes), as well as to identify the associated risk factors.

MATERIALS AND METHODS

1- Ethical approval.

This experiment was conducted in compliance with the institutional rules of the University of Baghdad's College of Veterinary Medicine's Animal Care and Use Committee, Iraq. Project Number: Pg. 1-50. The herd's owners gave their verbal agreement before samples were taken.

2- Animals and study area.

An epidemiological survey was conducted in two seasons, winter and spring. The study duration was between November 2023 and February 2024 for the winter group, and between February and April 2024 for the spring one. The investigated animals were rams (5 months-5 years of age) before slaughtering in abattoirs and dairy ewes (1–6 years). The study was carried out in Diyala province (X/Y coordinates: 33.883333, 45.066667), in the eastern region of Iraq. Rams were randomly selected from various slaughterhouses

across the province, while the grazing dairy ewes were sourced from multiple herds on the farm.

3- Samples.

Blood samples were collected randomly from 100 rams, in addition to 100 blood and milk samples from multiple herds of grassing dairy ewes during the active milking period. Data was obtained about the breed, either from local breeds in Iraq or imported from other countries, contact with cats, history of abortions, availability of drinking water, and other factors.

4- Samples preparation and processing.

Approximately 10 ml of jugular vein blood was obtained from all sheep, 5 ml in EDTA tubes with anticoagulant for molecular examination, and 5 ml in gel tubes without anticoagulant for separation of serum. Only 5 ml of blood was obtained from grassing dairy ewes in gel tubes without anticoagulant for obtaining serum. Each sample was labeled using codes describing the specific animal's gender and age. Blood collected in gel tubes was left at room temperature to clot, then centrifuged at 3000 rpm for 15 minutes. Serum was separated and used in the latex agglutination test. The blood collected in EDTA tubes was stored at (-20 °C) until used in the PCR test. 10 ml of milk samples were collected from grassing dairy ewe after cleaning and sanitizing teats and removal of the first few jets of milk. Milk samples were stored at (-20 °C) until used in the molecular analysis.

5-Serological examination.

Sheep and grassing dairy ewes' serum samples were tested using a toxoplasma rapid test (Toxo IgG and IgM combo rapid test, CTK Biotech, USA), according to the company's instructions for toxoplasmosis detection.

6-Molecular examination.

Only positive cases with the latex agglutination test were subjected to the presence of the *b1* gene of *T. gondii* using

polymerase chain reaction (PCR). Blood and milk samples were processed to obtain total genomic DNA. After the samples were pelleted, 200 µL of PBS were used for dilution, before extracting total genomic DNA. A 406-bp segment of the recurrent *b1* gene (35fold repeats/genome) was the target of the PCR using specific primers (Macro Gene Company, South Korea) (Table 1). The amplified DNA was electrophoresed in 1.5% Agarose gel.

Table 1: The primers designed to detect the *b1* gene of *T. gondii*.

Primer	Sequence
Forward	5-ATACAGGTGAAATGTACCTCC-3
Reverse	5-CGATCTTCTTCTCTCTGTCTT-3

7-Statistical analysis

The System-SAS (2018) program was used to statistically analyze the data to determine the impact of different factors on the research parameters. Also, the Chi-square test and differences were performed and considered statistically significant if P-values were below 0.05.

RESULTS

1-Seroprevalence of toxoplasmosis in sheep.

The overall prevalence of toxoplasmosis in rams from slaughterhouses was 18 out of 100 (18%) using the latex agglutination test (Table 2). While, the overall incidence in grassing dairy ewes was 14 out of 100 (14%).

2-Molecular detection.

Molecular examination of seropositive cases that confirmed the infection was in 9 (50%) of the rams cases, while it was confirmed in only 5 (35.7%) seropositive cases of dairy ewes (Table 2) by amplification of the *b1* gene of *T. gondii* (Figures 1 and 2).

3-Risk factors.

Seroprevalence of toxoplasmosis was statistically insignificantly higher in adult sheep and dairy ewes than juveniles. Moreover, there was no significant

variation between winter and spring seasons in both genders, although it was a little higher in winter groups (Tables 3 and 4).

By comparing the native sheep (Iraq) and the imported ones (Syria and Iran), the difference was not statistically significant (Table 3). Both of the Karadi and Hamdani

ewe's breeds had an insignificantly higher infection rate than other breeds (Table 4).

A non-significant seroprevalence was observed in ewes with a history of abortion compared to non-pregnant ones. According to our study, toxoplasmosis was significantly more common in sheep previously interacted with cats (Table 4).

Table 2: Toxoplasmosis prevalence rate in slaughterhouses' sheep and dairy ewes.

Animals	Latex agglutination test (screening)			Molecular detection (confirmation)		
	Tested no.	Positive no. (%)	Negative no. (%)	Tested no.	Positive no. (%)	Negative no. (%)
Slaughterhouses' sheep	100	18 (18)	82 (82)	18	9 (50)	9 (50)
Dairy ewes	100	14 (14)	86 (86)	14	5 (35.7)	9 (64.3)

Table 3: Seroprevalence of *T. gondii* antibody and the associated risk factors in slaughterhouses' sheep.

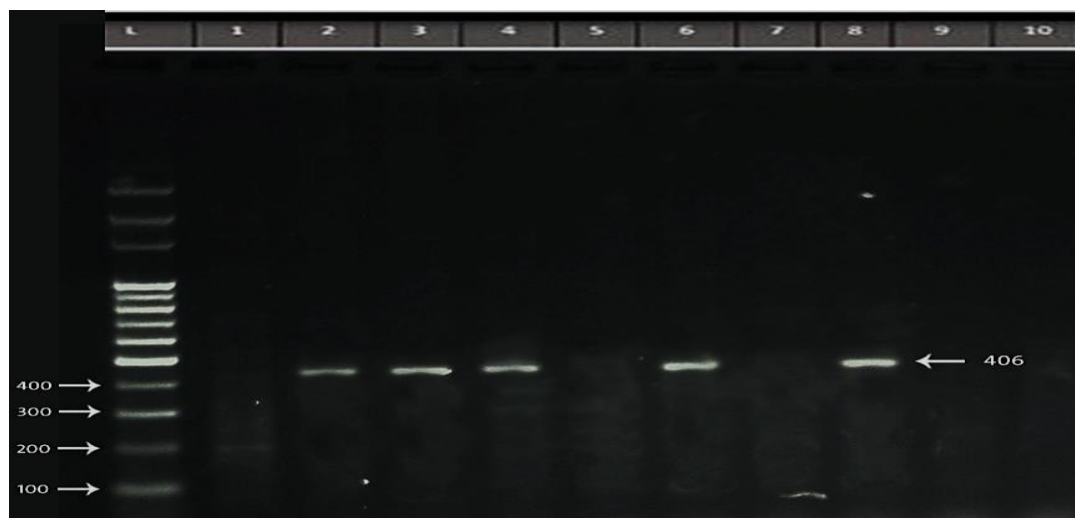
Factor	No. tested	Seroprevalence		P value
		Positive no. (%)	Negative no. (%)	
Age				
Less than one year	70	10 (14.3)	60 (85.7)	Reference
1 – 2 years	20	5 (25)	15 (75)	0.305
More than 2 years	10	3 (30)	7 (70)	0.353
Season				
Winter	50	10 (20)	40 (80)	0.795
Springe	50	8 (16)	42 (84)	
Importation				
Local from Iraq	49	6 (12.2)	43 (87.8)	Reference
Imported from Syria	19	4 (21.1)	15 (78.9)	0.448
Imported from Iran	32	8 (25)	24 (75)	0.228



Figure 1: Agarose gel electrophoresis of PCR after amplification of *the b1* gene of *T. gondii* infection in slaughterhouse sheep. Line 1: DNA ladder 100 bp, line 2 and 3: negative samples, and lines 4 to 11: positive DNA samples with amplified product at 406 bp.

Table 4: Seroprevalence of *T. gondii* antibody and the associated risk factors in grassing dairy ewes.

Factor	No. tested	Seroprevalence		P value
		Positive no. (%)	Negative no. (%)	
Age				
Less than one – 2 years	27	2 (7.4)	25 (92.6)	Reference
3 – 4 years	40	7 (17.5)	33 (82.5)	0.295
5 – 6 years	33	5 (15.2)	28 (84.8)	0.442
Season				
Winter	50	9 (18)	41 (82)	0.388
Springe	50	5 (10)	45 (90)	
Breed				
Awassi	49	5 (10.2)	44 (89.8)	Reference
Karadi	27	6 (22.2)	21 (77.8)	0.183
Niamey	18	2 (11.1)	16 (88.9)	1.000
Hamdani	6	1 (16.7)	5 (83.3)	0.518
Pregnancy status				
Non-pregnant	63	8 (12.7)	55 (87.3)	1.000
Pregnant	25	3 (12)	22 (88)	Reference
Aborted	12	3 (25)	9 (75)	0.367
Previous contact with cats				
Present	10	3 (30)	7 (70)	0.145
Absent	90	11 (12.2)	79 (87.8)	

**Figure 2:** Agarose gel electrophoresis of PCR after amplification of the *bl* gene of *T. gondii* infection in grassing dairy sheep. Line 0: DNA ladder 100 bp, line 1, 5, and 7: negative samples, and lines 2, 3, 4, 6, and 8: DNA samples with amplified product at 406 bp.

DISCUSSION

Being sheep an important reservoir for many pathogens with public health significance, this study was conducted to investigate the seroprevalence and

molecular recognition of sheep toxoplasmosis in Diyala province, Iraq. The overall seroprevalence of toxoplasmosis in rams was 18%, while it was 14% in grassing dairy ewes. This result agreed with Deyhimi *et al.* (2019), who recorded a

prevalence of 27.31% in Iran, while it disagreed with Abdel-Aziz *et al.* (2020) in Egypt, who recorded 45.8%, and Ishaku *et al.* (2018) in Nigeria, who documented 35.3% occurrence percentage. This variation may be related to the type of the collected sample, as Deyhimi *et al.* (2019) study was on blood samples and the two other studies were on meat samples. This confirms that *T. gondii* may persist in the blood for a short time but remain in tissues for a longer duration (Almeria and Dubey, 2021).

Molecular examination of seropositive cases had confirmed the infection in 9 (50%) of the ram cases, while it was confirmed in only 5 (35.7%) of the seropositive cases of dairy ewes. This result agreed with a study in Egypt (62.6%) by Elaadli *et al.* (2023). However, it disagreed with the result of 6.5% in Salvador (de Santana Rocha *et al.*, 2015), 5.4% in Brazil (Camossi *et al.*, 2011), and 28% of Slovak Republic-sourced sheep (Luptakova *et al.*, 2015). Various factors, such as farm management, environmental conditions, diagnostic techniques employed, and the presence of cats in the herds could be responsible for the discrepancy in results (Amairia *et al.*, 2016).

Seroprevalence of infection was higher in adult slaughterhouses' sheep and dairy ewes than juveniles, although it was statistically insignificant. This confirms previous investigations showing higher rates of exposure to toxoplasmosis infection with increasing age of sheep (Ibrahim *et al.*, 2017; Subedi *et al.*, 2018).

Additionally, there was no significant variation between winter and spring seasons in both rams and dairy ewes, and generally, the prevalence was a little higher in winter groups. This higher prevalence in winter may be related to climate conditions and the age of the sheep included in this study, because in this area people always slaughter old sheep during the winter season.

Comparing the native sheep (Iraq) and the imported ones (Syria and Iran), the difference was statistically insignificant. Sharif *et al.* (2015) documented the variation in prevalence of toxoplasmosis, which was at a higher rate in Iran than Iraq. Syrian sheep had an infection rate of 56.01%, which is likewise high. This clarifies why the infection rates of Syrian and Iraqi sheep differed (Adi and Alkhaled, 2011).

Dairy ewes of the Karadi and Hamdani breeds had an insignificant higher infection rate than other breeds. The two possible explanations for this could be as follows: first, sheep like Hamdani and Karadi mainly breed in regions close to Iran's borders, which puts them at high risk of infection. The second factor, currently under investigation, relates to how the immune competence of these breeds compares to the Iraqi Awassi breed.

A non-significant seroprevalence was observed in ewes with a history of abortion compared to non-pregnant ones. The animal might still be infected due to the chronic nature of the disease, the possibility that it had the parasite for a long time, and the fact that abortion did not completely eliminate the parasite (Ali *et al.*, 2019).

According to our study, toxoplasmosis was significantly more common in sheep previously interacted with cats. Cats, the definitive host of *T. gondii*, are a major source of infection for humans and animals due to shedding of oocysts into the environment (Frenkel *et al.*, 1970). Thus, transmission of infection can occur via drinking contaminated water and consumption of contaminated food (Robert-Gangneux and Dardé, 2012).

CONCLUSION

The study confirmed the presence of toxoplasmosis infection in Diyala province Iraqi sheep. The prevalence of the disease was higher in adult sheep imported from Iran and Syria, and in adult abortive Karadi and Hamdani ewes, especially in the winter months.

REFERENCES

- Abdel-Aziz, N.M.; Hassanien, A.A. and Arafa, M.I. (2020):* Detection of *Toxoplasma gondii* in aborted women and meat of slaughtered sheep and cattle in Sohag city, Upper Egypt. *Adv. Anim. Vet. Sci*, 8(6), 680-686.
- Abu-Madi, M.A.; Al-Molawi, N. and Behnke, J.M. (2008):* Seroprevalence and epidemiological correlates of *Toxoplasma gondii* infections among patients referred for hospital-based serological testing in Doha, Qatar. *Parasites & vectors*, 1, 1-9.
- Adi, E.Z. and Alkhaled, A. (2011):* Detection of *Toxoplasma gondii* in sheep and human in hama syria. *Assiut Veterinary Medical Journal*, 57(128), 1-26.
- Aguirre, A.A.; Longcore, T.; Barbieri, M.; Dabritz, H.; Hill, D.; Klein, P.N. and Sizemore, G.C. (2019):* The one health approach to toxoplasmosis: epidemiology, control, and prevention strategies. *EcoHealth*, 16(2), 378-390.
- Aleem, U.; Ullah, S.; Qasim, M. and Suliman, M. (2018):* Seroprevalence of Toxoplasmosis in Pregnant Women in Matta, Upper Swat, Khyber Pakhtunkhwa, Pakistan. *Journal of Saidu Medical College, Swat*, 82).
- Al-Hindawi, N.G. and Al-Shanawi, F.A. (2015):* Seroprevalence of *Toxoplasma gondii* and cytomegalovirus in aborted women in Baghdad-Iraq. *Iraqi journal of science*, 649-655.
- Ali, H.Z. and Al-Warid, H.S. (2021):* Changes in serum levels of lipid profile parameters and proteins in *Toxoplasma gondii* seropositive patients. *Iraqi Journal of Science*, 801-810
- Ali, S.; Zhao, Z.; Zhen, G.; Kang, J.Z. and Yi, P.Z. (2019):* Reproductive problems in small ruminants (sheep and goats): a substantial economic loss in the world. *Large Animal Review*, 25(6), 215-223.
- Al-Khafagi, A.M. and Zainab, R.Z. (2016):* Histopathological and diagnostic study of Toxoplasmosis in human and sheep by using ELISA in Kut city. *Iraqi J. Vet. Med*, 40(2), 94-9.
- Alkubaisi, S.A.M.S. and Al-Zubaidy, I.A.H.S. (2023):* Toxoplasmosis in females from Al-Anbar, Iraq. *Journal of the Faculty of Medicine Baghdad*, 65(1), 74-78.
- AL-Maamuri, S.D.; AL-Shanawi, F.A. and Melconian, A.K. (2014):* Seroprevalence of *Toxoplasma gondii* in Schizophrenic Patients in Iraq using ELISA test. *Iraqi Journal Science*, 55(3B), 1243-1248.
- Almeria, S. and Dubey, J.P. (2021):* Foodborne transmission of *Toxoplasma gondii* infection in the last decade. An overview. *Research in veterinary science*, 135, 371-385.
- Al-Taie, L.H. and Abdulla, S.H. (2011):* Seroprevalance of toxoplasmosis in sheep and goat: Iraq/Sulaimania. *The Iraqi Journal of Veterinary Medicine*, 35(1), 16-24.
- Amairia, S.; Rouatbi, M.; Rjeibi, M.R.; Nouasri, H.; Sassi, L.; Mhadhbi, M. and Gharbi, M. (2016):* Molecular prevalence of *Toxoplasma gondii* DNA in goats' milk and seroprevalence in Northwest Tunisia. *Veterinary Medicine and Science*, 2(3), 154-160.
- Asal, S.N. and Al Zubaidy, I.A. (2016):* Seroprevalance study of *Toxoplasma gondii* in horses and camels animal in Wasit province. *The Iraqi Journal of Veterinary Medicine*, 40(1), 177-182.

- Assim, M.M. and Saheb, E.J. (2018):* The association of severe toxoplasmosis and some cytokine levels in breast cancer patients. *Iraqi Journal of Science*, 1189-1194.
- Bawi, M.K and AL-ZUBAIDI, I.A.H. (2024):* Epidemiology of human toxoplasmosis by age, gender and location in Iraq. *Online Journal of Veterinary Research*. 28(6) 365-370.
- Camossi, L.G.; Greca-Júnior, H.; Corrêa, A.P.F.L.; Richini-Pereira, V.B.; Silva, R.C.; Da Silva, A.V. and Langoni, H. (2011):* Detection of *Toxoplasma gondii* DNA in the milk of naturally infected ewes. *Veterinary Parasitology*, 177(3-4), 256-261.
- Clune, T.; Beetson, S.; Besier, S.; Knowles, G.; Paskin, R.; Rawlin, G. and Jacobson, C. (2021):* Ovine abortion and stillbirth investigations in Australia. *Australian veterinary journal*, 99(3), 72-78.
- Daher, D.; Shaghilil, A.; Sobh, E.; Hamie, M.; Hassan, M.E.; Moumneh, M.B. and El Hajj, H. (2021):* Comprehensive overview of *Toxoplasma gondii*-induced and associated diseases. *Pathogens*, 10(11), 1351.
- Daryani, A.; Sarvi, S.; Aarabi, M.; Mizani, A.; Ahmadpour, E.; Shokri, A. and Sharif, M. (2014):* Seroprevalence of *Toxoplasma gondii* in the Iranian general population: a systematic review and meta-analysis. *Acta tropica*, 137, 185-194.
- Dawson, A.C.; Ashander, L.M.; Appukuttan, B.; Woodman, R.J.; Dubey, J.P.; Whiley, H. and Smith, J.R. (2020):* Lamb as a potential source of *Toxoplasma gondii* infection for Australians. *Australian and New Zealand Journal of Public Health*, 44(1), 49-52.
- De Santana Rocha, D.; De Sousa Moura, R.L.; Maciel, B.M.; Guimarães, L.A.; O'dwyer, H.N.; Munhoz, A.D. and Albuquerque, G.R. (2015):* Detection of *Toxoplasma gondii* DNA in naturally infected sheep's milk. *Genetics and Molecular Research*, 14(3), 8658-8662.
- Deyhimi, M.S.; Yousefidarani, H. and Soleimanifard, S. (2019):* Seroprevalence of *Toxoplasma gondii* in cattle and sheep in Isfahan, Iran. *Epidemiology and Health System Journal*, 6(3), 128-131.
- Diza, E.; Frantzidou, F.; Souliou, E.; Arvanitidou, M.; Gioula, G. and Antoniadis, A. (2005):* Seroprevalence of *Toxoplasma gondii* in northern Greece during the last 20 years. *Clinical microbiology and infection*, 11(9), 719-723.
- Dubey, J.P. (2016):* Toxoplasmosis of animals and humans. CRC press.
- Dubey, J.P. and Beattie, C.P. (1988):* Toxoplasmosis of animals and man (pp. 220-pp).
- Elaadli, H.; Abo El-Makarem, H.; Abd Elrahma, A.H.; Shaapan, R.M. and Bessat, M. (2023):* Prevalence and associated risk factors of *Toxoplasma gondii* infection in sheep and aborted women in Egypt. *Iraqi Journal of Veterinary Sciences*, 37(2), 437-445.
- Elfadaly, H.A.; Hassanain, N.A.; Shaapan, R.M.; Hassanain, M.A.; Barakat, A.M. and Abdelrahman, K.A. (2017):* Molecular detection and genotyping of *Toxoplasma gondii* from Egyptian isolates.
- Frenkel, J.K.; Dubey, J.P. and Miller, N.L. (1970):* *Toxoplasma gondii* in cats: fecal stages identified as coccidian oocysts. *Science*. 167 (3919): 893–896.
- Gharban, H.A. and Al-Kaabi, M.A. (2022):* Molecular and seroprevalence of toxoplasmosis in goats' blood and milk in Iraq. *Archives of Razi Institute*, 77(5), 1749.
- Hassanain, M.A.; Elfadaly, H.A.; Shaapan, R.M.; Hassanain, N.A. and Barakat, A.M. (2011):* Biological assay of *Toxoplasma gondii* Egyptian mutton isolates. *International Journal of Zoological Research*, 7(4), 330-337.

- Hill, D. and Dubey, J.P. (2002): *Toxoplasma gondii*: transmission, diagnosis and prevention. *Clinical microbiology and infection*, 8(10), 634-640.
- Ibrahim, H.M.; Mohamed, A.H.; El-Sharaawy, A.A. and El-Shqanqery, H.E. (2017): Molecular and serological prevalence of *Toxoplasma gondii* in pregnant women and sheep in Egypt. *Asian Pac. J. Trop. Med.* 10(10): 996–1001.
- Iqbal, A.; Arshad, M.; Elahi, A.; Hussain, K.; Kausar, M. and Javaid, Y. (2023): Epidemiology of Parasitic Diseases. In *Parasitism and Parasitic Control in Animals: Strategies for the Developing World* (pp. 40-55). GB: CABI.
- Ishaku, B.S.; Abdullahi, M.; Nalong, D.; Jonah, R. and Mayowa, O. (2018): Seroprevalence and risk factors for *Toxoplasma gondii* in pigs, sheep and goats at slaughter in Jos municipal abattoir, Nigeria. *Vet Sci Res Rev*, 4(2), 55-61.
- Karakavuk, M.; Can, H.; Çeltik, A.; Karakavuk, T.; Gül, C.; Erdem, H.A. and Döşkaya, A.D. (2024): Genetic characterization of *Toxoplasma gondii* strains isolated from humans living in İzmir, Türkiye. *Indian Journal of Medical Microbiology*, 49, 100571.
- Luptakova, L.; Benova, K.; Rencko, A. and Petrovova, E. (2015): DNA detection of *Toxoplasma gondii* in sheep milk and blood samples in relation to phase of infection. *Veterinary Parasitology*, 208(3-4), 250-253.
- Maggi, P.; Volpe, A.; Carito, V.; Schinaia, N.; Bind, S.; Basha, M. and Dentico, R. (2009): Surveillance of toxoplasmosis in pregnant women in Albania. *The new microbiologica*, 32(1), 89.
- Mahmood, S.; Ban, A.Q. and Zghair, K. (2013): Prevalence of toxoplasmosis of males blood donors in Baghdad. *Iraqi Journal of Science*, 54(4), 832-841.
- Mahmoud, M.A.; Ghazy, A.A. and Shaapan, R.M. (2021): Review of diagnostic procedures and control of some viral diseases causing abortion and infertility in small ruminants in Egypt. *Iraqi Journal of Veterinary Sciences*, 35(3), 513-521.
- McCall, J.; Rothfeldt, L.; Giesbrecht, K.; Hunt, A.; Bauck, L.; Scheftel, J. and Straily, A. (2022): Public Health Surveillance and Reporting for Human Toxoplasmosis-Six States, 2021. *Morbidity and Mortality Weekly Report*, 71(28), 889.
- Motoi, S.; Navolan, D.B.; Malita, D.; Ciohat, I.; Nemescu, D.; Manciu, C. and Dobrescu, A. (2020): A decreasing trend in toxoplasma gondii seroprevalence among pregnant women in Romania-results of a large scale study. *Experimental and Therapeutic Medicine*, 20(4), 3536-3540.
- Muñoz-Zanzi, C.; Tamayo, R.; Balboa, J. and Hill, D. (2012): Detection of oocyst-associated toxoplasmosis in swine from southern Chile. *Zoonoses and Public Health*, 59(6), 389-392.
- Musa, I.S.; Saheb, E.J. and Kuba, R.H. (2021): Toxoplasmosis and Its Potential Role to Change the Levels of C-reactive protein and Vitamin D3 in Atherosclerosis Patients. *Iraqi Journal of Science*, 1787-1792.
- Pereira, K.S.; Franco, R.M. and Leal, D.A. (2010): Transmission of toxoplasmosis (*Toxoplasma gondii*) by foods. *Advances in food and nutrition research*, 60, 1-19.
- Punda-Polić, V.; Tonkić, M. and Čapkun, V. (2000): Prevalence of antibodies to *Toxoplasma gondii* in the female population of the County of Split Dalmatia, Croatia. *European journal of epidemiology*, 16, 875-877.
- Robert-Gangneux, F. and Dardé, M.L. (2012): Epidemiology of and diagnostic strategies for

- toxoplasmosis. *Clin. Microbiol. Rev.* 25 (2): 264–96.
- SAS. (2018): Statistical Analysis System, User's Guide. Statistical. Version 9.6th ed. SAS. Inst. Inc. Cary. N.C. USA.
- Segbedzi, C.E.; Ansah, E.W. and Apaak, D. (2023): Compliance to food safety standards-Determining the barriers within the hotel industry. medRxiv, 2023-12.
- Shaapan, R.M.; El-Nawawi, F.A. and Tawfik, M.A.A. (2008): Sensitivity and specificity of various serological tests for the detection of *Toxoplasma gondii* infection in naturally infected sheep. *Veterinary Parasitology*, 153(3-4), 359-362.
- Shaapan, R.M.; Toaleb, N.I. and Abdel-Rahman, E.H. (2015): Significance of a common 65 kDa antigen in the experimental fasciolosis and toxoplasmosis. *Journal of Parasitic Diseases*, 39, 550-556
- Sharif, M.; Sarvi, S.; Shokri, A.; Hosseini Teshnizi, S.; Rahimi, M.T.; Mizani, A. and Daryani, A. (2015): *Toxoplasma gondii* infection among sheep and goats in Iran: a systematic review and meta-analysis. *Parasitology research*, 114, 1-16.
- Stelzer, S.; Basso, W.; Silván, J.B.; Ortega-Mora, L.M.; Maksimov, P.; Gethmann, J. and Schares, G. (2019): *Toxoplasma gondii* infection and toxoplasmosis in farm animals: Risk factors and economic impact. *Food and waterborne parasitology*, 15, e00037.
- Stopić, M.; Štajner, T.; Marković-Denić, L.; Nikolić, V.; Djilas, I.; Srzentić, S.J. and Bobić, B. (2022): Epidemiology of toxoplasmosis in Serbia: a cross-sectional study on blood donors. *Microorganisms*, 10(3), 492.
- Studeníčová, C.; Benčaiová, G. and Holková, R. (2006): Seroprevalence of *Toxoplasma gondii* antibodies in a healthy population from Slovakia. *European journal of internal medicine*, 17(7), 470-473.
- Subedi, S.; Sharma, B.; Singh, S. and Bindari, Y.R. (2018): Seroprevalence of *Toxoplasma gondii* in sheep in different geographical regions of Nepal. *Vet. Anim. Sci.* 5 7–9.
- Toaleb, N.I.; Shaapan, R.M.; Hassan, S.E. and El-Moghazy, F.M. (2013): High diagnostic efficiency of affinity isolated fraction in camel and cattle toxoplasmosis.
- Vieira da silva, A.R.I.S.T.E.U.; De Oliveira Mendonça, A.N.D.R.É.; Bergamaschi Pezerico, S.A.N.D.I.A.; Domingues, P.F. and Langoni, H. (2005): Genotyping of *Toxoplasma gondii* strains detected in pork sausage. *Parasitología latinoamericana*, 60(1-2), 65-68.
- Weiss, L.M. and Dubey, J.P. (2009): Toxoplasmosis: A history of clinical observations. *International journal for parasitology*, 39(8), 895-901.
- World Health Organization. (2015): WHO estimates of the global burden of foodborne diseases: foodborne disease burden epidemiology reference group 2007-2015. World Health Organization.

الكشف المصلي والجزئي لطفيل التوكسوبلازما جوندي *TOXOPLASMA GONDII* في الأغنام في محافظة ديالى، العراق

محمد خليل ، إبراهيم الزبيدي

Email: mohammed.bawi2204m@covm.uobaghdad.edu.iq Assiut University web-site: www.aun.edu.eg

يعد طفيل التوكسوبلازما جوندي هو السبب الشائع لداء المقوسات في كل من البشر والحيوانات. ويمثل داء المقوسات في الأغنام دوراً هاماً في الصحة العامة، حيث يسبب خسائر إنجابية واقتصادية بسبب الإجهاض ووفيات الأطفال حديثي الولادة عالمياً. كان الهدف من هذه الدراسة هو الانتشار المصلي والكشف الجزئي لداء المقوسات في الأغنام في محافظة ديالى، العراق. تم فحص مائة خروف من المسالخ و ١٠٠ نعجة حلوب وتم تقسيمها إلى مجموعات حسب العمر والموسم والسلالة والاستيراد ومخالطة القطط وحالة الحمل والإجهاض. تم التحقيق في عدوى التوكسوبلازما باستخدام اختبار التوكسوبلازما السريع واختبار تفاعل البلمرة المتسلسل. بلغت نسبة الانتشار المصلي لداء المقوسات في الكباش ١٨%، في حين بلغت ١٤% في النعاج الحلوب. أكد اختبار تفاعل البلمرة المتسلسل الإصابة الإيجابية للمصل في ٥٠% و ٣٥,٧% من الكباش والنعاج على التوالي. كان لدى الكباش المجازر والنعاج البالغة المنتجة للألبان معدل انتشار مصلي لداء المقوسات أكبر بشكل غير ملحوظ إحصائياً مقارنة بالأحداث. كان انتشار المرض أعلى قليلاً في المجموعات الشتوية سواء في ذكور الأغنام أو النعاج. وشهدت الأغنام المستوردة من سوريا وإيران معدل إصابة أعلى بمرتين من الأغنام المحلية من العراق. لم يكن معدل الإصابة بين النعاج الحلوب من سلالاتي الكرادي والهمداني أكبر بكثير من السلالات الأخرى. أظهرت الأغنام التي لديها تاريخ من الإجهاض معدل انتشار مصلي أعلى، كما وجد أيضاً أن داء المقوسات أكثر انتشاراً في الأغنام المختلطة سابقاً مع القطط. ولذلك، يوصى بشدة بالمراقبة المستمرة للكشف عن داء المقوسات في الأغنام لمزيد من الوقاية.