Egyptian Vegetables as Source for Lutein and its Role in Incidence of Cataract in Rats

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ABSTRACT

The last years the interesting of eye diseases increases and the relationship intervention with nutrients and the common diseases as well cataract. The aim of this study focus on the concern of the effect of lutein on cataract. Therefore, this research was designed to study the use of certain species of vegetables (spinach, leek, parsley and watercress), the most popular and rich in lutein and its relationship to reduce the incidence of cataract. It could be concluded that there is a relationship between lutein consumption and decreasing the risk of cataract. We must eat the fresh vegetables rich in vitamins specially lutein. It could was found that watercress, spinach, parsley and leek provide high content of lutein (12.72, 11.87, 11.03 and 7.30 mg /100g), respectively. Moreover these vegetables could be purchased with low price and available throughout the year. It is noticed that 10 mg lutein daily was effective dose for the beneficial effects of lutein on cataract. Also, it is noticed that feeding on lutein diets increased the serum antioxidant enzymes (SOD, GPx, GR, CAT and GSH) and reduced the proteases activities in rats lenses compared with cataract group and thus its role in reducing the incidence of cataract.

INTRODUCTION

Lutein is found in natural sources like squash, peaches, mangoes, papayas, yellow and dark green fruits and vegetables including carrots, apricots, cantaloupe, sweet potatoes, spinach, kale, broccoli and mustard greens (BCIEP, 1994).

Kale, turnip greens, spinach, collard greens, watercress, garden peas, broccoli, egg, carrot and avocado are considered good sources of lutein and zeaxanthin (SanGiovanni, *et al.*, 2007).

Lutein belongs to the xanthophyll family of carotenoids, which are synthesized on dark green leafy plants, such as spinach and kale (Subczynski, *et al.*, 2010).

USDA, (2015) stated the carotenoids composition of spinach as follows: $5626 \ \mu g \beta$ -carotene and 12197 $\ \mu g$ Lutein+Zeaxanthin (on fresh weight basis).

Perry, *et al.*, (2009) reported that leek contained 36.8 µg lutein /g fresh weight. Lutein and zeaxanthin are the most common xanthophylls in green leafy vegetables (e.g., kale, leek, spinach, broccoli, peas and lettuce) and egg yolks.

USDA, (2015) stated the carotenoids composition of leek as follows: $1000 \ \mu g \beta$ -carotene and $1900 \ \mu g Lutein + Zeaxanthin (on fresh weight basis).$

Parsley is a source of flavonoides, and antioxidants, apigenin, (Meyer, *et al.*, 2006) folic acid, vitamin K, vitamin C, and vitamin A. Half a of tablespoon (a gram) of dried parsley contains about $6.0 \ \mu g$ of lycopene and $10.7 \ \mu g$ of alpha carotene well as $82.9 \ \mu g$ of lutein+zeaxanthin and $80.7 \ \mu g$ of beta carotene (USDA, 2013).

USDA, (2015) stated the carotenoids composition of parsley as follows: $5054 \ \mu g \ \beta$ -carotene and $5561 \ \mu g \ Lutein+Zeaxanthin$ (on fresh weight basis).

(Food Standards Agency, 2002) told that watercress contained 18 Kcal/100g, iron, calcium, were: 1.6, 138 respectively and 2016 μ g β -carotene, 4614 μ g Lutein+Zeaxanthin.

Cataracts is a white cloud affect one or both eyes. Symptoms include blurry vision, difficulty seeing

at night, faded colors, and trouble with the bright lights and halos around light (NEI, 2015). Cataracts are the cause of half of blindness and 33% of poor vision worldwide (WHO, 2015).

Despite the good efficacy of surgical protocols for treating cataracts, there are limitations such as cost, time of diagnosis and inadequate service in some countries which decrease treatment outcome and leads to cataracts-induced inability and blindness (Miller, *et al.* 2005). Surgical intervention for the treatment of cataract is expensive and may be unavailable, so prevention is the use of foods rich in antioxidants can reduce the risk of cataracts. However, as far as lutein is concerned, interventional studies suggest that it might be effective against nuclear cataract but no other kinds of cataract (Ma *et al.*, 2014).

Many studies confirm that the profusion of lutein reduces the incidence of cataracts (SanGiovanni, *et al.*, 2007). Likely in studies that the effective dose in reducing the incidence of cataracts is 6 mg of lutein per day and the dose most commonly used in commercial products is 10 mg / day. Although the optimal dose of lutein supplementation has not been proven yet. (Harikumar, *et al.*, 2008).

Lutein is found in the lens of the human eye and they have two main functions there – as an antioxidant to reduce or scavenge free radicals and as a filter against high-energy and harmful blue light (*Landrum and Bone, 2001*). Exposure of the eye to high-energy blue light, results in free radical formation and oxidative stress (*Krinsky, et al., 2003*). Lutein by filtering the hurtful blue light reduces photo-induced oxidation of lens proteins thereby protecting against age-related eye diseases such as cataract.

So the main goal of this work was to study Egyptian vegetables that are sources for lutein and its role in cataract in rats.

MATERIALS AND METHODS

Materials:

Raw vegetables:

Spinach (Spinacia oleracea L.), Leek (Allium ampeloprasumvar. Porrum), Parsley (Petroselinum crispum), and Watercress (Nasturtium officinale) were obtained from local market in Mansoura city, El-Dakahlia, Egypt.

Standard lutein:

Standard lutein was purchased from Sigma Aldrich co., U.S.A.

Sodium selenite:

Sodium selenite was purchased from Sigma Aldrich co., U.S.A.

Methods:

Preparation of raw vegetables:

The vegetables were dried by oven according to Mary, 1994.

Determination of lutein (HPLC):

Lutein was determined in National Research Center (NRC), Giza, Egypt using HPLC.

Extraction of lutein:

high-performance By using liquid chromatography diode array detector lutein was extracted and analyzed. For 15 hours with 10 ml of methanol: tetrahydrofuran (1: 1, v / v), and for another 10 minutes at room temperature two hundred milligrams of the sample was extracted. And then subjecting the extract to room temperature. Then, the liquidation cruised through a filter 0.20 micron membrane and kept in the dark under nitrogen until HPLC analysis of lutein.

HPLC conditions:

Hentschel, et al., (2002), HPLC separation was accomplished according to a previously described protocol with modifications. HPLC analysis was performed using an Agilent 1100 HPLC system (Agilent Technologies, Palo Alto, CA). An Agilent 1100 series liquid chromatography (Agilent Technologies, Waldbronn, Germany), consisting of a vacuum solvent degassing unit,(a quaternary gradient pump), an automatic sample injector. Column: C18 5µm, 150mm×4.6mm i.d. Chromatogram were monitored at 450nm . In this process it was used at room temperature. lutein was eluted using a mobile phase of solvent A (hexane) and solvent B (1% i-PrOH in EtOAc). The injected volume was 10 µl.

Biological evaluation:

Experimental animals:

The experimental rats, (suckling white albino rats, Sprague Dawley strain) according to the following were obtained from the design. Albino rats experimental animal house of the Research National Center, Giza, Egypt

Experimental design:.

A number of 56 rats with a body weight of 20 to 55 g were divided into 8 groups each comprised 7 rats. Each rat in each group was housed individually into separate stainless steel cage in a room at temperature of 25°C. Water was admitted freely to rats from glass bottles mounted. Each group of rats was given one of the prepared diets. Group 1(control negative) and 2 (control positive) were given the control diet the others were given other formula . Part of the food provided to groups from G2 to G8 contained vegetable dryer has a recommended daily allowance of lutein (10 mg) and then completed the rest of the quantity by control diet.

Then, rats in group 1 to group 8 were fed as follows:

- G1- Rats fed on basal diet (Negative control or normal control).
- G2- Rats fed on basal diet (Positive control or cataract group).
- G3- Rats fed on spinach.
- G4- Rats fed on leek .
- G5- Rats fed on parsley.
- G6- Rats fed on watercress.
- G7- Rats fed on mixed vegetables (25% each).
- G8- Rats fed on mixed vegetables (25% each).

G2 to G7 were selenite cataract model from the first day while the G8 was injected by selenite after 15 days from the start of the experiment. An amount of food equal to 20 g was weighed and placed in the dish inside the cage. This was allowed for consumption over the day. Animals were weighted twice a week and the weights of the animal were recorded to follow their growth . The animal experiment lasted for 4 weeks.

Samples Collection:

At the end of experiment (one month), the rats were fasted overnight, anaesthetized and blood samples were withdrawn from the eye vein. Serum was prepared and kept in deep freezer at -30°C until used assessment of different biochemical parameters .The eyes were enucleated and lenses were excised, washed in saline solution and kept in 10% formalin.

Cataract model:

Sodium selenite used in studies and research since 1978, and a quick easy way to cataracts in rats (Ostadalova et al., 1978). suckling rats are injected under the skin at a rate of 19-30 µM/kg of body weight of sodium selenite (Shearer et al., 1997). Repeated injections of smaller doses of selenite (Huang et al., 1992) or oral administration (Shearer et al., 1983) are also cataractogenic.

Biochemical analysis:

Determination of serum triglycerides:

An enzymatic colorimetric method according to (Fassati and Prencipe, 1982).

Determination of total cholesterol:

The kits were provided form Biodiagnostic according to (Allain et. al., 1974).

Determination of high density lipoprotein cholesterol (HDL-c):

An enzymatic colorimetric method according to (Lopez, 1977).

Determination of LDL-cholesterol and vLDL-c:

LDL-cholesterol and vLDL-c in serum were performed following the method of (Lopez et.al., 1977). **Determination of lipid peroxide (MDA):**

Lipid peroxide was determined according to the method of (Satoh, 1978).

Determination of Proteolytic (proteases) activity:

Proteases activity was determined according to (Schoenberger, 1987).

Determination of catalase activity (CAT):

Catalase activity was determined according to the method of (Aebi, 1984).

Determination of reduced glutathione (GSH):

Reduced glutathione was determined according to the method of (Beutler, et al., 1963).

Determination of glutathione reductase activity:

Reductase glutathione was determined according to the method of (Goldberg and Spocner, 1983).

Determination of glutathione peroxidase (GPx) activity:

Glutathione peroxidase activity was determined according to the method of (Hafemann, *et al.*, 1974).

Determination of superoxide dismutase (SOD) activity:

Superoxide dismutase activity was determined according to the method of (Mc Cord and Fridovich, 1969).

Statistical analysis:

140

100

60

20

The data were subjected to statistical analysis using one way classification least significant differences (L.S.D) according to (Steel and Torrie, 1980).

Histopathological examinations:

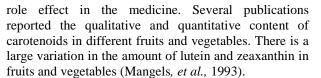
utein

The eyes were enucleated and lenses were excised, washed in saline solution and kept in 10% formalin, dehydrated in a graded ethanol series, cleared in xylene and embedded in paraffin wax. Then, sections (5 μ m thick) were stained with haematoxylin and eosin (H&E) (*Bancroft and Gamble, 2002*).

RESULTS AND DICSSIONS

Fruits and vegetables are the most important source of carotenoids in the diet, and the preventive

Figures of lutein HPLC chromatogram of raw vegetables.



So, at first, raw vegetables (spinach, leek parsley and watercress) lutein ratio must be determined as shown in table (1). From these data, it could noticed that watercress had the highest value of lutein and it was 12.72 mg /100g followed by spinach, parsley and leek as follows: 11.87, 11.04 and 7.31 mg /100g, respectively. These results go in parallel with those reported by (USDA, 2015).

These vegetables are available in the Egyptian market and is also cheap and easy in reach of the average consumer, who does not realize its potent effect of lutein and cataract.

Table 1. lutein ratio in raw vegetables

Vegetables	Lutein (mg/100g FW)
Spinach	11.87
Leek	7.31
Parsley	11.04
Watercress	12.72

Lutein concentration in serum for all groups of rats to know the lutein absorption as a result of eating vegetables rich in lutein was determined.

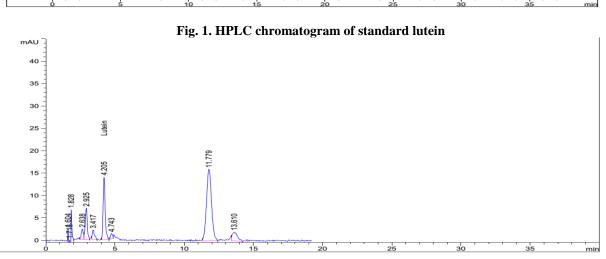


Fig. 2. HPLC chromatogram of lutein in spinach

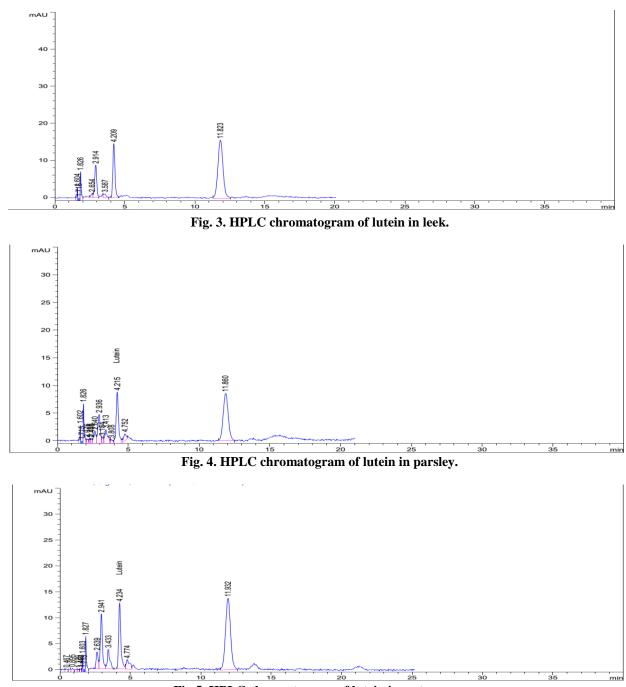


Fig.5. HPLC chromatogram of lutein in watercress.

Results in table (2) showed that lutein determined in blood serum in all groups of animals from group 1 to group 8 approximately were : (0.128), (0.114), (0.222), (0.218), (0.214), (0.211), (0.218) and (0.218) ug/ml serum, respectively. It could be noticed that G3 (spinach group) contained the highest level of lutein (0.222 ug/ml serum) while G2 (control positive) had the lowest level of lutein (0.114 ug/ml serum), This result shows that the oxidative stress reduces the bioavailability of lutein as a result of the presence of sodium selenite in group G2 (control positive) which had 0.114 ug/ml serum, while increasing the bioavailability of lutein as a result of its rich vegetable intake in groups from G3 to G8, the highest value in G3 (spinach group) which had 0.222 ug/ml serum, while the lowest value in G6 (watercress group) which had

0.211ug/ml serum. These results go in agreement with those found by (Leela *et al.*, 2014 and Mamatha and Baskaran 2011).

 Table 2.Lutein concentration in different groups of rats serum

Group No.	Lutein concentration(ug/ml Serum)		
G1 (Negative control)	0.128385096		
G2 (Positive control)	0.114007448		
G3 (spinach)	0.221551695		
G4 (leek)	0.218241766		
G5 (parsley)	0.214397977		
G6 (watercress)	0.210927891		
G7 (mixed vegetables)	0.218348538		
G8 (mixed vegetables)	0.217707906		

Long-term lutein supplementation could increase serum lutein concentration, macular pigment optical density (MPOD), and visual sensitivity in early agerelated macular degeneration (AMD) patients. The advisable long-term lutein dosage for early AMD treatment is 10 mg daily (*Huang et al.*, 2015).

In a research by (*Berendschot et al., 2000*) the influence of daily consumption with 10 mg lutein derived from marigold during 12 weeks on macular pigment density was investigated. This study also showed that plasma lutein concentrations reached a plateau after 4 weeks. Mean plasma lutein concentration increased from 0.18 to 0.90 μ mol/L within these 4 weeks and stayed at this level during the supplementation period.

Influence of feeding rats lutein diets on antioxidants enzymes is an important parameter when studying the effect of lutein on cataract.

There exists a group of oxygen eliminators in the lens, including reduced glutathione (GSH), superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (Gpx), glutathione reductase (GR), and glutathione S-transferase (GST), protecting crystallins from oxidative damage (Manikandan, et al., 2010). Selenite-induced cataract is a cataract model that is causally related to oxidative stress, where oxidation of the critical sulfhydryl groups is essential for the initiation of cataractogenesis. Primary defenses, including non enzymatic antioxidants and enzymatic antioxidants such as glutathione, SOD, CAT, GPx, GR, and GST, neutralize free radicals and repair, recover, or degrade molecules that are damaged (Shearer, et al., 1997). It has been demonstrated that antioxidant enzymes levels are altered in cataracts. Some reports showed that the activity of SOD, GSH, GPx, and CAT decreased in cataract (Ozmen, et al., 2002).

Effect of lutein on the antioxidant enzymes and glutathione in the serum of rats after administration for a period of 30 days is shown in Table (3).

The activity of superoxide dismutase (SOD), glutathione perodxidase (GPx), glutathione reductase (GR), catalase (CAT), and glutathione (GSH) were significantly increased in all groups of animals which treated with 10 mg lutein daily when compared with treated control (+) group G2 (selenite-induced cataract group).

Table 3. Effect of feeding lutein diets on antioxidants enzymes

Table 5. Effect of feeding futerin tiles on antioxidants enzymes							
GROUP	SOD(U/ml)	GPx(U/l)	GR(U/I)	Cat(U/l)	GSH(mg/dl)		
G1	32.57±4.96 ^a	179.29±20.70 ^a	210.00±15.81ª	43.40±7.32	29.43±8.06 ^a		
G2	18.86 ± 2.04^{d}	142.86±17.53°	175.71±25.40 ^b	30.57±5.35	20.29 ± 2.69^{d}		
G3	21.43±5.38 ^{cd}	148.57±17.96°	192.86±23.95 ^{ab}	38.57±9.11	22.17±3.18 ^{dc}		
G4	19.14±3.13 ^d	166.43±8.52 ^{ab}	190.00±13.23 ^{ab}	41.71±8.36	25.29±4.35a ^{bcd}		
G5	21.00±3.42 ^d	156.00±14.55bc	181.00±18.12 ^b	36.86±8.59	27.71±4.54 ^{ab}		
G6	19.71±3.35 ^d	169.29±13.97 ^{ab}	190.71±12.72 ^{ab}	32.57±6.65	23.86±3.58 ^{bcd}		
G7	26.14±5.08 ^{bc}	172.14±7.56 ^{ab}	188.57±18.19 ^b	38.33±9.07	26.14±4.26 ^{abc}		
G8	27.00±6.30 ^b	169.17±20.09 ^{ab}	187.86±19.33 ^b	37.14±7.69	25.40±6.67 ^{abcd}		
P value	0.0001	0.0008	0.0790	0.0668	0.0322		
LSD	4.7444	16.986	20.246	8.4494	5.3371		
$a_{\rm r}f_{\rm r}$ = Means with the same latter in each column are not significantly different P<0.05							

a-f = Means with the same letter in each column are not significantly different P≤0.05. LSD = Least Significant Difference SOD: Superoxide dismutase GPx: Glutathione peroxidase GR: Glutathione reductase Cat: Catalase GSH: reduced glutathion

The data in table (3) showed that G2 (seleniteinduced cataract group) has the lowest values in SOD, GPx, GR, CAT and GSH (18.86 ± 2.04^{d} U/ml, 142.86 ± 17.53^{c} U/l, 175.71 ± 25.40^{b} U/l, 30.57 ± 5.35 U/l and 20.29 ± 2.69^{d} mg/dl), respectively while G1 has the highest values (32.57 ± 4.96^{a} U/ml, 179.29 ± 20.70^{a} U/l, 210.00 ± 15.81^{a} U/l, 43.40 ± 7.32 U/l and 29.43 ± 8.06^{a} mg/dl), respectively.

It noticed that the values of SOD, GPx, GR, CAT and GSH were normal in G1(control negative), and then decreased in G2 (control positive) this is may be due to oxidative stress as a result of doses of sodium selenite then these values increased again in the remaining groups from G3 to G8 as a result of eating vegetables rich in lutein content.

These results are in parallel with those obtained by Seham, *et al.*, (2013), Shearer, *et al.*, (1997), Ozmen, *et al.*, (2002), and Chang, *et al.*, (2013).

Proteolytic (proteases) enzymes activity also important criteria when identifying the nature of the relationship between lutein and cataract. Cataract is accompanied by low lens protein (Kuck, J. and Kuck, K. 1983). This low protein content could partly be due to loss by proteolysis, and the evidence presented may suggest proteolysis due to increased enzyme activities in cataract lenses. This has also been shown in previous studies for human cataract lenses (Swanson, *et al.*, 1981).

(Gao, *et al.*, 2011), have shown that lutein or zeaxanthin supplementation protects lens protein, lipid and DNA from oxidative damage and improves intracellular redox status upon oxidative stress.

The data tabulated in table (4) showed that the G2 control positive (selenite-induced cataract group) showed the highest value it was $(3.200\pm0.798^{a} \text{ U/l})$, while G1 (control negative) was the lowest value of proteases activity $(1.414\pm0.414^{b} \text{ U/l})$. The mean activities of proteases was significantly increased in G2 cataract group (sodium selenite induced group) of animals. While, the enzyme activities were restored decreased on lutein treatment (G3 to G8). From these data ; it could be noticed that feeding on lutein diets reduced the proteases activities in rat lenses.

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Table 4. Proteases activity in rats lenses

Table 4.110 cases activity in rats relises				
Group No.	Proteases(U/l)			
G1 (Negative control)	1.414 ± 0.414^{b}			
G2 (Positive control)	3.200 ± 0.798^{a}			
G3 (spinach)	2.871±0.692ª			
G4 (leek)	2.029 ± 0.860^{b}			
G5 (parsley)	1.443±0.591 ^b			
G6 (watercress)	2.014 ± 0.626^{b}			
G7 (mixed vegetables)	1.729 ± 1.106^{b}			
G8 (mixed vegetables)	2.943±0.577 ^a			
P value	0.0001			
LSD	0.79			
a-f = Means with the same	letter in each column are not			

significantly different P≤0.05.

LSD = Least Significant Difference

These results are in parallel with those obtained by Gao, et al., (2011), Shearer, et al., (1997) and Swanson, et al., (1981).

Histopathological and microscopic examinations of lens show very clearly the absence of any change in the tissues of eyes and lenses in the first group G1(control negative ; untreated) normal (Fig. 1), while many changes (Fig, 2,3 and 4) have emerged in the second group G2 control positive (selenite induced cataract group). On other hand the rest of the groups (from G3 to G8), some of them had been influenced by factors oxidation and others had a protection from oxidation and various pathological changes are heading for a lutein.

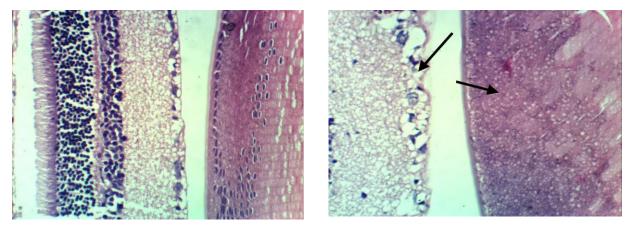
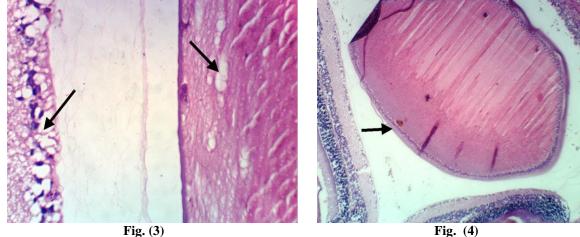


Fig. (1)





Finally, it could be concluded that there is a relationship between lutein consumption and decreasing the risk of cataract. We must eat the fresh vegetables rich in vitamins specially lutein. It could be also concluded that watercress, spinach, parsley and leek provide high content of lutein which were (12.72, 11.87, 11.03 and 7.30 mg /100g), respectively. Moreover these vegetables could be purchased with low price and available throughout the year

REFERANCES

- Aebi, H. (1984). Catalase in vitro. Methods Enzymol. (105): 121-126.
- Allain, C. C.; Richmond, N. and Rosechloy, P. (1974). Cholesterol enzymatic colormetric test. Clin. Chem., 19(20): 1350-1450.
- Bancroft, J. and Gamble, M. (2002): Theory and practice of histological techniques. 5th ed. Churchill Livingstone, Oxford, 139-162.

- BCIEP, (1994). Breast Cancer Information Exchange Project . Guide to unconventional cancer therapies. 1st ed. Toronto: Ontario Breast Cancer Information Exchange Project, 123-124.
- Berendschot, T.T.; Goldbohm, R.A.; Klopping, W.A.; van de Kraats, J.; van Norel, J. and van Norren, D. (2000). Influence of lutein supplementation on macular pigment, assessed with two objective techniques. Invest Ophthalmol. Vis. Sci., (41): 3322-3326.
- Beutler, E., Duron, O. and Kelly, M. (1963). Improved method for the determination of blood glutathione. J. Lab. Clin. Med., 61:882-8.
- Chang, D. ; Zhang, X. ; Rong, S. ; Sha, Q. ; Liu, P. ; Han, T. and Pan, H. (2013). Serum Antioxidative Enzymes Levels and Oxidative Stress Products in Age-Related Cataract Patients. Oxidative Medicine and Cellular Longevity Volume (2013), 7 pages.
- Fassati, P. and Precipe, L. (1982). Serum triglycerides determined colorimeteric-ally with an enzyme that produces hydrogen peroxide. Clin. Chem., (28): 2077-2080.
- Food Standards Agency (2002). McCance and Widdowson's The Composition of Foods, 6th Summary Edition. Cambridge: Royal Society of Chemistry.
- Gao, S.; Qin, Z.; Liu, M.; Caceres, C.; Ronchi, C.; Chen, K.; Yeum, A. Taylor, J.; Blumberg, Y and Shang, F. (2011). Lutein and zeaxanthin supplementation reduces H2O2-induced oxidative damage in human lens epithelial cells. Mol. Vis., (17): 3180-3190.
- Goldberg, C.M. and Spocner, J. (1983) . Methods of Enzymatic. Aruylg. Brgtn. ysn., (3): 258-265.
- Hafemann, D., Sunde, R. and Houestra, W. (1974). Effect of dietary selenium on erythrocyte and liver glutathione peroxidase in the rat. J. Nutr., (104): 580-584.
- Harikumar, K.B.; Nimita, C.V.; Preethi, K.C.; Kuttan, R.; Shankaranarayana, M.L. and Deshpande, J. (2008). Toxicity profile of lutein and lutein ester isolated from marigold flowers (Tagetes erecta). Int. J. Toxicol., (27): 1-9.
- Hentschel, V.; Kranl, K.; Hollmann, J.; Lindhauer, M. G.; Bohm, V. and Bitsch, R. (2002). Spectrophotometric determination of yellow pigment content and evaluation of carotenoids by highperformance liquid chromatography in durum wheat grain. J. Agric. Food Chem., (50): 6663-6668.
- Huang, Y.M.; Dou, H.L.; Huang, F.F.; Xu, X.R.; Zou, Z.Y. and Lin, X.M. (2015). Effect of supplemental lutein and zeaxanthin on serum, macular pigmentation, and visual performance in patients with early age-related macular degeneration. Biomed Res. Int., (2015): 564738.
- Krinsky, N. ; Landrum, J. and Bone, R. (2003). Biologic mechanisms of the protective role of lutein and zeaxanthin in the eye. Annu. Rev. Nutr., (23): 171-201.

- Kuck, J. and Kuck, K. (1983). The Emory Mouse cataract: loss of soluble protein, glutathione, protein sulfhydryl and other changes. Exp. Eye. Res., (36): 35.
- Landrum, J. and Bone, R. (2001). Lutein, zeaxanthin, and the macular pigment. Arch. Biochem. Biophys., (385): 28-40.
- Leela, S. ; Sunil, K. ; Dinesha, R. and Shankaranarayanan, J. (2014).Comparative single dose oral pharmacokinetics studies of lutein formulations in male wister albino rats . Asian Journal of Phytomedicine and Clinical Research. 2(3): 148 - 154.
- Lopez- Virella, M.F. (1977). High density lipoprotein cholesterol by selective precipitation. Clin. Chem.23(5): 882-884.
- Ma, L. ; Hao, Z. and Liu, R. (2014). A dose–response meta-analysis of dietary lutein and zeaxanthin intake in relation to risk of age-related cataract. Graefes Arch. Clin. Exp. Ophthalmol., (252): 63-70.
- Mamatha, B. and Baskaran, V. (2011). Effect of micellar lipids, dietary fiber and beta-carotene aged rats with lutein deficiency. Nutrition, (27): 960-966.
- Mangels, A.R. ; Holden, J.M. and Beecher, G.R. (1993). Carotenoid content of fruits and vegetables: an evaluation of analytical data. J. Am. Diet Assoc., (93): 284-296.
- Manikandan, R. ; Thiagarajan, R. ; Beulaja, S. ; Sudhandiran, G. and Arumugam, M. (2010). Effect of curcumin on selenite-induced cataractogenesis in Wistar rat pups. Curr. Eye Res., (35): 122-129.
- Mary, K., (1994). Drying Foods. A circular of Cooperative Extension Service University of Illinoise at Urbana Champaign College of Agriculture; 1994, 1227.
- McCord, J. and Fridovich, (1969). Superoxide dismutase. An enzymic function for erythrocuprein (hemocuprein). J. Biol. Chem., 244(22): 6049-6055.
- Meyer, H. ; Bolarinwa, A. ; Wolfram, G. , and Linseisen, J. (2006). Bioavailability of apigenin from apiin-rich parsley in humans. Annals of Nutrition and Metabolism, 50(3): 167-172.
- Miller, J. ; Scott, I. and Flynn, J. (2005). Acute-onset endophthalmitis after cataract surgery (2000– 2004): incidence, clinical settings, and visual acuity outcomes after treatment. Am. J. Ophthalmol., (139): 983-987.
- NEI, (2015). The National Eye Institute (NEI) is part of the National Institutes of Health (NIH). Facts About Cataract.
- Ostadalova, I. ; Babicky, A. and Obenberger, J. (1978). Cataract induced by administration of a single dose of sodium selenite to suckling rats. Experientia, (34): 222–223.
- Ozmen, B. ; Ozmen, D. and Erkin, E. (2002). Lens superoxide dismutase and catalase activity . Clinical Biochemistry, 35(1): 69-72.

- Perry, A. ; Rasmussen, H. and Johnson, E.J. (2009). Xanthophyll (lutein, zeaxanthin) content of fruits, vegetables and corn and egg products. J. Food Comp. Anal, (22): 9-15.
- SanGiovanni, J.P.; Chew, E.Y. and Clemons, T. (2007). The relationship of dietary carotenoid and vitamin A, E, and C intake with age-related macular degeneration in a case-control study: AREDS Report No.22 . Arch. Ophthalmol., 125(9): 1225-1232.
- Satoh, K. (1978). Serum lipid peroxide in cerebrovascular disorders determined by a new colorimetric method. Clin. Chim. Acta.; 90(1): 37-43.
- Schoenberger, O. (1987). Proteolytic activity of human tumor cell lines deriving from bronchial squamous cell carcinoma, pulmonary metastasis of rhabdomyosarcoma and pleural metastasis of mesothelioma. Eur. J. Respir. Dis., (71): 434-443.
- Seham, S. ; Margreet, A. ; Nora, M. ; Tahany, E. ; Mohamed S. ; Anhar, M. ; Fatma, H. and Hasnaa H. (2013). Modulation of Selenite-Induced Cataract by Dietary Supplement of Broccoli in Experimental Animals. World Applied Sciences Journal, 26(12): 1643-1652.

- Shearer TR, Anderson RS and Britton JL. (1983). Infl uence of selenite and fourteen trace elements on cataractogenesis in the rat. Invest. Ophthalmol. Vis. Sci., (24): 417-423.
- Shearer, T.R. ; Ma, H. ; Fukiage, C. and Azuma, M. (1997). Selenite nuclear cataract: review of the model. Mol. Vis., (3): 8-15.
- Steel, R.G. and Torrie, T.H. (1980). Principles and procedures of statistics. A biometrical approach. Mc Grow Hill Book Comp., Inc. New York, U.S.A.
- Subczynski, W.K. ; Wisniewska, A. and Widomska, J. (2010). Location of macular xanthophylls in the most vulnerable regions of photoreceptor outersegment membranes. Arch. Biochem. Biophys., (504): 61-66.
- Swanson, A. ; Davis R. ; Albers-Jackson, B. and McDonald, J. (1981). Lens exopeptidases. Exp. Eye Res., (32): 163-173.
- USDA, (2013). United States Department of Health and Human Services. Nutritional Data, Parsley, SR-21.
- USDA, (2015). United States Department of Health and Human Services. National Nutrient Database for Standard Reference Release 28.
- WHO (world health organization), 20015 .Visual impairment and blindness Fact. Sheet No, 282.

الخضروات المصرية كمصدر لليوتين و دوره فى الإصابة بالمياه البيضاء فى الفئران عبد الحميد إبراهيم عبد الجواد¹، شريف صلاح محمد² و ضياء محمد عباس هلال¹ ¹ قسم الصناعات الغذائية – كلية الزراعة – جامعة المنصورة – مصر ² قسم التغذية – المركز القومى للبحوث – مصر

تعتبر الخضروات الطازجة غذاءاً شعبياً تستهلكه يوميا الأعمار المختلفة من الأغنياء والفقراء لإنخفاض سعرها و طيب نكهتها. وبالرغم من السعر المنخفض لهذه الخضروات إلا أنها ذات قيمة غذائية عالية لاحتوائها على تركيزات عالية من الفيتامينات خاصة الليوتين و الألياف و الكالسيوم و البوتاسيوم و الماغنسيوم و الحديد إلى جانب أن هذه الخضروات نتوافر طول العام. في السنوات الأخيرة كانت زيادة أمراض العيون وتداخل العلاقة مع المواد الغذائية مثيرة للاهتمام والأمراض الشائعة خاصة المياه البيضاء. ولذلك تم تصميم هذا البحث لدراسة استخدام بعض أصناف الخضر (السبانخ و الكرات المصرى و البقدونس و الجرجير) الأكثر شعبية و الغنية في الليوتين و علاقتها فى الحد من الإصابة بالمياة البيضاء و لذلك تم تقدير نسبة الليوتين فى هذه الخضروات ثم تصميم التجربة البيولوجية لمدة شهر واحد و فى نهاية التجربة تم الإصابة بالمياة البيضاء و لذلك تم تقدير نسبة الليوتين فى هذه الخضروات ثم تصميم التجربة البيولوجية لمدة شهر واحد و فى نهاية التجربة تم الإصابة بالمياة البيضاء و لذلك تم تقدير نسبة الليوتين فى هذه الخضروات ثم تصميم التجربة البيولوجية لمدة شهر واحد و فى نهاية التجربة تم الحدام بعض ألم لتقدير القياسات المختلفة كما تم نزع العيون و إز الة العدسات لعمل الإختبارات الهستولوحية و الميكروسكوبية. أوضحت الدراسة أن الجرجير، السبانخ، البقدونس والكراث تحتوى على نسبة عالية من اللوتين (12.71)، 11.77)، على الدومن الارسات المراسة أن الجرجير، السباخ، البقدونس والكراث تحتوى على نسبة عالية من اللوتين (12.72)، 11.77)، على الدوم و الميكروسكوبية. أوضحت من الإصابة بالمياه البيضاء فى الفتران لذا توكره هذه الدوراسة على العربيات الميتولوحية و الميكروسكوبية. أوضحت من الإصابة بالمياه البيضاء فى الفتران لذا تعتوى على نسبة عالية من اللوتين (12.72)، 11.77)، على المولية المار من الإصابة بالمياه البيضاء فى الفتران لذا توكره الليوتين و كلامن إنزيمات الأكسدة و نشاط الإنزيمات المحالة البروتين و بالتالى الحد من الإصابة بالمياه البيضاء فى الفتران لذا توكد هذه الدواسة على العلاقة بين إستهلاك الليوتين ونظرا لإنخفاض سعرها و توفرها على مدار العام.