Effects of Pomegranate and Onion Peels Extracts on reducing weight and controlling diabetes in Obese Diabetic Rats

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Abstract

The current study aimed to evaluate the effects of pomegranate peel extract (PPE) and onion peel extract (OPE) on body and fat weights; serum levels of liver enzymes, lipid profile, glucose, insulin and leptin hormones and on activities of renal tissue antioxidant enzymes and histopathology of kidneys in obese diabetic rats. Forty nine male Sprague Dawley rats were randomized into 7 equal groups (n=7). Group 1 (negative control) was fed on basal diet, while the other 6 groups were fed on high-fat diet (HFD) for 4 weeks to induce acute obesity and hyperlipidemia. The obese rats were then rendered diabetic by intraperitoneal injection of alloxan (120 mg/kg/day) for 5 days. On becoming diabetic, group 2 was kept obese diabetic as positive control and groups 3, 4, 5 and 6 were orally given PPE in doses 100 and 200 mg /kg b.wt and OPE in doses 100 and 200 mg/kg b.wt, respectively for 6 weeks. Group 7 was orally treated with Glibenclamide (standard antidiabetic drug) in a daily dose 5 mg/kg b.wt for 6 weeks. Blood samples were collected for biochemical analyses. Kidneys were dissected out for histopathology and renal homogenates were prepared to assay activities of tissue antioxidant enzymes. The results showed that oral administration of PPE and OPE to obese diabetic rats significantly reduced body and fat weights; decreased serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), total cholesterol, triglycerides and low density lipoprotein. It also decreased blood urea, creatinine, glucose and leptin hormone and increased in insulin levels. Both extracts increased activities of superoxide dismutase, glutathione peroxidase and catalase antioxidant enzymes in renal tissues and alleviated kidney histopathological lesions induced by diabetes. The results denote that pomegranate and onion peel extracts possess anti-obesity, antidiabetic effects, and consequences in obese diabetic rats. The study recommends that pomegranate and onion peels as waste products may be economically used for manufacture of safe and beneficial product for trials on obese diabetics patient.

Keywords: Pomegranate, Onion, Obesity, Diabetes, Biochemical analysis, Antioxidant.

Introduction

Obesity represents a fast growing serious health problem due to its positive correlation with increased prevalence of chronic heart diseases, diabetes mellitus and hypertension (*Kumar et al., 2011 and Matsuda and Shimomura, 2013*). There is a strong association between obesity and diabetes mellitus. Obesity is a key component of the insulin resistance syndrome in type 2 diabetes. Insulin resistance, mainly in liver and skeletal muscle, is a major risk factor for incidence of diabetes mellitus (*Raza et al., 2013*) and obesity plays a crucial role in insulin resistance and pathogenesis and development of diabetes mellitus (*Hu et al. 2014*).

Diabetes mellitus is a chronic metabolic disease characterized by hyperglycemia due to insulin deficiency, or insulin resistance, or both. Hyperglycemia occurs when the cells become unable to utilize glucose and/or the liver and skeletal muscles cannot store glycogen (*Luis-Rodríguez et al., 2012*). The increased extracellular and intracellular

glucose concentrations lead to oxidative stress via increased production of reactive oxygen species (ROS) and sharp decrease in antioxidant body defenses (*Lucchesi et al., 2013*). The oxidative stress plays a key role in the onset and development of diabetes complications, mainly diabetic nephropathy (*Li et al., 2014*).

Pomegranate (*Punica granatum* L., Family *Punicaceae*) is an ancient fruit that grows on small trees cultivated throughout Mediterranean region of Southern Europe, the Middle East, tropical Africa and Southern Asia. Pomegranate peels (pericarps) are non edible waste parts of the fruit. Extracts of pomegranate peels were reported to attenuate hepatorenal toxicity induced by aluminum chloride in rats (*Abdel Moneim et al., 2013*) and to reduce oxidative stress induced by mercuric chloride in rats (*Kumar et al., 2013*). In addition, extracts of pomegranate peels were found to have anticancer against colon cancer in rats (*Waly et al., 2012*); antihyperglycemic, hepatoprotective and hypolipidemic (*Kam et al., 2013 and Middha et al., 2013*); antibacterial (*Mehta et al., 2014*); antiviral effects against human virus C (Reddey *et al., 2014*) and antioxidant and lipase inhibitory activities (*Hadrich et al., 2014*).

The extract of onion peels has been reported to enhance antioxidant status in aged rats (*Park et al., 2007*) and to produce vasorelaxant and hypotensive activities in rats (*Naseri et al., 2008*). Quercetin-rich onion peels extract when supplemented to diet induced anti-obesity effect via influencing adipokine expression in adipose tissues and inhibited adipogenesis in obese rats (*Kim et al., 2012 and Moon et al., 2013*). It was found that onion peels contain high amounts of quercetin which has been reported to have lipid lowering (anticholesterol), antithrombotic and insulin-sensitizing (antidiabetic) properties (*Bae et al., 2014*).

The main goal of this study was to assess the anti-obesity and antidiabetic activities of the ethanolic extracts of pomegranate and onion peels in obese diabetic rats.

Materials and Methods

Plant materials:

Pomegranate (*Punica granatum* L., Family *Punicaceae*) fruits and onion bulbs (*Allium cepa*, Family *Amaryllidaceae*) were purchased from a local market in Cairo, Egypt. The outer peels (pericarps) of pomegranate fruit and onion bulb were air dried, grinded using electric blender into a fine powder and kept till alcoholic extraction.

Alloxan, Glibenclamide and kits:

Alloxan was purchased from El-Gomhoryia Company for Chemicals, Egypt; as a white powder packed bottles containing 25 g alloxan monohydrate. Glibenclamide (Doanil[®] 5 mg) was purchased from Sanofi Synthelabo Company as tablets each containing 5 mg Glibenclamide (potent hypoglycemic drug). Glucose enzymatic kits for estimating blood glucose and radioimmunoassay kits for leptin and insulin hormones were purchased from Gamma Trade Company, Egypt. The other biochemical kits were procured from Biodiagnostics Company, Dokki, Egypt.

Rats:

Forty nine adult male Sprague Dawley rats of 175-185 g body weight and 8-10 weeks old were used in this study. Animals were obtained from the Laboratory Animal Colony, Agricultural Research Center, Egypt. Rats were housed in a well ventilated animal room under standard conditions (24 °C temperature and 50% relative humidity)). Basal diet and water were provided *ad libitum*. The experiment on rats was carried out according to the National regulations on animal welfare and Institutional Animal Ethical Committee (IAEC), NRC, Dokki, Egypt.

Preparation of basal diet:

The dietary supply of protein, fat, carbohydrates, vitamins and minerals was prepared recommended by *Reeves et al., (1993)*. Basal diet consisted of 20 % protein, 10 % sucrose, 4.7 % fat, 2% choline chloride, 1% vitamin mixture, 3.5 % salt mixture and 5% fibers (cellulose). The remainder was corn starch up to 100 %.

Preparation of ethanolic extracts:

Four hundred grams (400 g) of the dried powder of pomegranate peels and similar amount from onion peels were socked in 2 liter of 90 % ethanol and kept in a refrigerator with daily shaking for 5 days. This was followed by percolation for 5 to 7 times till complete exhaustion. The ethanolic extracts were concentrated under reduced pressure using a vacuum Rotatory evaporator at 50 °C temperature. Starting plant material of 400 g of pomegranate peel and onion peel yielded 41.8 and 38.5 g semisolid extract, respectively. Twenty grams of each semisolid extract were dissolved in 98 ml distilled water and 2 ml of Tween 80 (suspending agent) to obtain 20% ethanolic extract (concentration of 200 mg/ml) as described *by Shalaby and Hamowieh (2010)*. The tested doses of both extracts were 100 and 200 mg/kg b.wt, corresponding to 0.5 and 1 ml of the liquid extract, respectively.

Induction of obesity and diabetes:

Obesity was induced by feeding rats on high-fat diet (HFD) for 4 weeks which supplies 45 % calories from pig fat (lard). This obese model in rats closely resembles the reality of obesity in humans according to **Bhatt et al. (2006)**. The obese rats were rendered diabetic by intraperitoneal injection of alloxan in a dose of 120 mg/kg/day for 5 days according to **Ashok et al. (2007)**. Thereafter, fasting blood glucose levels were estimated, and rats with blood glucose level higher than 180 mg/dL were only used in the study.

Experiment design:

Forty nine adult Sprague Dawley rats were randomized into 7 groups, of 7 rats each. Group 1 was fed on basal diet (negative control), while the other 6 groups were fed on HFD for 4 weeks to induce obesity. There after, obese rats were rendered diabetic by intraperitoneal injection of alloxan (120 mg/kg/day) for 5 days. After induction of diabetes, group 2 was kept obese diabetic and continued as positive control, while groups 3, 4, 5 and 6 were orally given the ethanolic extract of pomegranate peel in doses of 100 and 200 mg/kg and onion peel extract in doses of 100 and 200 mg/kg, respectively, daily for 6 weeks. Group 7 was orally given Glibenclamide in a dose of 5 mg/kg b.wt for 6 weeks. At the end of experiment, the rats were weighed and body fats were carefully removed and weighed. The adiposity index(Ad.I) was calculated by dividing the total weight of mesenteric, visceral, epididymal and retroperitoneal adipose tissues by the body weight and multiplied by 100 i.e. Ad.I = fat weight/body weight x100 according to *Pichon et al. (2006)*. Rats were anethetized by prolonged exposure to ether and blood samples were withdrawn for separating the serum which was kept frozen until biochemical analyses. Kidneys were dissected out, divided into two portions; one portion was used for preparing renal homogenates to assay the activity of tissue antioxidant enzymes, the other portion of kidneys was preserved in 10 % formalin solution till processed for histopathological examination.

Biochemical analyses:

Serum aspartate aminotransferase and alanine aminotransferase (*Bergmeyer et al., 1978*), alkaline phosphatase (*Roy, 1970*), total cholesterol (*Richmond, 1973*), triglycerides (*Friedewald et al., 1972*) and high density lipoprotein cholesterol (*Richmond, 1973*) were chemically determined using specific diagnostic kits and measured on a spectrophotometer. Low density lipoprotein cholesterol was calculated according to Friedewald formula: LDL-c = TC – (TG/5) – HDL-c). Blood urea nitrogen (*Patton and Crouch, 1977*), uric acid (*Fossati et al., 1980*) and creatinine (*Husdan and Rapoport, 1968*) concentrations were chemically determined. Blood glucose was determined using glucose enzymatic kit according to *Siest et al. (1981*). Insulin was estimated using specific antibody radioimmunoassay (RIA) kit according to *Yallow and Bauman (1983*). Leptin hormone was determined using enzyme-linked immunosorbent assay (ELISA) according to *Xiong et al. (2005*).

Renal antioxidant enzymes:

One gram of kidney tissue was washed with ice-cooled 0.9% NaCl solution and homogenized in 100 ml of icecooled 1.5% solution of potassium chloride and 50 mMol of potassium phosphate buffer solution (pH 7.4) to yield 10% homogenate (W/V). Kidney homogenates were centrifuged at 8000 rpm for 10 minutes at 4°C and the supernatants were used to assay the activity of antioxidant enzymes superoxide dismutase (SOD), glutathione peroxidase (GPx), and catalase (CAT) according to *Spitz and Oberley (1989), Paglia and Valentaine (1979) and Sinha (1972)* respectively.

Histological procedure:

Kidneys of sacrificed rats were taken and fixed in 10 % neutral formalin solution. The fixed specimens were then trimmed, washed and dehydrated in ascending grades of alcohol. The specimens were then embedded in paraffin, cross sectioned (cs) at 4-6 microns thickness, stained with Hematoxylen and Eosin and then examined microscopically *(Carleton, 1976).*

Statistical analysis:

Data were presented as mean ± SE. Statistical comparisons between different groups were performed using oneway analysis of variance (ANOVA) test followed by Duncan multiple range tests (*Snedecor and Cochran, 1986*). Statistical analyses were performed using computerized program (SPSS version 15, Chicago, USA).

Results

Feeding rats on high-fat diet (HFD) for 4 weeks caused significant increases (P < 0.05) in body weight, fat weight and adiposity index when compared to the negative control group. Oral administration of pomegranate peel extract (PPE) at100 and 200 mg/kg and onion peel extract (OPE) at the same doses to obese diabetic rats for 6 weeks resulted in significant decreases (P < 0.05) in body weight, fat weight and adiposity index when compared to the positive control group. Glibenclamide also decreased body and fat weights and adiposity index as recorded in Table 1.

Table 1: Body weight (B.wt), fat weight (F.wt) and adiposity index (Ad.I) in obese diabetic rats at the end of the

		experiment. (n=	7 rats.)	
Parameters Groups	B.wt (gm)	F.wt (gm)	Ad.I (%)	
Group 1: Negative control	245.0 ± 8.0^{e}	6.55 ± 0.12^{d}	2.67 ± 0.10^{d}	
Group 2: Positive control	300.0 ± 9.5 ^a	14.50 ± 0.52^{a}	4.83 ± 0.17^{a}	
Group 3: PPE (100 mg/kg)	288.0 ± 7.5 ^b	8.80 ± 0.45^{c}	3.05 ± 0.14^{c}	
Group 4: PPE (200 mg/kg)	285.0 ± 6.0^{b}	8.00 ± 0.37^{c}	$2.80 \pm 0.12^{\circ}$	
Group 5: OPE (100 mg/kg)	280.0 ± 7.5^{c}	10.50 ± 0.30 ^b	3.75 ± 0.15 ^b	
Group 6: OPE (200 mg/kg)	283.0 ± 6.5^{c}	9.80 ± 0.15 ^b	3.46 ± 0.12^{c}	5. 5.
Group 7: Glibenclamide (5 mg/kg)	260.0 ± 5.0 ^d	7.00 ± 0.15 ^d	2.69 ± 0.10 ^d	

Means \pm SE with different letters superscripts (a, b, c, d) in the same column are significant at P < 0.05 using one way ANOVA test.

Rats fed on HFD for 4 weeks had significant increases (P < 0.05) in serum levels of liver enzymes aspartate aminotransferase (AST), alanine aminotransferase (ALT), and alkaline phosphatase (ALP) (recording 108 ± 0.17^{a} , 84 ± 0.35^{a} and 104 ± 0.73^{a} U/L, respectively) when compared with negative control rats fed on basal diet (recording 60 $\pm 0.43^{d}$, 38 ± 0.65^{d} and 84 ± 0.23^{d} U/L, respectively). Both PPE and OPE in doses 100 and 200 mg/kg when given to obese diabetic rats significantly lowered (P < 0.05) the high serum levels of AST, ALT and ALP enzymes when compared to the positive control group. PPE in doses 200 mg/kg have most significantly decrease (P < 0.05) of AST, ALT and ALP enzymes (recording 74 ± 0.15^{c} , 60 ± 0.30^{c} and 82 ± 0.47^{c} U/L, respectively) compared to OPE at the same doses 200 mg/kg (recording 89 ± 0.22^{b} , 72 ± 0.53^{b} and 89 ± 0.81^{b} U/L, respectively). Glibenclamide also significantly decrease (P < 0.05) serum levels of AST, ALT and ALP enzymes (recording 100 ± 0.20^{c} , 100 ± 0.30^{c} and 100 ± 0.105^{c} , 100 ± 0.005^{c} and 100 ± 0.005^{c

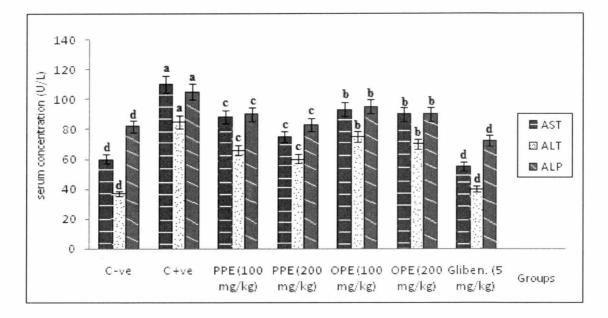


Fig. 1: Serum levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT) and alkaline phosphatase (ALP) enzymes in obese diabetic rats at the end of the experiment.

Feeding of rats on HFD for 4 weeks produced significant elevation at (P < 0.05) in serum levels of total cholesterol **TC** (136 ± 2.82^a mg/dL) and triglycerides **TG** (144 ± 3.13^a mg/dL) when compared to negative control rats fed on basal diet (recording 96 ± 1.25^d and 112 ± 2.591^d mg/dL, respectively). Oral administration PPE and OPE in doses of 100 and 200 mg/kg to obese diabetic rats for 6 weeks significantly decreased (P < 0.05) the elevated levels of serum TC and TG when compared to the positive control group. TC and TG (recording 120 ± 4.11^c and 100 ± 3.421^c mg/dL, respectively) for PPE at dose 200 mg/kg, while OPE (recording 130 ± 1.31^b and 114 ± 2.12^b mg/dL, respectively). Glibenclamide also induced significant decreases (P < 0.05) in high serum levels of TC and TG (recording 100 ± 4.11^d and 108 ± 2.721^d mg/dL, respectively) as demonstrated in Fig 2.

The results denoted that feeding of rats on HFD for 4 weeks caused a significant (P < 0.05) decrease in serum level of high density lipoprotein (HDL), increases significant (P < 0.05) in both low density lipoprotein (LDL), and atherogenic index (Al) when compared to negative control rats. Oral administration of PPE and OPE in doses of 100 and 200 mg/kg to obese diabetic rats for 6 weeks significantly increased (P < 0.05) serum HDL-c, significant decreased (P < 0.05) both LDL-c and Al when compared with the positive control groups. Glibenclamide also significant increased (P < 0.05) serum HDL-c and significant decreased (P < 0.05) both LDL-c and Al as depicted in Table 2.

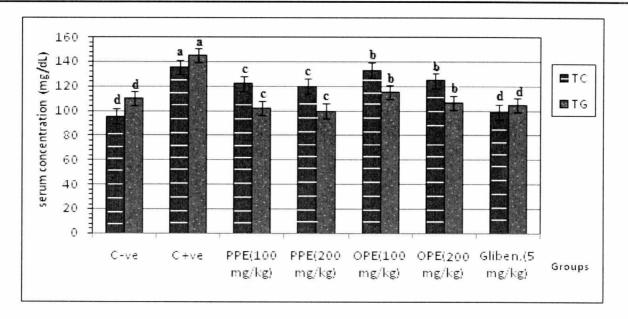


Fig. 2: Serum total cholesterol (TC) and triglycerides (TG) in obese diabetic rats at the end of the experiment.

 Table 2:

 Serum levels of high density lipoprotein (HDL-c), low density lipoprotein (LDL-c) cholesterol and atherogenic index (AI) in obese diabetic rats at the end of the experiment. (n= 7 rats).

Parameters	HDL-c	LDL-c	AI	
	(mg/dL)	(mg/dL)	(LDL-c / HDL-c)	
Groups				
Group 1:	79.00 ± 3.5^{a}	17.50 ± 0.2^{d}	0.221	
Negative control				
Group 2:	52.00 ± 2.1 ^d	39.50 ± 0.5^{a}	0.759	
Positive control				
Group 3:	763 ± 3.8 ^b	28.50 ± 0.1 ^b	0.373	
PPE (100 mg/kg)				
Group 4:	75.50 ± 4.7^{b}	27.00 ± 0.4^{b}	0.357	
PPE (200 mg/kg)				
Group 5:	$63.00 \pm 3.6^{\circ}$	$23.40 \pm 0.3^{\circ}$	0.371	
OPE (100 mg/kg)				
Group 6:	$65.00 \pm 2.5^{\circ}$	$21.40 \pm 0.1^{\circ}$	0.329	
OPE (200 mg/kg)				
Group 7:	78.50 ± 2.2^{b}	$19.00 \pm 0.3^{\circ}$	0.242	
Glibenclamide (5 mg	g/kg)			

Means \pm SE with different letters superscripts (a, b, c, d) in the same column are significantly different at P < -0.05 using one way ANOVA test.

Data in Table 3 showed that rats fed on HFD for 4 weeks had significantly (P < 0.05) increased blood glucose and leptin hormone and decreased insulin hormone levels when compared to rats fed on basal diet (negative control group).

PPE and OPE when orally given in doses 100 and 200 mg kg to obese diabetic rats for 6 weeks significantly decreased (P < 0.05) serum glucose and leptin hormone and increased insulin levels when compared with positive control rats. Glibenclamide also significantly decreased (P < 0.05) serum levels of glucose and leptin hormone and increased insulin levels.

Rats fed HFD for 4 weeks had significant increases (P < 0.05) in blood urea (BUN) and creatinine (Cr) levels when compared to control rats. PPE and OPE when given orally to obese diabetic rats in doses of 100 and 200 mg/kg significantly decreased (P < 0.05) the high serum levels of BUN and Cr as compared to the positive control group. Glibenclamide also significantly decreased (P < 0.05) BUN and Cr concentrations in the serum (Table 4).

	at the e	nd of the experimen	t. (n= 7 rats).	
Parameters	BG	Leptin	Insulin	
Groups	(mg/dl)	(ng/ml)	(ng/ml)	
Group 1:	200.5 ± 3.0 ^e	2.65 ± 0.12 ^d	2.88 ± 0.15 ^a	
Negative control				
Group 2:	255.0 ± 4.0^{a}	6.90 ± 0.13^{a}	0.89 ± 0.13^{d}	
Positive control				
Group 3:	165.5 ± 2.0 ^c	$3.55 \pm 0.16^{\circ}$	2.52 ± 0.24^{b}	
PPE (100 mg/kg)				
Group 4:	$163.5 \pm 3.0^{\circ}$	3.35 ± 0.17^{c}	2.33 ± 0.12 ^b	
PPE (200 mg/kg)				
Group 5:	189.5 ± 3.0 ^b	4.90 ± 0.12 ^b	$1.62 \pm 0.14^{\circ}$	
OPE (100 mg/kg)				
Group 6:	186.5 ± 4.0 ^b	4.75 ± 0.13 ^b	1.58 ± 0.14 ^c	
OPE (200 mg/kg)				
Group 7:	140.0 ± 3.0^{d}	3.45 ± 0.15 ^b	2.75 ± 0.14^{c}	
Glibenclamide				
(5mg/kg)				

Table 3: Blood glucose (BG) and leptin and insulin hormones levels in obese diabetic rats at the end of the experiment, (n= 7 rats).

Means \pm SE with different letters superscripts (a, b, c, d) in the same column are significantly different at P < 0.05 using one way ANOVA test.

Table 4:

Blood urea nitrogen (BUN), uric acid (UA) and creatinine (Cr) levels in obese diabetic rats at the end of the experiment. (n= 7 rats)

Parameters	BUN	UA	Cr	
	(mg/dL)	(mg/dL)	(mg/dL)	
Groups				
Group 1:	35.8 ± 2.1 ^d	1.60 ± 0.01 ^a	0.65 ± 0.01^{d}	
Normal control				
Group 2:	66.0 ± 4.3^{a}	1.62 ± 0.06^{a}	1.84 ± 0.04 ^a	
Positive control				
Group 3:	$47.1 \pm 2.6^{\circ}$	1.64 ± 0.02^{a}	$0.96 \pm 0.02^{\circ}$	
PPE (100 mg/kg)				
Group 4:	41.8 ± 3.3 ^c	1.63 ± 0.04^{a}	$0.82 \pm 0.02^{\circ}$	
PPE (200 mg/kg)				
Group 5:	56.5 ± 4.1 ^b	1.60 ± 0.01^{a}	1.27 ± 0.03 ^b	
OPE (100 mg/kg)				
Group 6:	55.4 ± 3.5 ^b	1.62 ± 0.05 ^a	1.24 ± 0.02 ^b	
OPE (200 mg/kg)				
Group 7:	40.6 ± 2.8^{c}	1.63 ± 0.01 ^a	$0.92 \pm 0.01^{\circ}$	
Glibenclamide				
(5mg/kg)				

Means ± SE with different letters superscripts (a, b, c, d) in the same column are significantly different at P < 0.05 using one way ANOVA test.

Feeding HFD to rats for 4 weeks significantly decreased (P < 0.05) renal tissue levels of superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) antioxidant enzymes when compared to rats fed on basal diet. Oral administration of PPE and OPE in doses of 100 and 200 mg/kg to obese diabetic rats for 6 weeks increased the activity of renal tissue of SOD, GPx and CAT enzymes when compared with the positive control group. Glibenclamide also significantly increased (P < 0.05) activities of SOD and CAT enzymes as depicted in Table 5.

Histopathological examination of renal cortex of normal control rats showed normal histological structure of renal parenchyma (renal corpuscles and tubules) as illustrated in Fig. (3). Kidneys of obese diabetic rats revealed marked necrosis and degeneration of renal tubules (Fig. 4). Examination of kidney section following oral administration of pomegranate peel extract in a dose of 200 mg/kg to obese diabetic rats for 6 weeks showed almost normal histological structure of renal parenchyma as demonstrated in Fig (5). In obese diabetic rats administered with 200 mg/kg of onion peel extract, the microscopical examination of kidney sections revealed moderate congestion of blood vessels in between renal tubules as shown in Fig (6).

Table 5:

Aof tissue superoxide dismutase (SOD), glutathione peroxidase (GPx) and catalase (CAT) antioxidant enzymes in obese diabetic rats at the end of the experiment. (n= 7 rats).

-	205		CAT	
Parameters	SOD	GPx	CAT	
	(U/mg protein)	(nmol/min/mg protein)	(nmol/min/mg protein)	
Groups				
Group 1:	59.50 ± 2.11^{a}	0.94 ± 0.04^{a}	0.188 ± 0.01^{a}	
Negative control				
Group 2:	36.50 ± 2.18 ^d	0.18 ± 0.01^{d}	0.135 ± 0.02 ^d	
Positive control				
Group 3:	53.74 ± 3.46 ^b	0.75 ± 0.03 ^b	0.175 ± 0.02 ^b	
PPE (100 mg/kg)				
Group 4:	54.95 ± 2.58^{b}	0.79 ± 0.03 ^b	0.178 ± 0.01 ^b	
PPE (200 mg/kg)				
Group 5:	$46.25 \pm 2.73^{\circ}$	0.57 ± 0.01^{c}	$0.145 \pm 0.02^{\circ}$	
OPE (100 mg/kg)				
Group 6:	$45.25 \pm 2.73^{\circ}$	$0.56 \pm 0.02^{\circ}$	0.147 ± 0.01°	
OPE (200 mg/kg)				
Group 7:	52.25 ± 2.73 ^b	0.29 ± 0.01^{d}	0.183 ± 0.01 ^b	
-	JZ.20 I 2.75	0.29 ± 0.01	0.100 ± 0.01	
Glibenclamide				
(5mg/kg)				

Means \pm SE with different letters superscripts (a, b, c, d) in the same column are Significantly different at P < 0.05 using one way ANOVA test.

Unit of GPx= nmol of GSH utilized/min/mg protein.

Unit of CAT= nmol of H₂O₂ utilized/min/mg protein.

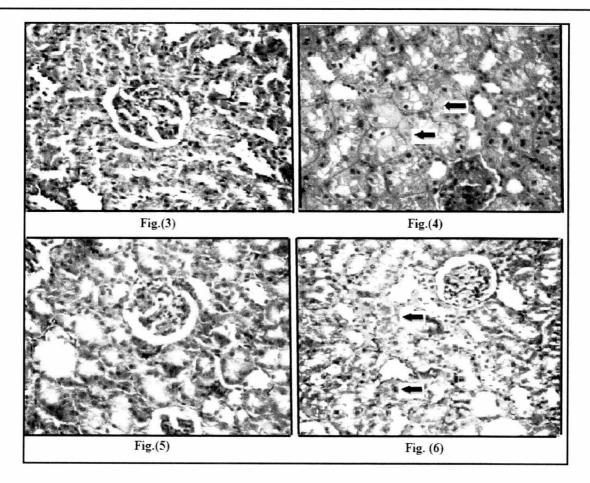


Fig. 3: C.S. of a kidney cortex of normal control rat showing normal architecture of renal parenchyma (glomeruli and tubules). (H&E X 200).

Fig. 4: C.S.in a kidney cortex of obese diabetic rat showing marked necrosis and fatty degeneration of renal tubules (arrows). (H&E X 200).

Fig. 5: C.S.in a kidney cortex of obese diabetic rat given orally pomegranate peel extract at 200 mg/kg showing almost normal histological structure of renal parenchyma. (H&E X 200).

Fig. 6: C.S.in a kidney cortex of obese diabetic rat given orally onion peel extract at 200 mg/kg showing moderate congestion of blood vessels in-between renal tubules (arrows). (H&E X 200).

Discussion

Fruits and vegetables which possess anti-obesity and antidiabetic effects have recently gained much attention, especially those with low toxicity. The biological value of fruits and vegetables depends on their bioactive constituents such as saponins, tannins, anthocyanins, flavonoids, polyphenols, diterpenes, triterpenes and other phytochemicals (*Patel et al., 2012*).

Results of the present study denoted that pomegranate peel extract (PPE) when given orally to obese diabetic rats reduced body and fat weights and decreased adiposity index. Measurement of adipose (fat) tissue weight has been used as a valid index in obesity studies. The anti-obesity activity of PPE is reported for the first time in this study. However, *Neyrinck et al. (2013)* reported that feeding pomegranate peel to obese mice counteracted the inflammatory markers in adipose tissue, denoting its anti-inflammatory activity. Onion peel extract (OPE) induced a potent reduction in body weight and fat in obese diabetic rats. The effect of OPE agreed with the previous reports of *Kim et al. (2012) and Moon et al., (2013)*. The previous authors concluded that OPE supplemented to diet induced its effect via influencing adipokine expression in adipose tissues in obese rats.

The mechanism(s) underlying the anti-obesity effect of PPE and OPE could be possibly explained by their hyperinsulinimic effect evident in the present study in obese diabetic rats. Hyperinsulinemia is a common features of obesity (*Raza et al., 2013*). Other mechanism of anti-obesity activity of PPE and OPE could be due to decreased level of leptin hormone reported in the present study. In this concern, *Friedman (2011)* mentioned that leptin hormone is secreted by adipose tissue in proportion to its weight (mass). When body fat mass decreases, the plasma leptin levels decreases till fat mass is restored. On this basis, the decreased fat weight in obese diabetic rats given PPE or OPE could be possibly attributed to the reported low serum leptin level (hypoleptinemia).

The hepatoprotection and antihyperlipidemia of PPE and OPE reported in this study were evident from the significant decreases in the elevated serum levels of liver enzymes (AST, ALT and ALP), total cholesterol and triglycerides in obese diabetic rats. These findings agreed with the previous reports of *Middha et al., (2013)* for PPE and of *Bae et al., (2014)* for OPE. The hepatoprotective activity of PPE and OPE could be possibly attributed to their antioxidant effect that previously reported by *Middha et al., (2013), Abdel Moneim et al., (2013) and Kumar et al., (2013)* for PPE and by *Park et al., (2007)* for OPE. The hypolipidemic effect of PPE could be due to its contents of polyphenols which inhibit the intestinal absorption of cholesterol and so reduce serum cholesterol levels (*Middha et al., 2013*). Concerning OPE, its hypolipidemic activity was explained by its high content of quercetin which has been reported to have lipid lowering properties (Bae *et al., 2014*).

In this study, blood glucose levels were decreased significantly by PPE. The hypoglycemic effect of PPE agreed with the previous reports of *Parmar and Kar (2008); Kam et al., 2013; Middha et al., (2013) and Benihani, et al., (2013).* This effect was attributed to the presence of punicalagin, ellagic and gallic acids in PPE which have been reported to possess antidiabetic action (*Parmar and Kar, 2008).* The hypoglycemic activity of OPE reported in this study was similar to that demonstrated by *Bae et al., (2014)* who concluded that onion peels contain high amounts of quercetin that induces insulin-sensitizing (antidiabetic) properties

The present results denoted that oral administration of PPE and OPE to obese diabetic rats caused hyperinsulinemia. The hyperinsulinemic effect of PPE and OPE was similar to that reported by *Parmar and Kar, (2008)* for PPE and by *Bae et al., (2014)* for OPE in rats. The increased insulin secretion following administration of PPE and OPE to obese rats, in this study, might be the possible mechanism of their antidiabetic activity.

The current results revealed that rats fed on HFD had high serum leptin hormone level when compared with rats fed on basal diet. This result was in accordance with that reported by *Huang et al. (2004)*. Leptin plays a key role in regulating energy intake and energy expenditure and the level of circulating leptin is proportional to the total amount of body fats. PPE and OPE significantly decreased serum leptin levels in obese diabetic rats. No references were available on the effect of PPE and OPE on serum leptin level in rats. The effect of PPE and OPE on serum leptin level in rats is performed for the first time in this study in obese diabetic rat model.

The activity of antioxidant enzymes (SOD, GPx and CAT) increased in renal tissues of obese diabetic rats fed on PPE and OPE. PPE and OPE given orally to obese diabetic rats caused an anti-oxidant activity. The antioxidant effect of PPE and OPE was in accordance with the previous findings of *Hadrich et al., (2014)* for PPE and of *Park et al.,* (2007) for OPE.

In conclusion, extracts of pomegranate and onion peels may reduce weight and helps to control diabetes and may have hypolipidemic, hepatoprotective, nephroprotective and antioxidant effects in obese diabetic rats. The study recommends that pomegranate and onion peels considered as waste products may be used for manufacture of a product that could be tried on obese diabetic patients.

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أثر خلاصات قشور الرمان وقشور البصل على انقاص الوزن وضبط مستوى السكر في الفئران البدينة المصابة بداء السكري

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الملخص العربى

استهدف هذا البحث تقييم تأثير خلاصات قشور الرمان والبصل على وزن الجسم، ووزن دهون الجسم، إنزيمات الكبد، صورة دهون وسكر الدم، ومستوى هرمون الإنسولين واللبتين بالمصل، وكذا على نشاط الإنزيمات المضادة للأكسدة بنسيج الكلي والفحص الهستوباتولوجي للكلي في ذكورالفنران البدينة المصابة بداء السكري. وتم توزيع عدد ٤٩ من ذكور الفنران البالغة بطريقة عشوانية إلى ىسبع مجموعات متساوية كل منها ٧ فنران. استخدمت المجموعة الأولى كمجموعة ضابطة سالبة وتم تغذيتها على الوجبة الأساسية، بينما تم تغذية فنران المجموعات الستة الاخرى على وجبة عالية في الدهون لمدة ٤ اسابيع لإحداث البدانة. وتم اصابة هذه الفنران بداء السكري عن طريق حقن مادة الألوكسان في التجويف البريتوني بجرعة ١٢٠ مجم/كجم يوميا لمدة ٥ أيام. وبعدها استخدمت المجموعة الثانية كمجموعة ضابطة موجبة (فنران بدينة مصابة بداء السكرى)، وتم إعطاء المجموعات الثالثة والرابعة والخامسة والسادسة عن طريق الفم خلاصة قشور الرمان بجرعتى ١٠٠و٢٠٠ مجم/كجم وخلاصة قشور البصل بجرعتى ١٠٠و٢٠٠ مجم/كجم على التوالي يوميا لمدة ستة اسابيع. وتم إعطاء المجموعة السابعة عن طريق الفم دواء جيبنكلاميد المخفض لسكر الدم بجرعة ٥ مجم/كجم/يوم لمدة ستة اسابيع . وبعد إنتهاء فترة التجربة تم تجميع عينات الدم لفصل المصل لاستخدامه في التحليلات البيوكيميانية ، وتم أخذ الكليتين لقياس نشاط الإنزيمات المضادة للأكسدة في نسيجها، وكذا إجراء الفحص الهستوباتولوجي لنسيج الكلي. وأظهرت النتائج أن إعطاء خلاصة قشور الرمان والبصل عن طريق الفم لمدة ستة اسابيع لذكور الفئران البدينة المصابة بداء السكري أدى إلى نقص معنوى في وزن الجسم ودهون الجسم ودليل البدانة، وكذا أدى إلى خفض مستوى إنزيمات الكبد المرتفعة (اسبرتات أمينو ترانسفيريز، الانين أمينو ترانسفيريز و الكالين فوسفاتيز)، ونقص في المستوى المرتفع من الكوليسترول الكلي، الدهون الثلاثية والبروتينات الدهنية قليلة الكثافة، مع نقص في مستوى سكر الدم و هرمون اللبتين وزيادة مستوى هرمون الإنسولين في المصل. وأدى كذلك إلى زيادة نشاط الإنزيمات المضادة للأكسدة في أنسجة الكلي، كما قلل من التغييرات الهستوباثولوجية التي سببها داء السكري بنسيج الكلي. وتدل النتائج أن لخلاصة قشور الرمان والبصل تأثيرات مضادة للبدانة ومضادة لداء السكرى وتوابعها في فئران التجارب. وتقترح الدراسة أن قشورالرمان والبصل يمكن استخدامها اقتصاديا في تصنيع منتجات طبيعية أمنة لتجربتها على المرضى البدناء ومصابي داء السكري.